

Figure 8. Radiograms of tungsten wires coiled around PMMA rods.

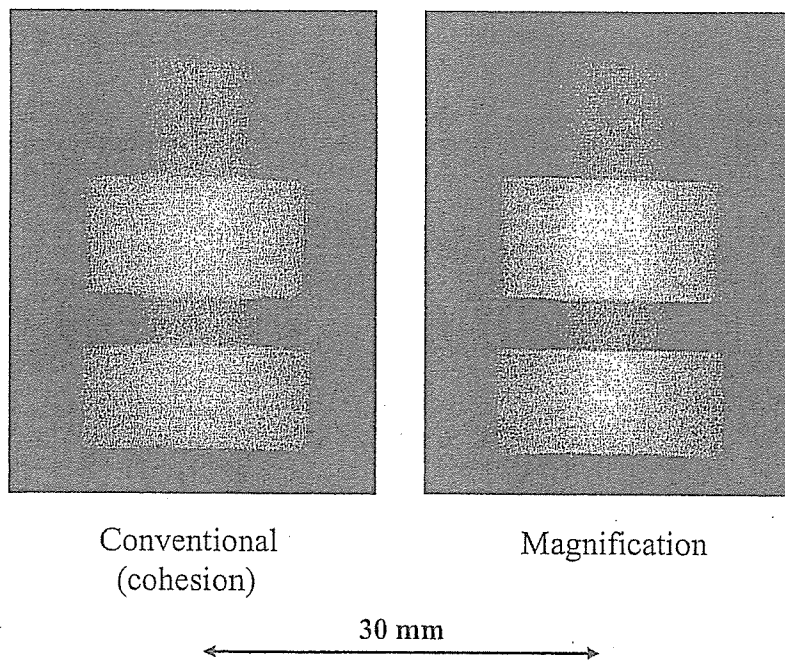


Figure 9. Radiograms of a set of a plastic bolt and a nut.

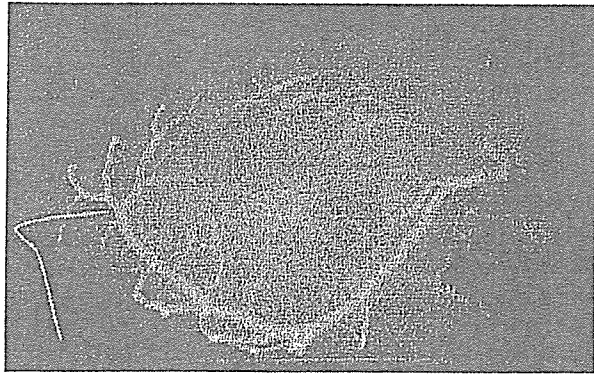
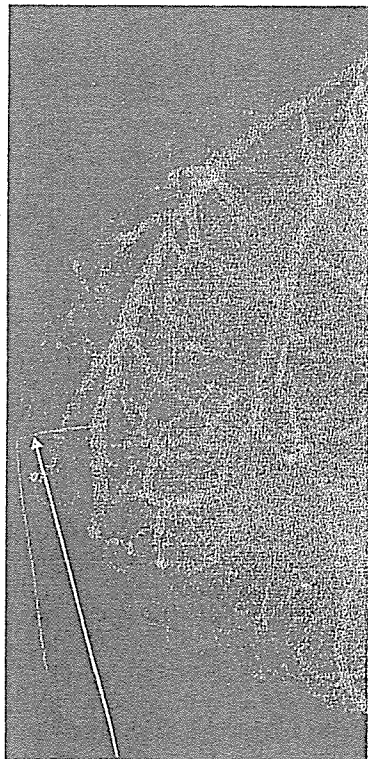
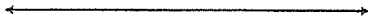


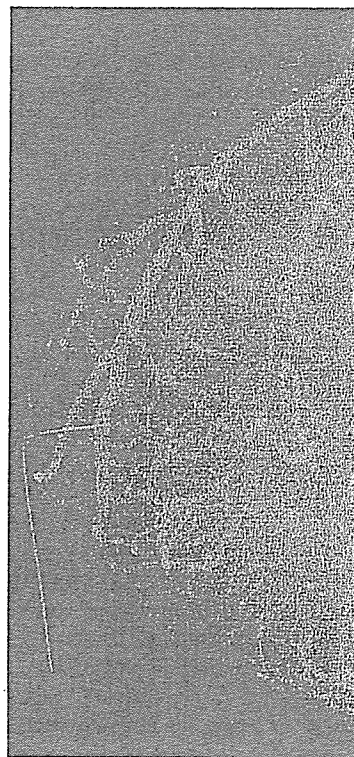
Figure 10: Angiogram of an extracted rabbit heart using iodine microspheres.

Magnification

20 mm



100 μm wire



Using a 100-mm-thick
PMMA plate

20 mm

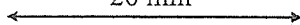


Figure 11. Angiograms of an extracted dog heart.

5. CONCLUSIONS AND OUTLOOK

In summary, we employed an x-ray generator with a 100- μm -focus tungsten tube and performed enhanced magnification angiography including phase-contrast effect using narrow-photon-energy bremsstrahlung x-rays with a peak photon energy of approximately 38 keV, which can be absorbed easily by iodine-based contrast media. The bremsstrahlung x-ray intensity substantially increased with increases in the tube voltage, and the tube voltage was determined as 60 kV in order to increase the image contrast. In enhanced angiography, although we obtained almost absorption-contrast images, phase-contrast effect may be added in cases where low-density media are employed.

Because the sampling pitch of the CR system is 87.5 μm , we obtained spatial resolutions of approximately 50 μm using 3-time magnification imaging even when a 100- μm -focus tube was employed. In order to observe fine blood vessels of less than 100 μm , the spatial resolution of the CR system should be improved to 43.8 μm (Konica Minolta Regius 190), and the iodine density should be increased.

At a tube voltage of 60 kV and a current of 0.50 mA, the maximum number of photons was approximately 4×10^7 photons/cm²·s at 1.0 m from the source, and the photon count rate can be increased easily using a rotating anode microfocus tube developed by Hitachi Medical Corporation. Recently, the maximum electric power of the microfocus x-ray tube has been increasing, and the kilowatt-range tube can be realized. Therefore, the dynamic magnification radiography is possible using a flat panel detector with a pixel size of less than 100 μm .

ACKNOWLEDGMENTS

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Conventional enhanced K-edge angiography utilizing cerium x-ray generator

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Abstract

The cerium-target x-ray tube is useful in order to perform cone-beam K-edge angiography because $K\alpha$ rays from the cerium target are absorbed effectively by iodine-based contrast media. The maximum tube voltage and current were 65 kV and 0.40 mA, respectively, and the focal-spot sizes were approximately 1×1 mm. Sharp cerium $K\alpha$ lines were left using a barium sulfate filter, and the x-ray

intensity was 16.8 $\mu\text{Gy/s}$ at 1.0 m from the source with a tube voltage of 60 kV and a current of 0.40 mA. Angiography was performed using iodine-based microspheres 15 μm in diameter. In angiography of non-living animals, we observed fine blood vessels of 100 μm or less.

1. Introduction

Synchrotrons generate monochromatic parallel x-ray beams using single crystals. These beams with photon energies of approximately 35 keV have been employed to perform enhanced K-edge angiography,¹⁻³ since the beams are absorbed effectively by iodine-based contrast media.

In order to perform high-speed medical radiography, although several different flash x-ray generators⁴⁻⁹ utilizing cold-cathode tubes have been developed, plasma flash x-ray generators¹⁰⁻¹³ are useful to produce quasi-monochromatic x rays without using a K-edge filter. Therefore, we have performed a demonstration of cone-beam K-edge angiography¹⁴ utilizing a cerium plasma generator, since K-series characteristic x rays from the cerium target are absorbed effectively by iodine. Recently, we have developed a steady-state x-ray generator utilizing a cerium-target tube, and have demonstrated enhanced K-edge angiography utilizing a barium sulfate filter.¹⁵ In this research, $K\alpha$ lines (34.6 keV) were left by absorbing $K\beta$ lines (39.2 keV).

