

Discussion

Our study had three major findings in Japanese obese subjects. First, SNP45 in the adiponectin gene was associated with body fat distribution. Second, SNP45 was associated with the development of carotid atherosclerosis. Moreover, SNP45 had an impact on the effect of visceral obesity for the progression of atherosclerosis. Third, there was a gender difference in the effect of SNP45.

First, we demonstrated that the G allele had higher VFA, lower SFA, and a significantly higher V/S ratio compared to the TT genotype in men. Multivariate regression analysis showed that SNP45 was an independent determinant of the V/S ratio. These results indicated that the G allele of SNP45 is a risky genotype of visceral adiposity, resulting in metabolic syndrome. To our knowledge, this is the first study to demonstrate the association of SNP45 with body fat distribution. Some reports have shown that SNP45 contributes to obesity, insulin resistance, or dyslipidemia^{10, 18, 19}. In contrast, Ukkola *et al.* reported that SNP45 was found in equal frequency among obese and non-obese Swedish subjects²⁰. In French Caucasians, the 45G allele frequency was similar in morbidly obese adults and control subjects²¹. The inconsistency between these reports suggested that SNP45 could not be associated simply with weight or prevalence of obesity, but might contribute to body fat distribution in the process of becoming obese. Since visceral adipose tissue is widely believed to play a key role in the pathogenesis of metabolic abnormalities, the G allele of SNP45 could be an independent risk factor for metabolic syndrome.

Second, another important finding of the present study was the significant association between SNP45 and carotid artery PS in men. A similar trend was observed in women. Multivariate regression analysis showed that SNP45 was an independent determinant of PS. These findings suggest that SNP45 may affect the development of carotid atherosclerosis not only by modulating visceral obesity but also by other pathways.

To the best of our knowledge, PS tends to be associated with the V/S ratio only in the G allele in both men and women. In a previous study, we described a strong association between the V/S ratio and carotid artery PS in Japanese males with metabolic syndrome²², but patients with the TT genotype were protected from the atherogenic effect of visceral obesity. We hypothesized that visceral obesity might exaggerate the dysregulation of adiponectin properties of the G allele. The mechanism was unclear, but this hypothesis needs confirmation by expression studies.

Third, in this study the degree of the effect of SNP45 on body fat distribution and PS was different between men and women. Adipose tissue is sexually dimorphic in humans, with gender-specific differences in body fat distribution^{23, 24}. Gonadal steroids are the major mediator of sex dimorphism of body composition in adults^{25, 26}. Estrogen regulates both the metabolism and location of adipose tissue and plays a role in adipogenesis, adipose deposition, lipogenesis, lipolysis, and adipocyte proliferation²⁷. Furthermore, in recent studies, Clegg *et al.* reported that gonadal steroids mediate body fat distribution and interact with the integrated adiposity messages conveyed to the brain²⁸. Taken together with previous studies, our findings suggest that estrogen may interact with the adiponectin gene in adipocyte and modulate the effect of SNP45.

In addition, estrogen is known to have a cardio-protective effect. *In vivo* evidence suggests that the effect of estrogen on adhesion molecules is mediated by the inhibition of nuclear factor (NF)- κ B DNA binding^{29, 30}. As adiponectin has been shown to suppress the expression of class A scavenger receptors in macrophages, to affect the NF- κ B pathway and to inhibit monocyte adhesion to aortic endothelial cells⁶⁻⁸, atherogenic properties of the G allele may be suppressed by the effect of estrogen. Estrogen could interact with SNP45 and modulate the atherogenic function of adiponectin, but further large studies are needed to confirm the mechanism of gender-specific differences in the effect of SNP45.

The mechanistic relationship between SNP45 and both body fat distribution and the progression of atherosclerosis is unclear. SNP45 is located in exon 2 of the adiponectin gene and does not cause an amino acid change (GGT to GGG, Gly15Gly). One possibility is that SNP45 may have linkage disequilibrium with other undiscovered SNPs of the adiponectin gene having an effect on adiponectin expression, secretion, structure, or action. Another possibility is that SNP45 located in exon 2 is relatively close to the exon-intron boundary which may affect splicing machinery and effect adiponectin expression. The G allele of SNP45 may act through decreased adiponectin expression, which may cause increased visceral adipose tissue. Indeed, in Japanese type 2 diabetes, SNP45 is reported to be associated with reduced adiponectin levels¹¹. Similar findings have been shown in an other study³¹. Furthermore, recent studies have reported various adiponectin functions as an adipocyte differentiation factor, helping to maintain equilibrium adipocyte size, as an autocrine/paracrine factor in adipose tissue and as a participating factor in the regulation of adipocyte metabolism and adipose tissue mass. In 3T3-L1 preadipocytes,

adiponectin overexpression accelerates cell proliferation and differentiation, while in mature adipocytes, autocrine adiponectin increases glucose uptake and lipid accumulation³²⁾. Transgenic overexpression of adiponectin in the physiological range induced morbid obesity without insulin resistance in ob/ob mice²¹⁾. These reports indicated that hyperadiponectinemia may induce simple obesity with more subcutaneous fat accumulation, while decreased adiponectin levels may induce visceral obesity. Interestingly, the present study showed that hypo-adiponectinemia was the third independent determinant of the V/S ratio. Due to these previous findings combined with our present study, the G allele might be genetically determined to have hypo-adiponectinemia, contributing to the progression of visceral obesity. In contrast, the TT genotype might favor the accumulation of subcutaneous adipose tissue through hyperadiponectinemia, preventing insulin resistance, and eventually metabolic syndrome.

Adiponectin exists largely as low molecular weight (LMW) hexamers and high molecular weight (HMW) multimers^{32, 33)}. Recent article showed that the ratio of HMW to total adiponectin was responsible for metabolic effects³⁴⁾. Another study showed that HMW adiponectin was an important factor in metabolic syndrome³⁵⁾. Therefore, the alternative possibility of the atherogenic effect of SNP45 is that the proportion of HMW adiponectin might decrease in the G allele of SNP45, leading to atherosclerosis. As we measured total adiponectin and did not assess multimeric forms of adiponectin, further study is needed.

In conclusion, we demonstrated that SNP45 was associated with body fat distribution and PS of carotid arteries. The TT genotype is a protective genotype from metabolic syndrome and atherosclerosis progression in Japanese obese subjects. The mechanism by which SNP45 affects body fat distribution and the development of atherosclerosis has not been clarified at present. Further investigations will be needed to elucidate the functional mechanism of this polymorphism.

Acknowledgements

This study was partially supported by the Second Department of Internal Medicine, Kanazawa University Hospital. We thank Mrs. Reiko Ikeda for her excellent technical assistance.

References

- 1) Maeda K, Okubo K, Shimomura I, Funahashi T, Matsuzawa Y, and Matsubara K: cDNA cloning and expression of a novel adipose specific collagen-like factor, apM1 (AdiPose Most abundant Gene transcript 1). *Biochem Biophys Res Commun*, 1996; 221:286-289
- 2) Scherer PE, Williams S, Fogliano M, Baldini G, and Lodish HF: A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem*, 1995; 270:26746-26749
- 3) Combs TP, Berg AH, Obici S, Scherer PE, and Rossetti L: Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *J Clin Invest*, 2001; 108:1875-1881
- 4) Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, Uchida S, Yamashita S, Noda M, Kita S, Ueki K, Eto K, Akanuma Y, Froguel P, Foufelle F, Ferre P, Carling D, Kimura S, Nagai R, Kahn BB, and Kadowaki T: Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med*, 2002; 8:1288-1295
- 5) Maeda N, Shimomura I, Kishida K, Nishizawa H, Matsuda M, Nagaretani H, Furuyama N, Kondo H, Takahashi M, Arita Y, Komuro R, Ouchi N, Kihara S, Tochino Y, Okutomi K, Horie M, Takeda S, Aoyama T, Funahashi T, and Matsuzawa Y: Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat Med*, 2002; 8:731-737
- 6) Ouchi N, Kihara S, Arita Y, Nishida M, Matsuyama A, Okamoto Y, Ishigami M, Kuriyama H, Ishida K, Nishizawa H, Hotta K, Muraguchi M, Ohmoto Y, Yamashita S, Funahashi T, and Matsuzawa Y: Adipocyte-derived plasma protein, adiponectin, suppresses lipid accumulation and class A scavenger receptor expression in human monocyte-derived macrophages. *Circulation*, 2001; 103:1057-1063
- 7) Ouchi N, Kihara S, Arita Y, Okamoto Y, Maeda K, Kuriyama H, Hotta K, Nishida M, Takahashi M, Muraguchi M, Ohmoto Y, Nakamura T, Yamashita S, Funahashi T, and Matsuzawa Y: Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF- κ B signaling through a cAMP-dependent pathway. *Circulation*, 2000; 102:1296-1301
- 8) Ouchi N, Kihara S, Arita Y, Maeda K, Kuriyama H, Okamoto Y, Hotta K, Nishida M, Takahashi M, Nakamura T, Yamashita S, Funahashi T, and Matsuzawa Y: Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin. *Circulation*, 1999; 100:2473-2476
- 9) Fumeron F, Aubert R, Siddiq A, Betoulle D, Pean F, Hadjadj S, Tichet J, Wilpart E, Chesnier MC, Balkau B, Froguel P, and Marre M: Adiponectin gene polymorphisms and adiponectin levels are independently associated with the development of hyperglycemia during a 3-year period: the epidemiologic data on the insulin resistance syndrome prospective study. *Diabetes*, 2004; 53:1150-1157
- 10) Menzaghi C, Ercolino T, Di Paola R, Berg AH, Warram JH, Scherer PE, Trischitta V, and Doria A: A haplotype at the adiponectin locus is associated with obesity and other features of the insulin resistance syndrome. *Diabetes*, 2002; 51:2306-2312
- 11) Hara K, Bautin P, Mori Y, Tobe K, Dina C, Yasuda K, Yamauchi T, Otabe S, Okada T, Eto K, Kadowaki H, Hagura R, Akanuma Y, Yazaki Y, Nagai R, Taniyama M, Matsubara K, Yoda M, Nakano Y, Tomita M, Kimura S, Ito C, Froguel P, and Kadowaki T: Genetic variation in the gene

- encoding adiponectin is associated with an increased risk of type 2 diabetes in the Japanese population. *Diabetes*, 2002; 51:536-540
- 12) Lacquemant C, Froguel P, Lobbens S, Izzo P, Dina C, and Ruiz J: The adiponectin gene SNP + 45 is associated with coronary artery disease in Type 2 (non-insulin-dependent) diabetes mellitus. *Diabetic Medicine*, 2004; 21:776-781
 - 13) The Examination Committee of Criteria for 'Obesity Disease' in Japan, Japan Society for the Study of Obesity: New criteria for 'obesity disease' in Japan. *Circ J*, 2002; 66:987-992
 - 14) Expert Committee on the Diagnosis and Classification of Diabetes Mellitus: Report of the Expert Committee on the Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care*, 1997; 20:1183-1197
 - 15) Matthews DR, Hosker JB, Rudenski AS, and Naylor BA: Homeostasis model assesment: insulin resistance and β -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*, 1985; 28:412-419
 - 16) Yoshizumi T, Nakamura T, Yamane M, Islam AHMW, Menju M, and Yamasaki K: Abdominal fat: Standardized technique for measurement at CT. *Radiology*, 1999; 211:283-286
 - 17) Nagai Y, Kitagawa K, Yamagami H, Kondo K, Hougaku H, Hori M, and Matsumoto M: Carotid artery intima-media thickness and plaque score for the risk assessment of stroke subtypes. *Ultrasound Med Biol*, 2002; 28:1239-1243
 - 18) Stumvoll M, Tschritter O, Fritsche A, Staiger H, Renn W, Weisser M, Machicao F, and Haring H: Association of the T-G polymorphism in adiponectin (exon 2) with obesity and insulin sensitivity: interaction with family history of type 2 diabetes. *Diabetes*, 2002; 51:37-41
 - 19) Kondo H, Shimomura I, Matsukawa Y, Kumada M, Takahashi M, Matsuda M, Ouchi N, Kihara S, Kawamoto T, Sumitsuji S, Funahashi T, and Matsuzawa Y: Association of adiponectin mutation with type 2 diabetes: a candidate gene for the insulin resistance syndrome. *Diabetes*, 2002; 51:2325-2328
 - 20) Ukkola O, Ravussin E, Jacobson P, and Bouchard C: Mutations in the adiponectin gene in lean and obese subjects from the Swedish obese subjects cohort. *Metabolism*, 2003; 52:881-884
 - 21) Bouatia-Naji N, Meyre D, Lobbens S, Seron K, Fumeron F, Balkan B, Heude B, Jourer B, Scherer P, Dina C, Weill J, and Froguel P: ACDC/Adiponectin polymorphism are associated with severe childhood and adult obesity. *Diabetes*, 2006; 55:545-549
 - 22) Murase Y, Asano A, Kobayashi J, Yamaaki N, and Mabuchi H: Impact of adiposity on carotid atherosclerosis in Japanese males with metabolic syndrome. *J Intern Med*, 2005; 257:311-312
 - 23) Butte NF, Hopkinson JM, Wong WW, Smith EO, and Ellis KJ: Body composition during the first 2 years of life: an updated reference. *Pediatr Res*, 2000; 47:578-585
 - 24) Taylor RW, Jones IE, Williams SM, and Goulding A: Body fat percentages measured by dual-energy X-ray absorptiometry corresponding to recently recommended body mass index cutoffs for overweight and obesity in child and adolescents aged 3-18 y. *Am J Clin Nutr*, 2002; 76:1416-1421
 - 25) Marin P and Bjorntorp: Endocrine-metabolic patterns and adipose tissue distribution. *Horm Res*, 1993; 39:81-85
 - 26) Rosenbaum M and Leibel RL: Role of gonadal steroids in the sexual dimorphisms in body composition and circulating concentrations of leptin. *J Clin Endocrinol Metab*, 1999; 84:1784-1789
 - 27) Cooke PS and Naaz A: Role of estrogens in adipocyte development and function. *Exp Biol Med*, 2004; 229:1127-1135
 - 28) Clegg D, Brown L, Woods S, and Benoit S: Gonadal hormones determine sensitivity to central leptin and insulin. *Diabetes*, 2006; 55:978-987
 - 29) Simoncini T, Maffei S, and Basta G: Estrogens and glucocorticoids inhibit endothelial vascular cell adhesion molecule-1 expression by different transcriptional mechanisms. *Circ Res*, 2000; 87:19-25
 - 30) Hsu SM, Chen YC, and Jiang MC: 17β -estradiol inhibits tumor necrosis factor- α -induced nuclear factor- κ B activation by increasing nuclear factor- κ B p105 level in MCF-7 breast cancer cells. *Biochem Biophys Res Commun*, 2000; 279:47-52
 - 31) Takahashi M, Arita Y, Yamagata K, Matsukawa Y, Okutomi K, Horie M, Shimomura I, Hotta K, Kuriyama H, Kihara S, Nakamura T, Yamashita S, Funahashi T, and Matsuzawa Y: Genomic structure and mutations in adipose-specific gene, adiponectin. *Int J Obes Relat Metab Disord*, 2000; 24:861-868
 - 32) Fu Y, Luo N, Klein R, and Garvey W: Adiponectin promotes adipocyte differentiation, insulin sensitivity, and lipid accumulation. *Journal of Lipid Research*, 2005; 46:1369-1379
 - 33) Pajvani UB, Du X, Combs TP, Berg AH, Rajala MW, Schulthess T, Engel J, Brownlee M, and Scherer PE: Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin: implications for metabolic regulation and bioactivity. *J Biol Chem*, 2003; 278:9073-9085
 - 34) Pajvani UB, Hawkins M, Combs TP, Rajala MW, Doebber T, Berger JP, Wagner JA, Wu M, Knopps A, Xiang AH, Utzschneider KM, Kahn SE, Olefsky JM, Buchanan TA, and Scherer PE: Complex distribution, not absolute amount of adiponectin, correlate with thiazolidinedione-mediated improvement in insulin sensitivity. *J Biol Chem*, 2004; 279:12152-12162
 - 35) Lara-Castro C, Luo N, Wallace P, Klein R, and Garvey W: Adiponectin multimeric complexes and the metabolic syndrome trait cluster. *Diabetes*, 2006; 55:249-259

High Frequency of a Retinoid X Receptor γ Gene Variant in Familial Combined Hyperlipidemia That Associates With Atherogenic Dyslipidemia

Atsushi Nohara, Masa-aki Kawashiri, Thierry Claudel, Mihoko Mizuno, Masayuki Tsuchida, Mutsuko Takata, Shoji Katsuda, Kenji Miwa, Akihiro Inazu, Folkert Kuipers, Junji Kobayashi, Junji Koizumi, Masakazu Yamagishi, Hiroshi Mabuchi

Objective—The genetic background of familial combined hyperlipidemia (FCHL) has not been fully clarified. Because several nuclear receptors play pivotal roles in lipid metabolism, we tested the hypothesis that genetic variants of nuclear receptors contribute to FCHL.

