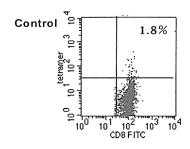
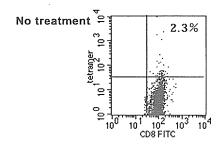
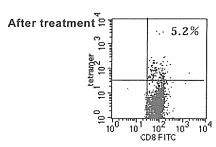
The Journal of Immunology 2329







**FIGURE 5.** Induction of CTL by peptide-liposome conjugates inoculated in combination with CpG and anti-IL-10. CD8-gated cells were analyzed with a tetramer-detecting  $OVA_{257-264}$  plus H-2K<sup>b</sup>-specific T cells. Spleen cells of normal mice (Control), tumor-bearing mice without treatment (no treatment), or tumor-bearing mice that received inoculation with peptide-liposome conjugates in combination with CpG and anti-IL-10 (after treatment) were stained with PE-conjugated, tetramer-detecting  $OVA_{257-264}$  plus H-2K<sup>b</sup>-specific T cells and FITC-conjugated anti-CD8 Ab. The experiment was repeated three times with similar results.

combined inoculation with peptide-liposome conjugates, CpG, and anti-IL-10 Abs.

We have investigated (25, 39) the potential ability of surfacelinked liposomal Ags for the application to vaccine development, whereas most of the investigations regarding liposomes as a drugdelivery system have been done by encapsulating Ags into liposomes. During the course of this investigation, several advantages of the liposome-coupled Ags over the liposome-encapsulated Ags became apparent. First, a predominant coupling efficiency of Ags to liposomes: following our previously reported procedure (20) for coupling Ags to liposomes, ~50% of the Ags bound to the surface of liposomes, whereas in the Ag encapsulation, a 60-fold higher volume of Ags was required to obtain the same amount of conjugates (our unpublished observation). Second, Ag-specific and IgEselective unresponsiveness induced by surface-linked liposomal Ags: Ags chemically coupled to the surface of liposomes induced Ag-specific IgG but not IgE Ab production in mice (19) and also in monkeys (40), suggesting the potential ability of surface-linked liposomal Ags for application to the development of vaccines with minimal allergic side effects. In addition, during the course of an investigation intended to clarify the mechanism of IgE-selective unresponsiveness induced by surface-linked liposomal Ag, we found the existence of an alternative mechanism, not involving T

cells, in the regulation of IgE synthesis (41). Third, an enhanced recognition of liposomal Ags by APCs: because liposomes basically consist of immunologically inert fatty acid, they are hardly recognized by APCs. Therefore, some contrivance, such as the introduction of mannose on the surface of liposomes (42), is required in Ag-encapsulated liposomes to enhance the recognition of liposomes by APCs. In contrast, in surface-linked liposomal Ags, Ags expressed on the surface of liposomes might be recognized more efficiently by APCs, which might result in an enhanced presentation to T cells. In fact, surface-linked liposomal Ags induced a significantly higher level of Ag-specific IgG production than that by liposome-encapsulated Ags in mice (our unpublished observation). In addition, a significant difference, which correlated closely with the adjuvant activity of liposomes, was observed in the recognition of liposomal Ags by APCs between liposomes with different lipid components; more Ags coupled to the unsaturated liposomes were engulfed by macrophages in vitro and a higher level of Ag-specific Ab production was induced in vivo than when saturated liposomes were used, suggesting that the adjuvant effects of liposomes are exerted at the beginning of the immune response, i.e., recognition of Ag by APCs (43). In addition to this quantitative difference between liposomes with differential lipid components, in the present study, a qualitative difference (i.e., the differential ability to induce cross-presentation) was observed between saturated and unsaturated liposomes. Although the precise mechanism underlying this difference is currently unclear, the significant difference in membrane mobility observed between these liposomes (23) might affect their ability to induce crosspresentation.

