

## Viremia Persists after Suppression by Antiretroviral Therapy

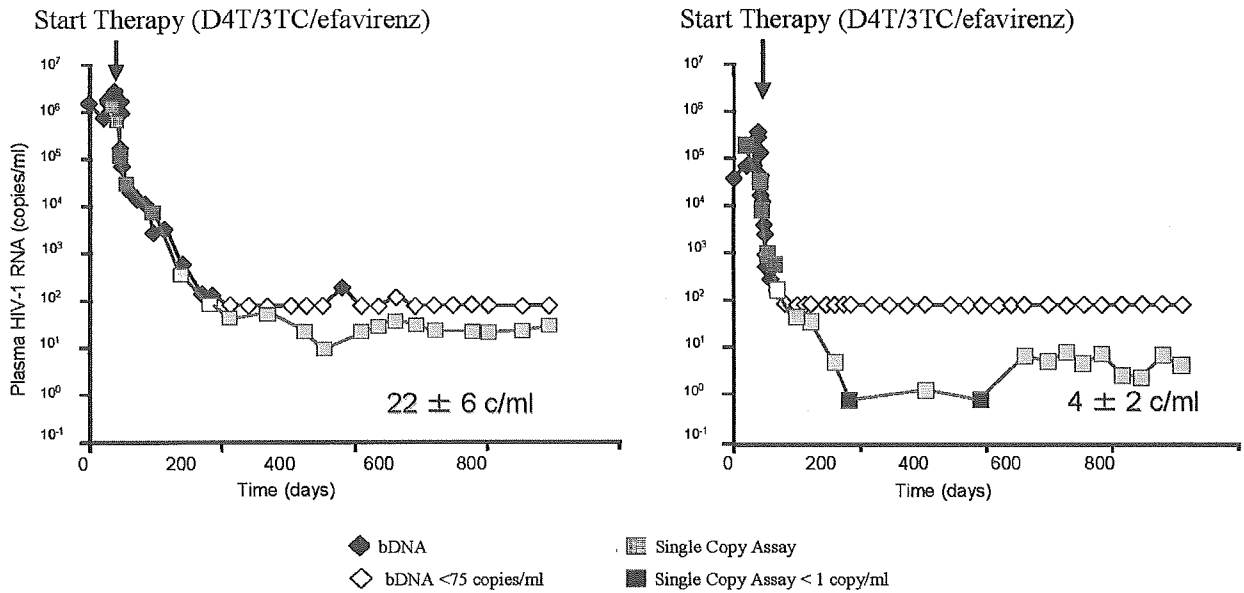


图 7

3. Both theory and experience imply that the virus in individuals who have been infected with HIV for more than a short time should have a low level of preexisting drug resistance mutations. We have therefore used allele-specific PCR to analyze levels of K103N in infected, untreated individuals. The background in the assay was about 0.02%, and the large majority of patient samples gave results were very close to this value, implying that the assay is not yet sufficiently sensitive to detect the true values in most patients. A few patients had values significantly larger than background, however, suggesting the possibility of stochastic fluctuations in frequency — such fluctuations — although rare — could have significant implications for the success of subsequent therapy.

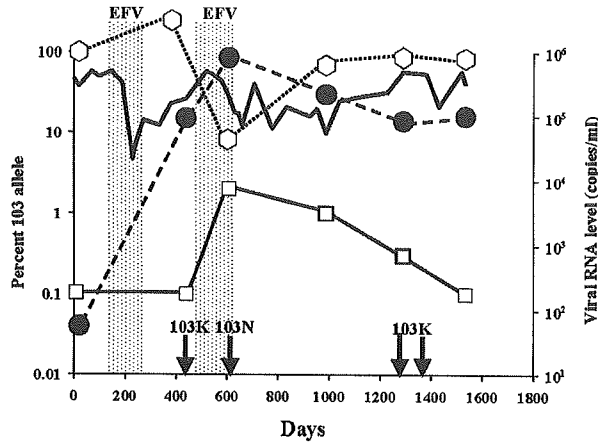
4. In individuals who have failed complex combinations of therapy, including the NNRTI efavirenz (EFV), the K103 N resistance mutation shows a wide variety of behaviors, including persistence at a level of nearly 100% for 5 years after the end of EFV treatment ; rapid reduction to about 10% of the virus population and persistence at that level, and a complete switch in the relevant codon in the virus from AAC to AAT and back again (图 9). Persistence is not due to linkage to other resistance mutations, but the codon switching is the result of linkage to the M184V mutation selected by

treatment with 3TC during part of the EFV therapy.

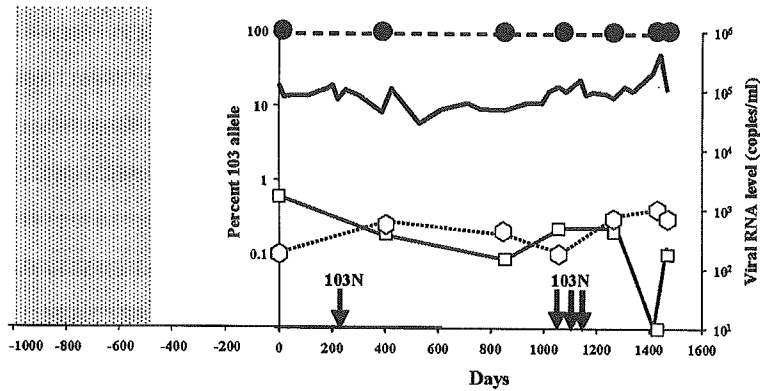
5. In patients who have failed combination antiretroviral therapy, it is standard to test for potentially active drugs by bulk sequence analysis of RT-PCR products derived from plasma virus. To test the ability of standard analysis to detect important resistance mutations, we compared bulk sequences with those of SGS products from plasma virus obtained from highly drug-experienced patients. SGS revealed the presence of resistance mutations that were not detected by standard bulk genotype analysis in 3–20% of genomes analyzed. In some cases, furthermore, the undetected mutations are linked on the same genomes. Since even minor populations of resistant virus are likely to cause rapid failure, bulk sequencing approaches, while useful for predicting resistance to specific drugs, are unlikely to be reliable in predicting sensitivity to them.

Conclusions. The tests we have developed are bringing new insights to the analysis of HIV in infected patients. We have uncovered a new therapeutic steady state viremia in most or all patients that explains our inability to cure the infection. We have found that the genetically highly diverse population of HIV in long-term infected patients is very stable in its diversity. We have learned that standard sequencing ap-

Patient 1 : Persistent K103N mutation



Patient 3 : Persistent K103N mutation:



Patient 5 : 103N codon switch

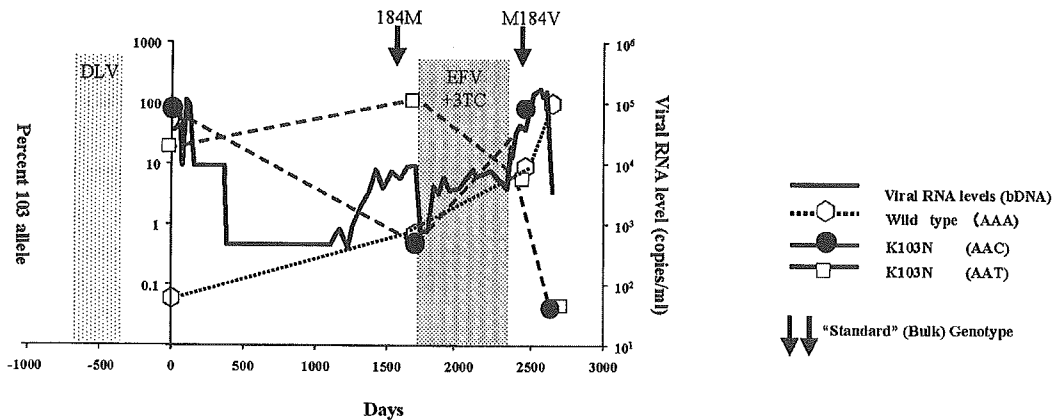


图 9

proaches miss a large fraction of resistance mutations in highly experienced patients, and we have uncovered a puzzling diversity of patterns of loss of drug resistant mutations

long after treatment has ceased. We expect further studies of these phenomena to be equally rewarding in their ability to uncover new aspects of the host-virus relationship.