Methods and Results—We screened all the coding regions of the PPAR α , PPAR γ 2, PPAR δ , FXR, LXR α , and RXR γ genes in 180 hyperlipidemic patients including 60 FCHL probands. Clinical characteristics of the identified variants were evaluated in other 175 patients suspected of coronary disease. We identified PPAR α Asp140Asn and Gly395Glu, PPAR γ 2 Pro12Ala, RXR γ Gly14Ser, and FXR $-1g \rightarrow t$ variants. Only RXR γ Ser14 was more frequent in FCHL (15%, $P < 0.05$) than in other primary hyperlipidemia (4%) and in controls (5%). Among patients suspected of coronary disease, we identified 9 RXR γ Ser14 carriers, who showed increased triglycerides (1.62 ± 0.82 versus 1.91 ± 0.42 [mean \pm SD] mmol/L, $P < 0.05$), decreased HDL-cholesterol (1.32 ± 0.41 versus 1.04 ± 0.26 , $P < 0.05$), and decreased post-heparin plasma lipoprotein lipase protein levels (222 ± 85 versus 149 ± 38 ng/mL, $P < 0.01$). In vitro, RXR γ Ser14 showed significantly stronger repression of the lipoprotein lipase promoter than RXR γ Gly14.

Conclusion—These findings suggest that RXR γ contributes to the genetic background of FCHL. (*Arterioscler Thromb Vasc Biol.* 2007;27:923-928.)

Key Words: apolipoproteins ■ gene mutations ■ lipoprotein lipase
■ familial combined hyperlipidemia ■ nuclear receptors

Familial combined hyperlipidemia (FCHL) is the most common form of inherited hyperlipidemia. FCHL shows strong genetic susceptibility resembling an autosomal dominant disease,¹⁻³ but most of the underlying causal mechanisms remain to be elucidated. Lipoprotein lipase (LPL) has been implicated as one of the genes that modify the lipid phenotype in FCHL.^{4,5} “Intra-individual variability” of the lipoprotein phenotype is often included as a criterion in diagnosis.⁶ However, a recent prospective study of FCHL families suggests that this variability may even include normolipidemic periods in affected subjects.⁷ This feature indicates that FCHL could be a “disease of regulation” rather than a genetic defect in certain peripheral components of lipid metabolism.

Nuclear receptors are transcription factors that can be activated by specific ligands. Recent studies have shown that nuclear receptors, especially retinoid X receptor (RXR) and its heterodimerization partners,⁸ play important roles in main-

tenance of lipid homeostasis on their activation by a variety of ligands derived from dietary cholesterol and fatty acids.⁹ The peroxisome proliferator-activated receptors (PPARs) family, the oxysterol sensor liver X receptor (LXR), and the bile acid sensor farnesoid X receptor (FXR) are all involved in control of plasma lipid concentrations.¹⁰ Thus, we tested the hypothesis that variants of these nuclear receptors, ie, PPAR α , PPAR γ 2, PPAR δ , LXR α , FXR, and RXR γ , could constitute part of the genetic background of atherogenic dyslipidemia, particularly of FCHL.

Methods

Subjects

The study design consists of 2 parts. First, we screened for frequent variants in the nuclear receptor candidate genes among 180 patients with primary hyperlipidemia, including 60 unrelated patients with FCHL (clinical characteristics are presented in supplemental Table I, available online at <http://atvb.ahajournals.org>). Patients with familial

Original received August 28 2006; final version accepted January 6, 2007.

From the Departments of Lipidology (A.N., M.M., J. Kobayashi, H.M.) and Cardiovascular Medicine (M.K., M. Tsuchida, M. Takata, S.K., K.M., M.Y.), Graduate School of Medical Science, Kanazawa University, Japan; Center for Liver, Digestive, and Metabolic Diseases, Laboratory of Pediatrics (T.C., F.K.), University Medical Center Groningen, Groningen, the Netherlands; School of Health Sciences, Faculty of Medicine (A.I.), and Department of General Medicine (J. Koizumi), Kanazawa University Hospital, Japan.

Correspondence to Atsushi Nohara, Department of Lipidology, Graduate School of Medical Science, Kanazawa University, Takara-machi 13-1, Kanazawa 920-8641, Japan. E-mail a-nohara@med.kanazawa-u.ac.jp

© 2007 American Heart Association, Inc.

Arterioscler Thromb Vasc Biol. is available at <http://www.atvbaha.org>

DOI: 10.1161/01.ATV.0000258945.76141.8a

hypercholesterolemia and secondary hyperlipidemia were excluded. Diagnosis of FCHL was based on the fulfillment of all of the following three criteria: (1) Phenotype IIb, IIa, or IV hyperlipidemia according to the Fredrickson classification; (2) Presence of phenotype IIb, IIa, or IV hyperlipidemia in a first-degree relative and at least one family member with phenotype IIb; (3) Exclusion of familial hypercholesterolemia. Two hundred ninety-eight anonymous samples from healthy males were used as controls for frequency analysis of identified mutations. All blood samples in this study were obtained after an overnight fast.

Second, we evaluated the clinical impact of potentially relevant variants in another 175 patients who were suspected of having coronary artery disease based on any of the following reasons: ECG abnormalities; cumulative coronary risk factors, and/or chest symptoms. The group included 105 patients who had undergone coronary angiography. Patients with familial hypercholesterolemia were excluded because of their clear genetic background for hyperlipidemia. The extent and severity of atherosclerotic changes in coronary angiography were assessed by assigning scores to each of the 15 segments, according to the classification of the American Heart Association Grading Committee. The coronary stenosis index (CSI) was defined as the sum of the following scores¹¹: A normal coronary angiogram was graded 0, stenosis of less than 25% was graded 1, 25% to 50% stenosis was graded 2, 50% to 75% stenosis was graded 3, and more than 75% stenosis was graded 4. CSI is a useful index for evaluating mild-moderate coronary atherosclerotic changes.

All the subjects and controls enrolled were inhabitants of the Hokuriku district of Japan. Written informed consent was obtained from each of the subjects. The study protocol was approved by the ethics committee of the Graduate School of Medical Science, Kanazawa University.

Laboratory Analyses

Total cholesterol (TC), triglycerides (TG), high-density lipoprotein (HDL)-cholesterol, apolipoproteins, glucose, and thyroid hormones were measured according to standard clinical laboratory techniques. HDL-cholesterol fractions were obtained by dextran sulfate-magnesium chloride precipitation and assayed using a commercial kit (Daiichi, Tokyo, Japan).¹² Separation of lipoproteins by ultracentrifugation was performed as described by Havel et al.¹³ Plasma remnant-like particle (RLP)-cholesterol was determined by immunoabsorption using the commercial RLP-C JIMRO kit.¹⁴ Plasma cholesteryl ester transfer protein (CETP) concentrations were determined by enzyme-linked immunosorbent assay using the monoclonal antibody TP2 and a rabbit polyclonal antibody raised against recombinant human CETP.¹⁵ For LPL assessment, blood samples were obtained 10 minutes after an intravenous injection of 30 IU heparin/kg body weight. LPL activity was measured using radiolabeled triolein emulsion after hepatic lipase (HL) inhibition by SDS as previously described.¹⁶ LPL mass was measured by sandwich enzyme-linked immunosorbent assay (ELISA) using specific monoclonal antibody against LPL (Daiichi Pure Chemicals Co Ltd, Tokyo, Japan).¹⁷

Genetic Analyses of Candidate Genes

Genomic DNA was isolated from peripheral white blood cells using standard phenol-chloroform extraction techniques. We screened all the coding regions of PPAR α (NM_032644), PPAR δ (NM_006238), PPAR γ 2 (NM_015869), LXR α (NM_005693), FXR (NM_005123), and RXR γ (NM_006917) genes with flanking exon-intron boundaries by polymerase chain reaction (PCR)-denaturing gradient gel electrophoresis (DGGE) using the DCode system (Bio-Rad), which is highly accurate in detecting changes in nucleic acids.¹⁸ The structural organization and nucleotide sequences of these genes were retrieved from the gene database of NCBI. Lists of all GC-clamped primers used in DGGE analysis are available online (supplemental Table II). Samples with a variant detected by DGGE analysis were directly sequenced on an ABI310 analyzer (Applied Biosystems). PCR-restriction-fragment-length polymorphisms analysis on the RXR γ Ser14 variant was performed with the primers 5'-AGCCGAGAGAGGCGGTAATA-3' (forward) and 5'-

TACAGGTCCACGCACTGAAG-3' (reverse) in patients suspected of coronary artery disease. Digestion with *AluI* resulted in a 76-bp fragment for Ser allele and a 120-bp fragment for Gly allele.

Cell Culture and Transfection Assays

Cos7 cells were grown in DMEM supplemented with 10% FCS, penicillin/streptomycin, sodium pyruvate, glutamine, and nonessential amino acids (Gibco BRL, Invitrogen). The medium was changed every 48 hours. Cos7 cells were transfected using FuGENE 6 reagent (Roche): 150 ng of the indicated LPL firefly luciferase reporter plasmid (a generous gift of Dr B. Staels, Institut Pasteur de Lille, France), that contains the proximal 466-bp of the human LPL promoter in front of the ATG cloned into the *HindIII* site of the pGL3 plasmid, was cotransfected with or without 100 ng of the human RXR γ expressing vector (a generous gift of Dr W. Lamph, Ligand Pharmaceuticals Inc, San Diego, Calif). After an overnight incubation, cells were incubated with medium containing 10% FCS with or without the retinoid LGD1069, (1 μ mol/L, Sigma) and luciferase activity was assayed 48 hours later using an Orion luminometer (Berthold). Transfection studies were performed at least 3 times in triplicate. Transfection efficiency was monitored by cotransfection of 150 ng of a SV40-driven β -galactosidase expression plasmid. A positive RXRE TKpGL3 construct was made by cloning 3 copies of the direct repeat AGGTCA spaced by 5 nucleotides in the TKpGL3 plasmid.

Plasmid Site-Directed Mutagenesis

Nucleotide substitution was introduced in the plasmid expressing human RXR γ using the QuikChange Site-Directed Mutagenesis Kit (Stratagene, The Netherlands) and the primer 5'-CATGAAGTTTCCCGCAAGCTATGGAGGCTCCCTGG C-3' in which the nucleotide in bold indicates the mutation.

Statistical Analysis

The frequency distribution of genotypes was compared using standard χ^2 tests. Student *t* test was used for normally distributed parameters and the Kruskal-Wallis test was used for non-normally distributed parameters: triglycerides levels, LPL levels, and CSI. JMP 5.1.2 software (SAS Institute Inc) was used for statistical calculation.

Results

Identified Variants in Nuclear Receptor Genes

With PCR-DGGE analysis, we identified 4 variants with amino acid changes, ie, Asp140Asn and Gly395Glu in the PPAR α gene, Pro12Ala in the PPAR γ 2 gene, Gly14Ser in the

TABLE 1. Frequencies of Nuclear Receptor Genes Variants Identified in This Study

	FCHL n=60	Other Hyperlipidemia n=120	General Population n=298	P Value
PPAR α Gly395Glu				
Glu395	3 (5%)	1 (0.8%)	6 (2%)	ns
PPAR α Asp140Asn				
Asn140	2 (3%)	1 (0.8%)	2 (0.6%)	ns
PPAR γ 2Pro12Ala				
Ala12	5 (8%)	10 (8%)	20 (7%)	ns
FXR -1g->t				
-1g/t	19 (32%)	34 (28%)	108 (36%)	ns
-1t/t	2 (3%)	6 (5%)	27 (9%)	ns
RXR γ Gly14Ser				
Ser14	9 (15%)	5 (4%)	15 (5%)	0.03