Because a detailed characterization of many tumor cell surface molecules that act as TAAs is now available (44), immunotherapy has become an increasingly essential component of cancer therapies (7). Emphasis to date has been placed on the development of cancer vaccines to enhance the immunogenicity of weak TAAs. In this context, surface-linked liposomal Ag might potentially serve as a candidate protocol for tumor vaccine preparation to present tumor Ags to APCs and induce effective antitumor responses.

## Disclosures

The authors have no financial conflict of interest.

## References

- Hanson, H. L., D. L. Donermeyer, H. Ikeda, J. M. White, V. Shankaran, L. J. Old, H. Shiku, R. D. Schreiber, and P. M. Allen. 2000. Eradication of established tumors by CD8<sup>+</sup> T cell adoptive immunotherapy. *Immunity* 13: 265–276.
- Shankaran, V., H. Ikeda, A. T. Bruce, J. M., White, P. E. Swanson, J. J. Old, and R. D. Schreiber. 2001. IFN-γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* 410: 1107–1111.
- Zhan, L., J. R. Conejo-Garcia, D. Katsaros, P. A. Gimotty, M. Massobrio, G. Regnani, A. Makrigiannakis, H. Gray, K. Schlienger, M. N. Liebman, S. C. Rubin, and G. Coukos. 2003. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. N. Engl. J. Med. 348: 203-213.
- Dunn, G. P., L. J. Old, and R. D. Schreiber. 2004. The three Es of cancer immunoediting. Annu. Rev. Immunol. 22: 329–360.
- Sznol, M., and D. R. Parkinson. 1993. Clinical applications of IL-2. Oncology 8: 61–67.
- Hu, J., W. Kindsvogel, S. Busby, M. C. Bailey, Y. Shi, and P. D. Greenberg. 1993. An evaluation of the potential to use tumor-associated antigens as targets or antitumor T cell therapy using transgenic mice expressing a retroviral tumor antigen in normal lymphoid tissues. J. Exp. Med. 177: 1681–1690.
- Blattman, J. N., and P. D. Greenberg. 2004. Cancer immunotherapy: a treatment for the masses. Science 305: 200–205.
- Heath, W. R., and F. R. Carbone. 2001. Cross-presentation, dendritic cells, tolerance and immunity. Annu. Rev. Immunol. 19: 47-64.
- Mark, A., B. Bjorksten, and M. Granstrom. 1995. Immunoglobulin E responses to diphtheria and tetanus toxoids after booster with aluminum-adsorbed and fluid DT-vaccines. Vaccine 13: 669-673.
- Aggerbeck, H., J. Wantzin, and I. Heon. 1996. Booster vaccination against diphtheria and tetanus in man: comparison of three different vaccine formulations— III. Vaccine 14: 1265-1272.