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# Both Regulatory T Cells and Antitumor Effector T Cells Are Primed in the Same Draining Lymph Nodes during Tumor Progression<sup>1</sup>

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The peripheral tolerance mechanism prevents effective antitumor immunity, even though tumor cells possess recognizable tumor-associated Ags. Recently, it has been elucidated that regulatory T cells (Treg) play a critical role in maintaining not only self-tolerance, but also tolerance of tumor cells. However, because the Treg that maintain self-tolerance arise naturally in the thymus and are thought to be anergic in peripheral, it is still unclear where and when Treg for tumor cells are generated. In this study we analyze tumor-draining lymph nodes (LNs) and demonstrate that both antitumor effector T cells and Treg capable of abrogating the antitumor reactivity of the effector T cells are primed in the same LNs during tumor progression. The regulatory activity generated in tumor-draining LNs exclusively belonged to the CD4<sup>+</sup> T cell subpopulation that expresses both CD25 and a high level of CD62L. Forkhead/winged helix transcription factor gene expression was detected only in the CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells. CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg and CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells, which possess effector T cell functions, had comparable expression of LFA-1, VLA-4, CTLA-4, lymphocyte activation gene-3, and glucocorticoid-induced TNFR. Thus, only CD62L expression could distinguish regulatory CD4<sup>+</sup>CD25<sup>+</sup> cells from effector CD4<sup>+</sup>CD25<sup>+</sup> cells in draining LNs as a surface marker. The Treg generated in tumor-draining LNs possess the same functional properties as the Treg that arise naturally in the thymus but recognize tumor-associated Ag. CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg contained a subpopulation that expressed CD86. Blocking experiments revealed that ligation of CTLA-4 on effector T cells by CD86 on Treg plays a pivotal role in regulating CD4<sup>+</sup> effector T cells. *The Journal of Immunology*, 2005, 175: 5058–5066.

The purpose of the immune system is to discriminate and eliminate invading nonself. To do this, it possesses not only an effector system to eliminate nonself, but also a regulatory system that abrogates the attack of effector cells against self-somatic cells. Thus, the balance between immunity and tolerance determines the outcome of an immune reaction. Although tumor cells have Ags altered by mutation, a lack of danger signals and antigenic similarity to self-somatic cells, from which tumor cells are derived, engage the peripheral tolerance mechanism (1, 2). This tolerance makes it difficult to obtain effective antitumor immunity.

Recent studies revealed CD4<sup>+</sup> T cells that constitutively express CD25 to play a critical role in maintaining peripheral tolerance during infection, transplantation, autoimmunity, and tumor immu-

nity (2–7). CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells (Treg)<sup>4</sup> that arise naturally in the thymus to maintain self-tolerance are considered anergic in peripheral (8). However, it has been demonstrated that Treg proliferate in peripheral tissues in response to antigenic stimulation and can be converted from naive CD4<sup>+</sup> T cells (9, 10). Thus, it is still unclear where and when Treg are generated for tumor cells.

APCs that acquire Ags migrate into secondary lymphoid organs, where the Ag information is converted to adaptive immune responses. Although CD4<sup>+</sup>CD25<sup>+</sup> is the best surrogate marker, it is difficult to distinguish Treg based on CD25 expression, especially in lymph nodes (LNs) where T cell priming is going on, because the expression of CD25 is also up-regulated on effector T cells upon TCR engagement before clonal expansion. This problem makes it impossible to analyze how regulatory T cells are primed in LNs. We reported that effector T cells, which are capable of mediating antitumor reactivity, are primed in LNs draining growing tumors and that these T cells exclusively belong to the CD62L<sup>low</sup> subpopulation (11). Additional CD40 stimulation as help signals for APC resulted in increased numbers of CD62L<sup>low</sup> T cells in draining LNs (12, 13). CD62L<sup>high</sup> T cells have been considered naive cells that have never been encountered by cognate Ag. However, our findings suggested that CD62L<sup>high</sup> tumor-draining LN T cells contained a regulatory subpopulation, because the elimination of CD62L<sup>high</sup> cells promotes the generation of highly potent antitumor T cells upon stimulation with CD3 (14).

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<sup>4</sup> Abbreviations used in this paper: Treg, regulatory T cell; CM, complete medium; DC, dendritic cell; Foxp3, forkhead/winged helix transcription factor gene; GITR, glucocorticoid-induced TNFR; LAG3, lymphocyte activation gene-3; LN, lymph node; m, murine.

Recent studies have demonstrated that CD62L<sup>high</sup> CD4<sup>+</sup>CD25<sup>+</sup> T cells possess superior suppressive activity (15–18).

In this study we demonstrate that the expression of CD62L distinguishes regulatory CD4<sup>+</sup>CD25<sup>+</sup> cells from effector CD4<sup>+</sup>CD25<sup>+</sup> T cells and that both antitumor effector T cells and regulatory T cells, which are capable of abrogating the therapeutic efficacy of the antitumor effector T cells *in vivo*, are primed in the same tumor-draining LNs with different kinetics. CTLA-4 ligation by CD86 exclusively expressed on regulatory CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> LN T cells plays a pivotal role in regulating effector CD4<sup>+</sup> T cell functions via direct T-T interaction.

## Materials and Methods

### Mice

Female C57BL/6J (B6) mice were purchased from CLEA Laboratory. They were maintained in a specific pathogen-free environment and used for experiments at the age of 8–10 wk. All animal experiments were conducted with the permission of the Niigata University ethics committee for animal experiments.

### Tumors

MCA 205 is a fibrosarcoma of B6 origin induced by *i.m.* injection of 3-methylcholanthrene (19). Single-cell suspensions were prepared from solid tumors by enzymatic digestion as described previously (20). An MCA 205 tumor cell line was established and maintained *in vitro*.

### mAbs and flow cytometry

Hybridomas producing mAbs against murine CD4 (GK1.5, L3T4), CD8 (2.43, Lyt-2), CD3 (2C11), and murine CD62L (MEL14) were obtained from American Type Culture Collection. Anti-CD4 mAb, anti-CD8 mAb, and anti-CD62L mAb were produced as ascites fluid from sublethally irradiated (500 cGy) DBA/2 mice. PE-conjugated anti-CD80 (16-10A), anti-CD86 (GL1), anti-CD62L (MEL14), anti-CTLA-4 (UC10-4F10-11), anti-lymphocyte activation gene-3 (anti-LAG3; C9B7W), anti-CD8 (2.43), and anti-CD25 (PC61) mAbs and FITC-conjugated anti-Thy1.2 (30-H12), and anti-CD4 (GK1.5) mAbs were purchased from BD Pharmingen. PE-conjugated anti-glucocorticoid-induced TNFR (anti-GITR; 108619) mAb was purchased from R&D Systems. Analyses of cell surface phenotypes were conducted by direct immunofluorescent staining of 0.5–1 × 10<sup>6</sup> cells with conjugated mAbs. In each sample, 10,000 cells were analyzed using a FACScan flow microfluorometer (BD Biosciences). PE-conjugated subclass-matched Abs used as isotype controls were also purchased from BD Pharmingen.