TABLE 2. Clinical Characteristics of Patients With RXR γ Variant

	RXR γ Gly14Ser		
	Gly/Gly	Gly/Ser	P Value
Number (M/F)	166 (78/88)	9 (5/4)	
Age, y	58 \pm 15	58 \pm 7	ns
BMI, kg/m ²	23.4 \pm 5	23.9 \pm 2	ns
Smoking, %	36	33	ns
Total cholesterol, mmol/L	5.98 \pm 1.4	5.96 \pm 1.55	ns
Triglycerides, mmol/L	1.62 \pm 0.82	1.91 \pm 0.42	<i>P</i> <0.05
HDL cholesterol, mmol/L	1.32 \pm 0.41	1.04 \pm 0.26	<i>P</i> <0.05
LDL cholesterol, mmol/L	3.94 \pm 1.27	4.07 \pm 1.45	ns
HDL2 cholesterol, mmol/L	0.78 \pm 0.28	0.54 \pm 0.10	<i>P</i> <0.05
HDL3 cholesterol, mmol/L	0.44 \pm 0.10	0.39 \pm 0.08	ns
ApoA-I, g/L	1.38 \pm 0.31	1.18 \pm 0.18	ns
ApoA-II, g/L	0.32 \pm 0.06	0.28 \pm 0.05	<i>P</i> <0.05
ApoB, g/L	1.31 \pm 0.38	1.35 \pm 0.31	ns
ApoC-II, g/L	0.06 \pm 0.02	0.05 \pm 0.02	ns
ApoC-III, g/L	0.11 \pm 0.05	0.10 \pm 0.03	ns
ApoE, g/L	0.06 \pm 0.02	0.05 \pm 0.01	ns
RLP cholesterol, mmol/L	0.15 \pm 0.10	0.21 \pm 0.10	<i>P</i> <0.01
CETP, mg/L	2.52 \pm 0.82	2.48 \pm 0.73	ns
Intraindividual lipoprotein phenotype variability, %	27	88	<i>P</i> <0.01
Fasting glucose, mmol/L	5.72 \pm 1.39	5.33 \pm 0.72	ns
HbA1c, %	5.6 \pm 1.0	5.8 \pm 1.0	ns
Fasting insulin, pmol/L	70.8 \pm 90.3	52.1 \pm 1.0	ns
HOMA-IR	2.28 \pm 2.1	2.19 \pm 1.7	ns
Diabetes, %	28	33	ns
HL activity, U/L	0.24 \pm 0.09	0.26 \pm 0.07	ns
LPL activity, U/L	0.11 \pm 0.06	0.08 \pm 0.03	<i>P</i> <0.05
LPL mass, ng/mL	222 \pm 85	149 \pm 38	<i>P</i> <0.01
FT3, pmol/L	0.42 \pm 0.01	0.044 \pm 0.01	ns
FT4, pmol/L	15.2 \pm 5.15	13.3 \pm 2.57	ns
TSH, μ U/mL	2.31 \pm 2.8	2.53 \pm 0.9	ns
Number (M/F)	100 (50/50)	5 (4/1)	
CSI	12.3 \pm 10	21.4 \pm 6	<i>P</i> <0.05

mean \pm SD

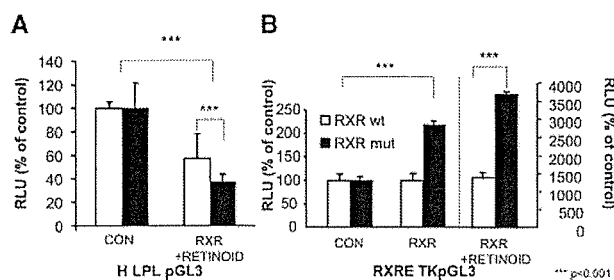
BMI indicates body mass index; HOMA-IR, homeostasis model assessment; FT3, free triiodothyronine; FT4, free thyroxine; TSH, thyroid stimulating hormone.

RXR γ gene, and 1 nucleotide substitution in a flanking coding region, ie, FXR $-1g \rightarrow t$ variant. The PPAR γ 2 Pro12Ala polymorphism has already been well-described,¹⁹ whereas the others represent novel variants identified in this study. In humans, variants in the RXR γ gene have been associated with elevated triglyceride levels in familial type 2 diabetes, but none of these variants showed an altered coding sequence.²⁰ Therefore, this is the first description of a RXR γ variant with an amino acid substitution. In the PPAR α gene, the Leu162Val variant has been reported in Western countries,²¹ but this variant was not identified in this study. We also identified some silent nucleotide substitutions, ie, 891C \rightarrow G (rs13306747) and 1431C \rightarrow T (rs1724155) in the PPAR γ 2 gene, 1233C \rightarrow T (rs9658166) in the PPAR δ gene,

and 1134A \rightarrow G (rs1131379) in the LXR α gene. We did not identify variants with amino acid changes in the PPAR δ and LXR α genes. We further investigated the variants with amino acid substitutions and the $-1g \rightarrow t$ FXR variant, because of the likelihood that these induced altered physiological function.

Higher Frequency of RXR γ Variant in FCHL

We evaluated the frequencies of the 5 identified polymorphisms in subjects with FCHL, subjects with other forms of primary hyperlipidemia and in the general population (Table 1). Only the RXR γ Ser14 variant was found to be significantly more frequent in FCHL patients (15%) compared with that in other forms of primary hyperlipidemia (4%) or the general population (5%).



A, Cos7 cells were cotransfected with RXR γ wild-type or the Ser14 variant and activated with retinoid in presence of the LPL promoter. B, Cos7 cells were cotransfected with RXR γ wild-type or the Ser14 variant and activated with retinoid in presence of a positive RXRE cloned in the TKpGL3 plasmid. *** $p < 0.001$.

Atherogenic Plasma Lipids Profiles and Coronary Atherosclerosis Associated With the RXR γ Ser14 Variant

To establish the impact of the identified RXR γ variant on metabolic parameters and on coronary atherosclerosis, we evaluated anthropometric parameters and laboratory data from 175 patients suspected of coronary disease. The RXR γ Ser14 variant was identified in 9 patients, all of whom were heterozygotes. Eight of the RXR γ Ser14 carriers had hyperlipidemia, while the remaining 1 demonstrated an isolated low HDL cholesterol level. Clinical characteristics of patients with or without the RXR γ Ser14 allele are shown in Table 2. There was no difference in age or body mass index between the two groups. In their lipid profiles, RXR γ Ser14 carriers had higher TG, lower HDL cholesterol especially in the HDL2 subfraction, and lower apolipoprotein A-II levels. There was no difference in CETP protein levels between the groups. Furthermore, we found that the RLP cholesterol level was significantly higher in the RXR γ Ser14 carriers than in the wild-type. Subjects with this variant also showed significantly lower LPL activities and protein levels in post-heparin plasma. Separation of lipoproteins demonstrated that the Ser14 carriers had higher TG levels in very low-density lipoprotein (VLDL) and low-density lipoprotein (LDL) fractions, higher cholesterol levels in VLDL, and lower cholesterol levels in HDL (supplemental Table III).

Two RXR γ Ser14 carriers were diagnosed as FCHL (22%), and 2 additional carriers were suspected of FCHL with hyperlipidemic siblings without information on first-degree relatives. Among non-carriers, 22 of 166 patients were diagnosed as FCHL (13%). One hundred twenty-five patients suspected of coronary disease showed hyperlipidemia and the intraindividual variability of lipoprotein phenotype was significantly more frequent in RXR γ Ser14 carriers (7 of 8 hyperlipidemic patients; 88%) than in wild-type (32 of 117 hyperlipidemic patients; 27%, Table 2).

There was no significant difference in the thyroid hormone levels between the two groups.

Four males and 1 female were identified as RXR γ variant carriers among 105 patients who underwent coronary angiography. The carriers of RXR γ Ser14 demonstrated significantly higher CSI than those with the wild-type (Table 2).

RXR γ Variant Represses More Efficiently the LPL Promoter Activity

Because RXR γ Ser14 carriers showed significantly lower LPL activities and protein levels in post-heparin plasma, we hypothesized that activated-RXR γ downregulates LPL gene expression by a transcriptional mechanism and that RXR γ variant is more effective in repressing the LPL promoter activity. Therefore, transfection assays were performed using the LPL promoter cotransfected with either wild-type RXR γ or the variant (Figure). Interestingly, RXR γ Gly14 significantly repressed ($\sim 40\%$) the LPL promoter activity, whereas the RXR γ Ser14 repressed even more strongly ($\sim 60\%$, $P < 0.001$, Figure A). Moreover, the RXR γ Ser14 was a more potent activator of a positive RXRE cloned in front of a TKpGL3 plasmid (note the different scales in Figure B). Taken together, our results indicate that RXR γ downregulates human LPL gene expression, at least partially by a transcriptional mechanism, and that the newly identified RXR γ variant is a more potent repressor than the wild-type in this respect, as well as a more potent transactivator of a positive RXR response element.

Gain of Function Variant of PPAR α and Increased LDL-C Levels

The carriers of the PPAR α variant Gly395Glu tended to have higher frequency in the FCHL population, although not statistically significant. Four subjects were identified as PPAR α Glu395 carriers in the coronary artery disease-suspected group and showed significantly higher LDL-cholesterol levels (supplemental Table IV). On in vitro functional analysis, Glu395 showed a moderately but significantly increased transcriptional activity compared with wild-type PPAR α (supplemental Figure I, available online at <http://atvb.ahajournals.org>). The previously described Leu162Val variant of the PPAR α gene has been shown to give gain of function in in vitro,²⁴ has been associated with raised LDL-cholesterol levels.^{21,22} Our results appear to be in accordance with these previous reports.

Discussion

The main findings of the present study are the following: (1) identification of novel polymorphisms in plasma lipid levels-associated nuclear receptor genes, (2) a higher frequency of the RXR γ gene variant Gly14Ser in subjects with FCHL, (3) RXR γ Ser14 variant carriers showed more atherogenic dyslipidemia associated with coronary atherosclerosis, (4) the RXR γ variant showed a stronger response to its ligand in repression of the LPL promoter than the wild-type RXR γ .

RXRs are major heterodimerization partners of nuclear receptors such as PPARs, LXRs, and FXR. Three RXR isotypes have been identified: RXR α , RXR β , and RXR γ . Synthetic RXR ligands induce hypertriglyceridemia through decreased clearance of VLDL by LPL-dependent pathways,^{23,24} except in 1 study.²⁵ In contrast to the embryonic lethality observed in RXR α - and RXR β -deficient mice, RXR γ -deficient mice develop apparently normal.²⁶ Yet, RXR γ -deficient mice showed reduced fasting plasma TG levels and increased skeletal muscle LPL activity when fed a high fat diet.²⁷ The human RXR γ gene is located on chro-

mosome 1q21-q23, ie, the so-called "FCHL locus",²⁸ and both linkage analysis and a twin study have indicated that the RXR γ gene is linked with dyslipidemia in Chinese and German families.^{29,30}

To our knowledge, there are only few data concerning the physiological roles and targets of RXR γ in humans. The RXR γ gene is mainly expressed in skeletal muscles, central nervous system, skin, intestine, and lung. In the present study, LPL protein mass and activity were significantly decreased in RXR γ variant carriers. Because LPL is mainly expressed in adipose tissues and in skeletal muscles, we assume that this is attributable to the fact that the presence of the RXR γ variant affects LPL expression in skeletal muscles. RXR γ mRNA is detectable in adipose tissue only at a low level,³¹ but it has been reported that RXR γ could replace RXR α in heterodimerization with PPAR γ in adipose tissue.³² Therefore, there is a possibility that RXR γ variant expression in adipose tissue contributes to the changes in LPL.

It has been reported that RXR γ -deficient mice show a 17% increase in serum thyroid hormone (T4) and a 20% increase in thyroid-stimulating hormone (TSH) levels.³³ In the present study, thyroid hormone levels did not appear to differ sufficiently between variant carriers and non-carriers to explain the differences observed in lipid levels.

It has been shown that low LPL levels contribute to disorders associated with TG-rich lipoprotein catabolism with low HDL, especially in HDL2,^{34,35} and are associated with increased risk for future coronary disease.³⁶ Thus, the low LPL could well contribute to the increase in TG and the decrease in HDL-cholesterol levels in subjects with the RXR γ variant.

We assessed the functional consequence of the RXR γ Ser14 variant in vitro. The activation function-1 (AF-1) domain of RXR γ is located between amino acids 1 and 103, and is required for optimal ligand-dependent transactivation of RXR response element.³⁷ Fourteen amino acids are located within the AF-1 domain and are conserved among humans, mice, and chickens. In a transfection assay, RXR γ Ser14 repressed LPL promoter activity more strongly than the wild-type RXR γ . In addition, the Ser14 variant was a more potent inducer of a positive RXR response element. Therefore, we speculate that the Ser14 variant induces a better recruitment and/or stabilization of RXR cofactors. Further studies will be required to understand the precise molecular mechanism(s) involved in the LPL regulation by RXR γ Ser14.

Within the so-called FCHL locus, on chromosome 1q21-q23, several genes have been reported to be associated with the FCHL phenotype^{28,30,38} and with type 2 diabetes.³⁹ First, the thioredoxin interacting protein gene was shown to be associated with combined hyperlipidemia in mice, but no disease-causing mutation has been found in humans so far.^{40,41} Currently upstream stimulatory factor 1 (USF1) is considered the most promising candidate gene of FCHL.⁴² In the USF1 gene, no amino acid substitution has been identified in the coding regions, but single nucleotide polymorphisms in the 3'untranslated region and in intron 7 have been reported to be associated with FCHL, metabolic syndrome, or type 2 diabetes mellitus quite reproducibly.⁴³⁻⁴⁵ However, popula-

tions did not show any such association have also been reported.⁴⁶⁻⁴⁸ These reports emphasize the complexity of phenotypic expression in multi-factorial diseases such as FCHL. RXR γ had been reported to show an association with TG and cholesterol levels on linkage analysis,^{29,30} and we identified novel RXR γ variant that associated with atherogenic dyslipidemia. However, the changes in lipid levels attributable to the RXR γ variant alone were not sufficient to cause FCHL. Thus, we suggest the RXR γ gene variant to be a strong modifier rather than a causative gene in development of the FCHL phenotype.

In conclusion, the present study suggests that a variant of RXR γ gene contributes to genetic dyslipidemia, including FCHL, based on the increased frequency of this variant in FCHL, its association with an atherogenic lipid profile, and initial functional studies.

Acknowledgments

The authors thank Sachio Yamamoto for technical assistance. Drs William W. Lamph and Bart Staels are kindly acknowledged for the generous gift of plasmids.

Sources of Funding

This work has been supported by a scientific research grant from the Ministry of Education, Science, and Culture of Japan (No.17790603) and ONO Medical Research Foundation. Thierry Claudel was supported by Grant 2002B017 from the Netherlands Heart Foundation.

Disclosures

None.

References

- Goldstein JL, Schrott HG, Hazzard WR, Bierman EL, Motulsky AG. Hyperlipidemia in coronary heart disease. II. Genetic analysis of lipid levels in 176 families and delineation of a new inherited disorder, combined hyperlipidemia. *J Clin Invest.* 1973;52:1544-1568.
- Rose HG, Kranz P, Weinstock M, Juliano J, Haft JI. Inheritance of combined hyperlipoproteinemia: evidence for a new lipoprotein phenotype. *Am J Med.* 1973;54:148-160.
- Nikkila EA, Aro A. Family study of serum lipids and lipoproteins in coronary heart-disease. *Lancet.* 1973;1:954-959.
- Hoffer MJ, Bredie SJ, Snieder H, Reymer PW, Demacker PN, Havekes LM, Boomsma DI, Stalenhoef AF, Frants RR, Kastelein JJ. Gender-related association between the -93T->G/D9N haplotype of the lipoprotein lipase gene and elevated lipid levels in familial combined hyperlipidemia. *Atherosclerosis.* 1998;138:91-99.
- Hoffer MJ, Bredie SJ, Boomsma DI, Reymer PW, Kastelein JJ, de Knijff P, Demacker PN, Stalenhoef AF, Havekes LM, Frants RR. The lipoprotein lipase (Asn291->Ser) mutation is associated with elevated lipid levels in families with familial combined hyperlipidaemia. *Atherosclerosis.* 1996;119:159-167.
- Gaddi A, Galetti C, Paucillo P, Arca M. Familial combined hyperlipoproteinemia: experts panel position on diagnostic criteria for clinical practice. Committee of experts of the Atherosclerosis and Dysmetabolic Disorders Study Group. *Nutr Metab Cardiovasc Dis.* 1999;9:304-311.
- Veerkamp MJ, de Graaf J, Bredie SJ, Hendriks JC, Demacker PN, Stalenhoef AF. Diagnosis of familial combined hyperlipidemia based on lipid phenotype expression in 32 families: results of a 5-year follow-up study. *Arterioscler Thromb Vasc Biol.* 2002;22:274-282.
- Mangelsdorf DJ, Evans RM. The RXR heterodimers and orphan receptors. *Cell.* 1995;83:841-850.
- Shulman AI, Mangelsdorf DJ. Retinoid x receptor heterodimers in the metabolic syndrome. *N Engl J Med.* 2005;353:604-615.
- Claudel T, Staels B, Kuipers F. The Farnesoid X receptor: a molecular link between bile acid and lipid and glucose metabolism. *Arterioscler Thromb Vasc Biol.* 2005;25:2020-2030.

