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References

- Bandi, C., La Rosa, G., Bardin, M.G., Damiani, G., Comincini, S., Tasciotti, L. and Pozio, E. 1995. Random amplified polymorphic DNA fingerprints of the eight taxa of *Trichinella* and their comparison with allozyme analysis. *Parasitology*., 110:401-407.
- Campbell, W.C. 1983. Historical introduction. In: *Trichinella and Trichinosis*, pp. 1-30, Campbell, W.C. ed., Plenum Press, New York.
- 3) Gamble, H.R., Bessonov, A.S., Cuperlovic, K., Gajadhar, A.A., van Knapen, F., Noeckler, K., Schenone, H. and Zhu, X. 2000. International Commission on Trichinellosis: recommendations on methods for the control of *Trichinella* in domestic and wild animals intended for human consumption. *Vet. Parasitol.*, 93:393-408.
- 4) Harrison, R.L. 1997. A comparison of gray fox ecology between residential and undeveloped rural landscapes. *J. Wildl. Manage.*, 61:112-122.
- 5) Hata, R. 1972. On the variation in number

- of the tooth and root in *Nyctereutes procyonoides albus. Jpn. J. Oral. Biol.*, 14; 118-122 (in Japanese).
- 6) Kanai, Y., Nonaka, N., Katakura, K. and Oku, Y. 2006. *Trichinella nativa* and *Trichinella* T9 in the Hokkaido island, Japan. *Parasitol. Int.*, 55: 313-315.
- 7) Kapel, C.M., Henriksen, S.A., Berg, T.B. and Nansen, P. 1995. *Trichinella* infections in arctic foxes from Greenland: studies and reflections on predilection sites of muscle larvae. *J. Helminthol.*, 69:325-330.
- 8) Kapel, C.M., Pozio, E., Sacchi, L. and Prestrud, P. 1999. Freeze tolerance, morphology, and RAPD-PCR identification of *Trichinella nativa* in naturally infected arctic foxes. *J. Parasitol.*, 85: 144-7.
- 9) Kudo, N., Arima, R., Ohtsuki, M. and Oyamada, T. 2001. The first host record of trichinosis in a red fox, *Vulpes vulpes japonica*, from Aomori Prefecture, northern Honshu, Japan. *J. Vet. Med. Sci.*, 63:823-826.
- 10) Maekawa, K., Yoneda, M. and Togashi, H. 1980. A preliminary study of the age structure of the fox in eastern Hokkaido. *Jpn. J. Ecol.*, 30:103-108.
- 11) Ohbayashi, M. and Satoh H. 1957. Discovery of a case of trichinosis in Japan. *Jpn. J. Vet. Res.*, 5: 39-42.
- 12) Oksanen, A., Lindgren, E. and Tunkkari, P. 1998. Epidemiology of trichinellosis in lynx in Finland. J. Helminthol., 72: 47-53.
- 13) Pozio, E. 2000. Factors affecting the flow among domestic, synanthropic and sylvatic cycles of *Trichinella*. *Vet Parasitol.*, 93: 241-262.
- 14) Pozio, E. 2005. The broad spectrum of Trichinella hosts: from cold-to warm-blooded animals. *Vet. Parasitol.*, 132: 3-11.
- 15) Pozio, E. and La Rosa, G. 2003. PCR-

- derived methods for the identification of *Trichinella* parasites from animal and human samples. *Methods Mol. Biol.*, 216: 299-309.
- 16) Pozio, E., La Rosa, L.A. and Gomez Morales M.A. 2000. Epidemiology of human and animal trichinellosis in Italy since its discovery in 1887. *Parasite*, 8: S106-108.
- 17) Pozio, E. and Zarlenga, D.S. 2005. Recent advances on the taxonomy, systematics and epidemiology of *Trichinella*. *Int. J. Parasitol.*, 35: 1191-1204.
- 18) Rah, H., Chomel, B.B., Follmann, E.H., Kasten, R.W., Hew, C.H., Farver, T.B., Garner, G.W. and Amstrup, S.C. 2005. Serosurvey of selected zoonotic agents in polar bears (*Ursus maritimus*). *Vet. Rec.*, 156: 7-13.
- 19) Roy, S.L., Lopez, A.S. and Schantz, P. M. 2003. Trichinellosis surveillance-United States, 1997-2001. MMRW Morb. Mortal. Whly. Rep., 52: SS1-6
- 20) Saito, S. and Yamaguchi, T. 1985. Trichinella spiralis in a Raccoon dog, Nyctereutes procyonoides viverrinus, from Yamagata prefecture, Honshu, Japan. Jpn. J. Parasitol., 34: 311-314.
- 21) Sasakawa, M. 1984. Growth of the skull and eruption sequences of permanent

- teeth in red fox, Vulpes vulpes. Jpn. J. Oral. Biol., 26: 1210-1227.
- 22) Yamaguchi, T. 1991. Present status of trichinellosis in Japan. Southeast Asian J. Trop. Med. Pub. Heal., 22: 295-301.
- 23) Yamaguchi, K., Yuzawa, M., Ichihara, S. and Hattori, K. 1987. Geographical distribution of *Trichinella spiralis* in wild bear (*Ursus arctos yezoensis*) in Hokkaido, Japan. *J. Hokkaido Vet. Med. Assoc.*, 31: 205-206 (in Japanese).
- 24) Yimam, A.E., Oku, Y., Nonaka, N., Sakai, H., Morishima, Y., Matsuo, K., La Rosa, G., Pozio, E., Yagi, K. and Kamiya, M. 2001. First report of *Trichinella nativa* in foxes (*Vulpes vulpes schrencki*) from Otaru City, Hokkaido, Japan. *Parasitol. Int.*, 50: 121-127.
- 25) Zarlenga, D.S., Chute, M.B., Martin, A. and Kapel, C.M. 1999. A multiplex PCR for unequivocal differentiation of all encapsulated and non-encapsulated genotypes of *Trichinella*. *Int. J. Parasitol.*, 29: 1859-1867.
- 26) Zarnke, R.L., Gajadhar, A.A., Tiffin, G.B. and Ver Hoef, J.M. 1995. Prevalence of Trichinella nativa in lynx (Felis lynx) from Alaska, 1988-1993. J. Wildl. Dis., 31: 314-318.

ORIGINAL PAPER

Epidemiology, histpathology, and muscle distribution of Trichinella T9 in feral raccoons (Procyon lotor) and wildlife of Japan

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Abstract The prevalences of Trichinella T9 in trapped raccoons (Procyon lotor) and several other potential mammalian reservoirs in Hokkaido, Wakayama, and Nagasaki Prefectures were investigated. Muscle samples were collected from 2003 to 2006 from 1,080 raccoons, 113 raccoon dogs including 2 species (Nyctereutes procyonoides albus and N. p. viverrinus), 41 wild boars (Sus scrofa leucomystax), 14 martens (Martes melampus), 10 badgers (Meles meles), 5 Siberian weasels (Martes sibirica coreana), 7 mink (Mustela vison), and 1 red fox (Vulpes vulpes japonica). The samples were digested, and the prevalence and mean intensity of infection with the Trichinella muscle larvae were determined. The prevalence and intensity of the muscle larvae were 0.9% and 93.3 larvae/g (range 0.4-201.8) in raccoons, and 1.6% and 61.6 larvae/g in raccoon dogs, respectively. The infected animals were captured in different areas in Hokkaido Prefecture. These results confirmed that raccoons, which have been introduced from North America since the 1970s, are involved in the sylvatic cycle of *Trichinella* in Japan. In raccoons, the muscle density of *Trichinella* T9 larvae was highest in the tongue, and larvae were not found in the heart muscle or diaphragm. This is the first report of *Trichinella* T9 infection of feral raccoons in Japan.

Introduction

The raccoon (*Procyon lotor*) is a medium-sized mammal that is widely distributed in North America. Previous studies have indicated that raccoons are one of the reservoirs of the sylvatic *Trichinella* cycle (Smith et al. 1985; Cole and Shoop 1987; Snyder 1987; Richardson et al. 1992). *Trichinella murrelli* have previously been detected in raccoons from Illinois and Indiana (Pozio and La Rosa 2000). Since the 1970s, a large number of raccoons have been imported to Japan as pets. As a result of both intentional release and escape, a well-established feral raccoon population now exists in Japan (Ikeda 2004).