### Fractionation of T cells

T cells in the LN cell suspension were concentrated by passing through nylon wool columns (Wako Pure Chemical Industries). To yield highly purified (>90%) cells with down-regulated CD62L expression (CD62L<sup>low</sup>), LN T cells were further isolated by a panning technique using T-25 flasks precoated with goat anti-rat Ig Ab (Jackson ImmunoResearch Laboratories)/anti-CD62L mAb (MEL14) and sheep anti-rat-Ig Ab/anti-CD62L mAb-coated Dynabeads M-450 (DynaL Biotech). T cells with high CD62L expression (CD62L<sup>high</sup>) were obtained as cells attached to flasks coated with goat anti-rat Ig Ab/anti-CD62L mAb. In some experiments cells were also separated into CD4<sup>+</sup> and CD8<sup>+</sup> cells by depletion using magnetic beads as described previously (14). For *in vitro* experiments, highly purified CD4<sup>+</sup> cells were obtained using anti-CD4 mAb-coated Dynabeads and Detachabeads (DynaL Biotech) according to the manufacturer's instructions. CD25<sup>+</sup> cells were isolated using PE-conjugated anti-CD25 mAb and anti-PE microbeads (Miltenyi Biotec) according to the manufacturer's directions. Cell purity was >90%.

### Bone marrow-derived dendritic cells (DCs)

DCs were generated from bone marrow cells as described previously. In brief, bone marrow cells obtained from femurs and tibias of naive mice were placed in T-75 flasks for 2 h at 37°C in complete medium (CM) containing 10 ng/ml rmGM-CSF (a gift from KIRIN). Nonadherent cells were collected by aspirating the medium and were transferred into fresh flasks. On day 6, nonadherent cells were harvested by gentle pipetting. CM consists of RPMI 1640 medium supplemented with 10% heat-inactivated LPS-qualified FCS, 0.1 mM nonessential amino acids, 1 μM sodium pyruvate, 100 U/ml penicillin, 100 μg/ml streptomycin sulfate (all from Invitrogen Life Technologies), and 5 × 10<sup>-5</sup> M 2-ME (Sigma-Aldrich).

### Tumor-draining LN cells

B6 mice were inoculated *s.c.* with 2 × 10<sup>6</sup> MCA 205 tumor cells in both flanks. Inguinal LN draining tumors were harvested. Single-cell suspensions were prepared mechanically as described previously (20).

### Adoptive immunotherapy

B6 mice were injected *s.c.* with 1.5 × 10<sup>6</sup> MCA 205 tumor cells in 100 μl of HBSS to establish *s.c.* tumors. Three days after inoculation, mice were sublethally irradiated (500 cGy) and then infused *i.v.* with T cells isolated from tumor-draining LNs. Perpendicular diameters of *s.c.* tumors were measured with calipers. The significance of differences in the diameters between groups was analyzed by Student *t* test. A value of *p* < 0.05 was considered significant.

### Cytokine ELISA

T cells were stimulated with immobilized anti-CD3 mAb or tumor Ag-pulsed bone marrow-derived DCs in CM. Supernatants were harvested and assayed for mouse IFN-γ content by a quantitative sandwich enzyme immunoassay using a mouse IFN-γ ELISA kit (Genzyme) according to the manufacturer's instructions.

### RT-PCR

Total RNA was isolated from T cells using Isogen (Nippon Gene) and used for cDNA synthesis. The cDNAs were used as templates for PCR (94°C for 2 min, 58°C for 30 s, and 72°C for 1.5 min), and 35 cycles were performed using primers specific for forkhead/winged helix transcription factor gene (*foxp3*; forward, 5'-GGCCCTTCTCCAGGACAGA-3'; 5'-GCTGATCATGGCTGGGTTGT-3'). To ensure the quality of the product, RT-PCR was also performed using primers specific for β<sub>2</sub>-microglobulin.

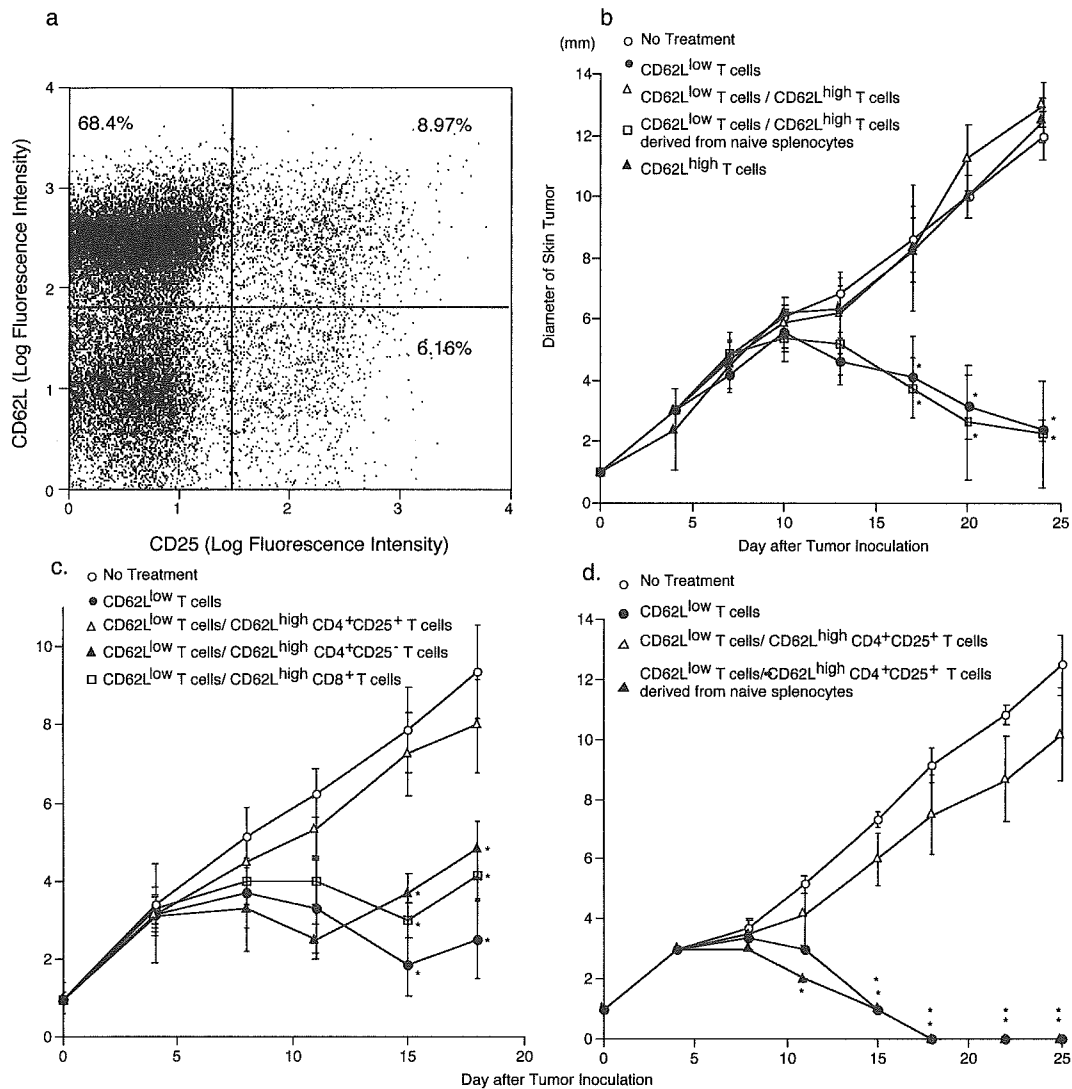
### Proliferation assay

T cells isolated from tumor-draining LNs were stimulated with immobilized anti-CD3 mAb for 48 h in 2 ml of CM on 24-well plates at 2 × 10<sup>6</sup>/ml. CD62L<sup>low</sup> T cells were labeled with 5 μM CFSE (Molecular Probes) in HBSS at 37°C for 15 min and washed twice before CD3 stimulation. The ratio of CD62L<sup>low</sup> T cells to CD62L<sup>high</sup> CD4<sup>+</sup>CD25<sup>+</sup> T cells was 2:1. After a 48-h stimulation, cells were counted and washed twice with HBSS. Then, T cells were cultured in CM supplemented with 10 U/ml human rIL-2 (gift from Shionogi) at 1 × 10<sup>5</sup>/ml. Three wells were analyzed for each condition.