In the present research, we describe a preliminary study on cone-beam K-edge angiography achieved with cerium $K\alpha$ rays using a barium sulfate filter.

2. Generator

Figure 1 shows the block diagram of the x-ray generator, which consists of a main controller, a cerium-target x-ray tube unit with a Cockcroft-Walton circuit and an insulation transformer, and a personal computer. The tube voltage, the current, and the exposure time can be controlled by both the controller and the computer. The main circuit for producing x rays is illustrated in Fig. 2, and employs the Cockcroft-Walton circuit in order to decrease the dimensions of the tube unit. In the x-ray tube, the negative high-voltage is applied to the cathode electrode, and the anode (target) is connected to the tube unit case (ground potential) to cool the anode and the target effectively. The filament heating current is supplied by an AC power supply in the controller in conjunction with an insulation transformer. In this experiment, the tube voltage applied was from 45 to 65 kV, and the tube current was regulated to within 0.40 mA (maximum current) by the filament temperature. The exposure time is controlled in order to obtain optimum x-ray intensity. Monochromatic $K\alpha$ lines were left using a 5-mm-thick barium sulfate filter in which barium sulfate powder was mixed with polymethyl methacrylate (PMMA) resin, since both the bremsstrahlung and the $K\beta$ rays were absorbed effectively by the filter. In designing the filter, the surface density of the barium sulfate powder is important, since the x rays are absorbed effectively by the powder as compared with the PMMA resin. In this case, the density was approximately 10 mg/cm^2 .

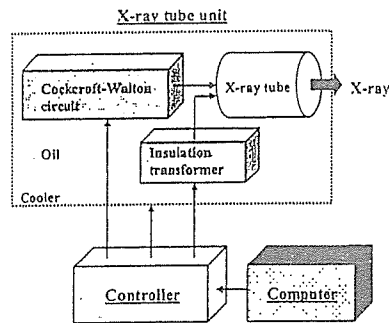


Fig. 1: Block diagram of compact x-ray generator with cerium-target tube.

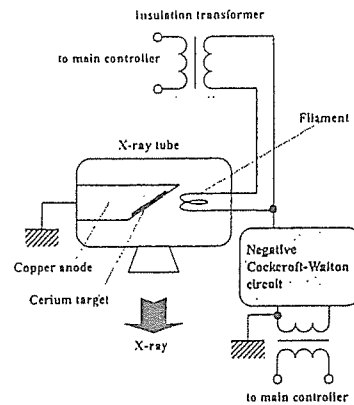


Fig. 2: Main circuit of x-ray generator.

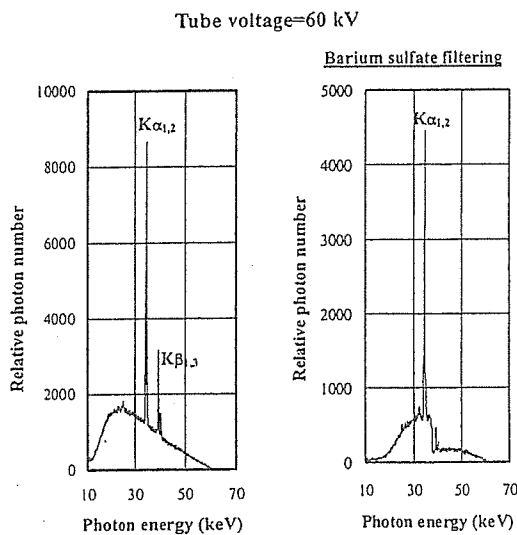


Fig. 3: X-ray spectra measured using germanium detector and filter.

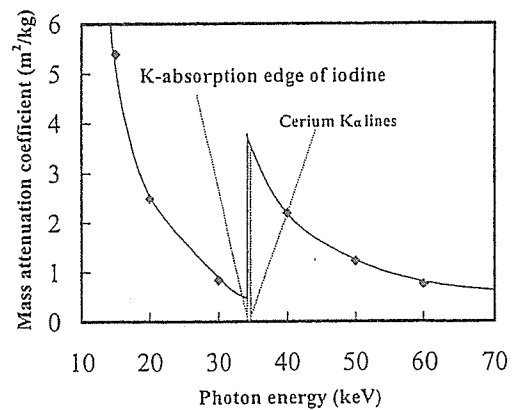


Fig. 4: Mass attenuation coefficients of iodine, and average photon energy of cerium $K\alpha$ lines.

3. Characteristics

The x-ray intensity rate was measured by a Victoreen 660 ionization chamber at 1.0 m from the x-ray source. At a constant tube current of 0.40 mA, the x-ray intensity increased when the tube voltage was increased. In this measurement, the intensity with a tube voltage of 60 kV and a current of 0.40 mA was $16.8 \mu\text{Gy/s}$ with errors of less than 0.2%.

In order to measure images of the x-ray source, we employed a pinhole camera with a hole diameter of $50 \mu\text{m}$ in conjunction with a Computed Radiography (CR) system¹⁶ with a sampling pitch of $87.5 \mu\text{m}$. When the tube voltage was increased, spot dimensions increased slightly and had values of

approximately 1×1 mm.

In order to measure x-ray spectra, we employed a germanium detector (GLP-10180/07-P, Ortec Inc.) (Fig. 3). When the tube voltage was increased, the $K\alpha$ intensity substantially increased, and both the maximum photon energy and the intensities of bremsstrahlung x rays increased.

4. Angiography

Figure 4 shows the mass attenuation coefficients of iodine at the selected energies; the coefficient curve is discontinuous at the iodine K-edge. The average photon energy of the cerium $K\alpha$ lines is shown just above the iodine K-edge. Cerium is a rare earth element and has a high reactivity; however, the average photon energies of $K\alpha$ is 34.6 keV, and iodine contrast mediums with a K-absorption edge of 33.2 keV absorb the lines easily. Therefore, blood vessels were observed with high contrasts.

The angiography was performed using the CR system (Konica Regius 150), iodine microspheres of 15 μm in diameter, and the filter. The distance (between the x-ray source and the imaging plate) was 1.5 m, and the tube voltage was 60 kV. Figure 5 shows angiograms of an extracted dog heart. Because the size of the dog heart is almost the same as human heart, human coronary arteries can be observed.

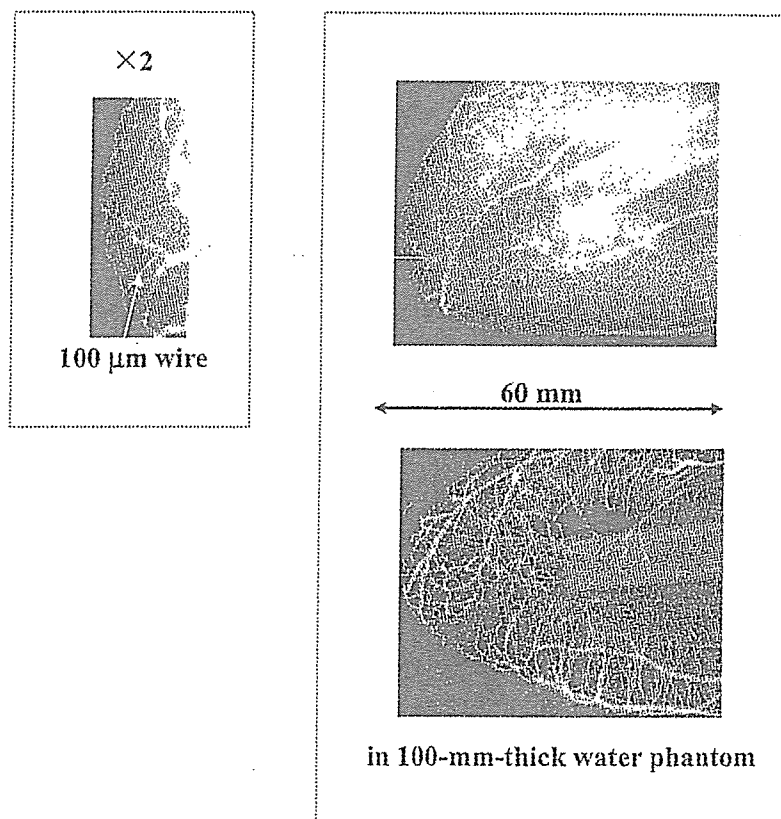


Fig. 5: Angiogram of extracted dog heart using iodine microspheres.