- Nothdurft, H. D., T. Jelinek, A. Marschang, H. Maiwald, A. Kapaun, and T. Loscher. 1996. Adverse reactions to Japanese encephalitis vaccine in travelers. J. Infect. 32: 119–122.
- Gregoriadis, G. 1990. Immunological adjuvants: a role for liposomes. *Immunol. Today* 11: 89–97.
- Wassef, N. M., C. R. Alving, and R. L. Richards. 1994. Liposomes as carriers for vaccines. *Immunomethods* 4: 217–222.
   Alving, C. R., V. Koulchin, G. M. Glenn, and M. Rao. 1995. Liposomes as
- Alving, C. R., V. Koulchin, G. M. Glenn, and M. Rao. 1995. Liposomes as carriers of peptide antigens: induction of antibodies and cytotoxic T lymphocytes to conjugated and unconjugated peptides. *Immunol. Rev.* 145: 5-31.
- Green, S., J. D. Fortier, J. Madsen, G. Swartz, L. Einck, E. Gubish, and C. Nancy. 1995. Liposomal vaccines. In *Immunology of Proteins and Peptides VIII*. M. Z. Atassi and G. S. Bixler., eds. Plenum, New York, pp. 83–92.
- 16. De Haan, A., J. F. C. Tomee, J. P. Huchshorn, and J. Wilschut. 1995. Liposomes as an immunoadjuvant system for stimulation of mucosal and systemic antibody responses against inactivated measles virus administered intranasally to mice. Vaccine 13: 1320-1324.
- Gluck, R. 1995. Liposomal presentation of antigens for human vaccines. Pharm. Biotechnol. 6: 325–345.
- Perrie, Y., P. M. Frederik, and G. Gregoriadis. 2001. Liposome-mediated DNA vaccination: the effect of vesicle composition. *Vaccine* 19: 3301–3310.
   Naito, S., A. Horino, M. Nakayama, Y. Nakano, T. Nagai, M. Mizuguchi,
- Naito, S., A. Horino, M. Nakayama, Y. Nakano, T. Nagai, M. Mizuguchi, K. Komuro, and T. Uchida. 1996. Ovalbumin-liposome conjugate induces IgG but not IgE antibody production. *Int. Arch. Allergy Immunol.* 109: 223–228.
- Nakano, Y., M. Mori, S. Nishinohara, Y. Takita, S. Naito, A. Horino, H. Kato, M. Taneichi, K. Komuro, and T. Uchida. 1999. Antigen-specific, IgE-selective unresponsiveness induced by antigen-liposome conjugates: comparison of four different conjugation methods for the coupling of antigen to liposome. *Int. Arch. Allergy Immunol.* 120: 199–208.
- Naito, S., M. Taneichi, H. Kato, Y. Ami, Y. Suzaki, M. Mori, Y. Nakano, H. Yamamura, K. Morokuma, K. Ohkuma, et al. 2002. Selective inhibition of systemic anti-OVA IgE production in response to oral preadministration with OVA-liposome conjugates. *Int. Arch. Allergy Immunol.* 129: 314-319.
   Yoshikawa, T., T. Uchida, S. Naito, K. Komuro, J. Chiba, T. Kurata, and
- Yoshikawa, T., T. Uchida, S. Naito, K. Komuro, J. Chiba, T. Kurata, and S. Tamura. 2000. Suppression of IgE antibody response to ovalbumin by intranasal administration of liposome-conjugated ovalbumin. *Int. Arch. Allergy Im*munol. 121: 108-115.
- Nakano, Y., M. Mori, S. Nishinohara, Y. Takita, S. Naito, H. Kato, M. Taneichi, K. Komuro, and T. Uchida. 2001. Surface-linked liposomal antigen induces IgEselective unresponsiveness regardless of the lipid components of liposomes. *Bioconjug. Chem.* 12: 391–395.
- Chang, E. Y., Y. Zheng, D. Holowka, and B. Baird. 1995. Alteration of lipid composition modulates FCsRI signaling in RBL-2H3 cells. *Biochemistry* 34: 4376-4384.
- Phillips, N. C., L. Gagne, N. Ivanoff, and G. Riveau. 1996. Influence of phospholipid composition on antibody responses to liposome encapsulated protein and peptide antigens. Vaccine 14: 898-904.
- Hincha, D. K., A. E. Oliver, and J. H. Crowe. 1999. Lipid composition determines the effect of arbutin on the stability of membranes. *Biophys. J.* 77: 2024-2034.
- Moody, D. B., V. Briken, T. Y. Cheng, C Roura-Mir, M. R. Guy, D. H. Geho, M. L. Tykocinski, G. S. Besra, and S. A. Porcelli. 2002. Lipid length controls