## Results

### CD62L<sup>high</sup> T cells derived from tumor-draining LNs, but not from naive spleens, abrogated antitumor reactivity of CD62L<sup>low</sup> LN T cells

It was believed that CD62L<sup>high</sup> T cells are naive T cells; however, we reported that the elimination of CD62L<sup>high</sup> T cells promotes the generation of highly potent antitumor CD4<sup>+</sup> T cells upon stimulation with CD3. To determine CD62L<sup>high</sup> LN T cells possess activity to abolish the antitumor reactivity of effector T cells primed in tumor-draining LNs, mice with established *s.c.* tumors were infused with 2 × 10<sup>6</sup> CD62L<sup>low</sup> LN T cells in the presence or the absence of 10 × 10<sup>6</sup> CD62L<sup>high</sup> T cells. T cells were isolated from LNs draining growing MCA 205 tumors for 12 days or from spleens of naive mice. The ratio of CD62L<sup>low</sup> to CD62L<sup>high</sup> T cells was approximately the same as that in LNs, because 15–25% of all T cells were CD62L<sup>low</sup> in 12-day tumor-draining LNs. As shown in Fig. 1*b*, 2 × 10<sup>6</sup> CD62L<sup>low</sup> T cells alone successfully mediated the antitumor efficacy to regress *s.c.* tumor growth. In contrast, the *s.c.* tumor growth curve of the mice infused with 2 × 10<sup>6</sup> CD62L<sup>low</sup> T cells in the presence of 10 × 10<sup>6</sup> CD62L<sup>high</sup> T cells derived from tumor-draining LNs was identical with that of the no treatment group. CD62L<sup>high</sup> T cells derived from naive splenocytes did not affect the antitumor reactivity of CD62L<sup>low</sup> tumor-draining LN T cells. Thus, CD62L<sup>high</sup> T cells of tumor-draining LNs contain a subpopulation that is capable of abrogating the antitumor reactivity of effector T cells primed in the same tumor-draining LNs.



**FIGURE 1.** *a*, CD62L and CD25 expression of CD4<sup>+</sup> T cells in 12-day tumor-draining LNs. LN cells were stained with FITC-conjugated anti-CD4 mAb, PE-conjugated anti-CD62L mAb, and PerCP-conjugated anti-CD25 mAb. Gated CD4<sup>+</sup> cells were analyzed with a microfluorometer. *b–d*, Subcutaneous tumor growth of mice infused with CD62L<sup>low</sup> T cells in the presence or the absence of CD62L<sup>high</sup> T cells. Two million MCA 205 tumor cells were injected s.c. along the midline of the abdomen to establish s.c. tumors. Three days later, mice were adoptively infused i.v. with  $2 \times 10^6$  CD62L<sup>low</sup> T cells alone or with  $10 \times 10^6$  CD62L<sup>high</sup> T cells after sublethal whole body irradiation (500 cGy; *b*). CD62L<sup>low</sup> T cells were isolated from LNs draining growing MCA 205 s.c. tumors for 12 days. CD62L<sup>high</sup> T cells were isolated from tumor-draining LNs or naive splenocytes. *c* and *d*, CD62L<sup>high</sup> T cells were further fractionated according to CD4, CD8, and CD25 expression using magnetic beads. One million fractionated CD62L<sup>high</sup> T cells were infused into mice bearing established 3-day s.c. tumors with  $2 \times 10^6$  (*c*) or  $4 \times 10^6$  (*d*) CD62L<sup>low</sup> LN T cells. Diameters of s.c. tumors were measured twice weekly with calipers, and size was recorded as the average of two perpendicular diameters. Statistical analyses were performed with Student's *t* test. \*,  $p < 0.01$  compared with the no treatment group. Each group contained five mice.

#### CD4<sup>+</sup>CD25<sup>+</sup> subpopulation of CD62L<sup>high</sup> tumor-draining LN T cells mediated regulatory functions

Because CD4<sup>+</sup>CD25<sup>+</sup> is the best surrogate marker for regulatory T cells identified to date, CD4<sup>+</sup>CD25<sup>+</sup>, CD4<sup>+</sup>CD25<sup>-</sup>, or CD8<sup>+</sup>CD62L<sup>high</sup> T cells isolated from tumor-draining LNs were infused with CD62L<sup>low</sup> LN T cells into mice bearing established s.c. tumors to determine which subpopulation of CD62L<sup>high</sup> T cells mediates regulatory functions. In Fig. 1*c*, the antitumor efficacy of  $2 \times 10^6$  CD62L<sup>low</sup> T cells was not enough to cure the s.c. tumor, which, after starting to regress, eventually grew in the mice. Neither CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>-</sup> nor CD62L<sup>high</sup>CD8<sup>+</sup> T cells showed any additive antitumor or regulatory activity. The s.c. tumor growth curves showed no significant differences from the curve for the mice who received CD62L<sup>low</sup> T cells alone. In contrast, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells abolished the antitumor efficacy

of effector T cells, resulting in a growth curve identical with that of the no treatment group. Thus, the CD4<sup>+</sup>CD25<sup>+</sup> subpopulation of CD62L<sup>high</sup> LN T cells is probably made up of Treg cells, whereas CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>-</sup> and CD8<sup>+</sup> cells are functionally irrelevant naive T cells. Moreover, Fig. 1*d* shows that  $1 \times 10^6$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> tumor-draining LN T cells were capable of abrogating the antitumor reactivity of  $4 \times 10^6$  CD62L<sup>low</sup> LN T cells, which was sufficient to cure established 3-day s.c. tumors. In contrast, the same number of CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells derived from naive spleens, which are considered resident Treg (21), did not influence the therapeutic efficacy of antitumor effector T cells in vivo.

In 12-day MCA 205 s.c. tumor-draining LNs, ~20% of T cells are CD62L<sup>low</sup>, and 5–7% are CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> (Fig. 1*a*). The ratio of CD62L<sup>low</sup> to CD62L<sup>high</sup> CD4<sup>+</sup>CD25<sup>+</sup> T cells is

3–4:1. Thus, it seems that Treg cells sufficient to abolish the antitumor reactivity of primed effector T cells are generated in tumor-bearing hosts.

*CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> inhibited IFN- $\gamma$  production by either CD4<sup>+</sup> or CD8<sup>+</sup> CD62L<sup>low</sup> T cells via direct T-T interaction*

To test whether CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells generated in tumor-draining LNs influence cytokine production, we measured the amount of IFN- $\gamma$  produced by  $1 \times 10^5$  CD62L<sup>low</sup> T cells in the presence or the absence of  $5 \times 10^4$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells in 200  $\mu$ l of CM on 96-well plates. As shown in Fig. 2a, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells in tumor-draining LNs abolished the Ag-specific production of IFN- $\gamma$  by tumor-draining LN effector T cells stimulated with  $5 \times 10^4$  tumor Ag-loaded DCs. As shown in Fig. 2b, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells derived from naive splenocytes did not affect the production of IFN- $\gamma$  stimulated by tumor-associated Ag, although they inhibited IFN- $\gamma$  production in the presence of nonspecific stimulation with immobilized anti-CD3 mAb (data not shown).

Next, we examined whether this suppression of cytokine production can be reproduced without APC and tested whether it is cell-cell contact dependent, because the Treg naturally arise in the thymus to maintain self-tolerance. One million CD62L<sup>low</sup> LN T cells on the bottom of 24-well plates were cocultured with  $0.5 \times 10^6$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells on either 0.4- $\mu$ m pore size Transwell inserts (Costar) or the bottom of plates in 0.5 ml of CM. Both 24-well plates and Transwell inserts were coated with anti-CD3 mAb. As shown in Fig. 2, c and d, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells abrogated IFN- $\gamma$  production by either CD8<sup>+</sup> or CD4<sup>+</sup>

effector T cells in the absence of APC upon stimulation with CD3, and the suppression was dependent on cell-cell contact.