5. Discussion and Conclusions

In summary, we developed a new x-ray generator with a cerium-target tube and succeeded in producing cerium $K\alpha$ lines, which can be absorbed easily by iodine-based contrast media. Both the characteristic and bremsstrahlung x-ray intensities increased with increases in the tube voltage, and $K\beta$ lines were absorbed effectively by the barium sulfate filter.

In this preliminary experiment, although the maximum tube voltage and current were 65 kV and 0.40 mA, respectively, the voltage and current could be increased. Subsequently, the generator produced maximum number of $K\alpha$ photons was approximately 3×10^7 photons/cm²·s at 1.0 m from the source, and the photon count rate can be increased easily by improving the target.

Acknowledgment

This work was supported by Grants-in-Aid for Scientific Research (13470154, 13877114, and 16591222) and Advanced Medical Scientific Research from MECSST, Health and Labor Sciences Research Grants (RAMT-nano-001, RHGTEFB-genome-005 and RHGTEFB-saisei-003), Grants from Keiryō Research Foundation, The Promotion and Mutual Aid Corporation for Private Schools of Japan, Japan Science and Technology Agency (JST), and New Energy and Industrial Technology Development Organization (NEDO, Industrial Technology Research Grant Program in '03).

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X-ray spectra from a cerium target and their application to cone beam K-edge angiography

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Abstract. The cerium-target x-ray tube is useful for performing cone beam K-edge angiography, because K-series characteristic x-rays from the cerium target are absorbed effectively by iodine-based contrast media. The x-ray generator consists of a main controller and a unit with a high-voltage circuit and a fixed anode x-ray tube. The tube is a glass-enclosed diode with a cerium target and a 0.5-mm-thick beryllium window. The maximum tube voltage and current are 65 kV and 0.4 mA, respectively, and the focal-spot sizes are 1.3×0.9 mm. Cerium K-series characteristic x-rays are left, using a 3.0-mm-thick aluminum filter, and the x-ray intensity is $19.9 \mu\text{Gy/s}$ at 1.0 m from the source with a tube voltage of 60 kV and a current of 0.40 mA. Angiography is performed with a computed radiography system using iodine-based microspheres $15 \mu\text{m}$ in diameter. In angiography of nonliving animals, we observe fine blood vessels of approximately $100 \mu\text{m}$ with high contrasts. © 2005 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.2049268]

Subject terms: x-ray tube; cerium target; quasimonochromatic x-rays; characteristic x-rays; K-edge angiography.

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1 Introduction

Monochromatic parallel x-ray beams are the basis of radiography using synchrotrons in conjunction with single crystals, and these beams have been employed to perform enhanced K-edge angiography¹⁻³ and x-ray phase imaging.⁴⁻⁶ In angiography, the beams with photon energies of approximately 35 keV are absorbed effectively by iodine-based contrast media. However, it is difficult to obtain sufficient machine times for various research projects, including medical applications. Subsequently, monochromatic cone beams with energies of approximately 35 keV are useful for increasing the irradiation field for K-edge angiography.

To perform high-speed medical radiography, although several different flash x-ray generators⁷⁻¹³ utilizing cold-cathode tubes have been developed, plasma flash x-ray generators¹⁴⁻¹⁸ are useful for producing quasimonochromatic x-rays without using a K-edge filter. Therefore, we have performed a demonstration of cone beam K-edge

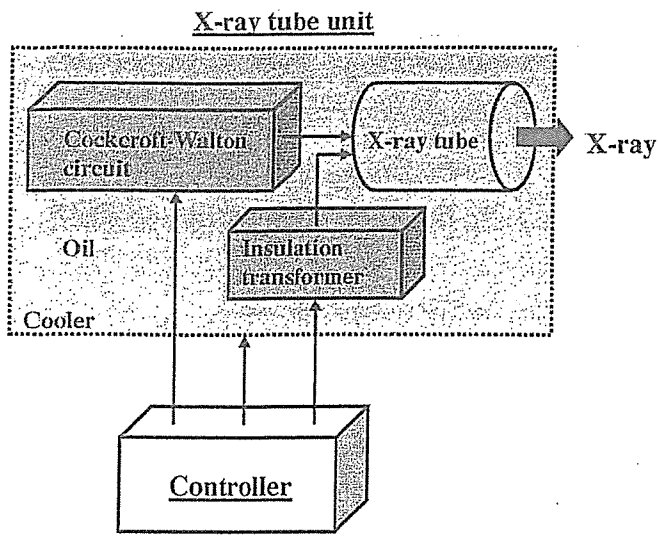


Fig. 1 Block diagram of the compact x-ray generator with a cerium-target radiation tube, which is used specially for K-edge angiography using iodine-based contrast media.

angiography¹⁹ utilizing a cerium plasma generator, since K-series characteristic x-rays from the cerium target are absorbed effectively by iodine.

Recently, we have developed a steady-state x-ray generator utilizing a cerium-target tube, and have demonstrated enhanced K-edge angiography utilizing cerium $K\alpha$ lines (34.6 keV).²⁰ In this research, $K\alpha$ lines were left by absorbing $K\beta$ lines (39.2 keV) using a barium sulfate filter with a barium K edge of 37.4 keV. However, because cerium $K\beta$ lines are also absorbed effectively by iodine, both $K\alpha$ and

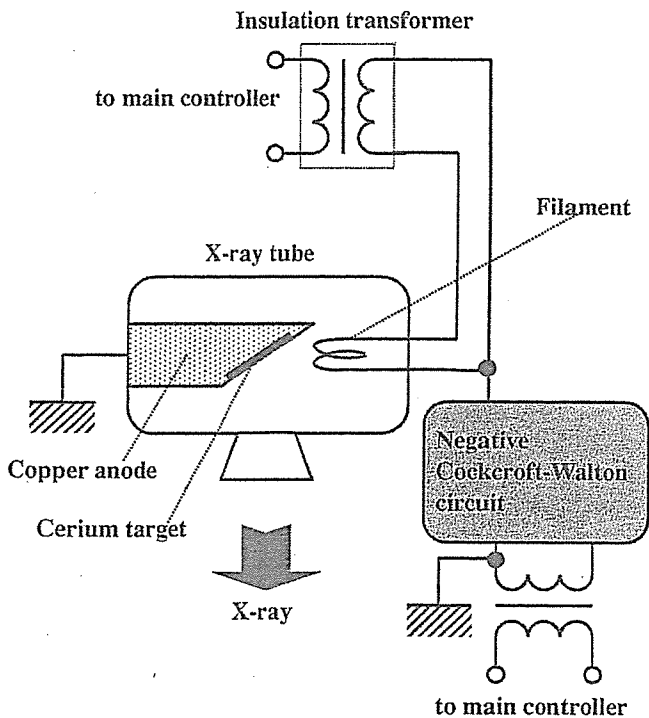


Fig. 2 Main circuit of the x-ray generator.