11. Mabuchi H, Koizumi J, Shimizu M, Takeda R. Development of coronary heart disease in familial hypercholesterolemia. *Circulation*. 1989;79:225-232.
12. Talameh Y, Wei R, Naito H. Measurement of total HDL, HDL2, and HDL3 by dextran sulfate-MgCl₂ precipitation technique in human serum. *Clin Chim Acta*. 1986;158:33-41.
13. Havel RJ, Eder HA, Bragdon JH. The distribution and chemical composition of ultracentrifugally separated lipoproteins in human serum. *J Clin Invest*. 1955;34:1345-1353.
14. Nakajima K, Saito T, Tamura A, Suzuki M, Nakano T, Adachi M, Tanaka A, Tada N, Nakamura H, Campos E, et al. Cholesterol in remnant-like lipoproteins in human serum using monoclonal anti apo B-100 and anti apo A-I immunoaffinity mixed gels. *Clin Chim Acta*. 1993;223:53-71.
15. Kiyohara T, Kiriyaama R, Zamma S, Inazu A, Koizumi J, Mabuchi H, Chichibu K. Enzyme immunoassay for cholesteryl ester transfer protein in human serum. *Clin Chim Acta*. 1998;271:109-118.
16. Baginsky ML, Brown WV. A new method for the measurement of lipoprotein lipase in postheparin plasma using sodium dodecyl sulfate for the inactivation of hepatic triglyceride lipase. *J Lipid Res*. 1979;20:548-556.
17. Kobayashi J, Hashimoto H, Fukamachi I, Tashiro J, Shirai K, Saito Y, Yoshida S. Lipoprotein lipase mass and activity in severe hypertriglyceridemia. *Clin Chim Acta*. 1993;216:113-123.
18. Grompe M. The rapid detection of unknown mutations in nucleic acids. *Nat Genet*. 1993;5:111-117.
19. Stumvoll M, Haring H. The peroxisome proliferator-activated receptor-gamma2 Pro12Ala polymorphism. *Diabetes*. 2002;51:2341-2347.
20. Wang H, Chu W, Hemphill C, Hasstedt SJ, Elbein SC. Mutation screening and association of human retinoid X receptor gamma variation with lipid levels in familial type 2 diabetes. *Mol Genet Metab*. 2002;76:14-22.
21. Flavell DM, Pineda Torra I, Jamshidi Y, Evans D, Diamond JR, Elkeles RS, Bujac SR, Miller G, Talmud PJ, Staels B, Humphries SE. Variation in the PPARalpha gene is associated with altered function in vitro and plasma lipid concentrations in Type II diabetic subjects. *Diabetologia*. 2000;43:673-680.
22. Tai ES, Demissie S, Cupples LA, Corella D, Wilson PW, Schaefer EJ, Ordovas JM. Association between the PPARA L162V polymorphism and plasma lipid levels: the Framingham Offspring Study. *Arterioscler Thromb Vasc Biol*. 2002;22:805-810.
23. Vahlquist C, Lithell H, Michaelsson G, Selinus I, Vahlquist A, Vessby B. Plasma fat elimination tissue lipoprotein lipase activity and plasma fatty acid composition during sequential treatment with etretinate and isotretinoin. *Acta Derm Venereol*. 1987;67:139-144.
24. Davies PJ, Berry SA, Shipley GL, Eckel RH, Hennuyer N, Crombie DL, Ogilvie KM, Peinado-Onsurbe J, Fievret C, Leibowitz MD, Heyman RA, Auwerx J. Metabolic effects of retinoids: tissue-specific regulation of lipoprotein lipase activity. *Mol Pharmacol*. 2001;59:170-176.
25. Mukherjee R, Davies PJ, Crombie DL, Bischoff ED, Cesario RM, Jow L, Hamann LG, Boehm MF, Mondon CE, Nadzan AM, Paterniti JR Jr, Heyman RA. Sensitization of diabetic and obese mice to insulin by retinoid X receptor agonists. *Nature*. 1997;386:407-410.
26. Krezel W, Dupe V, Mark M, Dierich A, Kastner P, Chambon P. RXR gamma null mice are apparently normal and compound RXR alpha +/-RXR beta +/-RXR gamma -/- mutant mice are viable. *Proc Natl Acad Sci U S A*. 1996;93:9010-9014.
27. Haugen BR, Jensen DR, Sharma V, Pulawa LK, Hays WR, Krezel W, Chambon P, Eckel RH. Retinoid X receptor gamma-deficient mice have increased skeletal muscle lipoprotein lipase activity and less weight gain when fed a high-fat diet. *Endocrinology*. 2004;145:3679-3685.
28. Pajukanta P, Nuotio I, Terwilliger JD, Porkka KV, Ylitalo K, Pihlajamaki J, Suomalainen AJ, Syvanen AC, Lehtimaki T, Viikari JS, Laakso M, Taskinen MR, Ehnholm C, Peltonen L. Linkage of familial combined hyperlipidaemia to chromosome 1q21-q23. *Nat Genet*. 1998;18:369-373.
29. Knoblauch H, Busjahn A, Muller-Myhsok B, Faulhaber HD, Schuster H, Uhlmann R, Luft FC. Peroxisome proliferator-activated receptor gamma gene locus is related to body mass index and lipid values in healthy nonobese subjects. *Arterioscler Thromb Vasc Biol*. 1999;19:2940-2944.
30. Pei W, Baron H, Muller-Myhsok B, Knoblauch H, Al-Yahyaee SA, Hui R, Wu X, Liu L, Busjahn A, Luft FC, Schuster H. Support for linkage of familial combined hyperlipidemia to chromosome 1q21-q23 in Chinese and German families. *Clin Genet*. 2000;57:29-34.
31. Kamei Y, Kawada T, Kazuki R, Sugimoto E. Retinoic acid receptor gamma 2 gene expression is up-regulated by retinoic acid in 3T3-L1 preadipocytes. *Biochem J*. 1993;293(Pt 3):807-812.
32. Metzger D, Imai T, Jiang M, Takukawa R, Desvergne B, Wahli W, Chambon P. Functional role of RXRs and PPARgamma in mature adipocytes. *Prostaglandins Leukot Essent Fatty Acids*. 2005;73:51-58.
33. Brown NS, Smart A, Sharma V, Brinkmeier ML, Greenlee L, Camper SA, Jensen DR, Eckel RH, Krezel W, Chambon P, Haugen BR. Thyroid hormone resistance and increased metabolic rate in the RXR-gamma-deficient mouse. *J Clin Invest*. 2000;106:73-79.
34. Patsch JR, Prasad S, Gotto AM Jr, Patsch W. High density lipoprotein2. Relationship of the plasma levels of this lipoprotein species to its composition, to the magnitude of postprandial lipemia, and to the activities of lipoprotein lipase and hepatic lipase. *J Clin Invest*. 1987;80:341-347.
35. Blades B, Vega GL, Grundy SM. Activities of lipoprotein lipase and hepatic triglyceride lipase in postheparin plasma of patients with low concentrations of HDL cholesterol. *Arterioscler Thromb*. 1993;13:1227-1235.
36. Rip J, Nierman MC, Wareham NJ, Luben R, Bingham SA, Day NE, van Miert JN, Hutten BA, Kastelein JJ, Kuivenhoven JA, Khaw KT, Boekholdt SM. Serum lipoprotein lipase concentration and risk for future coronary artery disease: the EPIC-Norfolk prospective population study. *Arterioscler Thromb Vasc Biol*. 2006;26:637-642.
37. Dowhan DH, Muscat GE. Characterization of the AB (AF-1) region in the muscle-specific retinoid X receptor-gamma: evidence that the AF-1 region functions in a cell-specific manner. *Nucleic Acids Res*. 1996;24:264-271.
38. Coon H, Myers RH, Borecki IB, Arnett DK, Hunt SC, Province MA, Djousse L, Leppert MF. Replication of linkage of familial combined hyperlipidemia to chromosome 1q with additional heterogeneous effect of apolipoprotein A-I/C-III/A-IV locus. The NHLBI Family Heart Study. *Arterioscler Thromb Vasc Biol*. 2000;20:2275-2280.
39. Elbein SC, Hoffman MD, Teng K, Leppert MF, Hasstedt SJ. A genome-wide search for type 2 diabetes susceptibility genes in Utah Caucasians. *Diabetes*. 1999;48:1175-1182.
40. van der Vleuten GM, Hijmans A, Kluijtmans LA, Blom HJ, Stalenhoef AF, de Graaf J. Thioredoxin interacting protein in Dutch families with familial combined hyperlipidemia. *Am J Med Genet A*. 2004;130:73-75.
41. Bodnar JS, Chatterjee A, Castellani LW, Ross DA, Ohmen J, Cavalcoli J, Wu C, Dains KM, Catanese J, Chu M, Sheth SS, Charugundla K, Demant P, West DB, de Jong P, Lusis AJ. Positional cloning of the combined hyperlipidemia gene Hyplip1. *Nat Genet*. 2002;30:110-116.
42. Pajukanta P, Lilja HE, Sinsheimer JS, Cantor RM, Lusis AJ, Gentile M, Duan XJ, Soro-Paavonen A, Naukkarinen J, Saarela J, Laakso M, Ehnholm C, Taskinen MR, Peltonen L. Familial combined hyperlipidemia is associated with upstream transcription factor 1 (USF1). *Nat Genet*. 2004;36:371-376.
43. Coon H, Xin Y, Hopkins PN, Cawthon RM, Hasstedt SJ, Hunt SC. Upstream stimulatory factor 1 associated with familial combined hyperlipidemia, LDL cholesterol, and triglycerides. *Hum Genet*. 2005;117:444-451.
44. Huertas-Vazquez A, Aguilar-Salinas C, Lusis AJ, Cantor RM, Canizales-Quinteros S, Lee JC, Mariana-Nunez L, Riba-Ramirez RM, Jokiah A, Tusie-Luna T, Pajukanta P. Familial combined hyperlipidemia in Mexicans: association with upstream transcription factor 1 and linkage on chromosome 16q24.1. *Arterioscler Thromb Vasc Biol*. 2005;25:1985-1991.
45. Putt W, Palmen J, Nicaud V, Tregouet DA, Tahri-Daizadeh N, Flavell DM, Humphries SE, Talmud PJ. Variation in USF1 shows haplotype effects, gene : gene and gene : environment associations with glucose and lipid parameters in the European Atherosclerosis Research Study II. *Hum Mol Genet*. 2004;13:1587-1597.
46. Ng MC, Miyake K, So WY, Poon EW, Lam VK, Li JK, Cox NJ, Bell GI, Chan JC. The linkage and association of the gene encoding upstream stimulatory factor 1 with type 2 diabetes and metabolic syndrome in the Chinese population. *Diabetologia*. 2005;48:2018-2024.
47. Gibson F, Hercberg S, Froguel P. Common polymorphisms in the USF1 gene are not associated with type 2 diabetes in French Caucasians. *Diabetes*. 2005;54:3040-3042.
48. Zeggini E, Damcott CM, Hanson RL, Karim MA, Rayner NW, Groves CJ, Baier LJ, Hale TC, Hattersley AT, Hitman GA, Hunt SE, Knowler WC, Mitchell BD, Ng MC, O'Connell JR, Pollin TI, Vaxillaire M, Walker M, Wang X, Whittaker P, Xiang K, Jia W, Chan JC, Froguel P, Deloukas P, Shuldiner AR, Elbein SC, McCarthy MI. Variation within the gene encoding the upstream stimulatory factor 1 does not influence susceptibility to type 2 diabetes in samples from populations with replicated evidence of linkage to chromosome 1q. *Diabetes*. 2006;55:2541-2548.

CETP (cholesteryl ester transfer protein) promoter – 1337 C > T polymorphism protects against coronary atherosclerosis in Japanese patients with heterozygous familial hypercholesterolaemia

Mutsuko TAKATA*, Akihiro INAZU†, Shoji KATSUDA*, Kenji MIWA*, Masa-aki KAWASHIRI*, Atsushi NOHARA‡, Toshinori HIGASHIKATA*, Junji KOBAYASHI§, Hiroshi MABUCHI‡ and Masakazu YAMAGISHI*

*Molecular Genetics of Cardiovascular Disorders, Graduate School of Medical Science, Kanazawa University, Takara-machi 13-1, Kanazawa 920-8641, Japan, †Department of Laboratory Science, Division of Health Sciences, Graduate School of Medical Science, Kanazawa University, Takara-machi 13-1, Kanazawa 920-8641, Japan, ‡Department of Lipidology, Kanazawa University, Takara-machi 13-1, Kanazawa 920-8641, Japan, and §Department for Lifestyle-related Diseases, Graduate School of Medical Science, Kanazawa University, Takara-machi 13-1, Kanazawa 920-8641, Japan

A B S T R A C T

CETP (cholesteryl ester transfer protein) and HL (hepatic lipase) play a role in the metabolism of plasma lipoproteins, but the effects of *CETP* and *LIPC* (gene encoding HL) genotypes on coronary atherosclerosis may be dependent on LDL (low-density lipoprotein)-receptor activity. Recently, the – 1337 C > T polymorphism in the *CETP* gene has been reported in REGRESS (Regression Growth Evaluation Statin Study) to be a major determinant of promoter activity and plasma CETP concentration. In the present study, we have investigated the effects of the *CETP* promoter – 1337 C > T and *LIPC* promoter – 514 C > T polymorphisms on serum lipid profiles and risk of coronary atherosclerosis in 206 patients (154 males) with heterozygous FH (familial hypercholesterolaemia). To evaluate coronary atherosclerosis, we used CSI (coronary stenosis index) calculated from coronary angiograms. The *CETP* – 1337 T allele was less frequent in subjects with a CSI ≥ 14 (mean value) in the group with coronary artery disease ($P = 0.04$, as determined by χ^2 test). ANOVA revealed that HDL-C (high-density lipoprotein-cholesterol) and triacylglycerol (triglyceride) levels were not significantly higher in the presence of the *CETP* promoter – 1337 T allele. Combined with *LIPC* promoter polymorphisms, HDL-C levels were highest and CSI were lowest with *CETP* – 1337 CT + TT and *LIPC* – 514 CC genotypes, but a significant interaction was not shown. A multiple logistic regression analysis revealed that, in patients with coronary atherosclerosis, the *CETP* – 1337 CC genotype was a significant genetic risk factor in FH (odds ratio = 2.022; $P = 0.0256$). These results indicate that the *CETP* promoter – 1337 C > T polymorphism is associated with the progression of coronary atherosclerosis in Japanese patients with FH, independent of HDL-C and triacylglycerol levels.