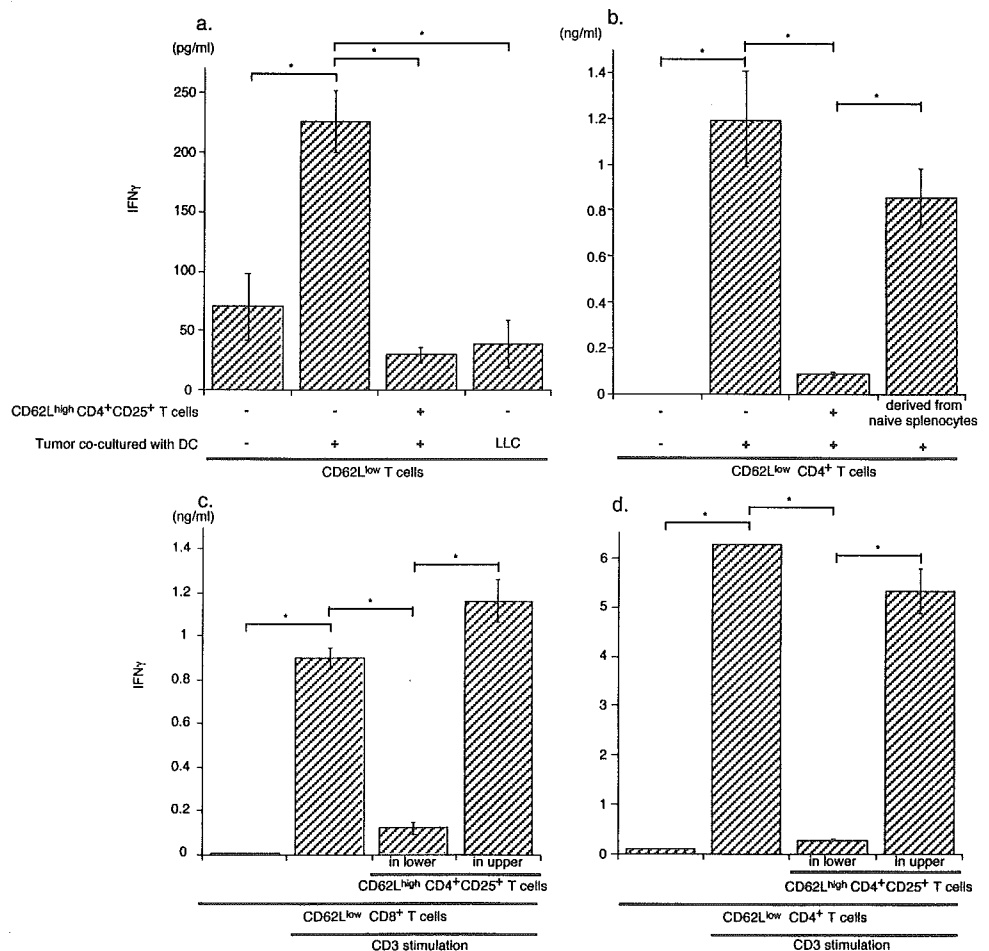
*CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells abrogated proliferation of both CD4<sup>+</sup> and CD8<sup>+</sup>CD62L<sup>low</sup> T cells*

To elucidate whether CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells generated in tumor-draining LNs inhibit cell proliferation, a T cell proliferation assay was performed as described in *Materials and Methods*. CFSE-labeled CD62L<sup>low</sup> T cells stimulated with immobilized anti-CD3 mAb increased the total number of cells by 7-fold during a 3-day culture period accompanied by a reduction in the intensity of CFSE (Fig. 3, a and b). CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells did not affect CD62L<sup>low</sup> T cell proliferation, because the total number of cells increased and the reduction in intensity of CFSE intensity during the 3-day culture was identical with that of CD62L<sup>low</sup> T cells alone. In contrast, CD62L<sup>low</sup> T cells stimulated in the presence of CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells did not proliferate at all. CFSE intensity did not change during the 3-day culture. Fig. 3c demonstrates the relative number of CD8<sup>+</sup> or CD4<sup>+</sup> cells according to phenotypic analysis. Thus, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells generated in tumor-draining LNs have the ability to abrogate the proliferation of both CD4<sup>+</sup> and CD8<sup>+</sup> T cells.

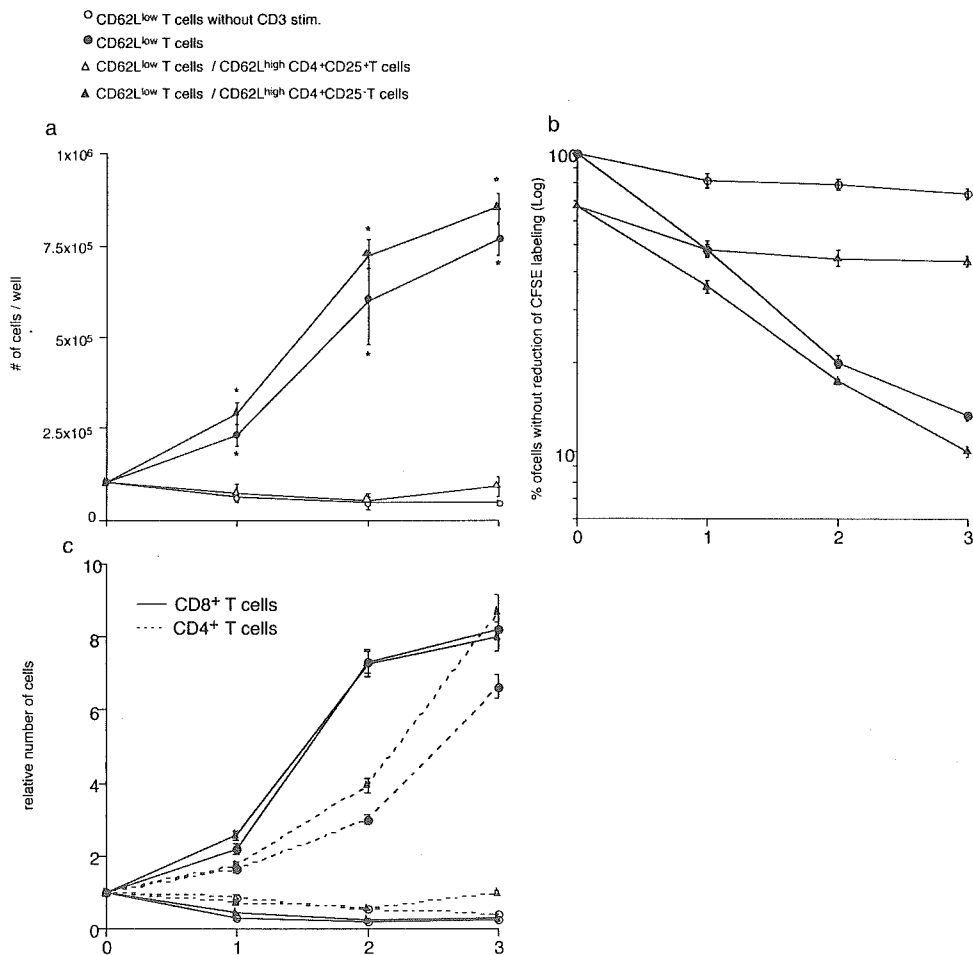
*CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> LN T cells had effector, but not regulatory, functions*

To examine the properties of CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells, which comprise 20–30% of the CD62L<sup>low</sup> T cell population in 12-day tumor-draining LNs, we tested whether CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> LN T cells affect IFN- $\gamma$  production by CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T

**FIGURE 2.** Measurement by ELISA of IFN- $\gamma$  secreted in the medium. CD62L<sup>low</sup> T cells  $1 \times 10^5$  were cocultured with  $0.5 \times 10^5$  DCs in the presence or the absence of  $0.5 \times 10^5$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells for 72 h in 200  $\mu$ l of CM in 96-well plates (a and b). Before coculture with T cells, DCs were incubated with 5000 cGy-irradiated MCA 205 or LLC tumor cells overnight. The amount of IFN- $\gamma$  produced by  $1 \times 10^6$  CD62L<sup>low</sup>CD8<sup>+</sup> or CD4<sup>+</sup> T cells cultured with  $0.5 \times 10^6$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells upon CD3 stimulation is shown in c and d. CD62L<sup>low</sup> T cells on the bottom of 24-well plates were cocultured with CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells either on 0.4- $\mu$ m pore size Transwell inserts or on the bottom. Both the Transwell inserts and 24-well plates were coated with anti-CD3 mAb. T cells were isolated from LNs draining growing MCA 205 s.c. tumors for 12 days. Three wells were analyzed for each condition. ELISA was performed in duplicate. Statistical analyses were performed by Student's *t* test. \*, *p* < 0.01.