In Japan, although several surveys on the helminth fauna of raccoons have been reported (Miyashita 1993; Asakawa et al. 1999, 2000; Matoba et al. 2002; Sato and Suzuki 2005a,b, 2006), there is no information concerning *Trichinella* infection in raccoons. Infections caused by the consumption of the raw meat of black bear (*Ursus thibetanus*) and brown bear (*U. arctos*) have been reported since 1974 in Japan (Yamaguchi 1991). *Trichinella* infection in the wildlife has also been detected in a raccoon dog (*Nyctereutes procyonoides*) from Yamagata Prefecture

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K. Suzuki Hikiiwa Park Center, Tanabe, Wakayama 646-0051, Japan (Saito and Yamaguchi 1985), in red foxes (*Vulpes vulpes schrencki*) from Hokkaido Prefecture (Yimam et al. 2001; Kanai et al. 2006), and in a red fox (*V. v. japonica*) from Aomori Prefecture (Kudo et al. 2001).

Because of the potential public health and epidemiological significance of this parasite, the current study was conducted to assess the prevalence of *Trichinella* infection in feral raccoons and in several other potential mammalian reservoirs in Japan.

Materials and methods

Feral raccoons were trapped from April 2003 to March 2006 in Hokkaido, Wakayama, and Nagasaki Prefectures by each municipality as a part of the local governmental control and eradication programs (Fig. 1). The animals were caught using box traps (Havahart model 1089, Woodstream, Lititz, PA, USA, or handmade by hunters), and were euthanized by humanitarian methods (AVMA Panel on Euthanasia 2001). Tongues or diaphragms were collected from 1,080 raccoons (678 from Hokkaido, 390 from Wakayama, 12 from Nagasaki) and frozen at -20°C at the Wild Animal Medical Center (WAMC) at Rakuno Gakuen University. Individual numbers (AS nos.) were given to animals examined at WAMC. Using 5 g of muscle per sample, the presence of *Trichinella* larvae was

determined by an artificial digestion method (Henriksen 1978) with the following changes: Ten individual samples were digested at a time in 0.5 L of 1% pepsin-HCl solution for at least 4 h at 37°C with constant gentle stirring. After the muscle tissues had been digested, the sediment was allowed to settle and was washed several times. The sediment from the last washing was examined for larvae under a dissection microscope. When the sediment contained *Trichinella* larvae, the procedure was repeated using the individual samples.

Portions of the tongue, diaphragm, heart muscle, and superficial pectoral muscle of 469 raccoons from Hokkaido Prefecture were collected and fixed in 10% neutral-buffered formalin for 24 h and processed by routine histological methods for examination by light microscopy. Paraffinembedded tissues were sectioned at 6 µm, stained with haematoxylin and eosin (H & E) and examined with the aid of a light microscope to assess the host reaction to muscle larvae.

To compare the prevalence of *Trichinella* larvae in raccoons and other animals, tongues were collected from 113 raccoon dogs (*Nyctereutes procyonoides albus* and *N. p. viverrinus*), 41 wild boars (*Sus scrofa leucomystax*), 14 martens (*Martes melampus*), 10 badgers (*Meles meles*), 5 Siberian weasels (*Martes sibirica coreana*), 7 mink (*Mustela vison*), and 1 red fox (*Vulpes vulpes japonica*) in the same study areas, and these were examined using the same methods as for raccoons.

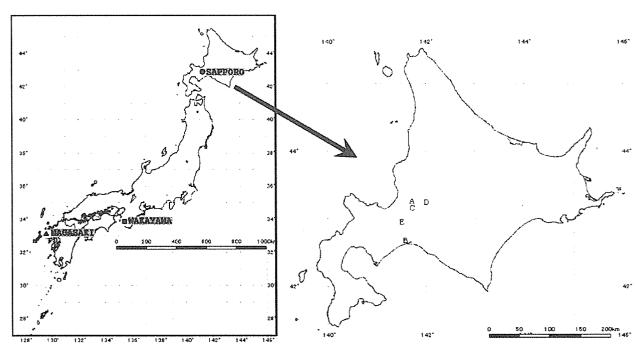


Fig. 1 Locations of the study areas in Japan. The raccoons positive for *Trihinella* were captured in the following areas (no. of the raccoon captured): a Kurisawa, 43°7′N, 141°44′E no. 4324; b Tomakomai, 42°

37'N, 141°36'E nos. 5342 and 4529; **c** Iwamizawa, 43°11'N, 141°46'E no. 5417; **d** Mikasa, 43°13'N, 141°50'E no.5498; **e** Kitahirosima, 42° 59'N, 141°33'E no.5601

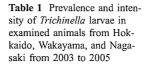


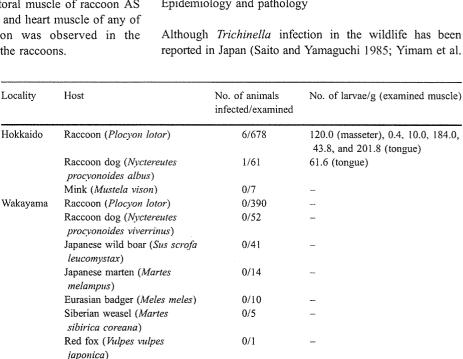
The age of infected raccoons was determined by counting the annual incremental lines that occur in the tooth cementum of canines (Grau et al. 1970).

Results

Trichinella T9 larvae were detected in six (0.8%) raccoons and one (1.6%) raccoon dog in Hokkaido Prefecture (Table 1). The intensity of infection in these raccoons ranged from 0.4 to 201.8 larvae/g of muscle tissue. All the samples from Wakayama and Nagasaki Prefectures were negative for Trichinella larvae.

Histopathological examination was carried out on the infected muscle of 469 raccoons. Cysts and brightly staining eosinophilic cyst walls were found in the tongue muscle fibers of four raccoons (AS 4324, 5342, 5417, and 5498; these are same raccoons that were found to be infected using the digestion technique). In the cysts, a single coiled larva and nurse cells were observed (Fig. 2). Inflammatory cells consisted of mononuclear cells, and eosinophils sparsely infiltrated the tongue of the raccoon AS 4324. On the other hand, indistinct capsules and a prominent inflammatory infiltration consisting of eosinophils and lymph cells were observed in raccoon AS 5342. In the other two raccoons (AS 5417 and 5498), the inflammatory infiltrate surrounding the capsules was prominent, and both distinct and indistinct capsules were observed. Muscle larvae were also observed in the superficial pectoral muscle of raccoon AS 4324, but not in the diaphragm and heart muscle of any of the raccoons. No calcification was observed in the investigated sections of any of the raccoons.





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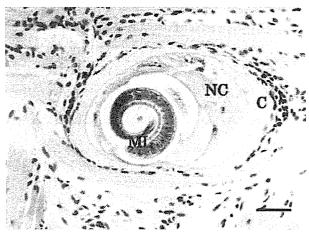


Fig. 2 A Trichinella larva encysted in the tongue muscle of a raccoon. Inflammatory cells are observed around the capsule (tongue of the raccoon AS 4324). C Capsule, L larva, NC nurse cell. HE bar = 100 µm

The results of multiplex PCR-test and COI (partial mitochondrial cytochrome oxidase subunit I) sequence analysis (Zarlenga et al. 1999; Kanai et al. 2006) of Trichinella isolates in the present study will be reported separately (Kobayashi et al. submitted).

Discussion

Epidemiology and pathology

Raccoon (Plocyon lotor)

Nagasaki

2001; Kanai et al. 2006; Kudo et al. 2001), this is the first report of *Trichinella* infection in feral raccoons.

Yimam et al. (2001) reported a high prevalence (11.6%) of *Trichinella nativa* in red foxes in Otaru. The distribution of *Trichinella* species in wild animals is strongly influenced by environmental temperature, and therefore, their distribution is bounded by isotherms (Pozio et al. 1992a,b). Geographically, *Trichnella* T9 and *T. nativa* were present in wild animals in Hokkaido.

Histopathologic responses of encapsulated *Trichinella* larvae of raccoons in the present study were different from those of the *T. nativa* in raccoon dogs (Sukura et al. 2002). The raccoon dog is a medium-sized omnivore that is as widely distributed in Hokkaido as the raccoon. In the experimental infection of raccoon dogs with *T. nativa*, a cellular inflammation reaction developed, and cyst walls became thicker over time. In one of the cases of raccoon trichinellosis in the present study, thickened cyst walls along with a minimum host reaction were observed in the same section. Although there have been no reports of experimental infection of *T. nativa* in raccoons, these histopathological features indicate that *Trichinella* larvae from raccoons were distinct from *T. nativa*.