Key words: cholesteryl ester transfer protein (CETP), coronary artery disease, familial hypercholesterolaemia, hepatic lipase, single nucleotide polymorphism.

Abbreviations: AP, angina pectoris; Apo, apolipoprotein; BMI, body mass index; CAD, coronary artery disease; CETP, cholesteryl ester transfer protein; CSI, coronary stenosis index; FH, familial hypercholesterolaemia; HDL, high-density lipoprotein; HDL-C, HDL-cholesterol; HL, hepatic lipase; IDL, intermediate-density lipoprotein; LDL, low-density lipoprotein; LDL-C, LDL-cholesterol; MI, myocardial infarction; NCBI, National Center for Biotechnology Information; REGRESS, Regression Growth Evaluation Statin Study.

Correspondence: Dr Mutsuko Takata (email mutsuko9447@yahoo.co.jp).

INTRODUCTION

CETP (cholesteryl ester transfer protein) is a key player in the metabolism of major plasma lipoproteins. CETP mediates the transfer of cholesteryl esters from HDL (high-density lipoprotein) to Apo (apolipoprotein) B-containing lipoproteins in exchange for triacylglycerols (triglycerides) [1]. CETP activities are known to be highly affected by genetic factors. For example, individuals with homozygous CETP deficiency have high HDL-C (HDL-cholesterol) levels and low LDL-C [LDL (low-density lipoprotein)-cholesterol] levels, and have no evidence of premature atherosclerosis [2]. Also, *CETP* gene polymorphisms, especially the *TaqIB* polymorphism identified in intron 1, is reported to be highly associated with plasma CETP concentrations and HDL-C levels. Moreover, recent meta-analyses revealed that this polymorphism is associated with the incidence of CAD (coronary artery disease) [3–7]. However, this polymorphism is unlikely to be functional by itself, instead representing a surrogate marker of functional variants of the *CETP* gene [8]. Indeed, previous studies have shown that the *CETP* promoter –629 A > C polymorphism has almost complete linkage disequilibrium with the *TaqIB* polymorphism [9,10], and that this polymorphism is associated with CAD [11]. On the other hand, we have reported [8] that the haplotype block consisting of –2668 G/A, –2505 C/A, –1337 C/T and the shortest (gaaa) repeat had a stronger association than *TaqIB2* or –629 A/C with low plasma CETP concentrations and high HDL-C levels in healthy Japanese males. Moreover, functional interaction between –629 C/A, –971 G/A and –1337 C/T polymorphisms in the *CETP* gene is a major determinant of promoter activity and plasma CETP concentration in REGRESS (Regression Growth Evaluation Statin Study) [12].

In addition to CETP, HL (hepatic lipase) also plays a crucial role in the metabolism of plasma lipoproteins. HL is involved in the hydrolysis of triacylglycerol and phospholipids in IDL (intermediate-density lipoprotein) and large LDL particles to form smaller and denser LDL particles, and also plays a major role in promoting the conversion from HDL₂ into HDL₃ particles [13]. The effects of the *LIPC* genotype (the gene encoding HL) on atherosclerosis have been controversial [14], and may be dependent on LDL-receptor activity.

FH (familial hypercholesterolaemia) is an autosomal-dominant disorder characterized by primary hypercholesterolaemia with tendon xanthomas and premature CAD caused by mutations in the LDL receptor [15,16]. Mortality from CAD is reported to be several times higher in subjects with heterozygous FH than in the general population [15,16]. There are several reports that polymorphisms or mutations in the *CETP* gene influence the clinical characteristics of FH subjects

[17,18]. Carmena-Ramon et al. [17] reported that in FH the *TaqIB2* allele was associated with higher HDL-C and ApoAI levels. On the other hand, our previous study [18] showed that increased HDL-C levels caused by a heterozygous CETP deficiency was insufficient to prevent CAD in FH.

With this background, the present study investigated the effects of *CETP* promoter –1337 C > T and *LIPC* promoter –514 C > T polymorphisms on coronary atherosclerosis in Japanese patients with heterozygous FH.

METHODS

Study participants

We enrolled 206 consecutive Japanese patients with heterozygous FH (26–83 years old; 154 males) who attended our hospital. FH was diagnosed when one of the following two criteria was met: (i) primary hypercholesterolaemia [> 5.96 mmol/l (> 230 mg/dl) in any age group] in a patient with tendon xanthomas, or (ii) primary hypercholesterolaemia with a definitive diagnosis of FH in any first-degree relative [19]. All the females were post-menopausal, as defined by the absence of menstruation for > 6 months or having attained an age of ≥ 60 years. Those with surgical menopause were excluded. For patients with MI (myocardial infarction), the age at the first event was recorded, whereas for patients with AP (angina pectoris), the age at which coronary angiography was performed was recorded. Inclusion criteria for this study were FH patients who were examined by coronary angiography because of chest symptoms and/or a positive exercise test before lipid-lowering therapy was initiated. Individuals who had thyroid disease, levels of triacylglycerol ≥ 4.52 mmol/l (> 400 mg/dl) or who received lipid-lowering agents, corticosteroid or oestrogen hormone replacement therapy were excluded. All patients provided informed consent for participation in the present study, which was approved by the Ethical committee of Kanazawa University Graduate School of Medical Science.

Assessment of CAD

For the evaluation of CAD, we used CSI (coronary stenosis index) to quantify the severity of coronary atherosclerosis. The severity of stenotic changes was assessed by a score assigned to each of the 15 segments according to the classification of the American Heart Association Grading Committee. A normal coronary angiogram was graded as 0, stenosis of $< 25\%$ was graded as 1, 25–50% stenosis was graded as 2, 50–75% stenosis was graded as 3, and $> 75\%$ stenosis was graded as 4. CSI was defined as the sum of these scores in all 15 segments, producing a maximal value of 60 [15]. In the present study, MI was diagnosed in 56 subjects with

heterozygous FH (48 male), and AP was diagnosed in 53 subjects with heterozygous FH (all male). The mean CSI was 14.0 ± 11 . The mean CSI in subjects who were diagnosed with MI and AP was 20, whereas the mean CSI in those subjects who were without clinical symptoms of CAD was 8. In our previous study [15], we observed that the age of coronary artery stenosis detectable by angiogram occurs after 17–25 years of age in male and female subjects with heterozygous FH. In the present study, 86 % of the subjects with MI and AP had a CSI > 14, whereas 80 % of subjects without clinical symptoms of CAD had a CSI < 14. Therefore we diagnosed CAD as being present when CSI was > 14.

Assessment of conventional risk factors

Data for BMI (body mass index), smoking history, alcohol drinking, blood pressure, diabetes status and lipid profile were collected. Hypertension was considered to be present if any antihypertensive treatment had been instituted, if systolic blood pressure was > 160 mmHg or diastolic blood pressure > 95 mmHg. Diabetes mellitus was diagnosed if fasting plasma glucose was ≥ 6.70 mmol/l (> 120 mg/dl) or ≥ 11.10 mmol/l (> 200 mg/dl) at 120 min after 75 g of oral glucose loading, or if HbA_{1c} (glycated haemoglobin) was ≥ 6.5 %. For smoking status, we defined subjects who smoked ≤ 10 cigarettes/day as non-smokers, past smokers as ex-smokers and current smokers.

Laboratory analysis

Blood samples were collected from subjects after 12 h of fasting before starting lipid-lowering agents. Total cholesterol, triacylglycerols and HDL-C levels were determined by standard enzymatic methods. LDL-C levels were calculated using the Friedewald formula [20]. Plasma CETP levels were determined by sandwich ELISA, as described previously [21].

Determination of CETP and LIPC promoter polymorphisms

Genomic DNA was isolated and purified from peripheral white blood cells. The *CETP* promoter –1337 C > T polymorphism and the *LIPC* promoter –514 C > T polymorphism (–480 in older reports) were analysed by PCR-RFLP (restriction-fragment-length polymorphism) methods, as described previously [8,22]. Accession numbers are as follows: *CETP*, gene ID 1071 [NCBI (National Center for Biotechnology Information) Entrez Gene database], nucleotide sequence NM_000078 (NCBI Entrez Nucleotide database) and –1337C/T SNP rs17231506 (NCBI SNP database); and *LIPC*, gene ID 3990 (NCBI Entrez Gene database), nucleotide sequence NM_000236 (NCBI Entrez Nucleotide database), –514 C/T SNP rs1800588 (NCBI SNP database) and –514 C/T USF binding site ccttttgaca(c/t)gggggtgaag.

Table 1 Characteristics of subjects in this study

Values are means \pm S.E.M. HDL-C* was adjusted by multiple linear regression analysis, including gender, alcohol intake, smoking and BMI.

Parameter	CAD	non-CAD	P value
Gender (male/female)	77/17	77/35	0.0303
Age (years)	52 \pm 12	50 \pm 12	0.3001
BMI (kg/m ²)	23.7 \pm 3.0	23.9 \pm 2.7	0.5000
Total cholesterol (nmol/l)	8.34 \pm 1.74	8.37 \pm 1.63	0.9131
Triacylglycerol (nmol/l)	1.64 \pm 0.69	1.65 \pm 0.80	0.8618
HDL-C (nmol/l)	1.04 \pm 0.28	1.09 \pm 0.34	0.2052
HDL-C* (nmol/l)	1.17 \pm 0.28	1.22 \pm 0.31	0.3888
LDL-C (nmol/l)	6.55 \pm 1.79	6.53 \pm 1.66	0.8658
ApoA1 (g/l)	1.01 \pm 0.25	1.08 \pm 0.24	0.1222
ApoB (g/l)	1.77 \pm 0.53	1.78 \pm 0.44	0.9052
ApoE (g/l)	0.06 \pm 0.03	0.06 \pm 0.02	0.7330
Hypertension (n)	32 (34.0 %)	20 (17.9 %)	0.0070
Diabetes mellitus (n)	34 (36.2 %)	22 (19.6 %)	0.0079
Smokers (n)	54 (57.4 %)	63 (56.2 %)	0.8629
Alcohol drinkers (n)	36 (38.3 %)	44 (39.3 %)	0.8848
CSI	23.7 \pm 7.4	5.8 \pm 4.3	< 0.0001

Statistical analyses

All values are expressed as means \pm S.D. unless otherwise noted. The allele frequency was estimated by gene counting. One-way ANOVA was performed, followed by multiple comparisons using Fisher's protected least significant difference. Serum HDL-C was adjusted by multiple linear regression analysis. The prevalence of patients with hypertension, diabetes mellitus, current and past smoking, and alcohol drinking were compared between different groups using a χ^2 test. A multiple logistic regression analysis was used to predict CAD from the genotype of polymorphism, with conventional risk factors as covariates. A probability value of $P < 0.05$ was considered to be significant. All tests were performed with StatView software (version 5.0; SAS Institute).

RESULTS

Characteristics of study subjects

The clinical and biochemical characteristics of the study population either with CAD or without CAD (non-CAD) are summarized in Table 1. A total of 94 the subjects with heterozygotes FH were suffering from CAD. There were significantly more males and subjects with hypertension and diabetes mellitus in the CAD group compared with the non-CAD group.

Association between –1337 C > T polymorphism and CSI

The frequency of the *CETP* promoter –1337 T allele was 0.20 in both males and females; lower than in Caucasians [12]. A few subjects in the present study had

Table 2 *CETP* – 1337 C > T polymorphism and plasma *CETP* levels*P* value was determined using χ^2 test.

	<i>CETP</i> genotype				
	– 1337 CC		– 1337 CT + TT		<i>P</i> value
	<i>n</i>	<i>CETP</i> ($\mu\text{g/ml}$)	<i>n</i>	<i>CETP</i> ($\mu\text{g/ml}$)	
All	31	3.1 \pm 1.1	13	2.6 \pm 0.6	0.1364
Male	17	2.6 \pm 0.6	8	2.4 \pm 0.6	0.3139
Female	14	3.6 \pm 1.3	5	3.0 \pm 0.5	0.2831

the *CETP* promoter – 1337 TT genotype (11 males and two females), and the T allele was less frequent in subjects with a CSI \geq 14. The distribution of the *CETP* promoter – 1337 CC genotype differed significantly between those with a CSI \geq 14 and those with a CSI < 14 ($P = 0.0426$, as determined by a χ^2 test).

CETP promoter polymorphism and *CETP* concentrations

We compared plasma *CETP* concentrations between the – 1337 CC and – 1337 CT + TT genotypes in a subset of 44 subjects (25 males; Table 2). The *CETP* concentration tended to be lower in the presence of the T allele ($P = 0.14$).

CETP promoter polymorphism, lipid profile and development of CAD

The characteristics of subjects according to *CETP* promoter polymorphism are summarized in Table 3. As there were only two females with the TT genotype, we analysed men and women combined. HDL-C levels were not significantly higher in TT genotype, and the CSI tended to be lower in patients carrying the T allele ($P = 0.19$).

Effects of *CETP* and *LIPC* promoter polymorphisms on lipid profile and CSI

The frequency of the *LIPC* promoter – 514 T allele was 0.53 in males and 0.50 in females, which is similar to the frequencies previously reported in Japanese subjects, but higher than those in Caucasians [22,23]. To investigate the effects of *CETP* and *LIPC* promoter polymorphisms on lipid profile, we compared four subgroups stratified by high *CETP* genotype CC compared with low *CETP* CT + TT, and high *LIPC* genotype CC compared with low *LIPC* genotype CT + TT. Figure 1 shows that the HDL-C level was significantly higher in – 514 CC/– 1337 CT + TT than in – 514 CC/– 1337 CC [1.22 ± 0.36 mmol/l (47 ± 14 mg/dl) compared with 0.98 ± 0.30 mmol/l (38 ± 10 mg/dl) respectively; $P < 0.02$], and it was significantly higher in – 514 CC/– 1337 CT + TT than in – 514 CT + TT/– 1337 CC or in both CT + TT ($P < 0.05$). LDL-C

Table 3 Characteristics of the subjects according to *CETP* genotype statusValues are means \pm S.E.M. HDL-C* was adjusted by multiple linear regression analysis, including gender, alcohol intake, smoking and BMI.