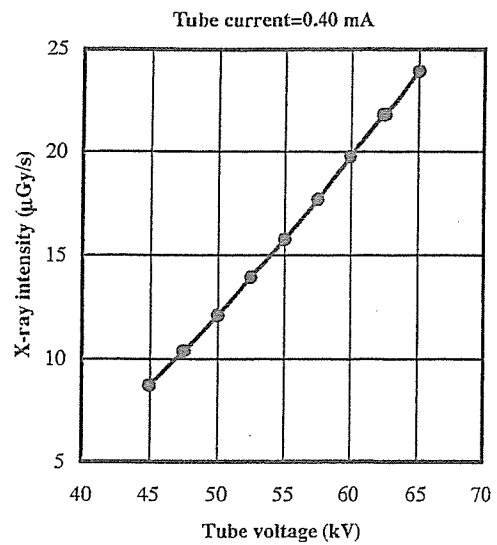


Fig. 3 The x-ray intensity ($\mu\text{Gy/s}$) as a function of tube voltage (kV) with a tube current of 0.40 mA.

$K\beta$ lines can be selected to increase the x-ray intensity for angiography. In measurements of x-ray spectra, although we usually employed a cadmium telluride detector with a photon energy resolution of 1.7 keV, the resolution should be minimized to measure the characteristic x-ray intensity.

In the present research, we measured the x-ray spectra from a cerium-target tube using a germanium detector, and performed a preliminary study on cone beam K-edge angiography achieved with cerium characteristic x-rays without using a K-edge filter.

2 Experimental Setup

Figure 1 shows the block diagram of the x-ray generator, which consists of a main controller and an x-ray tube unit with a Cockcroft-Walton circuit and a cerium-target tube. The tube voltage, the current, and the exposure time can be controlled by the controller. The main circuit for producing x-rays is illustrated in Fig. 2, and employs the Cockcroft-Walton circuit to decrease the dimensions of the tube unit. In the x-ray tube, the negative high voltage is applied to the cathode electrode, and the anode (target) is connected to the tube unit case (ground potential) to cool the anode and the target effectively. The filament heating current is supplied by an AC power supply in the controller in conjunction with an insulation transformer. In this experiment, the tube

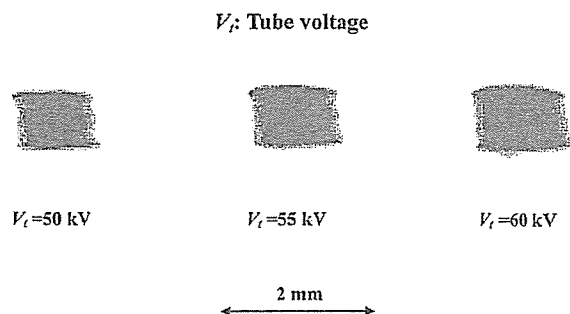


Fig. 4 Effective focal spots with changes in the tube voltage.

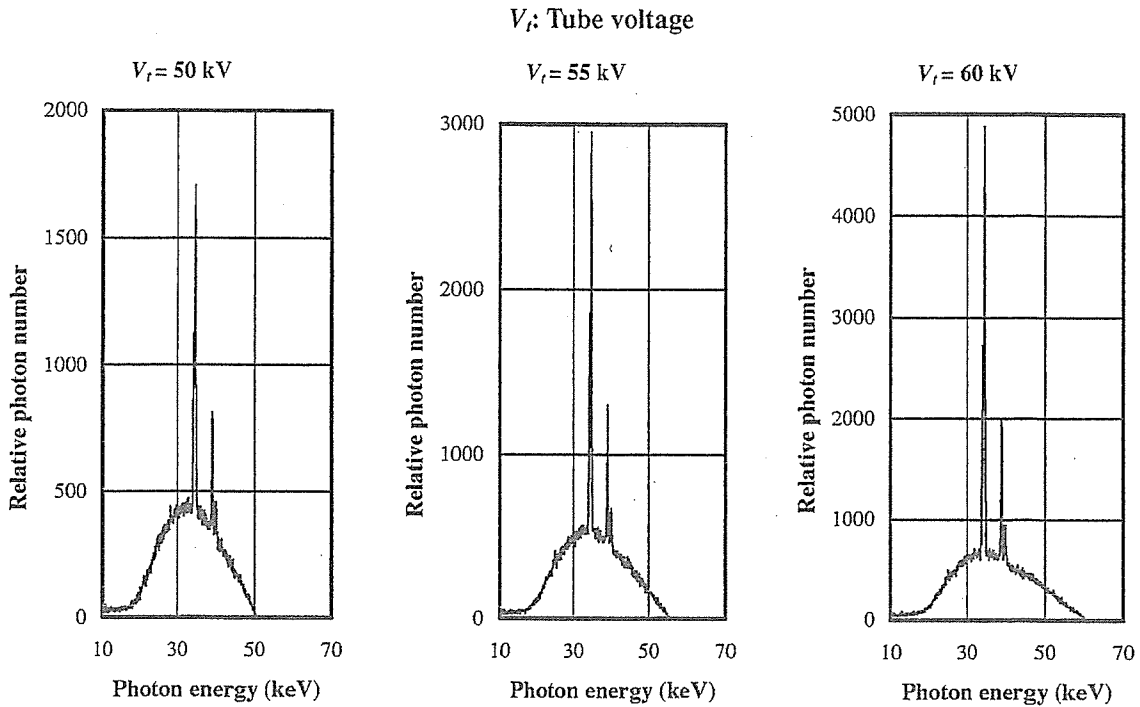


Fig. 5 X-ray spectra measured using a germanium detector with changes in the tube voltage.

voltage applied was from 45 to 65 kV, and the tube current was regulated to within 0.40 mA (maximum current) by the filament temperature. The exposure time is controlled to obtain optimum x-ray intensity. Quasimonochromatic x-rays are produced using a 3.0-mm-thick aluminum filter for absorbing soft bremsstrahlung rays.

3 Results and Discussion

3.1 X-ray Intensity

X-ray intensity was measured by a Victoreen 660 ionization chamber at 1.0 m from the x-ray source using the filter (Fig. 3). At a constant tube current of 0.40 mA, the x-ray intensity increased when the tube voltage was increased. In this measurement, the intensity with a tube voltage of 60 kV and a current of 0.40 mA was $19.9 \mu\text{Gy/s}$ at 1.0 m from the source, with errors of less than 0.2%.

3.2 Focal Spot

To measure images of the x-ray source after the aluminum filtration, we employed a pinhole camera with a hole diameter of $50 \mu\text{m}$ (magnification ratio of 1:2) in conjunction with a computed radiography (CR) system²¹ with a sampling pitch of $87.5 \mu\text{m}$. When the tube voltage was increased, spot dimensions increased slightly and had values of $1.3 \times 0.9 \text{ mm}$ (Fig. 4).

3.3 X-ray Spectra

To measure x-ray spectra, we employed a germanium detector (GLP-10180/07-P, Ortec Incorporated) with a photon energy resolution of approximately 0.12 keV (Fig. 5). When the tube voltage was increased, the characteristic x-ray intensities of $K\alpha$ and $K\beta$ lines substantially increased, and both the maximum photon energy and the intensities of bremsstrahlung x-rays increased. Because the widths of the lines were approximately 1 keV, the photon energy resolution of this detector was an optimum value. In an empirical equation, because the characteristic x-ray in-

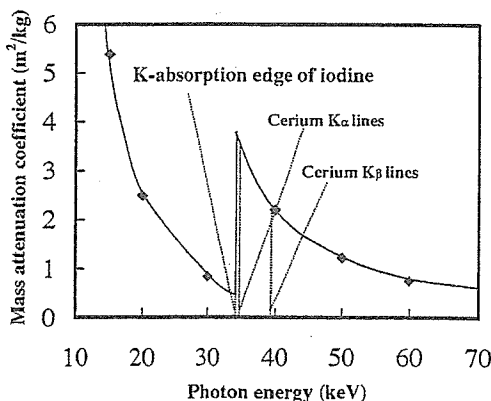


Fig. 6 Mass attenuation coefficients of iodine, and the average photon energies of cerium $K\alpha$ and $K\beta$ lines.