Host ethology and public health

Smith et al. (1985) and Cole and Shoop (1987) reported that *Trichinella* showed a significant prevalence in the male raccoons, and in our study, the infections occurred only in adult males. The reason for this difference between males and females is not apparent, but there are several reports of a relationship between host sex and *Trichinella spiralis* infection (Mankau and Hamilton 1972; Mascaro-Lazcano et al. 1978; Klein et al. 1999). One possible interpretation of the sex differences in *Trichinella* infection is that testosterone may suppress immune function, although empirical evidence is lacking.

The foods consumed by raccoons in the Nopporo Forest Park, Hokkaido have been identified. In winter (November to March), rodents (Clethrionomys rufocanus bedfordiae, Apodemus speciosus, and Rattus norvegicus), which were the most probable paratenic reservoir for Trichinella, represented approximately 50% by frequency of occurrence of all food items in the gastrointestinal tract of raccoons (Matoba et al., personal communication). This observation suggests that there is a greater opportunity for infection in winter. In Hokkaido, the mating season for raccoons reaches a peak in February, and offspring are born between March and May (Asano et al. 2003). The explanation for the absence of infected juvenile raccoons is that they were captured and euthanized before their first winter and, therefore, had no chance to acquire paratenic reservoirs. On the other hand, Kanai et al. (2006) examined 344 rodents and 27 insectivores by the artificial digestion method and found no evidence of infection with *Trichinella* larvae. This suggests the existence of another host cycle such as foxraccoon, raccoon dog-raccoon, or bear-raccoon cycles.

In the present study, all samples from Wakayama and Nagasaki Prefectures were negative for Trichinella larvae. This result suggested that there is a focal distribution of this parasite in the Japanese archipelago. In fact, until now, the reports of trichinellosis in wild animals of Japan were limited in Hokkaido Prefecture and Northern Honshu Prefecture (Yamaguchi 1989; Yimam et al. 2001; Kanai et al. 2006; Kudo et al. 2001). However, the expansion of the distribution of feral raccoon populations is remarkable, and well-established feral raccoon population now exists in Japan (Ikeda 2004). In addition, the raccoons utilized the residential areas positively to get food or to keep their covers (Ikeda et al. 2000). Although the prevalence of feral raccoon trichinellosis in Japan is lower than in North America, these habitual characters may increase the area potentially contaminated by this parasite and the risk of human trichinellosis. To better understand the epizootiology of trichinellosis, more information is needed concerning the distribution of this parasite in various regions in Japan.

Acknowledgments The present survey was supported in part by a grant-in-aid (nos.14560271, 18510205) and by the High Technological Research Center (Rakuno Gakuen University) of the Ministry of the Education, Science, and Culture of Japan. We are grateful for the raccoon samples provided to us by the local government offices, EnVision, Hokkaido Forest Management, Nopporo Natural Forest Park Office, Japan Wildlife Research Center and Raccoon Research Group. We are also grateful for Y. Asakawa, G. Abe, M. Sashika, and K. Tanida for the sampling and histological analysis.

References

Asakawa M, Kurachi T, Wildlife Ecological Society (1999) Parasitic helminths of raccoons in Hokkaido, Japan. Jpn J Zoo Wildl Med 4:101-103 (in Japanese with English abstract)

Asakawa M, Matoba Y, Yamada D, Kamiyama T (2000) Review of the parasitological state of feral raccoons captured in Nopporo National Park and its proximity, Hokkaido. Nat Sci 25:1–8 (in Japanese with English abstract)

Asano M, Matoba Y, Ikeda T, Suzuki M, Asakawa M, Ohtaishi N (2003) Reproductive characteristics of the feral raccoon (*Procyon lotor*) in Hokkaido, Japan. J Vet Med Sci 65:369–373

AVMA Panel on Euthanasia (2001) 2000 Report of the AVMA panel on euthanasia. J Am Vet Med Assoc 218:669-696

Cole RA, Shoop WL (1987) Helminthes of the raccoon (*Procyon lotor*) in Western Kentucky. J Parasitol 73:762-768

Grau GA, Sanderson GC, Rogers JP (1970) Age determination of raccoons. J Wildl Manage 34:364-372

Henriksen SA (1978) A new technique for demonstration of Trichinella spiralis larvae in suspensions of digested muscle tissue. Acta Vet Scand 19:466-468

Ikeda T (2004) Present status of invasive alien raccoon and its impact in Japan. Glob Environ Res 8:125-131



- Ikeda T, Endo M, Murano N (2000) Home range of invasive raccoons in Nopporo Forest Park. Journal of Rakuno Gakuen University 25:311-319
- Kanai Y, Nonaka N, Katakura K, Oku Y (2006) Trichinella nativa and Trichinella T9 in Hokkaido island, Japan. Parasitol Internatl 55:313-315
- Klein SL, Gamble HR, Nelson RJ (1999) Role of steroid hormones in Trichinella spiralis infection among voles. Am J Physiol 277: R1362–R1367
- Kudo N, Arima R, Ohtsuki M, Oyamada T (2001) The first host record of trichinosis in a red fox, *Vulpes vulpes japonica*, from Aomori Prefecture, Northern Honshu, Japan. J Vet Med Sci 63:823–826
- Mankau SK, Hamilton R (1972) The effect of sex and sex hormones on the infection of rats by *Trichinella spiralis*. Can J Zool 50:597–602
- Mascaro Lazcano C, Osuna Carrillo A, Guevara Pozo D (1978) The effect of the administration of testosterone propionate and dihydro stilboestrol diacetate on the susceptibility of the mouse infection with *Trichinella spiralis*. Rev Iber Parasitol 38:427–433
- Matoba Y, Asano M, Masubuchi H, Asakawa M (2002) First records of the genera *Eimeria* and *Isospora* (protozoa: *Eimeriidae*) obtained from feral raccoons (*Procyon lotor*) alien species in Japan and prevalence of serum antibodies to *Toxoplasma gondii* among the raccoons. Jpn J Zoo Wildl Med 7:87–90 (in Japanese with English abstract)
- Miyashita M (1993) Prevalence of *Baylisascaris procyonis* in raccoons in Japan and experimental infections of the worm in laboratory animals. Seikatsu-eisei 37:137–151 (in Japanese with English abstract)
- Pozio E, La Rosa G (2000) *Trichinella murrelli* n. sp. etiological agent of sylvatic trichinellosis in temperate areas of North America. J Parasitol 86:134–139
- Pozio E, La Rosa G, Rossi P, Murrell KD (1992a) Biological characterization of *Trichinella* isolates from various host species and geographical regions. J Parasitol 78:647-653
- Pozio E, La Rosa G, Murrell KD, Lichtenfels JR (1992b) Taxonomic revision of the genus *Trichinella*. J Parasitol 78:654-659

- Richardson DJ, Owen WB, Snyder DE (1992) Helminth parasites of the raccoon (*Procyon lotor*) from north-central Arkansas. J Parasitol 78:163–166
- Saito S, Yamaguchi T (1985) *Trichinella spiralis* in a raccoon dog, *Nyctereutes procyonoides viverrinus*, from Yamagata Prefecture, Honshu, Japan. Jpn J Parasitol 34:311–314
- Sato H, Suzuki K (2005a) Gastrointestinal helminths of feral raccoons (*Procyon lotor*) in Wakayama Prefecture, Japan. J Vet Med Sci 68:311-318
- Sato H, Suzuki K, Uni S, Kamiya H (2005b) Recovery of the everted cystacanth of seven acanthocephalan species of birds from feral raccoons (*Procyon lotor*) in Japan. J Vet Med Sci 67:1203–1206
- Sato H, Suzuki K, Osanai A, Kamiya H, Furuoka H (2006) Identification and characterization of the threadworm, Strongyloides procyonis, from feral raccoons (Procyon lotor) in Japan. J Parasitol 92:63–68
- Smith RA, Kennedy ML, Wilhelm WE (1985) Helminth parasites of the raccoon (*Procyon lotor*) from Tennessee and Kentucky. J Parasitol 71:599-603
- Snyder DE (1987) Prevalence and intensity of *Trichinella spiralis* infection in Illinois wildlife. J Parasitol 73:874-875
- Sukura A, Nareaho A, Mikkonen T, Niemi M, Oivanen L (2002) Trichinella nativa and T. spiralis induce distinguishable histopathologic and humoral responses in the raccoon dog (Nyctereutes procyonoides). Vet Pathol 39:257–265
- Yamaguchi T (1989) *Trichinella* and trichinellosis in Japan. Nankodo (in Japanese)
- Yamaguchi T (1991) Present status of trichinellosis in Japan. Southeast Asian J Trop Med Public Heal 22:295-301
- Yimam AE, Oku Y, Nonaka N, Sakai H, Morishima Y, Matsuo K, Rosa G la, Pozio E, Yagi K, Kamiya M (2001) First report of *Trichinella nativa* in red foxes (*Vulpes vulpes schrencki*) from Otaru City, Hokkaido, Japan. Parasitol Int 50:121–127
- Zarlenga DS, Chute MB, Martin A, Kapel CMO (1999) A multiplex PCR for unequivocal differentiation of all encapsulated and nonencapsulated genotypes of *Trichinella*. Int J Parasitol 29:1859– 1867

Detecting Clostridium botulinum

To the Editor: In the October 2005 issue of Emerging Infectious Diseases, Song et al. described a fiber-optic, microsphere-based, high-density array composed of 18 species-specific probe microsensors, used to identify biological warfare agents, including Clostridium botulinum (1). Although the researchers used multiple probes for C. botulinum, we doubt that this approach is suitable for this organism.