	<i>CETP</i> genotype		
	CC	CT	TT
<i>n</i>	127	66	13
Total cholesterol (nmol/l)	8.50 \pm 1.71	8.18 \pm 1.66	7.87 \pm 1.27
Triacylglycerol (nmol/l)	1.62 \pm 0.72	1.73 \pm 0.82	1.48 \pm 0.57
HDL-C (nmol/l)	1.04 \pm 0.28	1.09 \pm 0.37	1.17 \pm 0.37
HDL-C* (mmol/l)	1.19 \pm 0.28	1.22 \pm 0.37	1.23 \pm 0.37
LDL-C (nmol/l)	6.71 \pm 1.81	6.29 \pm 1.61	6.03 \pm 1.24
ApoA1 (g/l)	1.02 \pm 0.25	1.09 \pm 0.25	1.08 \pm 0.21
ApoB (g/l)	1.81 \pm 0.50	1.76 \pm 0.47	1.66 \pm 0.36
ApoE (g/l)	0.07 \pm 0.03	0.07 \pm 0.03	0.05 \pm 0.02
Age (years)	50 \pm 11	53 \pm 13	48 \pm 11
BMI (kg/m^2)	23.7 \pm 2.7	24.3 \pm 3.2	22.8 \pm 2.5
Smokers (%)	69 (54.3)	38 (57.6)	10 (76.9)
Hypertension (%)	30 (23.6)	18 (27.3)	4 (30.8)
Diabetes mellitus (%)	35 (27.6)	19 (28.8)	2 (15.4)
CSI	15.0 \pm 10.7	12.4 \pm 10.6	11.6 \pm 9.6

levels did not differ significantly between the four groups. CSI was significantly lower in – 514 CC/– 1337 CT + TT than in – 514 CC/– 1337 CC (9.6 compared with 17.2 respectively; $P = 0.02$), suggesting an interaction between *CETP* and *LIPC* genotype on CSI.

Multiple logistic regression analysis

A multiple logistic regression analysis was performed to determine the association of CAD and *CETP* promoter polymorphism and other conventional risk factors. Gender, hypertension, diabetes mellitus and *CETP* – 1337 CC genotype exhibited significantly higher odds ratios; however, age, smoking, HDL-C and triacylglycerol levels, and the presence of *LIPC* – 514 C > T were not significant variates (Table 4).

DISCUSSION

The present study investigated the effects of *CETP* and *LIPC* promoter polymorphisms on serum lipid profiles and risk of coronary atherosclerosis in subjects with heterozygous FH. None of the other coronary risk factors differed significantly between *CETP* genotypes; however, multiple logistic regression analysis revealed that coronary atherosclerosis was associated with the *CETP* – 1337 CC genotype. An interaction between the *CETP* and *LIPC* genotypes for plasma HDL-C and CAD has also been shown.

To our knowledge, this is the first study on the effects of the *CETP* promoter – 1337 C > T polymorphism

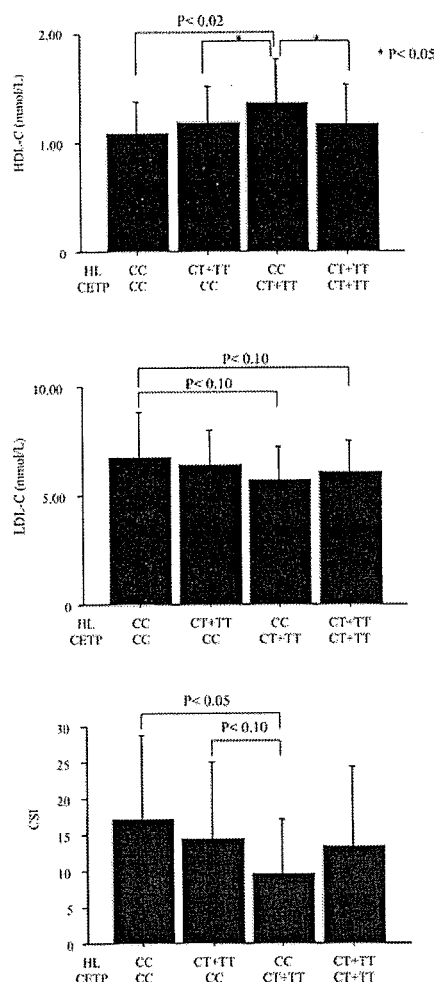


Figure 1 Effects of *CETP* and *LIPC* promoter polymorphisms on LDL-C, HDL-C and CSI

Subjects: -514 CC/-1337 CT+TT ($n=29$); -514 CT+TT/-1337 CC ($n=98$); -514 CC/-1337 CT+TT ($n=17$); -514 CT+TT/-1337 CT+TT ($n=62$). The HDL-C level was significantly higher in -514 CC/-1337 CT+TT than in -514 CC/-1337 CC (1.22 ± 0.36 compared with 0.98 ± 0.03 mmol/L; $P < 0.02$), and it was significantly higher in -514 CC/-1337 CT+TT than in -514 CT+TT/-1337 CC or CT+TT ($P < 0.05$). The LDL-C levels did not differ significantly between the four groups. CSI was significantly lower in -514 CC/-1337 CT+TT subjects than in -514 CC/-1337 CC subjects (9.6 compared with 17.2 ; $P = 0.02$).

in coronary atherosclerosis and, therefore, the first to suggest that the *CETP* promoter -1337 C>T polymorphism is associated with the severity of coronary atherosclerosis in heterozygous FH. In a previous study [8], this polymorphism was associated with low plasma CETP concentrations and high HDL-C levels more strongly than with the *TaqIB2* allele in elderly Japanese males and, recently, this polymorphism has been reported to be a major determinant of promoter activity and plasma CETP concentration in REGRESS [12]. Therefore we

Table 4 Multivariate adjusted relative prevalence odds ratio of coronary atherosclerosis by multiple logistic regression analysis

For sex, male = 1 and female = 0; for hypertension, yes = 1 and no = 0; for diabetes mellitus, yes = 1 and no = 0; for *CETP* -1337 C>T polymorphism, CC = 1 and CT+ = 0; for *LIPC* -514 C>T polymorphism, CC = 2, CT = 1 and TT = 0.

Variate	Odds ratio	P value
Age	1.021 (0.993–1.050)	0.1431
Sex	4.283 (1.788–10.259)	0.0011
Hypertension	2.628 (1.252–5.519)	0.0107
Diabetes mellitus	2.136 (1.081–4.218)	0.0289
Smoking	0.992 (0.969–1.015)	0.3261
<i>CETP</i> -1337 C>T polymorphism	2.022 (1.090–3.754)	0.0256
<i>LIPC</i> -514 C>T polymorphism	0.856 (0.562–1.305)	0.4698

investigated this -1337 site rather than the well-known *TaqIB* polymorphism. As subjects with FH have a high risk of premature CAD, we determined the existence of early stage coronary atherosclerotic changes by using CSI. Our present data suggest that the association of the *CETP* genotype with cardiovascular risk is independent of serum HDL-C levels. As indicated in Table 3, there was no significant difference in HDL-C/adjusted HDL-C levels between *CETP* genotypes. The *CETP* *TaqIB2* allele was associated with HDL-C, especially HDL₂-C, in Japanese subjects [24] and, therefore, if we had assessed HDL₂-C, this might have revealed a significant difference between the genotypes.

There are conflicting reports as to whether *CETP* is pro- or anti-atherogenic. Humans with homozygous *CETP* deficiency have markedly high HDL-C levels and decreased LDL-C levels, with no clear evidence of premature atherosclerosis [2]. A *CETP* gene mutation (D442G) was shown to be associated with increased LDL particle size [25], suggesting that *CETP* is pro-atherogenic. In contrast, Hirano et al. [26] have reported that the prevalence of *CETP* deficiency was lower in individuals older than 80 years of age residing in a district of northern Japan, suggesting that *CETP* deficiency is not association with longevity, and the same investigators have shown that reduced *CETP* activity in conjunction with reduced HL activity is associated with an increased risk of CAD [27]. On the other hand, Moriyama et al. [28] found in a cross-sectional analysis that HDL-C elevation (≥ 80 mg/dl) was protective against coronary heart disease, regardless of *CETP* genotype, in 19044 male and 29487 female Japanese subjects. In addition, a recent prospective study in the Honolulu Heart Program has shown the protective effects of heterozygous *CETP* deficiency against CAD, although the effect was not statistically significant [29].

At lower CETP concentrations, LDL-receptor activity is up-regulated, causing a reduction in serum LDL levels and leading to atheroprotection. Lowering CETP activity may be beneficial in an affluent environment, where high-fat and cholesterol-rich diets increase plasma LDL-C levels and down-regulate hepatic LDL-receptors, such as in FH. We presume that individuals with FH have higher CETP activity or concentration than normolipidaemic controls [30,31], which would be less pro-atherogenic when they carry the *CETP* promoter -1337 T allele. De Grooth et al. [32] reported a significant positive correlation between carotid intima-media thickness and CETP levels in FH, suggesting that plasma CETP would be pro-atherogenic in FH. There are also some reports on the *CETP* TaqIB polymorphism and impaired glucose tolerance [33], suggesting that CETP could be pro-atherogenic independently of lipid metabolism. In the present study, however, there was no significant difference between *CETP* promoter -1337 C > T polymorphism and serum glucose levels (5.99 ± 1.94 mmol/l in -1337 CC compared with 5.72 ± 1.33 mmol/l in -1337 CT + TT; $P = 0.20$), and no difference in diabetes prevalence (results not shown).

In addition to CETP, HL also plays a crucial role in the metabolism of plasma lipoproteins, but the effects of CETP and HL activity on lipid profile and CAD are unclear [14,34]. The present study found no association between the *LIPC* promoter -514 C > T polymorphism and CAD and HDL-C levels; however, CSI with the *CETP* -1337 T allele and *LIPC* -514 CC was lowest in the subgroup. In another study from our laboratory (M. Takata and A. Inazu, unpublished work), HL activity was significantly higher in -514 CC than CT + TT (0.282 ± 0.011 compared 0.231 ± 0.005 mmol/l respectively, $P < 0.001$) in hyperlipidaemic patients ($n = 325$, of which 183 were male). In human studies, HL activity tends to be elevated in the presence of smoking [35], insulin resistance in Type II diabetes mellitus [36], in females with omental fat mass [37] and males in general. These reports suggest that HL is pro-atherogenic. On the other hand, it has been reported that HL activity is lower in patients with CAD than in those without CAD [38]. Another group found that HL activity did not differ between subjects with and without CAD in REGRESS [39]. In an environment of low HL activity, IDL increases and it may be pro-atherogenic [40]. HL also promotes the formation of small and dense atherogenic LDL particles [13]. Lowering HL activity in hypertriglyceridaemia may decrease the pro-atherogenic risk due to an improved lipid profile, notably an increased LDL size [14]. In conditions where LDL-receptor activity is low, as in FH, HL activity appears to be inversely associated with CAD in subjects with low CETP concentrations (Figure 1), suggesting that the flux of cholesterol through the system of HDL-C transport may be more important in preventing atherosclerosis.

The main limitations of the present study were the relatively small sample size and the absence of data on HDL subclass and LDL particle size.

In conclusion, the *CETP* promoter -1337 C > T polymorphism is associated with the progression of coronary atherosclerosis in Japanese patients with FH, independent of HDL-C and triacylglycerol levels. We believe that this genetic variant of the *CETP* gene promoter could be an important determinant of coronary atherosclerosis in FH, and genotype differences between promoter variants and missense mutations need to be clarified in future investigations.

ACKNOWLEDGMENTS

We express special thanks to the lipidologists and cardiologists at the Second Department of Internal Medicine, Kanazawa University, and also to Sachio Yamamoto, Mihoko Mizuno and Mayumi Yoshida for their technical assistance. This work was supported by the Scientific Research Grant from the Ministry of Education, Science and Culture of Japan (no. 10770568 and 0907010).

REFERENCES

- 1 Tall, A. R. (1993) Plasma cholesteryl ester transfer protein. *J. Lipid Res.* **34**, 1255–1274
- 2 Inazu, A., Brown, M. L., Hesler, C. B. et al. (1990) Increased high-density lipoprotein levels caused by a common cholesteryl-ester transfer protein gene mutation. *N. Engl. J. Med.* **323**, 1234–1238
- 3 Kuivenhoven, J. A., Jukema, J. W., Zwilander, A. H. et al. (1998) The role of a common variant of the cholesteryl ester transfer protein gene in the progression of coronary atherosclerosis. The Regression Growth Evaluation Statin Study Group. *N. Engl. J. Med.* **338**, 86–93
- 4 Kuivenhoven, J. A., de Knijff, P., Boer, J. M. et al. (1997) Heterogeneity at the CETP gene locus. Influence on plasma CETP concentrations and HDL cholesterol levels. *Arterioscler., Thromb., Vasc. Biol.* **17**, 560–568
- 5 Ordovas, J. M., Cupples, L. A., Corella, D. et al. (2000) Association of cholesteryl ester transfer protein-TaqIB polymorphism with variations in lipoprotein subclasses and coronary heart disease risk: the Framingham study. *Arterioscler., Thromb., Vasc. Biol.* **20**, 1323–1329
- 6 Carlquist, J. F., Muhlestein, J. B., Horne, B. D. et al. (2003) The cholesteryl ester transfer protein TaqIB gene polymorphism predicts clinical benefit of statin therapy in patients with significant coronary artery disease. *Am. Heart J.* **146**, 1007–1014
- 7 Boekholdt, S. M., Sacks, F. M., Jukema, J. W. et al. (2005) Cholesteryl ester transfer protein TaqIB variant, high-density lipoprotein cholesterol levels, cardiovascular risk, and efficacy of pravastatin treatment: individual patient meta-analysis of 13 677 subjects. *Circulation* **111**, 278–287
- 8 Lu, H., Inazu, A., Moriyama, Y. et al. (2003) Haplotype analyses of cholesteryl ester transfer protein gene promoter: a clue to an unsolved mystery of TaqIB polymorphism. *J. Mol. Med.* **81**, 246–255
- 9 Dacher, C., Poirier, O., Cambien, F., Chapman, J. and Rouis, M. (2000) New functional promoter polymorphism, CETP/-629, in cholesteryl ester transfer protein (CETP) gene related to CETP mass and high density lipoprotein cholesterol levels: role of Sp1/Sp3 in transcriptional regulation. *Arterioscler., Thromb., Vasc. Biol.* **20**, 507–515