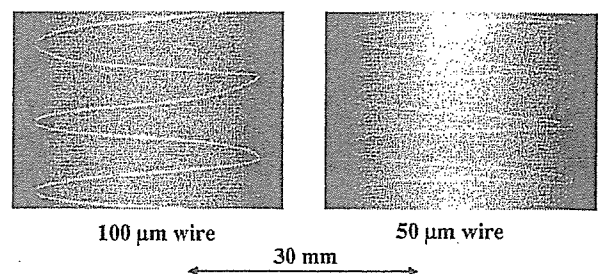


Fig. 7 Radiograms of tungsten wires coiled around PMMA rods with a tube voltage of 60 kV.

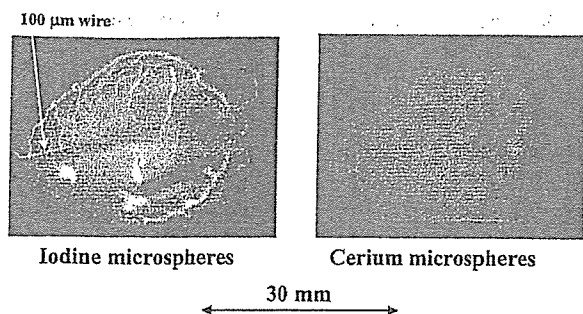


Fig. 8 Angiograms of extracted rabbit hearts using iodine and cerium microspheres with a tube voltage of 60 kV.

tensity is proportional to approximately 1.5 power of the voltage difference between the tube voltage and the critical excitation voltage, the measured intensities of the characteristic x-rays corresponded well to the equation.

3.4 K-Edge Angiography

Figure 6 shows the mass attenuation coefficients of iodine at the selected energies; the coefficient curve is discontinuous at the iodine K edge. The average photon energy of the cerium $K\alpha$ and $K\beta$ lines are shown just above the iodine K edge. The average photon energies of $K\alpha$ and $K\beta$ lines are 34.6 and 39.2 keV, respectively, and iodine contrast media with a K-absorption edge of 33.2 keV absorb the lines easily. Therefore, blood vessels were observed with high contrasts.

The angiography was performed by a CR system (Konica Minolta Regius 150) using the filter, and the distance (between the x-ray source and the imaging plate) was 1.5 m. First, rough measurements of spatial resolution were made using wires. Figure 7 shows radiograms of tungsten wires coiled around a rod made of polymethyl methacrylate. Although the image contrast decreased somewhat with decreases in the wire diameter, due to blurring of the image caused by the sampling pitch of 87.5 μm , a 50- μm -diam wire could be observed.

Angiograms of rabbit hearts are shown in Fig. 8. These two images were obtained using iodine and cerium microspheres of 15 μm in diameter at a tube voltage of 60 kV.

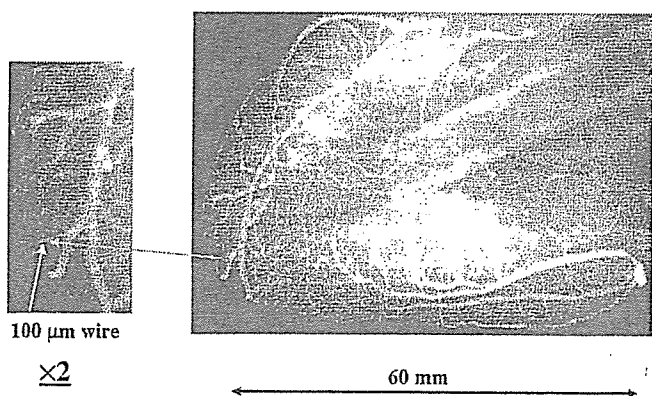


Fig. 9 Angiograms of an extracted dog heart using iodine microspheres with a tube voltage of 60 kV.

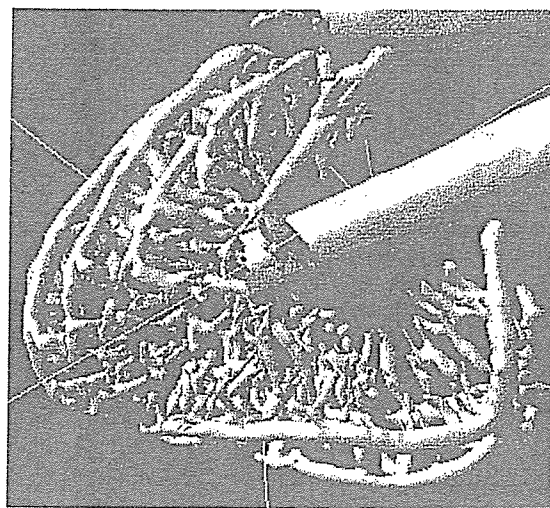


Fig. 10 3-D image of coronary arteries constructed from x-ray CT images by Pascal.

The microspheres are very useful for making phantoms of nonliving animals used for angiography. The iodine plastic spheres contained 37% iodine by weight, and the cerium plastic spheres were chemically stable and contained 18% cerium by weight. In the case where the cerium spheres were employed, the coronary arteries were barely visible, since the cerium spheres transmitted cerium characteristic x-rays easily. Figure 9 shows an angiogram of a larger dog heart at a tube voltage of 60 kV using iodine spheres. For comparison, we show a 3-D image of the coronary arteries constructed from x-ray CT images by Pascal (Digital Culture Technical Corporation) with a tungsten x-ray tube (Fig. 10). Using this imaging technique, fine blood vessels were not observed at all.

4 Conclusion and Outlook

In summary, we employ an x-ray generator with a cerium-target tube and succeed in producing cerium characteristic x-rays, which can be absorbed easily by iodine-based contrast media. The characteristic x-ray intensities increase with increases in the tube voltage, and low-photon-energy bremsstrahlung rays are absorbed effectively by the filter.

Although the cerium x-ray generator used in this research produces both the characteristic and the bremsstrahlung x-rays, bremsstrahlung intensity can be decreased effectively by considering the angle dependence without using the filter, since bremsstrahlung rays are not emitted in the opposite direction to that of electron trajectory in Sommerfeld's theory.²² Subsequently, the generator-produced maximum number of characteristic photons is approximately 3.5×10^7 photons/cm²·s at 1.0 m from the source, and the photon count rate can be increased easily by improving the target.

The x-ray intensity is limited because the thermal contact between the target and the anode is not good. However, the intensity can be increased by welding the target or using a cerium-alloy target. In addition, a rotation anode tube can be developed by the sputtering of cerium.

Compared to the 3-D blood images constructed from x-ray CT images by Pascal, fine blood vessels are visible.

Because the sampling pitch of the CR system is $87.5 \mu\text{m}$, we obtain spatial resolutions of approximately $100 \mu\text{m}$. To observe fine blood vessels of less than $100 \mu\text{m}$, the spatial resolution of the CR system should be improved to approximately $50 \mu\text{m}$ (Konica Minolta Regius 190). In addition, the spatial resolution can be improved easily to approximately $50 \mu\text{m}$ or less in cases where an x-ray film (Fuji Ix 100) is employed.

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EXOGENOUS NITRIC OXIDE CENTRALLY ENHANCES PULMONARY REACTIVITY IN THE NORMAL AND HYPERTENSIVE RAT

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SUMMARY

1. Chronic hypoxia causes sustained pulmonary hypertension and, although impairment of the pulmonary endothelial nitric oxide (NO) pathway has been implicated, no study has described the central role of NO in modulating pulmonary vascular tone and reactivity. Centrally, NO inhibits sympathetic outflow, so we hypothesised that central NO would modulate pulmonary vascular tone and its reactivity to acute hypoxia, especially in the hypertensive state.

2. Male adult Sprague-Dawley rats were exposed to normoxia (N) or chronic hypoxia (CH; 12% O₂) for 14 days. Mean pulmonary arterial pressure (MPAP), systemic mean arterial blood pressure (MABP), cardiac output and heart rate were then measured in pentobarbitone-anaesthetized, artificially ventilated rats. The N and CH rats were exposed to acute hypoxia (10% O₂ for 4 min) after the intracerebroventricular (i.c.v.) administration of artificial cerebrospinal fluid (control) and then again after either i.c.v. N^G-nitro-L-arginine methyl ester (L-NAME; 150 µg in 10 µL) or 3-morpholino-sydnonimine hydrochloride (SIN-1; 100 µg in 10 µL).