C. botulinum comprises a heterogenous group of subspecies that produce botulinum neurotoxin (BoNT); identification and characterization usually rely on animal testing that focuses on antigenetically distinct toxins (2). Although strains of C. botulinum that do not produce toxins are sometimes isolated from wound infections not related to botulism, some strains of C. butyricum and C. baratii are also able to produce BoNTs.

The mouse bioassay is currently the accepted method for detecting BoNT. In this assay, mice that receive an intraperitoneal injection containing a sample with more than a minimum lethal dose show symptoms of botulinum intoxication and die. ELISAs, which recognize protein antigenic sites, are still less sensitive than the mouse bioassay (3).

Because the mouse bioassay requires euthanizing many animals, and results are not available for several hours, new diagnostic methods are needed. For *C. botulinum*, an organism widely dispersed in the environment, DNA-based methods may not provide the ultimate solution. Rapid methods to detect and differentiate active BoNTs, such as the rapid, mass spectrometry-based, functional method, are promising candidates to substitute for animal testing in the near future (4).

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References

- Song L, Ahn S, Walt DR. Detecting biological warfare agents. Emerg Infect Dis. 2005;11:1629-32.
- Grif K, Dierich MP, Much P, Hofer E, Allerberger F. Identifying and subtyping species of dangerous pathogens by automated ribotyping. Diagn Microbiol Infect Dis. 2003;47:313-20.
- Ferreira JL, Eliasberg SJ, Edmonds P, Harrison MA. Comparison of the mouse bioassay and enzyme-linked immunosorbent assay procedures for the detection of type A botulinal toxin in food. J Food Prot. 2004;67:203-6
- Barr JR, Moura H, Boyer AE, Woolfitt AR, Kalb SR, Pavlopoulos A, et al. Botulinum neurotoxin detection and differentiation by mass spectrometry. Emerg Infect Dis. 2005;11:1578–83.

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Echinococcus multilocularis in Dogs, Japan

To the Editor: Alveolar echinococcosis in humans is endemic in Japan; however, the causal agent, Echinococcus multilocularis, has been restricted to the northernmost insular prefecture of Hokkaido, where the Tsugaru Strait acts as a natural physical barrier against migration to the mainland. Two E. multilocularis invasions into Hokkaido have occurred (1). The first invasion to the offshore island of Rebun in the mid-1920s was successfully controlled; however, the second invasion, sup-

posedly in the 1940s, led to the current epidemic on the main island of Hokkaido. Both invasions were entirely or partly caused by humans who removed foxes from disease-endemic areas without taking the necessary precautions.

The finding of 19 autochthonously acquired cases of alveolar echinococcosis in prefectures other than Hokkaido (2) implies that the parasite exists in other areas, although the source of infection has yet to be identified. In many countries, studies of the increased spread of the parasite have traditionally focused on the contribution of foxes (3); however, these cases may also have been spread by domestic dogs from disease-endemic areas. Dogs are susceptible to infection with the parasite from rodents. Although the prevalence of E. multilocularis among dogs in Hokkaido is certainly lower than that in foxes (4-6), dogs can traverse considerably greater distances by various modes of transport. The number of dogs that travel from Hokkaido to other prefectures has been estimated at >12,000 per year (7). Although dogs may carry the parasite to remote areas, surveys of population dynamics have not been undertaken. We therefore studied the extent of E. multilocularis infection in dogs being transported by their owners from 4 ferry ports in Hokkaido (Hakodate, Muroran, Otaru, and Tomakomai) from September 2003 through October 2004.

We tested 183 fecal samples from 41 resident (in Hokkaido) and 142 nonresident dogs. We screened for the *Echinococcus*-specific coproantigen by using a commercial enzyme-linked immunosorbent assay kit (CHEKIT-Echinotest, Bommeli Diagnostics, Liebefeld-Bern, Switzerland) and following the manufacturer's recommendations. One dog from each group had the *Echinococcus* coproantigen. To confirm the specificity of the results, these 2 dogs were treated with 1 oral dose of praziquantel, 5 mg/kg.

Subsequent fecal samples were subjected to coproantigen testing and specific PCR amplification according to the method of Dinkel et al. (8). The coproantigen test showed a significant reduction in the optical density value for both dogs, which can be interpreted as effective deworming for Echinococcus. However, different results were obtained for the PCR test, in which assays of fecal samples from the nonresident dog during the second round of nested PCR produced a single band of the expected size (Figure). Direct sequencing showed that the band was the same as bands obtained for E. multilocularis isolates from Hokkaido (GenBank accession no. AB243207). Conversely, fecal samples from the resident dog did not yield any positive PCR results.

The reason for the discrepancy is unclear, but it may be a false reaction in either test. Given that a reduced optical density value was obtained after administration of the taeniacidal drug, the false-positive result of the coproantigen test might have been caused by another taeniid species.

Such cross-reaction has been reported previously with this test (9). However, no worm debris was found in the fecal samples. Alternatively, sexual maturation or low infection intensity of *E. multilocularis* may produce false-negative results in PCR assays (8). Thus, because the owner stated that the dog was allowed to roam freely and frequently preyed on rodents, this coproantigen-positive but coproDNA-negative dog was highly suspected of being infected with *E. multilocularis*.

Infection among wild foxes can spread to domestic dogs by way of highly contaminated rodent hosts (10). A nonresident dog became infected with E. multilocularis despite staying in Hokkaido for only 5 days and being permitted to roam freely for just a few hours. This finding suggests a high infection pressure of E. multilocularis to domestic dogs within the area. In addition, the increased popularity of keeping dogs as companions, greater frequency of dogs' traveling with their owners, and high prevalence in foxes from urban and rural areas in Hokkaido (5,6) all contribute to the possibility that *E. multilocularis* could emerge in unsuspected locations. Thus, to prevent this parasite from spreading, measures such as those used by the Pet Travel Scheme of the United Kingdom should be applied to ensure that dogs from disease-endemic areas are pretreated before entry to the main island of Japan.

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References

- Yamashita J. Echinococcus and echinococcosis. In: Morishita K, Komiya Y, Matsubayashi H, editors. Progress of medical parasitology in Japan. Volume 5. Tokyo: Meguro Parasitological Museum. p. 65–123
- Doi R, Kanda E, Nihei N, Uchida A. Occurrence of alveolar hydatid disease (multilocular echinococcosis) outside of Hokkaido and a proposal for its prevention. Nippon Koshu Eisei Zasshi. 2000;47:111–26.
- Eckert J, Conraths FJ, Tackmann K. Echinococcosis: an emerging or re-emerging zoonosis? Int J Parasitol. 2000;30: 1283-94.
- 4. Kamiya M, Morishima Y, Nonaka N, Oku Y. Epidemiologic surveys of Echinococcus multilocularis in the definitive hosts by coproantigen detection between 1996–2000. In: Proceedings of the Joint Meeting of the 70th Annual Meeting of Japanese Society of Parasitology and the 53rd Annual Meeting of the Japan Society of Medical Entomology and Zoology; 2001. Apr 4–6; Yamagata City, Yamagata Prefecture, Japan. Abstract P30W2–1.

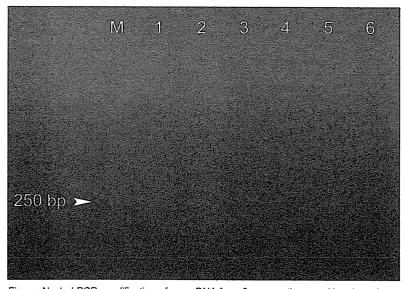


Figure. Nested PCR amplification of coproDNA from 2 coproantigen-positive dogs. Lane M, size marker (100-bp ladder); lane 1, nonresident dog (before treatment); lane 2, nonresident dog (1 day after treatment); lane 3, resident dog (before treatment); lane 4, resident dog (1 day after treatment); lane 5, positive control; lane 6, negative control. Arrowhead shows the expected band in a positive result.