- 10 Klerkx, A. H., Tanck, M. W., Kastelein, J. J. et al. (2003) Haplotype analysis of the CETP gene: not TaqIB, but the closely linked -629C→A polymorphism and a novel promoter variant are independently associated with CETP concentration. *Hum. Mol. Genet.* **12**, 111–123
- 11 Blankenberg, S., Rupprecht, H. J., Bickel, C. et al. (2003) Common genetic variation of the cholesteryl ester transfer protein gene strongly predicts future cardiovascular death in patients with coronary artery disease. *J. Am. Coll. Cardiol.* **41**, 1983–1989
- 12 Frisdal, E., Klerkx, A. H., Le Goff, W. et al. (2005) Functional interaction between -629C/A, -971G/A and -1337C/T polymorphisms in the CETP gene is a major determinant of promoter activity and plasma CETP concentration in the REGRESS Study. *Hum. Mol. Genet.* **14**, 2607–2618
- 13 Kuusi, T., Saarinen, P. and Nikkila, E. A. (1980) Evidence for the role of hepatic endothelial lipase in the metabolism of plasma high density lipoprotein 2 in man. *Atherosclerosis* **36**, 589–593
- 14 Jansen, H., Verhoeven, A. J. and Sijbrands, E. J. (2002) Hepatic lipase: a pro- or anti-atherogenic protein? *J. Lipid Res.* **43**, 1352–1362
- 15 Mabuchi, H., Koizumi, J., Shimizu, M. and Takeda, R. (1989) Development of coronary heart disease in familial hypercholesterolemia. *Circulation* **79**, 225–232
- 16 Goldstein, J. L., Hobbs, H. H., Brown, M. S. et al. (2001) Familial hypercholesterolemia. In *The Metabolic and Molecular Bases of Inherited Disease*, vol. 2, 8th ed. (Scriver, C. R., Beaudet, A. L., Sly, W. S. et al., eds.), pp. 2863–2913, McGraw-Hill, New York
- 17 Carmena-Ramon, R., Ascaso, J. F., Real, J. T., Najera, G., Ordovas, J. M. and Carmena, R. (2001) Association between the TaqIB polymorphism in the cholesteryl ester transfer protein gene locus and plasma lipoprotein levels in familial hypercholesterolemia. *Metab., Clin. Exp.* **50**, 651–656
- 18 Haraki, T., Inazu, A., Yagi, K., Kajinami, K., Koizumi, J. and Mabuchi, H. (1997) Clinical characteristics of double heterozygotes with familial hypercholesterolemia and cholesteryl ester transfer protein deficiency. *Atherosclerosis* **132**, 229–236
- 19 Mabuchi, H., Higashikata, T., Nohara, A. et al. (2005) Cutoff point separating affected and unaffected familial hypercholesterolemic patients validated by LDL-receptor gene mutants. *J. Atheroscler. Thromb.* **12**, 35–40
- 20 Friedewald, W. T., Levy, R. I. and Fredrickson, D. S. (1972) Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin. Chem.* **18**, 499–502
- 21 Kiyohara, T., Kiriya, R., Zamma, S. et al. (1998) Enzyme immunoassay for cholesteryl ester transfer protein in human serum. *Clin. Chim. Acta* **271**, 109–118
- 22 Inazu, A., Nishimura, Y., Terada, Y. and Mabuchi, H. (2001) Effects of hepatic lipase gene promoter nucleotide variations on serum HDL cholesterol concentration in the general Japanese population. *J. Hum. Genet.* **46**, 172–177
- 23 Carr, M. C., Brunzell, J. D. and Deeb, S. S. (2004) Ethnic differences in hepatic lipase and HDL in Japanese, black, and white Americans: role of central obesity and LIPC polymorphisms. *J. Lipid Res.* **45**, 466–473
- 24 Ikewaki, K., Mabuchi, H., Teramoto, T. et al. (2003) Japan CETP Study Group. Association of cholesteryl ester transfer protein activity and TaqIB polymorphism with lipoprotein variations in Japanese subjects. *Metab., Clin. Exp.* **52**, 1564–1570
- 25 Wang, J., Qiang, H., Chen, D., Zhang, C. and Zhuang, Y. (2002) CETP gene mutation (D442G) increases low-density lipoprotein particle size in patients with coronary heart disease. *Clin. Chim. Acta* **322**, 85–90
- 26 Hirano, K., Yamashita, S., Nakajima, N. et al. (1997) Genetic cholesteryl ester transfer protein deficiency is extremely frequent in the Omagari area of Japan. Marked hyperalphalipoproteinemia caused by CETP gene mutation is not associated with longevity. *Arterioscler., Thromb., Vasc. Biol.* **17**, 1053–1059
- 27 Hirano, K., Yamashita, S., Kuga, Y. et al. (1995) Atherosclerotic disease in marked hyperalphalipoproteinemia. Combined reduction of cholesteryl ester transfer protein and hepatic triglyceride lipase. *Arterioscler., Thromb., Vasc. Biol.* **15**, 1849–1856
- 28 Moriyama, Y., Okamura, T., Inazu, A. et al. (1998) A low prevalence of coronary heart disease among subjects with increased high-density lipoprotein cholesterol levels, including those with plasma cholesteryl ester transfer protein deficiency. *Prev. Med.* **27**, 659–667
- 29 Curb, J. D., Abbott, R. D., Rodriguez, B. L. et al. (2004) A prospective study of HDL-C and cholesteryl ester transfer protein gene mutations and the risk of coronary heart disease in the elderly. *J. Lipid Res.* **45**, 948–953
- 30 Inazu, A., Koizumi, J., Mabuchi, H., Kajinami, K. and Takeda, R. (1992) Enhanced cholesteryl ester transfer protein activities and abnormalities of high density lipoproteins in familial hypercholesterolemia. *Horm. Metab. Res.* **24**, 284–288
- 31 Hogue, J. C., Lamarche, B., Gaudet, D. et al. (2004) Relationship between cholesteryl ester transfer protein and LDL heterogeneity in familial hypercholesterolemia. *J. Lipid Res.* **45**, 1077–1083
- 32 de Grooth, G. J., Smilde, T. J., Van Wissen, S. et al. (2004) The relationship between cholesteryl ester transfer protein levels and risk factor profile in patients with familial hypercholesterolemia. *Atherosclerosis* **173**, 261–267
- 33 Weitgasser, R., Galvan, G., Malaimare, L. et al. (2004) Cholesteryl ester transfer protein TaqIB polymorphism and its relation to parameters of the insulin resistance syndrome in an Austrian cohort. *Biomed. Pharmacother.* **58**, 619–627
- 34 de Grooth, G. J., Klerkx, A. H., Stroes, E. S., Stalenhoef, A. F., Kastelein, J. J. and Kuivenhoven, J. A. (2004) A review of CETP and its relation to atherosclerosis. *J. Lipid Res.* **45**, 1967–1974
- 35 Kong, C., Nimmo, L., Elatrozy, T. et al. (2001) Smoking is associated with increased hepatic lipase activity, insulin resistance, dyslipidaemia and early atherosclerosis in Type 2 diabetes. *Atherosclerosis* **156**, 373–378
- 36 Baynes, C., Henderson, A. D., Anyaoku, V. et al. (1991) The role of insulin insensitivity and hepatic lipase in the dyslipidaemia of type 2 diabetes. *Diabetic Med.* **8**, 560–566
- 37 Carr, M. C., Hokanson, J. E., Zambon, A. et al. (2001) The contribution of intraabdominal fat to gender differences in hepatic lipase activity and low/high density lipoprotein heterogeneity. *J. Clin. Endocrinol. Metab.* **86**, 2831–2837
- 38 Dugi, K. A., Brandauer, K., Schmidt, N. et al. (2001) Low hepatic lipase activity is a novel risk factor for coronary artery disease. *Circulation* **104**, 3057–3062
- 39 Jansen, H., Verhoeven, A. J., Weeks, L. et al. (1997) Common C-to-T substitution at position -480 of the hepatic lipase promoter associated with a lowered lipase activity in coronary artery disease patients. *Arterioscler., Thromb., Vasc. Biol.* **17**, 2837–2842
- 40 Hodis, H. N., Mack, W. J., Dunn, M. et al. (1997) Intermediate-density lipoproteins and progression of carotid arterial wall intima-media thickness. *Circulation* **95**, 2022–2022

Received 20 April 2006/21 June 2006; accepted 6 July 2006

Published as Immediate Publication 6 July 2006, doi:10.1042/CS20060088

Cardiac Resurrection After Bone-Marrow-Derived Mononuclear Cell Transplantation During Left Ventricular Assist Device Support

Satoshi Gojo, MD, PhD, Shunei Kyo, MD, PhD,
Shigeyuki Nishimura, MD, PhD,
Nobuyuki Komiyama, MD, PhD,
Nobutaka Kawai, MD, PhD, Masami Bessho, MD, PhD,
Hiroshige Sato, MD, PhD, Toshihisa Asakura, MD, PhD,
Motonobu Nishimura, MD, PhD,
and Kenji Ikebuchi, MD, PhD

Department of Cardiovascular Surgery, Saitama Medical Center,
and Departments of Cardiovascular Surgery, Cardiology,
Hematology, and Transfusion and Cell Therapy, Saitama
Medical School, Saitama, Japan

We describe a novel therapy of mononuclear cell transplantation combined with a left ventricular assist device (LVAD) for severe ischemic heart failure. Significant myocardial recovery by the LVAD rarely occurs in the severely failing heart. We undertook successful mononuclear cell transplantation in a patient who sustained an acute myocardial infarction that had resulted in the LVAD therapy. The heart regained good function after cell transplantation, and the LVAD was explanted 6 weeks later. These results suggest that this novel therapy could be an alternative to cardiac transplantation for severe ischemic heart failure.

(Ann Thorac Surg 2007;83:661–2)

© 2007 by The Society of Thoracic Surgeons

The discovery of pluripotent stem cells in an adult has opened a novel clinical research field, regenerative medicine [1]. Many studies have demonstrated that bone-marrow-derived progenitor cells can differentiate into cardiomyocytes and endothelial cells, and they can be involved in repairing injured hearts [2]. Several clinical trials of autologous bone-marrow-derived mononuclear cell transplantation after acute myocardial infarction revealed the steady improvement in cardiac function [3]. We report a successful myocardial recovery with mononuclear cell transplantation and left ventricular assist device (LVAD) support after cardiogenic shock due to acute myocardial infarction.

A 61-year-old man who had diabetes mellitus was transferred in a shock state due to acute myocardial infarction. Emergency cardiac catheterization demonstrated the diagnosis of complete occlusion of the #7 left anterior descending artery (LAD) and 90% stenosis of the #2 right coronary artery (RCA). The culprit lesion, #7LAD, was not eligible for percutaneous coronary intervention because the wire could not cross it. Thereafter, the patient's shock state was worse, and percutaneous cardiopulmonary support was initiated.

Accepted for publication June 23, 2006.

Address correspondence to Dr Gojo, Department of Cardiovascular Surgery, Saitama Medical Center 1981 Kamoda, Kawagoe, Saitama 350-8550, Japan; e-mail: satoshi@saitama-med.ac.jp.

Despite maximum pharmacologic support, the patient lapsed into multiple organ failure. The decision was made to implant a Toyobo LVAD (Toyobo, Inc, Osaka, Japan), and simultaneously perform a coronary artery bypass graft to the LAD and RCA with saphenous veins.

The status of multiple organ failure was gradually improved, but an ejection fraction (EF) by echocardiography was 0.13 on day 97 after LVAD implantation (Fig 1). Scintigraphy demonstrated complete infarctions in the anteroapical and inferior walls (Fig 2). The patient was briefed in detail about mononuclear cell transplantation. This clinical study had been approved by the Ethics Committee of the Saitama Medical School, Saitama, Japan.

Bone marrow (600 mL) was aspirated under general anesthesia from both posterior ilia and enriched to the mononuclear cell fraction. The mononuclear cells were implanted in the infarcted zone through the saphenous grafts to the LAD and RCA on day 99 after LVAD implantation. During the procedure, the electrocardiogram was monitored and did not demonstrate any significant changes to suggest ischemic events. The possibility of microemboli was also negative on the basis of the stable normal values for creatine kinase-MB fraction and troponin-T after the procedure.

The patient's cardiac function became gradually better with time after the cell transplantation. The LVAD was removed on day 43 after mononuclear cell transplantation. The EF increased from 0.064 to .40 and remained stable, as assessed by echocardiography. Analysis of LV function by scintigraphy demonstrated a sustained improvement in blood perfusion and regional EF in the apical and inferior walls and growing thickness of the septal and inferior walls in the time course (Fig 2). The patient was discharged 58 days after explantation of the LVAD. After mononuclear cell transplantation, there were no complications, including acute inflammatory response, novel infarction, malignant arrhythmias, or ectopic differentiation.

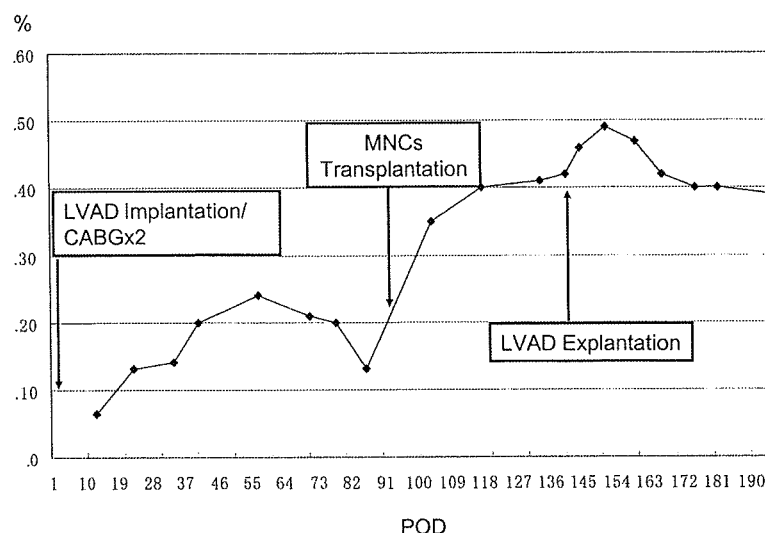
Comment

Our aim was to extend the target of cell transplantation from the mild to the severely failing heart. The requirements were (1) a donor cell type, (2) an implantation procedure, including duration, route, and targets for cell delivery, and (3) a timing of cell transplantation after LVAD implantation. We chose bone-marrow-derived mononuclear cells to contain all of the cell fractions because (1) there is a controversy about the cell source to be grafted, (2) nobody can deny the possibility that the enrichment procedure will cause the useful cell population to be lost based on the current experimental data, and (3) pure, dense stem cells in a site might induce an ectopic differentiation.

As a grafting route, an antegrade intracoronary infusion through the saphenous vein grafts was chosen to avoid the isolated islet-like formation of grafted cells by direct injection into the myocardium, which could not effectively induce neogenesis of either cardiomyocytes or coronary capillaries.

The final important issue of the protocol was the timing of the cell transplantation after LVAD implantation. Although the LVAD improves cardiac milieu interne in the

Fig 1. Left ventricular ejection fraction (EF) in transthoracic echocardiography. After left ventricular assist system (LVAS) implantation, the first evaluation showed an EF of 6.4%. In the time course under LVAS support, EF gradually improved, but the value began decreasing 2 months later. (POD = postoperative day; CABG = coronary artery bypass grafting; MNCs = mononuclear cells; LVAD = left ventricular assist device.)



early phase, long-term LVAD support induces ventricular atrophy so that the LVAD is a double-edged sword. We had been examining EF, LV wall thickness, and motions by echocardiography. Because the decline of EF commenced on day 72 after LVAD implantation, we judged that the global effects of LVAD for cardiac recovery had turned from benefits to drawbacks and performed mononuclear cell transplantation on day 99 after LVAD implantation.