3. Chronic hypoxia caused pulmonary hypertension (MPAP 20 ± 1 vs 30 ± 1 mmHg in N and CH rats, respectively) and attenuated acute hypoxic pulmonary vasoconstriction (HPV). Central inhibition of NO synthesis (by L-NAME) did not alter baseline MPAP or the acute HPV in either N or CH rats, but it did elevate MABP. The NO donor SIN-1 did not alter baseline MPAP, but it did enhance (N rats) or restore (CH rats) the HPV and decreased MABP.

4. The results of the present study indicate that central NO has a limited role in the tonic modulation of MPAP during normoxia and after chronic hypoxia. However, the acute HPV seems to be enhanced by exogenous NO.

Key words: chronic hypoxia, nitric oxide, pulmonary vasoconstriction, sympathetic nervous system.

INTRODUCTION

Acute alveolar hypoxia causes pulmonary vasoconstriction that is reversible upon re-oxygenation. However, during chronic hypoxia, as is the case with several chronic pulmonary pathological conditions, elevated shear stress within the pulmonary vasculature causes endothelial cell injury/dysfunction,¹ smooth muscle cell proliferation² and, inevitably, the irreversible remodelling of the pulmonary vasculature.^{3,4} The consequential sustained increase in pulmonary arterial pressure (PAP) increases the workload of the heart and is therefore closely associated with heart failure and increased mortality.

Although the exact cellular mechanisms responsible for the pathogenesis of chronic hypoxia-induced pulmonary arterial hypertension are unknown, alterations in the endothelial nitric oxide (NO) pathway have been implicated as a significant contributing factor.^{5,6} Nitric oxide is a potent vasodilator and an inhibitor of vascular remodelling.^{7–9} Nitric oxide is produced from L-arginine in a reaction catalysed by isoenzymes of NO synthase (NOS). The synthesis of NO is blocked by several L-arginine analogues, including N^G-nitro-L-arginine methyl ester (L-NAME), which competitively and stereoselectively inhibit the generation of NO from L-arginine.¹⁰

A reduction in the production of NO has been implicated in the pathophysiology of pulmonary hypertension.^{6,8} Endothelial NOS (eNOS)-knockout mice have an increased risk of pulmonary hypertension,^{11–13} whereas the hypoxia-induced pulmonary hypertension is attenuated in transgenic mice that overexpress eNOS.¹⁴ In addition, chronic hypoxia limits endogenous NO synthesis,¹⁵ despite an increase in NOS expression.^{16–18}

Nitric oxide is produced not only peripherally, but also centrally within various parts of the brain, including the nucleus tractus solitarius (NTS) of the medulla oblongata and the ventrolateral medulla, the cardiovascular regulatory centres.^{19–21} Therefore, NO is considered to be involved in the neural regulation of blood pressure, independent of its direct effects on the endothelium of blood vessels.²² A review by Patel *et al.*²³ reported that the general consensus in the literature was that NO acts as a sympatho-inhibitory substance within the central nervous system. Therefore, central NO inhibition increases sympathetic outflow and, subsequently, arterial blood pressure.^{24,25}

The central role of NO in modulating pulmonary vasculature, especially in pathological conditions (e.g. pulmonary hypertension), is still incompletely understood. Yet, as evidence accumulates in support of central NO as an important regulator

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of cardiovascular function, the central role of NO in the modulation of the pulmonary vasculature needs to be addressed. Therefore, our first aim in the present study was to elucidate the role of NO in the central modulation of pulmonary vascular tone under normoxic conditions and after the development of chronic hypoxia-induced pulmonary hypertension. Nitric oxide is thought to have an important role in modulating the hypoxic pulmonary vasoconstriction (HPV) in response to acute hypoxic challenges.²⁶ However, there are conflicting reports as to whether the HPV response is accentuated or attenuated by pulmonary hypertension.^{6,27} Therefore, a second aim of the present study was to assess the acute HPV before and after chronic hypoxia and to determine whether central NO has a role in modulating this response.

METHODS

Animals

Experiments were conducted on 23 male Sprague-Dawley rats (8 weeks old; bodyweight approximately 200–280 g). Rats were divided into four groups, as follows: (i) group 1, normoxia (N) + L-NAME; (ii) group 2, chronic hypoxia (CH) + L-NAME; (iii) group 3, N + 3-morpholino-sydnominine hydrochloride (SIN-1); and (iv) group 4, CH + SIN-1. The CH rats were housed in a hypoxic chamber ($12.0 \pm 0.1\%$ O₂) continuously for 2 weeks, except for a 10 min interval each day when the chamber was cleaned. The N rats were housed in similar housing, except that N rats breathed room air. The hypoxic gas mixture was delivered to the hypoxic chamber (30 L capacity) at a flow rate of approximately 8 L/min. All rats were on a 12 h light/dark cycle at $25 \pm 1^\circ\text{C}$ and were provided with food and water *ad libitum*. All experiments were approved by the local Animal Ethics Committee and conducted in accordance with the guidelines of the Physiological Society of Japan (<http://www.soc.nii.ac.jp/psj/>).

Anaesthesia and surgery

Rats were initially anaesthetized with pentobarbitone sodium (45 mg/kg, i.p.) and supplementary doses of anaesthetic were administered periodically throughout the experimental protocol (15–30 mg/kg per h, i.p.) so as to maintain a constant surgical level of anaesthesia (assessed by using the limb withdrawal reflex test). Rectal temperature was maintained at 38°C using a rectal thermistor coupled to a thermostatically controlled heating pad.

Using a stereotaxic frame, the tip of a 27 gauge stainless steel cannula was positioned in the right lateral cerebral ventricle based on the coordinates of Paxinos and Watson²⁸ (0.8 mm posterior to the bregma, 1.5 mm lateral to the midline and 5.0 mm ventral to the skull surface). The distal end of the cannula was connected to a 0.5 mL syringe for subsequent drug administration. Correct positioning of the intracerebroventricular (i.c.v.) catheter was confirmed after each experiment by staining with Evans blue dye (10 μL).

The trachea was cannulated and the lungs ventilated with a rodent ventilator (SN-480-7; Shinano, Tokyo, Japan). The inspirate gas was enriched with O₂ (approximately 50% O₂) and the ventilator settings were adjusted (tidal volume (V_T) approximately 3.5 mL; frequency approximately 80 /min) to maintain arterial P_{CO₂} normocapnic. The femoral artery and vein were cannulated for the measurement of systemic arterial blood pressure and drug administration, respectively. The arterial line contained heparinized saline (50 U/mL). A right thoracotomy was made between the second and third ribs and the conus of the right ventricle was exposed. A 23 gauge needle was used to pierce the ventricle wall and then the gel-filled sensing catheter of a telemetric transmitter (model TA11PA-C40; Data Sciences, St Paul, MN, USA) was inserted anteriorly into the right ventricle and advanced into the pulmonary artery. The catheter was fixed in position with a 7.0 Prolene suture to the pericardium. The aorta was bluntly dissected free from the pulmonary artery and a transonic perivascular flowprobe

(model 2RB; Transonic Systems, Ithaca, NY, USA) was positioned around the ascending aorta for the continuous measurement of cardiac output.

Drugs

Artificial cerebrospinal fluid (aCSF) was used as the vehicle for administering the NOS inhibitor L-NAME and the slow-releasing NO donor, 3-[4-morpholinyl]-sydnominine-hydrochloride (SIN-1). The aCSF (pH 7.36–7.43) was comprised of 150 mmol/L NaCl, 3.0 mmol/L KCl, 0.8 mmol/L MgCl₂, 1.4 mmol/L CaCl₂ and 1.0 mmol/L Na phosphate. Intracerebroventricular injections were given as a 10 μL bolus over 30 s. Intravenous (i.v.) injections were administered as a 0.2 mL bolus over 15 s, followed by a 0.1 mL saline flush.