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- Morishima Y, Tsukada H, Nonaka N, Oku Y, Kamiya M. Coproantigen survey for Echinococcus multilocularis prevalence of red foxes in Hokkaido, Japan. Parasitol Int. 1999;48:121-34.
- Tsukada H, Morishima Y, Nonaka N, Oku Y, Kamiya M. Preliminary study of the role of red foxes in *Echinococcus multilocularis* transmission in the urban area of Sapporo, Japan. Parasitology. 2000;120:423–8.
- Doi R, Matsuda H, Uchida A, Kanda E, Kamiya H, Konno K, et al. Possibility of invasion of *Echinococcus multilocularis* into Honshu with pet dogs from Hokkaido and overseas. Nippon Koshu Eisei Zasshi. 2003;50:639-49.
- Dinkel A, von Nickisch-Rosenegk M, Bilger B, Merli M, Lucius R, Romig T. Detection of *Echinococcus multilocularis* in the definitive host: coprodiagnosis by PCR as an alternative to necropsy. J Clin Microbiol. 1998;36:1871-6.
- Manfredi MT, Genchi C, Deplazes P, Trevisiol K, Fraquelli C. Echinococcus multilocularis infection in red foxes in Italy. Vet Rec. 2002;150:757.
- Gottstein B, Saucy F, Deplazes P, Reichen J, Demierre G, Busato A, et al. Is high prevalence of *Echinococcus multilocularis* in wild and domestic animals associated with disease incidence in humans? Emerg Infect Dis. 2001;7:408-12.

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New World Hantavirus in Humans, French Guiana

To the Editor: Hantaviruses are etiologic agents for hemorrhagic fever with renal syndrome in Europe and Asia and for hantavirus pulmonary syndrome (HPS) in the Americas. These viruses belong to the family Bunyaviridae, genus Hantavirus. The natural reservoir of these viruses is wild or domestic rodents. HPS was

first described in 1993 in the Four Corners region of the United States (1). It is a respiratory illness associated with the inhalation of aerosolized rodent excreta (urine and feces) contaminated with hantavirus particles. Sin Nombre virus (SNV) was the first etiologic agent of this syndrome. Since 1993, HPS has also been reported and confirmed in 6 countries in South America: Argentina, Bolivia, Brazil, Chile, Paraguay, Uruguay (2,3). Several distinct hantaviruses have been associated with HPS, including Juquituba virus in Brazil (4), Andes virus in Southern Argentina (5), and Laguna Negra virus in Paraguay (6).

French Guiana, an overseas French Administrative Unit in the Amazonian forest complex, is located on the northeastern coast of the South America between Brazil and Suriname. Ninety percent of its surface is tropical rain forest; the remaining 10% is a coastal plain, where 90% of the 200,000 inhabitants live. Cayenne and 2 adjacent towns, Remire and Matoury, constitute the main urban centers, with 80,000 inhabitants, ≈40% of the population. People live mainly in individual houses and small buildings. Many houses are built near forests, except those in the center of Cayenne. The outskirts of Remire and Matoury are surrounded by secondary rain forest, and those of Cayenne by wooded hills, where wild mammals such as rodents live in large numbers.

The prevalence of antibodies to New World hantavirus is unknown in French Guiana. Several cases of atypical pneumonia not linked to other etiologic agents (*Coxiella burnetii*, *Histoplasma boydii*), combined with identification of hantavirus rodent reservoirs in neighboring countries, prompted us to determine the seroprevalence of hantavirus in this area (7,8).

To estimate the prevalence of antibodies to New World hantavirus, we

conducted a retrospective serologic survey of patients with symptoms compatible with HPS. Patients were from all areas of French Guiana: 64% from the urban centers, 7% from rural regions, and 30% from unspecified regions. From April 2002 through April 2004, a total of 420 serum samples were collected from patients with acute-phase febrile illness, unexplained acute respiratory syndrome, or bilateral interstitial pulmonary infiltrates. Diagnosis of Q fever was excluded by negative serologic results for immunoglobulin M (IgM), IgG, or both to C. burnetii (bioMérieux, Marcy-l'Etoile, France).

To detect patients with IgG antibodies to SNV, the ELISA described by Feldmann et al. was used (9). Briefly, an SNV-positive serum provided by the Centers for Disease Control and Prevention (CDC, Atlanta, GA, USA) was used as a positive control. Negative controls were obtained by random sampling of all previously negative samples. A sample was considered positive if the net absorbance values (after subtraction of absorbance values with and without antigen) were >0.2 for dilutions of 1:100 and 1:400 and the sum of 4 net absorbance values was >0.95. Seropositive samples were confirmed at CDC.

Antibodies reactive with SNV antigen indicate infection with a New World hantaviruses. However, because SNV is broadly cross-reactive with most New World hantavirus, the specific hantavirus cannot be identified.

The seroprevalence of IgG antibody to hantavirus was 1.42% (6/420) in the selected population. Three other samples showed borderline positivity. Antibody prevalence was not significantly different among the 7 age classes used (0–9, 10–19, 20–29, 30–39, 40–49, 50–59, and >60 years of age, p = 0.36, degrees of freedom = 6, by χ^2 test) or by sex (p = 0.22, by Fisher exact test).

Laboratory and Epidemiology Communications

The First Reported Case of a Dog Infected with Echinococcus multilocularis in Saitama Prefecture, Japan

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Echinococcus multilocularis is a causative agent of human alveolar hydatidosis. The distribution of the parasite in Japan is thought to be limited to Hokkaido, the northernmost insular prefecture. The life cycle of this parasite is predominantly

sylvatic. In Hokkaido, the red fox (*Vulpes vulpes*) is a definitive host that excretes *E. multilocularis* eggs into its feces. The gray-sided vole (*Clethrionomys rufocanus*) is a principal intermediate host that becomes infected by ingesting the eggs, which develop into larvae in the host viscera. Domestic dogs also become definitive hosts of the parasite after eating voles harboring the larvae. Since human infections follow the accidental ingestion of *E. multilocularis* eggs, a survey of *E. multilocularis* infection in dogs is a crucial for disease

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prevention. Data obtained from an examination of 9,907 dogs in Hokkaido between 1966 and 2004 by the prefectural government revealed that 99 (1.0%) were infected with the adult parasites. The estimated number of dogs that travel from Hokkaido to other prefectures is over 12,000 per year (1). Some of these dogs might constitute important carriers, transmitting the parasite to remote areas. Morishima et al. (2) recently studied the extent of *E. multilocularis* infection in 183 dogs transported by their owners from Hokkaido to other prefectures and found that two of the dogs were infected. Here we describe the first case of a dog infected with *E. multilocularis* in Saitama Prefecture, which neighbors northern metropolitan Tokyo.

The dogs examined in this survey were abandoned and pound animals. A total of 550 dogs kept in the Saitama Prefectural Pet Owner's Guidance Center were examined between April 1999 and September 2005. Staff members at the center collected fecal samples. These were routinely checked for the presence of helminth eggs and protozoan oocysts by microscopy using direct smear, formalin-ether and sucrose flotation techniques. Taeniid eggs (Fig. 1) were found in a fecal sample from a female mongrel dog that was captured in northern Saitama in June 3, 2005. Because Echinococcus eggs are morphologically indistinguishable from those of other tapeworms of the family Taeniidae (3), they were examined by PCR according to the method of Dinkel et al. (4). The second round of nested PCR produced a single band of the predicted size at 250 bp. Direct sequencing showed that the band was the same as those found in E. multilocularis

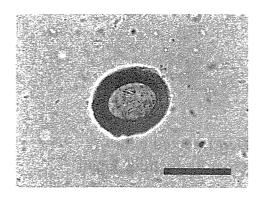


Fig. 1. Echinococcus multilocularis egg found in the fecal sample of a dog captured in Saitama Prefecture, Japan. Scale bar equals 30 $\,\mu\mathrm{m}$.

isolates from Hokkaido (GenBank accession no. AB244598). These results showed that this dog was infected with *E. multilocularis*.

The source of infection was not identified. No convincing evidence yet supports the notion that wild animals in Saitama and its neighboring prefectures are infected. Since some dogs that move from Hokkaido to the mainland of Japan are infected (2), the dog described herein seems to have been infected in Hokkaido, then taken to Saitama by her owner, and then either abandoned or allowed to escaped. Regardless, an extensive epizootiological survey should be conducted on wild animals in suspect areas of the prefecture. The Infectious Diseases Control Law of Japan has classified the significance of this disease to public health as Category IV, which has required the mandatory reporting of not only infected humans but also infected dogs since October 1, 2004. In accordance with this law, this is the first reported instance of a dog infected with $E.\ multilocularis$ in a Japanese prefecture other than Hokkaido.