The structural and functional improvements in last scintigraphy compared with that 1 month after mononuclear cell transplantation suggest that the cells engrafted, survived, and functioned in recipient heart. Grafted mononuclear cells release a wide array of cytokines related to the regeneration process. Mononuclear cell transplantation might stimulate the native environment to promote angiogenesis and cardiomyogenesis through the paracrine fashion in addition to vasculogenesis by the mononuclear cells themselves.

Many reports demonstrated that the LVAD support could induce reverse remodeling. In addition to relief of myocardium from the mechanical stretch of LVAD, the reverse remodeling of diseased heart could facilitate the engraftment, survival, and differentiation process of the grafted cells. Taken together, we think that mononuclear cell transplantation could fully work to repair the end-stage failing heart under the resting state created by LVAD support. This synergy effect of mononuclear cell transplantation and LVAD could be an explanation of this cardiac resurrection.

In conclusion, mononuclear cell transplantation for the treatment of ischemic cardiomyopathy with LVAD led to successful recovery of the failing heart and the LVAD to be unnecessary. We believe that this combination therapy might be an alternative to cardiac transplantation in the treatment of ischemic end-stage heart failure.

This work was supported in part by a Research Grant for Cardiovascular Diseases (16C-6) from the Ministry of Health, Labour and Welfare.

Regional EF Motion Thickening

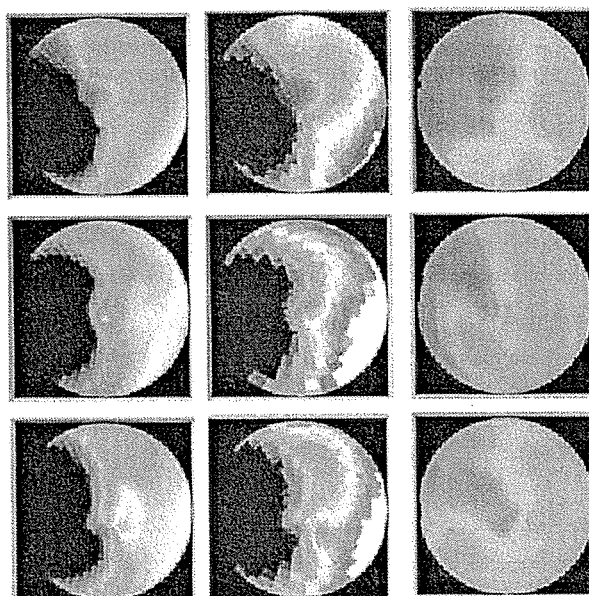


Fig 2. Technetium (Tc 99m)-tetrofosmin-gated single photon emission computed tomography. (Upper panels) Thirty-seven days after left ventricular assist device (LVAD) implantation and two coronary artery bypass grafts. (Middle panels) Twenty-nine days after mononuclear cell transplantation. (Lower panels) Twenty-five days after LVAD explantation (57 days after mononuclear cell transplantation).

References

- Pittenger MF, Mackay AM, Beck SC, et al. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999;284:143-7.
- Gojo S, Umezawa A. Plasticity of mesenchymal stem cells—regenerative medicine for diseased hearts. *Hum Cell* 2003;16:23-30.
- Wollert KC, Meyer GP, Lotz J, et al. Intracoronary autologous bone-marrow cell transfer after myocardial infarction: the BOOST randomised controlled clinical trial. *Lancet* 2004;364:141-8.

Stent Deformity Caused by Coronary Artery Spasm

Toshihiko Yoshida, MD; Yoshio Kobayashi, MD; Takashi Nakayama, MD;
Nakabumi Kuroda, MD; Nobuyuki Komiyama, MD*; Issei Komuro, MD

Previous studies have shown that coronary stents have radial strength above the pressure induced by coronary artery spasm. This case report describes a stent deformity caused by coronary artery spasm during percutaneous coronary intervention. (*Circ J* 2006; 70: 800–801)

Key Words: Angioplasty; Stent; Vasospasm

Despite full medical treatment with calcium channel blockers and nitrates, some patients with coronary artery spasm continue to have recurrent episodes of angina and myocardial infarction, and arrhythmic sudden death might occur.^{1–6} Previous reports demonstrate the usefulness of coronary stents to prevent vasospasm refractory to medical therapy.^{7–9} However, coronary stent implantation is sometimes ineffective in patients with multivessel spasm.¹⁰ This case report describes another limitation of coronary stent implantation to prevent coronary artery spasm.

Case Report

A 53-year-old man had been well until he had severe chest pain caused by acute myocardial infarction. He was referred 2 weeks after the onset of acute myocardial infarction. Coronary angiography revealed a 90% stenosis in the proximal obtuse marginal artery (Fig 1A). There was no significant stenosis in the left anterior descending coronary artery and right coronary artery. A 0.014-inch Skipper guidewire (Asahi Intecc, Nagoya, Japan) was placed across the lesion into the distal obtuse marginal artery. Predilation was performed by using a 3.0-mm OMNIPASS balloon catheter (Cordis, Miami, FL, USA) inflated to 6 atm. Intravascular ultrasound (IVUS) imaging was performed in the obtuse marginal artery using a 30-MHz 3.2F Ultracross catheter (Boston Scientific, Natick, MA, USA). The IVUS image showed a significant stenosis with fibrofatty plaque. A 25 mm NIR stent premounted on a 3.0-mm balloon catheter (Medinol, Tel Aviv, Israel) was deployed in the proximal obtuse marginal artery using an inflation pressure of 14 atm. Angiography and IVUS showed a good result (Figs 1B, 2A). After the guidewire was withdrawn, the patient complained of chest pain; electrocardiogram demonstrated ST-segment elevation in lead I and aVL and the systolic blood pressure dropped from 140 to 80 mmHg. Angiography demonstrated coronary artery spasm at the proximal stented segment and distal reference (Fig 1C).

Thereafter, stent deformity was observed (Fig 2B). Intravenous norepinephrine (0.2 mg) and intracoronary nitroglycerine (200 µg) were administered. The systolic blood pressure increased to 100 mm Hg and the coronary artery spasm was relieved (Fig 1D). Further coronary intervention was not performed because there was no flow disturbance (TIMI 3) in the obtuse marginal artery. The patient received oral aspirin (100 mg daily), ticlopidine (100 mg twice daily for 4 weeks), diltiazem (100 mg daily), and isosorbide mononitrate (20 mg twice daily). There was no in-hospital event. During follow-up, no adverse event was observed. Six months later, follow-up angiography was performed. The deformed stent (Fig 2C) and a 25% stenosis at the distal stented segment (Fig 1E) were observed.

Discussion

Calcium antagonists and nitrates are effective in preventing coronary artery spasm in most cases. However, coronary artery spasm refractory to the treatment with these drugs is observed in some cases.^{1–6} Alpha-1 blocking agents,² magnesium,³ benzhexol hydrochloride,⁴ denopamine,⁵ and nicorandil⁶ have been reported as alternatives. Previous reports demonstrated the efficacy of coronary stenting in patients with clinically severe coronary artery spasm refractory to aggressive pharmacologic management.^{7–9} Gaspardone et al evaluated the usefulness of coronary stent placement in 9 patients with vasospastic angina refractory to medical treatment.¹⁰ The NIR stent was used in 6 patients. During follow-up, 3 patients developed recurrent episodes of angina at rest. Holter monitoring demonstrated ST-segment elevation associated with angina. Repeat coronary angiography showed coronary artery spasm after the administration of methylergometrine in these patients. Coronary artery spasm occurred proximally to the previously implanted stent in 2 patients and in other coronary arteries in 1 patient.

Coronary artery spasm is sometimes observed during percutaneous coronary intervention. It is usually relieved by the intracoronary administration of nitroglycerin. Balloon inflation at a low pressure may be used to treat it. Agrawal et al calculated the minimum acceptable collapse pressure for stents using arterial strain caused by experimentally induced artery spasm.¹¹ They reported 0.4 atm as the minimum acceptable limit for collapse pressure. Almost all coronary stents have more radial strength.^{12,13} The NIR stent is one of the stents with strong radial strength.¹² An in-vitro study reported that the NIR stent expanded to 3 mm

(Received January 10, 2006; revised manuscript received March 10, 2006; accepted March 28, 2006)

Department of Cardiovascular Science and Medicine, Chiba University Graduate School of Medicine, Chiba, *Division of Cardiovascular Medicine, Saitama Medical College, Saitama, Japan

Mailing address: Yoshio Kobayashi, MD, Department of Cardiovascular Science and Medicine, Chiba University Graduate School of Medicine, 1-8-1 Inohana, Chuo-ku, Chiba 260-8670, Japan. E-mail: yoshio.kobayashi@wonder.ocn.ne.jp

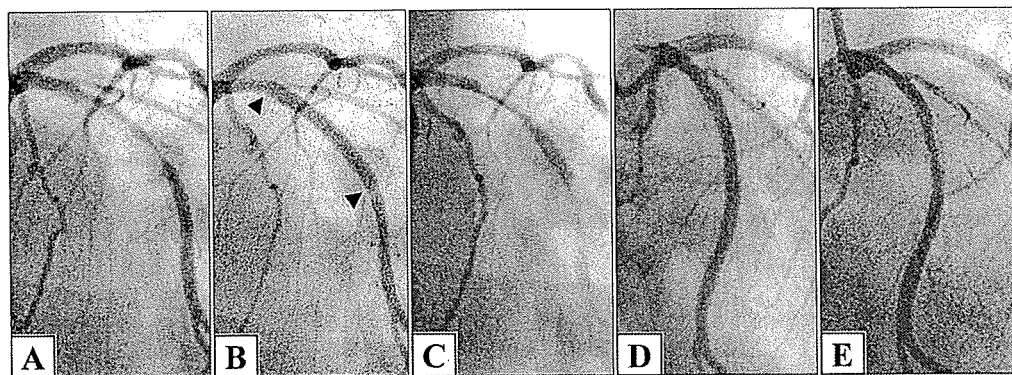


Fig 1. Coronary angiography showing a 90% stenosis in the proximal obtuse marginal artery (A). After deployment of a NIR stent, angiography demonstrates a good result (B). Arrowheads indicate the edges of the stent. Angiography demonstrates coronary artery spasm at the proximal stented segment and distal reference (C). After intracoronary administration of nitroglycerine, coronary artery spasm is relieved (D). Follow-up angiography shows a 25% stenosis at the distal stented segment (E).

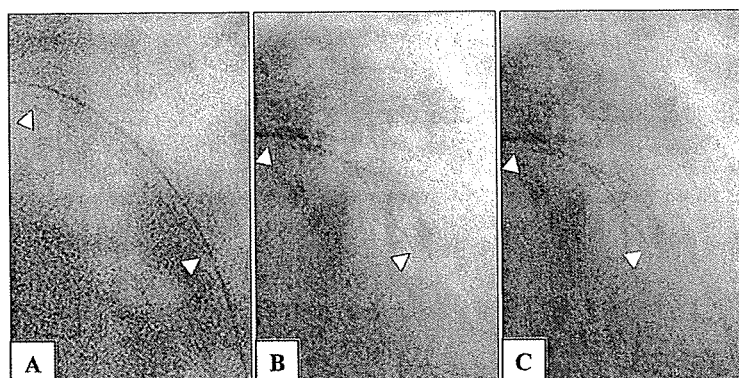


Fig 2. Fluoroscopy demonstrates a fully expanded stent (A). After coronary artery spasm, stent deformity is observed (B). Fluoroscopy shows the deformed stent at follow up (C). Arrowheads indicate the edges of the stent.

collapsed at a compressive strength of 1.05 atm.¹² This case report demonstrates unusually severe coronary artery spasm because the NIR stent was deformed. Coronary stenting may be ineffective in some patients with severe coronary artery spasm as well as in those with multivessel spasm. These are the limitations of stent implantation for coronary artery spasm. Thus, alternative medical treatment such as α -1 blocking agents,² magnesium,³ benzhexol hydrochloride,⁴ denopamine,⁵ and nicorandil⁶ should be tried for coronary artery spasm refractory to calcium antagonists and nitrates before stent implantation is considered. Stent implantation would be the last resort. In some patients, stent implantation for refractory coronary artery spasm might be performed. However, intensive medical treatment should be continued in those patients even after stent implantation.

References

1. Takagi S, Goto Y, Hirose E, Terashima M, Sakuragi S, Suzuki S, et al. Successful treatment of refractory vasospastic angina with corticosteroids: Coronary arterial hyperactivity caused by local inflammation? *Circ J* 2004; **68**: 17–22.
2. Tzivoni D, Keren A, Benhorin J, Gottlieb S, Atlas D, Stern S. Prazosin therapy for refractory variant angina. *Am Heart J* 1983; **105**: 262–266.
3. Miyagi H, Yasue H, Okumura K, Ogawa H, Goto K, Oshima S. Effect of magnesium on anginal attack induced by hyperventilation in patients with variant angina. *Circulation* 1989; **79**: 597–602.
4. Joy M, Haywood GA, Webb-Peploe MM. Management of a case of refractory variant angina with benzhexol hydrochloride (trihexyphenidyl hydrochloride). *Br Heart J* 1985; **54**: 448–451.
5. Shimizu H, Lee JD, Ogawa KB, Sugiyama T, Yamamoto M, Hara A, et al. Refractory variant angina relieved by denopamine. *Jpn Circ J* 1991; **55**: 692–694.
6. Noguchi T, Nonogi H, Yasuda S, Daikoku S, Morii I, Itoh A, et al. Refractory coronary spasm relieved by intracoronary administration of nicorandil. *Jpn Circ J* 2000; **64**: 396–398.
7. Lopez JA, Angelini P, Leachman DR, Lufschanowski R. Gianturco-Roubin stent placement for variant angina refractory to medical treatment. *Cathet Cardiovasc Diagn* 1994; **33**: 161–165.
8. Nakamura T, Furukawa K, Uchiyama H, Seo Y, Okuda S, Ebizawa T. Stent placement for recurrent vasospastic angina resistant to medical treatment. *Cathet Cardiovasc Diagn* 1997; **42**: 440–443.
9. Kultursay H, Can L, Payzin S, Turkoglu C, Altintig A, Akin M, et al. A rare indication for stenting: Persistent coronary artery spasm. *Heart Vessels* 1996; **11**: 165–168.
10. Gaspardone A, Tomai F, Versaci F, Ghini AS, Polisca P, Crea F, et al. Coronary artery stent placement in patients with variant angina refractory to medical treatment. *Am J Cardiol* 1999; **84**: 96–98.
11. Agrawal CM, Haas KF, Leopold DA, Clark HG. Evaluation of poly (L-lactic acid) as a material for intravascular polymeric stents. *Biomaterials* 1992; **13**: 176–182.
12. Schrader SC, Beyar R. Evaluation of the compressive mechanical properties of endoluminal metal stents. *Cathet Cardiovasc Diagn* 1998; **44**: 179–187.
13. Rieu R, Barragan P, Masson C, Fuseri J, Garitey V, Silvestri M, et al. Radial force of coronary stents: A comparative analysis. *Catheter Cardiovasc Interv* 1999; **46**: 380–391.