Measurement of right ventricular weight

Immediately following the experiment, rats were killed by anaesthetic overdose, the heart was excised, the atria were removed and the right ventricle wall was separated from the left ventricle, including the septum. Tissues were blotted and weighed. Right and left ventricular weights were expressed as the ratio of the right ventricle to the left ventricular + septum weight ($W_{RV}/W_{LV+septum}$; Fulton's ratio).

Experimental protocols

Once all variables had stabilized after surgery (approximately 20 min), baseline values were recorded for 10 min in response to the i.c.v. administration of aCSF (10 μL). The inspirate was then switched to hypoxia (10% O₂ balanced by N₂) for 4 min. Upon recovery from acute hypoxia, rats received a bolus i.c.v. injection of either L-NAME (150 μg ; groups 1 and 2) or SIN-1 (100 μg ; groups 3 and 4). After 5 (L-NAME) or 10 min (SIN-1), the acute hypoxic test was repeated. After a further 10 min, once variables had recovered to prehypoxic values, rats (groups 1 and 2 only) received a bolus i.v. injection of L-NAME (50 mg/kg) and acute hypoxia was tested 10 min later.

Data acquisition and analysis

Pulmonary arterial pressure was measured using telemetry. The signal from the implanted transmitter (model TA11PA-C20; Data Sciences) was calibrated in reference to an input from an ambient-pressure monitor (C11PR; Data Sciences) and subsequently relayed to a personal computer. Cardiac output (CO) was measured continuously from the ascending aorta using a Transonic small animal blood flowmeter (model T206; Transonic Systems) with a flowprobe (model 2RB). Arterial blood pressure (ABP) was measured continuously with a Deltran pressure transducer (Utah Medical Products, Midvale, UT, USA) and the signal was relayed to a Powerlab bridge amplifier (ML117; ADInstruments, Tokyo, Japan).

The signals for ABP, CO and PAP were relayed from their respective units and sampled continuously at 200 Hz with an eight-channel MacLab/8s interface hardware system (ADInstruments) and recorded on a Macintosh Power Book G4 using Chart (v. 5.0.1; ADInstruments). Heart rate (HR) was derived from the arterial systolic peaks. Cardiac output was normalized (off-line) to 100 g bodyweight. Total systemic vascular resistance (SVR) and total pulmonary vascular resistance (PVR) were calculated by dividing mean ABP (MABP) and mean PAP (MPAP) by the normalized CO. A 10 block of data was analysed –60, –30 s and immediately before (time '0') acute hypoxic exposure and then after 20, 40, 60, 90, 120, 180 and 240 s of exposure to 10% O₂. Normoxic data for individual rats in each group were averaged from values acquired at –60, –30 s and at time 0. An arterial blood sample (0.1 mL) was extracted 5 min after the completion of surgery for analysis of arterial P_{CO₂} using an ABL 605 blood gas analyser (Radiometer, Copenhagen, Denmark). An additional 0.1 mL was extracted to measure haematocrit.

Statistical analysis

All statistical analyses were conducted using Statview (v. 5.01; SAS Institute, Cary, NC, USA). All results are presented as the mean \pm SEM. Two-way ANOVA (repeated measures) was used to test significance for: (i) changes in each variable in response to 4 min of acute hypoxia; and (ii) whether the hypoxic response was altered by drug administration (i.c.v. or i.v.).

One-way ANOVA (factorial) was used to test for differences in baseline values for normoxic and chronic hypoxic groups of rats. Where statistical significance was reached, post hoc analyses were incorporated using the paired or unpaired *t*-test with Dunnett's correction for multiple comparisons. $P \leq 0.05$ was predetermined as the level of significance for all statistical analyses.

RESULTS

Chronic hypoxia induced pulmonary hypertension (Table 1), as evidenced by the observation that the MPAP and PVR of CH rats ($n = 11$) was 50 and 34% higher, respectively, than that of normoxic (N) rats ($n = 12$; $P < 0.01$). Consequently, CH rats developed right ventricular hypertrophy (Δ RV/LV + Sep ratio of 0.23). Chronic hypoxia did not significantly change MABP or CO, but it did cause a significant reduction in HR (Δ HR 37 b.p.m.; $P < 0.01$), suggesting that the development of right ventricular hypertrophy resulted in an increase in stroke volume. An increase in haematocrit (Δ Hct 18%; $P < 0.001$) also occurred in CH.

Responses to acute hypoxia

In N rats, acute exposure to 10% O₂ provoked an increase in MPAP, after a latency of 65–88 s, which reached a plateau after approxi-

mately 180 s (Fig. 1a). By 4 min, MPAP had increased significantly by $31 \pm 5\%$, in spite of a small, albeit not significant, decline in both HR ($6 \pm 1\%$) and CO ($16 \pm 6\%$), reflecting substantial pulmonary vasoconstriction ($67 \pm 13\%$ increase in PVR). Acute hypoxia also provoked a $56 \pm 2\%$ decrease in MABP by the end of the 4th min, which was primarily due to systemic vasodilatation ($46 \pm 3\%$ decrease in SVR; Fig. 1a). Although CH slightly altered baseline values (detailed above), the magnitude of the response to 4 min hypoxia for MABP, HR, CO and SVR was essentially the same for the CH and N groups of rats (Fig. 1a,b). However, the potent pulmonary vasoconstriction seen in N rats was abolished in CH rats. Consequently, exposure to 10% O₂ did not cause a significant increase in MPAP or PVR in CH rats.

Administration of L-NAME

The maximum cardiovascular responses following a single i.c.v. injection of L-NAME are given in Table 2. In N rats, L-NAME caused a small (<3% increase) but consistent increase in PVR ($P < 0.05$), although MPAP was not significantly affected. In addition, L-NAME caused a maximal $15 \pm 4\%$ increase in MABP after 35 ± 8 s, which was solely attributed to systemic vasoconstriction ($16 \pm 4\%$ increase in SVR). All other variables were unaltered. The PVR, MABP and SVR responses were brief, returning to pre-L-NAME values within 4.5 min. Consequently, because acute hypoxia was tested 5 min after L-NAME injection, the prehypoxia baseline data were similar before and after L-NAME treatment. Furthermore, the magnitude of the cardiovascular responses to L-NAME for CH rats was statistically similar to that of N rats (Table 2). The i.c.v. administration of L-NAME did

Table 1 Baseline cardiovascular variables in rats exposed to normoxic or chronic hypoxia

	N rats	CH rats
MPAP (mmHg)	20.2 \pm 1.1	30.4 \pm 1.2**
MABP (mmHg)	106 \pm 6	118 \pm 4
CO (mL/min/100 g)	11.17 \pm 0.75	12.32 \pm 0.53
HR (b.p.m.)	416 \pm 8	379 \pm 11*
SVR (mmHg/mL per min per 100 g)	9.73 \pm 0.64	9.72 \pm 0.53
PVR (mmHg/mL per min per 100 g)	1.88 \pm 0.14	2.52 \pm 0.15*
RV/LV + Septum	0.31 \pm 0.01	0.55 \pm 0.02**
Haematocrit (%)	47 \pm 1	65 \pm 1**

Data are the mean \pm SEM. * $P < 0.01$, ** $P < 0.001$ compared with normoxia values.

N, normoxic rats (groups 1 and 3; $n = 12$); CH, chronic-hypoxic rats (12% O₂ for 2 weeks; groups 2 and 4; $n = 11$); MPAP, mean pulmonary arterial pressure; MABP, mean arterial blood pressure; CO, cardiac output; HR, heart rate; SVR, systemic vascular resistance; PVR, pulmonary vascular resistance; RV, right ventricle; LV, left ventricle.