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REFERENCES

- 1. Doi, R., Matsuda, H., Uchida, A., Kanda, E., Kamiya, H., Konno, K., Tamashiro, H., Nonaka, N., Oku, Y. and Kamiya, M. (2003): Possibility of invasion of *Echinococcus multilocularis* into Honshu with pet dogs from Hokkaido and overseas. Jpn. J. Public Health, 50, 639-649 (in Japanese with English summary).
- Morishima, Y., Sugiyama, H., Arakawa, K. and Kawanaka, M. (2006): *Echinococcus multilocularis* in dogs, Japan. Emerg. Infect. Dis., 12, 1292-1294.
- 3. Thompson, R.C.A. and McManus, D.P. (2001): Aetiology: parasites and life-cycles. p 1-19. *In* Eckert, J., Gemmell, M.A., Meslin, F.-X., Pawlowski, Z.S. (ed.), WHO/OIE Manual on Echinococcosis in Humans and Animals: a Public Health Problem of Global Concern. World Organization for Animal Health, Paris.
- 4. Dinkel, A., von Nickisch-Rosenegk, M., Bilger, B., Merli, M., Lucius, R. and Romig, T. (1998): Detection of *Echinococcus multilocularis* in the definitive host: coprodiagnosis by PCR as an alternative to necropsy. J. Clin. Microbiol., 36, 1871-1876.

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Intestinal helminths of dogs in northern Japan

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HUMAN beings can become infected by a number of canine intestinal helminths, some of which are considered to be potential public health problems worldwide. However, intestinal worms of dogs currently receive less attention than their protozoan counterparts. This may partly be due to the decline in endemicity reported in developed countries, where dogs are likely to receive more attention and anthelmintic treatment from their owners. To illustrate the importance of veterinary care, a longitudinal study in Japan revealed that progress in prophylactic drug administration against dirofilariosis had reduced the prevalence of several intestinal nematodes in dogs (Asano and others 2004). The low levels of prevalence among canine hosts translate into a reduction in potential zoonotic risk for human beings in a given area. However, endemicity has been found to vary markedly from one region to another, and is also influenced by aspects of the survey protocols, such as subject choice and the diagnostic techniques that are employed (Robertson and others 2000).

This short communication is part of an ongoing study on the zoonotic helminths of Japan. It describes a coprological survey of dogs in Aomori, the northernmost prefecture on the mainland, where no such information is currently available. The analyses focused on hunting dogs, as the way in which they are managed and their activities in fields makes them more susceptible to acquiring a variety of zoonotic helminths, either directly or indirectly from wildlife. Faecal samples were collected from 134 hunting dogs and 86 companion dogs between December 2003 and March 2005. Two different diagnostic techniques were employed: a centrifugal flotation technique (Ito 1980) with a sucrose solution with a specific gravity of 1-27, and a formalin-ethyl acetate sedimentation technique (Young and others 1979).

With the exception of Trichuris species and Taeniidae, all eggs were identified to genus or genus and species on the basis of morphological characteristics. The eggs of T vulpis are morphologically similar to those of other Trichuris species (Bundy and Cooper 1989), and parasitic eggs may be excreted after being acquired from other animals through coprophagy (Traub and others 2002). A total of 100 randomly selected trichurid eggs from positive faecal specimens were therefore re-examined according to the dimensions described by Yoshikawa and others (1989). Given the difficulty of distinguishing between taeniid tapeworm eggs on the basis of shape and size (Thompson 1995), Taenia and Echinococcus species were differentiated using nested PCR (Dinkel and others 1998). Pearson's χ^2 test was used to assess differences in the egg prevalence of each helminth between both groups. McNemar's χ^2 test was used to compare the diagnostic efficiency of the coprological techniques. All statistical analyses were performed using a statistical package (S-PLUS 6.1J for Windows; Mathematical System).

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The results from both the hunting dogs and companion dogs are summarised in Table 1. T vulpis was the most common helminth in both groups, and the prevalence was significantly higher in hunting dogs (P<0.001). Although comparison of results is difficult because a previous study determined prevalence by postmortem examination (Yagisawa 1978), the occurrence of T vulpis in dogs in Aomori appears to have decreased, and the numbers of other intestinal worms have also exhibited an appreciable decline. The reason for the remaining prevalence, particularly in hunting dogs, is uncertain, but these dogs are thought to be reservoir hosts; T vulpis is prevalent among wild canids in Aomori (Sato and others 1999a,b). The hunting dogs may have become infected with this parasite in fields that were contaminated by the sylvatic hosts, and have then transmitted the parasite to the local dog population.

In addition, the choice and/or usage of drugs is particularly important. Several anthelmintics are used to prevent canine dirofilariasis. Of these, ivermectin (Cardomec; Mweial) is the most commonly used for this purpose in Japan. However, ivermectin has no effect on intestinal trichuriasis at the usual chemoprophylactic dosage (6 μ g/kg bodyweight), and an increase in concentration of more than 16-fold is required (Anderson and Robertson 1982). Ivermectin at this concentration also showed an effect on other intestinal helminths such as *Toxocara canis*.

Regarding the relative sensitivities of each diagnostic method (Table 2), the sedimentation technique had a significantly higher positive rate of trichurid egg detection than did the centrifugal flotation technique (P=0·016). Notably, the centrifugal flotation technique failed to detect instances of low egg density/abundance (less than 10 eggs per mount in the competitor). Because of the relatively low fecundity of T vulpis (Miller 1941), the sedimentation technique appears to be a preferable means of detection. Despite its apparent zoonotic potential (Dunn and others 2002), the prevalence of T vulpis has been somewhat overlooked. Consequently, underestimation or underscoring the potential of this parasite is likely to occur if inadequate diagnostic tests are adopted.

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References

- ANDERSON, D. L. & ROBERTSON, E. L. (1982) Activity of ivermectin against canine intestinal helminths. *American Journal of Veterinary Research* 43, 1691, 1692
- ASANO, K., SUZUKI, K., MATSUMOTO, T., SAKAI, T. & ASANO, R. (2004) Prevalence of dogs with intestinal parasites in Tochigi, Japan in 1979, 1991 and 2002. Veterinary Parasitology 120, 243-248
- BUNDY, D. A. P. & COOPER, E. S. (1989) *Trichuris* and trichuriasis in humans.

 Advances in Parasitology 28, 107-173

 DINKEL, A., VON NICKISH-ROSENEGK, M., BILGER, B., MERLI, M.,
- DINKEL, A., VON NICKISH-ROSENEGK, M., BILGER, B., MERLI, M., LUCIUS, R. & ROMIG, T. (1998) Detection of *Echinococcus multilocularis* in the definitive host: coprodiagnosis by PCR as an alternative to necropsy. *Journal of Clinical Microbiology* 36, 1871-1876
- DUNN, J. J., COLUMBUSM, S. T., ALDEEN, W. E., DAVIS, M. & CARROLL, K. C. (2002) *Trichuris vulpis* recovered from a patient with chronic diarrhoea in five dogs. *Journal of Clinical Microbiology* 40, 2703-2704