- antigen entry into endosomal and nonendosomal pathways for CD1b presentation. Nat. Immunol. 3: 435-442.
- Uchida, T., S. T. Ju, A. Fay, Y. N. Liu, and M. E. Dorf. 1985. Functional analysis
  of macrophage hybridomas: I. Production and initial characterization. J. Immunol. 134: 772-778
- 29. Hermel, E., J. Yuan, and J. J. Monaco. 1995. Characterization of polymorphism within the H2-M MHC class II loci. *Immunogenetics* 42: 136–142.
- Ishida, H., R. Hastings, J. Kearney, and M. Howard. 1992. Continuous antiinterleukin-10 antibody administration depletes mice of LY-1 B cells but not conventional B cells. J. Exp. Med. 175: 1213–1220.
- Rock, K. L. 1996. A new foreign policy: MHC class I molecules monitor the outside world. *Immunol. Today* 17: 131–137.
- Carbone, F. R., and M. J. Bevan. 1990. Class I-restricted processing and presentation of exogeneous cell-associated antigen in vivo. J. Exp. Med. 171: 377–387.
- Liu, T., X. Zhou, C. Orvell, E. Leferer, H. G. Ljunggren, and M. Jondal. 1995. Heat-inactivated Sendai virus can enter multiple MHC class I processing pathways and generate cytotoxic T lymphocyte responses in vivo. *J. Immunol.* 154: 3147–3155.
- Zinkernagel, R. M. 2002. On cross-priming of MHC class I-specific CTL: rule or exception? Eur. J. Immunol. 232: 2385–2392.
- Houde, M., S. Berholet, E. Gagnon, S. Brunet, G. Goyette, A. Laplante, M. F. Princlotta, P. Thibault, D. Sacks, and M. Desjardins. 2003. Phagosomes are competent organelles for antigen cross-presentation. *Nature* 425: 402-406.
- Ackerman, A. L., and P. Cresswell. 2004. Cellular mechanisms governing crosspresentation of exogenous antigens. Nat. Immunol. 5: 678-684.
- Dermine, S., D. E. Gilham, D. M. Shaw, E. J. Davidson, E. K. Meziane, A. Armstrong, R. E. Hawkins, and P. Stern. 2004. Vaccine and antibody-directed T cell tumor immunotherapy. *Biochem. Biophys. Acta* 1704: 11–35.
- Vicari, A. P., C. Chiodoni, C. Vaure, S. Ait-Yahia, C. Dercamp, F. Matsos, O. Reynard, C. Taverne, P. Merle, M. P. Colombo, et al. 2002. Reversal of tumor-induced dendritic cell paralysis by CpG immunostimulatory oligonucleotide and anti-interleukin 10 receptor antibody. J. Exp. Med. 196: 541-549.
- Gregoriadis, G. 1994. Liposomes as immunoadjuvants and vaccine carriers: antigen entrapment. *Immunomethods* 4: 210–216.
- Suzaki, Y., Y. Ami, N. Nagata, S. Naito, H. Kato, M. Taneichi, M. Takahashi, T. Komiya, S. Satoh, F. Gondaira, et al. 2002. Protection of monkeys against Shiga toxin induced by Shiga toxin-liposome conjugates. *Int. Arch. Allergy Im*munol. 127: 294-298.
- Taneichi, M., S. Naito, H. Kato, K. Komuro, and T. Uchida. 2002. T cell-independent regulation of IgE antibody production induced by surface-linked liposomal antigen. J. Immunol. 169: 4246–4252.
- Garcon, N., G. Gregoriadis, M. Taylor, and J. Summerfield. 1988. Mannosemediated targeted immunoadjuvant action of liposomes. *Immunology* 64: 743-745.
- Tanaka, Y., M. Kasai, M. Taneichi, S. Naito, H. Kato, M. Mori, M. Nishida, N. Maekawa, H. Yamamura, K. Komuro, and T. Uchida. 2004. Liposomes with differential lipid components exert differential adjuvanticity in antigen-liposome conjugates via differential recognition by macrophages. *Bioconjug. Chem.* 15: 35-40.
- Forni, G., P. L. Lollini, P. Musiani, and M. P. Colombo. 2000. Immunoprevention of cancer: is the time ripe? Cancer Res. 60: 2571–2575.