**FIGURE 3.** T cell proliferation was evaluated in the presence of 10 U/ml IL-2 after stimulation with CD3. CD62L<sup>low</sup> T cells isolated from tumor-draining LNs were labeled with CFSE and cultured alone or with CD62L<sup>high</sup>CD4<sup>+</sup> LN T cells at  $2 \times 10^6$ /ml in 2 ml of CM on 24-well plates coated with anti-CD3 mAb for 48 h. The ratio of CD62L<sup>low</sup> to CD62L<sup>high</sup> cells was 2:1. After CD3 stimulation, T cells were harvested and counted, then recultured at  $0.1 \times 10^6$ /ml in 2 ml of CM supplemented with 10 U/ml IL-2 on new 24-well plates. T cells were counted and analyzed with a microfluorometer every 24 h. *a*, Total number of cells per well. *b*, Percentage of T cells that still possess high levels of CFSE labeling and represent cells without division. *c*, Relative number of CD8<sup>+</sup> or CD4<sup>+</sup> T cells. Three wells were analyzed for each condition. \*,  $p < 0.01$  compared with the no stimulation group.



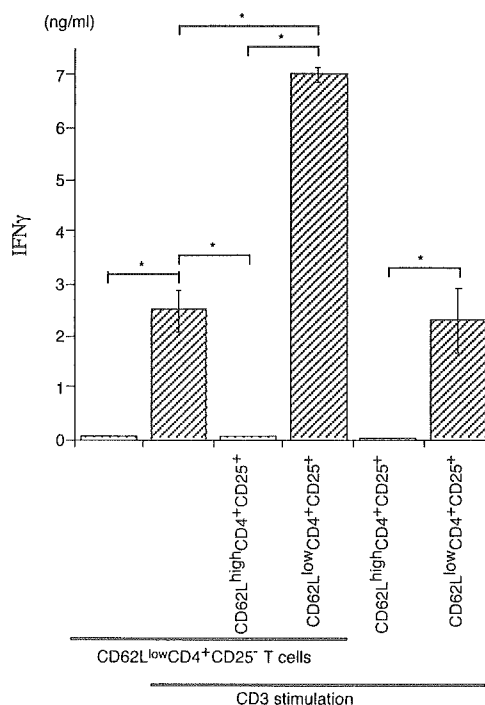
cells. CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells completely inhibited the production of IFN- $\gamma$  by CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells (Fig. 4), whereas the addition of CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells increased production. Moreover, CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells alone produced the same amount of IFN- $\gamma$  as CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells. In contrast, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells produced no IFN- $\gamma$ . Thus, it is likely that CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> cells possess effector T cell function, but not regulatory activity.

#### Foxp3 mRNA expression was specific to CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells

It has been reported that mutation of Foxp3 is responsible for immune dysregulation, polyendocrinopathy, enteropathy, and X-linked inheritance, a syndrome of systemic autoimmunity in humans (22, 23). It is now believed that Foxp3 is a master switch of regulatory functions (9, 24, 25). Thus, we tested whether fractionated T cells derived from tumor-draining LNs express mRNA for Foxp3. As depicted in Fig. 5A, only CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells expressed *foxp3* mRNA.

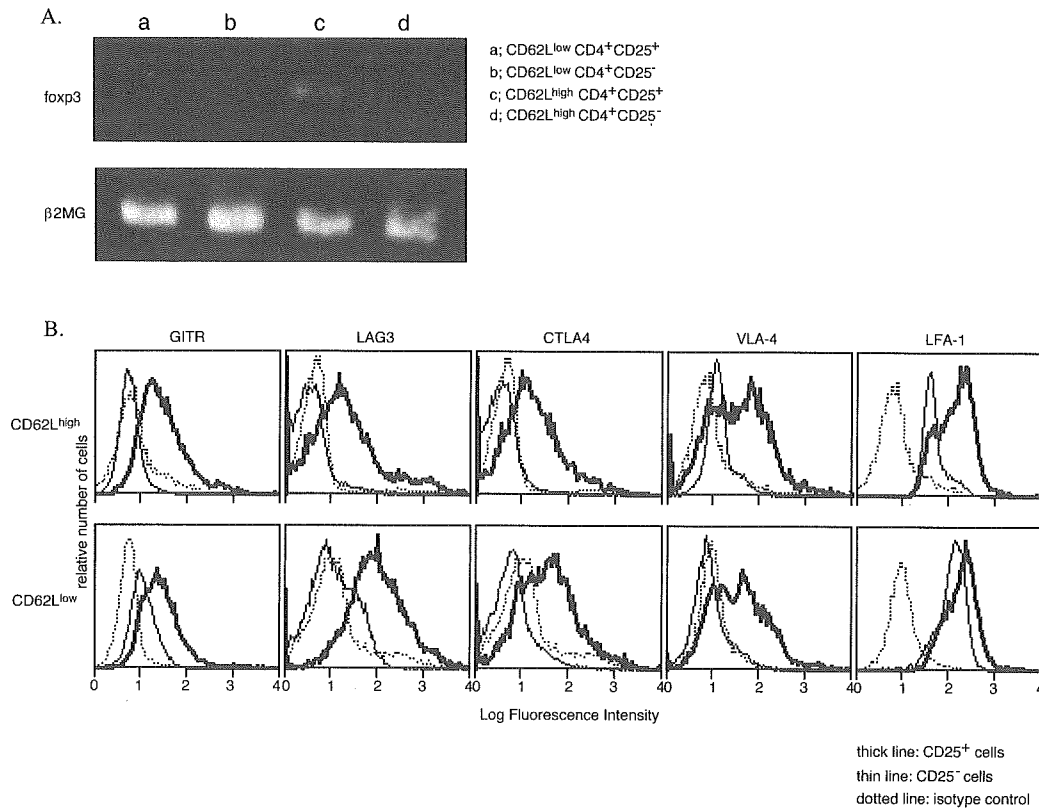
#### Functionally distinct CD62L<sup>high</sup> and CD62L<sup>low</sup> CD4<sup>+</sup>CD25<sup>+</sup> T cells expressed comparable levels of GITR, CTLA-4, LAG3, VLA-4, and LFA-1

Next, we analyzed the phenotype of fractionated T cells derived from tumor-draining LNs. Because it was demonstrated that CD4<sup>+</sup>CD25<sup>+</sup> Treg cells express GITR, CTLA-4, and LAG3 (26–29), we tested for these molecules and adhesion molecules that were important for T cell migration. Although CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> regulatory LN T cells have an up-regulated expression of



**FIGURE 4.** Measurement by ELISA of IFN- $\gamma$  secreted in the medium. CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells alone ( $1 \times 10^5$ ) or with  $0.5 \times 10^5$  T cells fractionated according to the expression of CD62L and CD25 were stimulated with immobilized anti-CD3 mAb in 200  $\mu$ l of CM on 96-well plates for 48 h. \*,  $p < 0.01$ .