FOR PROOFING PURPOSES ONLY. PLEASE PRINT OUT THIS PDF AND RETURN YOUR CORRECTED PROOF TO THE ADDRESS SHOWN ON PAGE 1

- ITO, S. (1980) Modified Wisconsin sugar centrifugal-flotation technique for nematode eggs in bovine faeces. Journal of the Japan Veterinary Medical Association 33, 424-429
- $MILLER, M. J. (1941) \ Quantitative studies on \it Trichocephalus \it vulpis infections in dogs. \it American Journal of Hygiene 33, 58-70$
- ROBERTSON, I. D., IRWIN, P. J., LYMBERY, A. J. & THOMPSON, R. C. A. (2000) The role of companion animals in the emergence of parasitic zoonoses. *International Journal for Parasitology* 30, 1369-1377
- SATO, H., IHAMA, Y., INABA, T., YAGISAWA, M. & KAMIYA, H. (1999a) Helminth fauna of carnivores distributed in north-western Tohoku, Japan, with special reference to Mesocestoides paucitesticulus and Brachylaima tokudai. Journal of Veterinary Medical Science 61, 1339-1342
- SATO, H., INABA, T., IHAMA, Y. & KAMIYA, H. (1999b) Parasitological survey on wild carnivores in north-western Tohoku, Japan. *Journal of Veterinary Medical Science* 61, 1023-1026
- THOMPSON, R. C. A. (1995) Biology and systematics of *Echinococcus*. In *Echinococcus* and Hydatid Disease. 1st edn. Eds R. C. A. Thompson, A. J. Lymbery. Wallingford, CAB International. pp 1-50
- TRAUB, R. J., ROBERTSON, I. A., IRWIN, P., MENCKE, N. & THOMPSON, R. C. A. (2002) The role of dogs in transmission of gastrointestinal parasites in a remote tea-growing community in northeastern India. *American Journal of Tropical Medicine & Hygiene* 67, 539-545
- YAGISAWA, M. (1978) Studies on zoonotic helminths from mammals in northern Honshu, Japan. *Hirosaki Medical Journal* 30, 239-284
- YOSHIKAWA, H., YAMADA, M., MATSUMOTO, Y. & YOSHIDA, Y. (1989) Variations in egg size of *Trichuris trichiura*. Parasitology Research 75, 649-654
- YOUNG, K. H., BULLOCK, S. L., MELVIN, D. M. & SPRUILL, C. L. (1979) Ethyl acetate as a substitute for diethyl ether in the formalin-ether sedimentation technique. *Journal of Clinical Microbiology* 10, 852-853

| | Hunting dog | S | Companion dogs | | | |
|--------------------|---------------------|-----------|---------------------|-----------|--|--|
| Helminth species | Number (%) positive | 95% CI | Number (%) positive | | | |
| Toxocara canis | 0 (0) | 0-2-2 | 4 (4-7) | 1-3-11-5 | | |
| Trichuris vulpis | 52 (38-8) | 31-0-47-3 | 14 (16-3) | 10-0-25-5 | | |
| Dipylidium caninum | 0 (0) | 0-2-2 | 2 (2-3) | 0-3-8-2 | | |
| Taenia species | 1 (0-7) | 0.02-4-1 | 0 (0) | 0-3-4 | | |
| Metagonimus specie | s 2 (1:5) | 0-2-5-3 | 0 (0) | 0-3-4 | | |

TABLE 2: Comparison of the sensitivity of centrifugal flotation and formalin-ethyl acetate sedimentation techniques for the detection of *Trichuris vulpis* eggs in 220 canine faecal samples

| | S | edimentation | |
|-------------|------------|--|-------------|
| | Posi | tive Nega | ative Total |
| Centrifugal | Positive 5 | 5 | 56 |
| flotation | | TAKE - 10 TO TO TO THE OWNER OF THE PARTY OF | 4 164 |
| Total | 6 | 5 15 | 5 220 |

神奈川県におけるアライグマの駆除と アライグマ回虫の調査

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Key Words: アライグマ, アライグマ回虫, 神奈川県, 外来生物法, 監視体制

はじめに

アライグマ回虫 (Baylisascaris procyonis) は、アライグマの原産地である北米大陸において普通に認められる腸管内寄生虫で、基本的にアライグマ以外の動物では成虫になることはないが、ヒトがその虫卵を経口摂取すると幼虫移行による致死的な中枢神経障害を引き起こすことがある。米国では、1984年と1985年に相次いで幼児死亡例が確認され、最悪の「動物由来寄生虫症」と目されており、現在までに5名の死亡例を含む少なくとも13名の重症例が報告されている10。

近年、わが国では展示用動物やペットに由来するアライグマの野生化が、農業被害や在来種の絶滅危惧(生物多様性に及ぼす悪影響)などの問題を引き起こし、特に、北海道、神奈川、岐阜、愛知、和歌山などの各県では深刻な事態となっている。このような状況から、2005年6月に施行された「外来生物法」ではアライグマを「特定外来生物」に指定し、野生

個体の積極的な駆除と、輸入、飼育、販売の原則禁止が求められることとなった。それと共に、野生アライグマの駆除作業に伴うアライグマ回虫による健康被害の予防対策も重要となっている。

われわれは 1999 年からアライグマ回虫に関する 全国調査を開始し、動物園等の展示施設のアライグ マ、ならびに捕獲された野生アライグマについての 調査報告の収集や糞便検査等を実施した ^{2) 3)}。 その 結果、動物園等での飼育群からはアライグマ回虫の 寄生例が少なからず確認されたが、全国の野生アラ イグマからは、現在のところアライグマ回虫の寄生 例は発見されていない。神奈川県は、首都圏にあっ て最も野生アライグマ問題が先鋭化している地域で あることから、この調査を開始して以来 7 ヵ年間 にわたりアライグマの生息状況をフォローすると共 に、直接アライグマ回虫の検査を実施してきた。「外 来生物法」の施行に伴って野生アライグマの駆除が 全国的に実施されつつある現在、神奈川県を国内で のアライグマ回虫問題の一典型として、現在の問題

Surveillance of *Baylisascaris procyonis* Infection among Wild Raccoons in Kanagawa Prefecture, Japan

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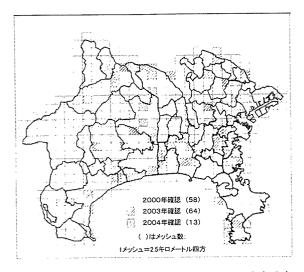


図1 神奈川県における野生化アライグマの生息分布の拡大状況(計画書5)に依る)

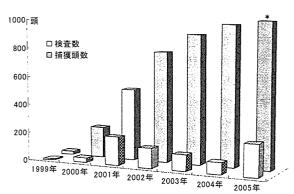


図2 捕獲した野生化アライグマの数と糞便検査実施 状況

*:捕獲頭数は計画書5)に依り、2005年は推定頭数.

点の整理をする。

県内動物園で展示されているアライグマのアライグ マ回虫検査

神奈川県には、社団法人日本動物園水族館協会に 所属している動物園が5施設、その他小規模のいわ ゆる「ふれあい系動物園」が少なくとも14施設存 在する。アライグマの展示がなされているのは、こ のうちの5施設で、個体数は合わせて13頭であっ た。2001年に、「ふれあい系動物園」の1施設にお いて、飼育されている複数のアライグマからアラ イグマ回虫卵が検出された。調査の結果、その前年 に他県の動物園から導入したアライグマがアライグ

表 1 神奈川県における捕猟アライグマ数と糞便検査 実施状況

| LE WELD TO | 捕獲年 | | | | | 地区別 | | |
|------------|------|------|------|------|------|------|------|-----|
| 捕獲場所 | 1999 | 2000 | 2001 | 2002 | 2003 | 2004 | 2005 | 検査数 |
| 鎌倉市 | 8 | 26 | 140 | 80 | 104 | 46 | 73 | 477 |
| 横須賀市 | | | | 9 | | 4 | 96 | 109 |
| 藤沢市 | | 3 | 38 | 22 | 5 | 5 | 9 | 82 |
| 横浜市 | | | 5 | 4 | 4 | 19 | 20 | 52 |
| 逗子市 | | | 5 | 8 | | 5 | 23 | 41 |
| 相模原市 | | 1 | 8 | | | | | 9 |
| 三浦市 | | | | 5 | | 2 | | 7 |
| 茅ヶ崎市 | | | | 5 | | | l | 6 |
| 城山町 | • | | 2 | | | | | 2 |
| 津久井町 | | 2 | | | | | | 2 |
| 寒川町 | | 1 | | | | | | 1 |
| 小田原市 | | | 1 | | | | | 1 |
| 不明 | | | | 6 | | | | 7 |
| 年別検査総計 | 8 | 33 | 199 | 139 | 113 | 81 | 222 | 796 |

マ回虫に感染していたものと推定され、その個体によって以前からの飼育アライグマにも感染が広がったものと判明した。この施設では、木造什器の焼却処分、ならびに飼育区域の床、壁などの熱湯処理による虫卵徐染作業を行い、全飼育個体について糞便検査による確認を行いつつ数ヵ月にわたる継続的な駆虫を実施することによって汚染を終息させた。そして、その後も獣医師による定期的な監視が続けられている。

神奈川県における野生アライグマのアライグマ回虫 検査

神奈川県における野生アライグマは、鎌倉市において1988年頃に目撃されたものが最初である⁴⁾。その後分布を拡げてゆき、三浦半島とその周辺域で農業被害が増大し始めたのが1999年であった。さらには、三浦半島から鎌倉、湘南までの地区で高密度に繁殖しつつ、その他の地区でもさまざまな被害情報や捕獲例が増加するなど生息域を急速に拡大していることが確認された(図1)。有害(鳥)獣として捕獲されたアライグマは1998年(平成10年度)では僅かに4頭であったが、以後、急速に増加し、2003年は11市町で903頭、2005年には1,000頭近くになっている。そして2006年3月に発表された「神奈川県アライグマ防除実施計画」では、この時点での県内の野生アライグマの推定生息数を4,030