Table 2 Maximum responses to intracerebroventricular N^G-nitro-L-arginine methyl ester (150 μ g in 10 μ L) in normoxic rats ($n = 7$) and chronic hypoxic rats ($n = 5$)

	Pre-L-NAME	Normoxia Max L-NAME	Pre-L-NAME	Chronic hypoxia Max L-NAME
MPAP (mmHg)	18.6 \pm 0.6	19.0 \pm 0.6	30.4 \pm 1.6	31.1 \pm 1.9
MABP (mmHg)	102 \pm 10	116 \pm 8**	110 \pm 5	120 \pm 5*
CO (mL/min per 100 g)	10.12 \pm 1.10	10.09 \pm 1.13	12.24 \pm 1.07	11.98 \pm 1.00
HR (b.p.m.)	413 \pm 10	416 \pm 9	373 \pm 13	376 \pm 14
SVR (mmHg/mL per min per 100 g)	10.79 \pm 1.85	12.39 \pm 2.05**	9.14 \pm 0.55	10.19 \pm 0.66*
PVR (mmHg/mL per min per 100 g)	1.94 \pm 0.20	2.00 \pm 0.21*	2.53 \pm 0.18	2.64 \pm 0.21*

Data are the mean \pm SEM. * $P < 0.05$, ** $P < 0.01$ compared with pre-N^G-nitro-L-arginine methyl ester (L-NAME).

MPAP, mean pulmonary arterial pressure; MABP, mean arterial blood pressure; CO, cardiac output; HR, heart rate; SVR, systemic vascular resistance; PVR, pulmonary vascular resistance.

not alter the acute hypoxic response for any of the variables recorded in either N or CH rats (Fig. 1a,b).

Cardiovascular responses to the i.v. administration of L-NAME (50 mg/kg, i.v.) were also recorded. In N rats, a bolus i.v. dose of L-NAME provoked robust systemic vasoconstriction ($123 \pm 20\%$ increase in SVR; $P < 0.001$), as well as, to a lesser extent, pulmonary vasoconstriction ($80 \pm 10\%$ increase in PVR; $P < 0.01$). The baroreflex decrease in HR and CO (NS; see baseline data in Fig. 1a,b) meant that the changes in vascular resistance were accompanied by smaller, but still significant, increases in MABP ($42 \pm 18\%$ increase in MABP; $P < 0.001$) and MPAP ($13 \pm 9\%$ increase in MPAP; NS) in N rats. After 2 weeks of chronic hypoxia, L-NAME (i.v.) had a similar effect on the systemic vasculature and caused a similar depression of HR (NS) and CO ($P < 0.001$), but it had a more pronounced effect on the pulmonary vasculature ($24 \pm 5\%$ increase in MPAP (NS); $116 \pm 18\%$ increase in PVR ($P < 0.01$)).

Treatment of N rats with L-NAME (i.v.) exacerbated the reduction in MABP and SVR in response to acute hypoxia and accentuated the increase in MPAP and PVR (Fig. 1a,b). Despite a depression in baseline values for CO and HR, i.v. administration of L-NAME did not change the transient responses to hypoxia. In CH rats, the transient MABP and SVR responses to acute hypoxia were elevated by L-NAME (i.v.). In addition, L-NAME restored the potent MPAP (and PVR) response to acute hypoxia (observed in N rats), which had been absent prior to L-NAME treatment (Fig. 1a,b).

Administration of SIN-1

Following a single bolus i.c.v. injection of SIN-1 (100 μ g) in N rats, baseline MPAP and PVR were not significantly altered. However, MABP declined slowly before stabilizing $38 \pm 7\%$ below the pre-SIN-1 baseline (Table 3). Preliminary studies

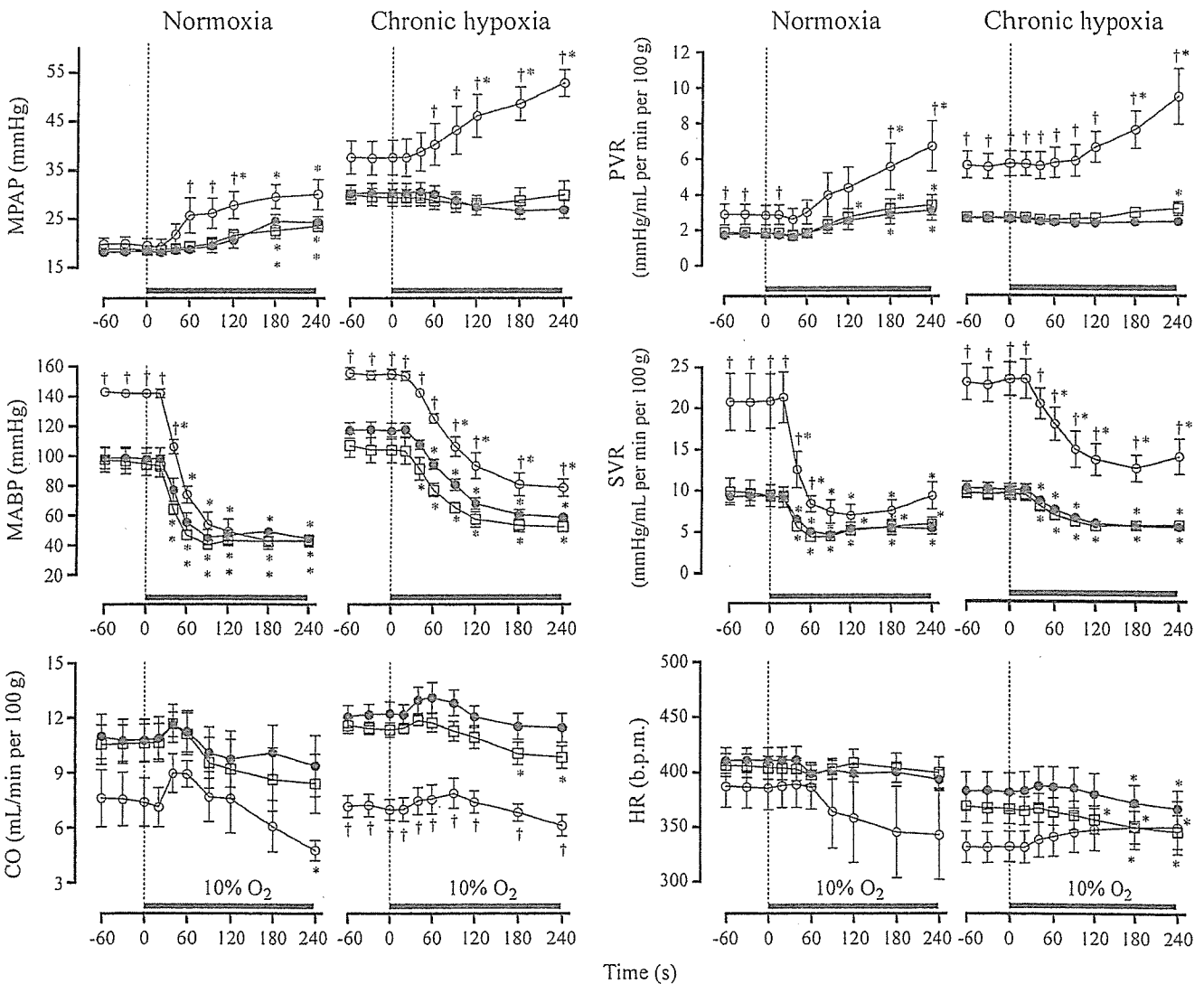


Fig. 1 Mean pulmonary arterial pressure (MPAP), mean arterial blood pressure (MABP), cardiac output (CO), pulmonary vascular resistance (PVR), systemic vascular resistance (SVR) and heart rate (HR) responses to acute hypoxia (10% O₂ for 4 min) in normoxic (N) rats ($n = 7$) and chronic hypoxic (CH) rats ($n = 5$), before (●) and after the administration of *N*^G-nitro-L-arginine methyl ester (L-NAME; □, 150 μ g in 10 μ L, i.c.v.; ◇, 50 mg/kg, i.v.). * $P < 0.05$ compared with pre-acute hypoxia values; † $P < 0.05$ compared with L-NAME values.