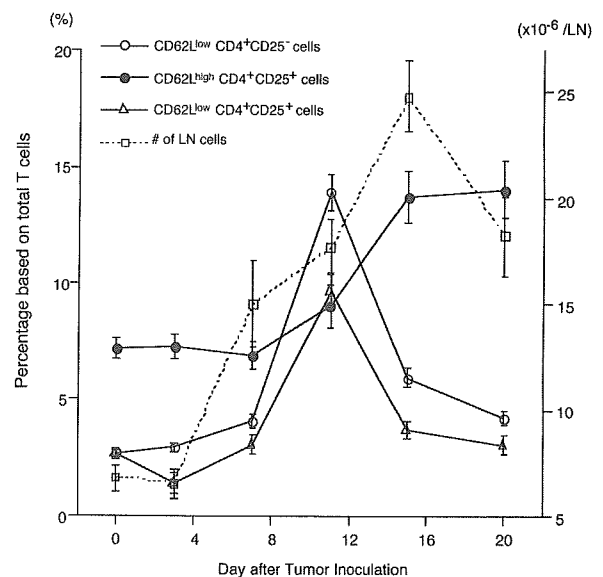
**FIGURE 5.** A, *Foxp3* mRNA expression in isolated  $CD62L^{low} CD4^{+} CD25^{+}$ ,  $CD62L^{low} CD4^{+} CD25^{-}$ ,  $CD62L^{high} CD4^{+} CD25^{+}$ , or  $CD62L^{high} CD4^{+} CD25^{-}$  T cells derived from LNs draining MCA 205 s.c. tumors for 12 days. Total RNA was isolated from T cells and analyzed by RT-PCR for *foxp3*.  $\beta 2$ -Microglobulin gene expression is shown to confirm that equal amounts of RNA were used in each RT-PCR. Results shown are representative of three separate experiments. B, GITR, LAG3, CTLA-4, VLA-4, and LFA-1 expression on isolated T cells derived from MCA 205 tumor-draining LNs. Immediately after fractionation, T cells were double stained with PE-labeled anti-CD25 and FITC-conjugated anti-GITR, anti-LAG3, anti-CTLA-4, anti-VLA-4, anti-LFA-1, or isotype control Ab. Either  $CD25^{+}$  or  $CD25^{-}$  cells were gated for analyses. A total of  $10^6$  cells were analyzed for each sample. Each frame consists of 10,000 cells. Dotted lines indicate the isotype control.

GITR, CTLA-4, and LAG3, it is difficult to distinguish  $CD62L^{high} CD4^{+} CD25^{+}$  T cells from  $CD62L^{low} CD4^{+} CD25^{+}$  T cells, which possess effector T cell properties, from these molecules (Fig. 5B). Furthermore,  $CD62L^{high} CD4^{+} CD25^{+}$  Treg cells and  $CD62L^{low} CD4^{+} CD25^{+}$  effector T cells had a comparable up-regulated expression of VLA-4 and LFA-1. In contrast,  $CD62L^{high} CD4^{+} CD25^{-}$  T cells possessed the naive cell phenotype, such as no VLA-4, GITR, or CTLA-4, and a low level of LFA-1.

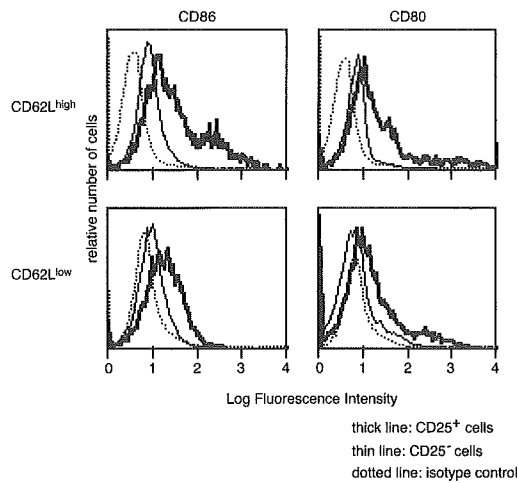
*Different kinetics of  $CD62L^{high} CD4^{+} CD25^{+}$  and  $CD62L^{low} CD4^{+}$  T cell priming in LNs draining growing s.c. tumors*

To address the priming of  $CD62L^{high} CD4^{+} CD25^{+}$  and  $CD62L^{low} CD4^{+}$  T cells in LNs draining growing MCA 205 s.c. tumors, we examined the number and phenotype of LN cells. Kinetic analysis revealed that the proportion of both  $CD62L^{low} CD4^{+} CD25^{+}$  and  $CD4^{+} CD25^{-}$  T cells started to increase 7 days after s.c. tumor inoculation and peaked on the 11th day (Fig. 6). The percentage of  $CD62L^{low}$  T cells rapidly decreased, reaching the starting level by the 14th day. Although the proportion of  $CD62L^{high} CD4^{+} CD25^{+}$  T cells started to increase 7 days after s.c. tumor inoculation, like that of  $CD62L^{low}$  cells, it kept increasing until the total number of LN cells started to decrease. The increase in  $CD62L^{high} CD4^{+} CD25^{+}$  T cells was not caused by a nonspecific accumulation of  $CD62L^{high}$  cells, because the proportion of  $CD62L^{high} CD4^{+} CD25^{-}$  naive T cells decreased in tumor-draining

LNs. Hence, it is likely that  $CD62L^{high} CD4^{+} CD25^{+}$  Treg cells underwent clonal expansion in tumor-draining LNs during tumor progression.



**FIGURE 6.** Kinetics of cellularity and the ratio of the indicated T cell subpopulations based on total T cells in LNs draining growing MCA 205 s.c. tumors. MCA 205 tumor cells ( $1.5 \times 10^6$ ) were inoculated s.c. into both flanks of mice. Inguinal LNs were harvested from three mice serially 0, 3, 7, 11, 15, and 20 days after s.c. injection and analyzed.



**FIGURE 7.** T cells isolated from 12-day tumor-draining LNs were analyzed for the expression of CD80 or CD86. CD4<sup>+</sup> T cells positively selected with magnetic beads were three-color-stained with FITC-labeled anti-CD80 mAb or FITC-labeled anti-CD86 mAb in the presence of PE-labeled anti-CD62L mAb and PerCP-labeled anti-CD25 mAb. Gated CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup>, CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>-</sup>, CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup>, or CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells were analyzed for CD80 or CD86 expression. A total of 10<sup>7</sup> cells were analyzed for each sample.

*CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells suppress effector CD4<sup>+</sup> T cell functions via CD86/CTLA-4 T-T interactions*

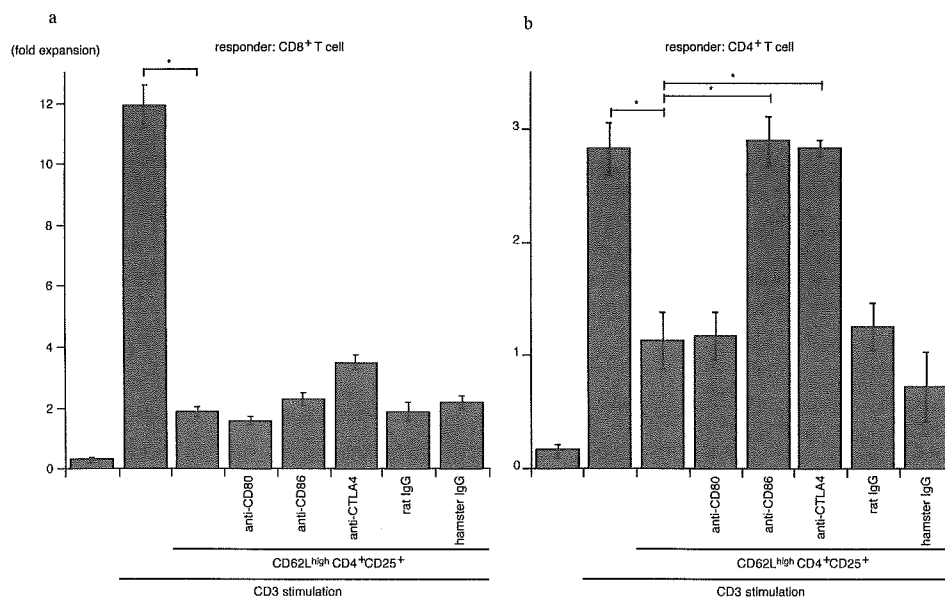
It has been reported that an inhibitory Ab against CTLA-4 could abrogate the induction of suppression by CD4<sup>+</sup>CD25<sup>+</sup> Treg, and CTLA-4 expressed on Treg was thought to be important for this phenomenon (27). However, the mechanism by which Treg deliver a regulatory signal to effector T cells is still unclear. We examined the expression of CD80 and CD86 on fractionated tumor-draining LN T cells. Unexpectedly, only the CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells had a subpopulation that expressed CD86 (Fig. 7).