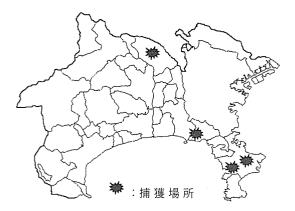


図3 神奈川県内のタヌキ回虫寄生アライグマ 捕獲場所

頭としている⁵⁾。

表1は、現在までの捕獲アライグマの糞便検査の 実績を地区別にまとめたものである。検査サンプル は、鎌倉市、横須賀市、藤沢市、横浜市、逗子市で 捕獲されたものから採取されたものが全体の95% を占める。また、図2はこのようにして実施したア ライグマ回虫の検査が、この期間に捕獲されたアラ イグマに対してどれ程の比率にあるかを示したもの である。結論的に言えば、1999 年以来、県内で捕 獲された野生アライグマは約4,400 頭であり、その うちの約800 頭 (17%) について糞便検査を実施し たが、これらの糞便からはアライグマ回虫卵は検出 されなかった。

捕獲アライグマからのタヌキ回虫の検出

これまでの調査において、捕獲アライグマからはアライグマ回虫は検出されなかったが、タヌキ回虫(Toxocara tanuki)の寄生例が見つかっている。最初の例は、2001年に相模原市で捕獲された野生アライグマからタヌキ回虫の虫卵と成虫が検出されたものである。アライグマ回虫卵の形態はタヌキ回虫卵と類似しているため、これらの種鑑別には成虫を採取してその形態的特徴を確認することが望ましい。しかし成虫が採集できなかった場合は、虫卵由来の DNA による PCR を実施し遺伝子を解析することで同定が可能である プ。タヌキ回虫は、その後、図3に示す様に鎌倉市からの1頭、横須賀市からの2頭を加えて、現在までに捕獲アライグマ4頭から

その寄生例が確認されている。

考察

野生アライグマに関する行政的な対応は、2005年に「外来生物法」が施行されたことにより大きな転換期を迎えた。神奈川県においては、環境省からアライグマ対策モデル事業の指定を受けて2005年の夏に横須賀市内の繁殖地を重点調査域として生態調査を行い、その調査結果を踏まえて2006年3月には10ヵ年で県全域からの完全排除を目標とした「神奈川県アライグマ防除実施計画」が策定されたが(http://www.pref.kanagawa.jp/osirase/ryokusei/ysi/keikakuhonbun.)。しかし、この計画は県の環境農政部門が推進し市町村が実施母体となるために衛生行政との連携が不十分であり、アライグマ回虫を始めとする動物由来感染症への対策が十分に実施され難いという問題点がある。

現在まで、神奈川県の野生アライグマからはアラ イグマ回虫検出例は1例もないが,動物園でアラ イグマ回虫が検出された事例があり、今後も何らか の機会に感染個体が野生化する可能性を否定できな い。アライグマ回虫に関しては、1頭でも感染獣の 侵入があれば非感染獣への伝播は容易に起こりうる こと、しかも虫卵汚染の除去を徹底して行わない限 り, 容易に再感染を繰返すことが確認されている3)。 また, アライグマ回虫卵は粘着性が強いため除去が 容易でなく,また自然環境下での耐熱耐寒性が強く, クレゾールなどの消毒薬にも強い抵抗性を示す上, 数年にわたって感染力を保持する。そのため一旦, 野生アライグマの中に感染獣が発見された場合、そ の地域における虫卵汚染対策は困難を極めることが 予想され、何よりも迅速な状況把握と対応が求めら れる。

一方、これまでに捕獲アライグマから4例のタヌキ回虫が検出されたが、これはタヌキの生息域にアライグマが侵入したために起きたことであり、今後はタヌキ回虫の伝播がアライグマ個体間でも容易に起こりうると推定される。タヌキ回虫は、イヌ回虫(Toxocara canis)と同じトキソカラ属で、ヒトに対してイヌ回虫程度の幼虫移行症を起こしうると考えられているが、アライグマ回虫ほどの重篤な障害を

起こす可能性は低い。従ってこれからは、捕獲アライグマから回虫卵を検出した際には、行政的な対応の必要性を判断するためにも必ずタヌキ回虫卵との鑑別を行うことが必要になったと云える。これらの事実を踏まえて、現在進行中のアライグマ駆除事業に、自治体レベルでのアライグマ回虫監視と緊急時に対応できる連絡体制を組み込むことが急務であると考える。

謝辞:稿を終えるに当たり、検査サンプルの蒐集にご協力を頂いた神奈川県内の自治体担当職員とアライグマの捕獲に携わっている民間委託業者の方々にお礼を申し上げます。

文 献

1) Gavin, P. J. et al. (2005) : Baylisascariasis. Clin

- Microbiol Rev, 18, 703-718.
- 2) 川中正憲, 他 (2001): アライグマとアライグマ回 虫 (*Baylisascaris procyonis*) に関する全国調査. Clin Parasitol, 12, 121-125.
- 川中正憲,他(2002):動物園,観光施設でのアライグマ回虫卵汚染問題.病原微生物検出情報, Vol. 23, No. 8, 202-203.
- 4) 中村一恵 (1991): 神奈川県におけるアライグマの 野生化. 神奈川県自然誌資料 12, 17-19.
- 5) 神奈川県環境農政部緑政課野生生物班(2006):神 奈川県アライグマ防除実施計画(平成18年3月).
- 6) 森嶋康之,他(2002):神奈川県で捕獲された野生 化アライグマから検出された回虫について.第71 回日本寄生虫学会大会プログラム抄録集,pp152.
- 7) 杉山 広,他(2003):幼虫移行症の原因としての アライグマ回虫、獣医寄生虫学会誌,2,13-19.



エキノコックス (幼虫) エキノコックス (幼虫)

図 1 エキノコックス症の感染経路

埼玉県で捕獲犬一頭からエキノコック ス虫卵を検出

2005年9月9日,"「エキノコックス」埼玉で確認","野犬一頭からエキノコックス,北海道から来る?"といったニュースが,新聞やテレビで報じられたことをご存知だろうか。埼玉県の動物指導センターと衛生研究所が,調査事業のなかで埼玉県北部を放浪していたイヌを捕獲して糞便を検査し,国内では北海道でしかみつからないはずのエキノコックス(多包条虫)の虫卵を検出したというものである。詳しい経過は,国立感染症研究所(感染研)で出している「病原微生物検出情報」で報告されている1).

本稿では、イヌのエキノコックスはヒトの病気とどんな関係にあるのか?ということと、近年進歩したイヌのエキノコックス検査法について述べる。1999年に施行された感染症の予防及び感染症の患者に対する医療に関する法律(感染症法)により、ヒトのエキノコックス症は「4類感染症」として医師からの届け出を要する疾病となり、2004年10月より、イヌのエキノコックス感染事例も獣医師による届け出を要するものとされているからである²¹.

イヌのエキノコックスはヒトの病気とどんな関係にあるのか?

日本国内では、エキノコックス(多包条虫)は、

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現在のところ北海道にだけ分布する寄生虫である。成虫はキツネの腸管に寄生し、排泄する糞に混じって外へ出た虫卵が野ネズミに摂食されると、肝臓などで幼虫(包虫)が増殖する。こうした感染野ネズミがキツネに捕食され、腸管内でエキノコックスが幼虫から成虫になることで生活環が維持されている(図 1).

ヒトは野ネズミと同様に虫卵を飲み込むことでエキノコックスに感染するが、中間宿主として好適ではないので幼虫の増殖には長期間を要する. 感染から発症に至るのは5~20年後であり、肝腫大、腹痛、黄疸などの症状から始まり、有効な治療が施されなければ死亡率は90%ときわめて高い.

イヌの終宿主としての好適性はキツネと差異はないと考えられるが、生活圏と食性からみて、イヌが野ネズミを摂食することは一般にまれとされていた。それでも最近の調査では、北海道で飼育されているイヌの1~3%がエキノコックスに感染していることがわかっている。イヌは感染してもとくに症状はなく元気なので、症状からその感染の有無を知ることはできない。感染犬からの虫卵の排出は、感染後4週から始まりおおむね半年間に及ぶ。飼育犬はヒトとの関係が密接なので、周囲の人々はこの期間、感染の危険に曝されるの

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