CD62L<sup>low</sup>CD4<sup>+</sup> or CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells did not express a detectable level of CD86 even after stimulation with CD3 (data not shown). Next, we tested whether CD86 on regulatory T cells participates in the suppression of T cell functions. CD62L<sup>low</sup> T cells and CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells were stimulated with immobilized anti-CD3 mAb in the presence or the absence of an inhibitory mAb against CD80, CD86, or CTLA-4. Anti-CTLA-4 as well as anti-CD86 inhibitory mAb prevented the suppression of CD62L<sup>low</sup>CD4<sup>+</sup> T cells induced by CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells, whereas anti-CD80 inhibitory mAb or isotype-matched irrelevant Abs did not have any influence (Fig. 8a). However, blockade of CTLA-4 or CD86 could not reverse the suppression of CD62L<sup>low</sup>CD8<sup>+</sup> LN T cells (Fig. 8b).

## Discussion

In this study we demonstrated that both effector T cells and Treg cells are primed in the same LNs that drain growing MCA 205 s.c. tumors. The antitumor effector T cells in tumor-draining LNs belong to a subpopulation that down-regulated CD62L expression, as we previously demonstrated (11, 14). In contrast, the CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> subpopulation in tumor-draining LNs abrogate the in vivo antitumor therapeutic efficacy of CD62L<sup>low</sup> antitumor LN T cells (Fig. 1). Kinetic analyses indicate that CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg proliferated in tumor-draining LNs during tumor progression (Fig. 6). Data obtained in vitro reveal that the Treg generated in tumor-draining LNs to abrogate antitumor reactivity possess the same functional properties and level of *foxp3* expression as the Treg that naturally arise in the thymus to maintain self-tolerance (Figs. 2–5).

CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells derived from spleens of naive mice could not abrogate antitumor reactivity in vivo (Fig. 1). Moreover, Treg cells from naive mice did not inhibit the production of IFN- $\gamma$  by antitumor effector T cells upon Ag stimulation by DCs acquired from apoptotic tumor cells (Fig. 2b), even though they had comparable suppressive activity as the Treg cells generated in tumor-draining LNs upon nonspecific CD3 stimulation.



**FIGURE 8.** Proliferation assay of T cells in the presence or the absence of a blockade between CD86 and CTLA-4. CD62L<sup>low</sup>CD4<sup>+</sup> or CD8<sup>+</sup> T cells ( $1 \times 10^5$ ) were cocultured with  $5 \times 10^4$  CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells in 200  $\mu$ l of CM on 96-well plates with immobilized anti-CD3 mAb for 48 h. T cells were isolated from LNs draining MCA 205 s.c. tumors for 12 days. To inhibit the interaction between CD86 and CTLA-4, anti-CD86 mAb or anti-CTLA-4 mAb was added to the medium at 10  $\mu$ g/ml during the 48 h of stimulation. T cells were recultured in 1 ml of CM supplemented with 10 U/ml IL-2 on 24-well plates at  $1 \times 10^5$ /ml. As a control, anti-CD80 mAb, rat IgG, or hamster IgG was added at the same concentration. Three wells were examined for each condition, and cells were enumerated every 24 h. The y-axis indicates the fold increase in number on day 2. \*,  $p < 0.01$ .

Hence, it is likely that Treg are required to receive TCR/CD3 signaling to interfere with the function of effector T cells and that the Treg generated in tumor-draining LNs recognize tumor-associated Ag.

Although it has been demonstrated that CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells possess superior regulatory activity in several systems, CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells were still considered Treg cells (15, 16, 18). However, the differences in *foxp3* expression and priming kinetics suggest that the CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cell subpopulation in tumor-draining LNs is distinct from CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells (Figs. 5A and 6). CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells produced even more IFN- $\gamma$  than CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells upon CD3 stimulation (Fig. 4). Furthermore, CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> LN T cells mediated antitumor efficacy in vivo (Fig. 1). These results indicated that CD62L<sup>low</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells are effector T cells that express CD25 because of recent TCR stimulation before clonal expansion in LNs.

Little is known about the trafficking of Treg cells; however, our study shows that Treg cells generated in tumor-draining LNs have an up-regulated expression of CD62L, VLA-4, and LFA-1 (Figs. 1 and 5B). CD62L recognizes specific ligands on high endothelial venules and is considered the homing receptor for secondary lymphoid tissues (30). In contrast, VLA-4 and LFA-1 are thought to play a central role in T cell trafficking to inflammatory sites by recognizing VCAM-1 and ICAM-1 on endothelial cells. Thus, it is likely that CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells can follow either CD62L<sup>high</sup> naive T cells to suppress priming in secondary lymphoid organs or activated CD62L<sup>low</sup> effector T cells that express VLA-4 and LFA-1 to suppress immune reactions in the effector phase.

CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells suppressed either CD4<sup>+</sup> or CD8<sup>+</sup> effector T cell functions, including cytokine production and cell proliferation, in the absence of APC upon stimulation with CD3 in a cell-cell contact-dependent manner (Fig. 2, *c* and *d*, and Fig. 3). Hence, the regulatory signals sent to either CD4<sup>+</sup> or CD8<sup>+</sup> effector T cells were received directly from CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells and were not mediated by APC. It is postulated that ligation of CD80 and/or CD86 expressed on effector T cells by CTLA-4 on Treg cells causes outside-in signaling and results in suppression (31), although it is well documented that p56<sup>lck</sup>-induced tyrosine phosphorylation, which is the major signal pathway of the TCR/CD3 complex, can be reversed by CTLA-4 ligation (32). We found that the CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells contained a subpopulation that expressed CD86 in tumor-draining LNs (Fig. 7). Effector CD62L<sup>low</sup>CD4<sup>+</sup> or naive CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>-</sup> T cells derived from tumor-draining LNs did not express CD86 or CD80 even after 48-h stimulation with immobilized anti-CD3 mAb (data not shown). The inhibitory mAb against CTLA-4 or CD86, but not CD80, completely reversed the inhibitory effect on the proliferation of CD4<sup>+</sup> effector LN T cells by CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells (Fig. 8). In contrast, the same treatment could not reverse the suppression of CD8<sup>+</sup> effector LN T cells. CD62L<sup>low</sup>CD4<sup>+</sup> T cells, but not CD8<sup>+</sup> T cells, isolated from tumor-draining LNs expressed CTLA-4 (Fig. 5B). Our data suggested that CD86 exclusively expressed on Treg cells plays a pivotal role in regulating CD4<sup>+</sup> effector T cells by interacting with CTLA-4 on effector T cells. This is a novel mechanism by which Treg cells suppress effector T cell function. This finding is compatible with recent reports demonstrating that CD4<sup>+</sup>CD25<sup>+</sup> T cells derived from CTLA-4-deficient mice mediated suppression (33) and that T cells transfected with cDNA encoding CD86 suppressed graft-vs-host disease (34). The reason why the bioactivity of CD86 on T cells is different from that on APC is unclear; however, DCs are capable of selecting the receptor of B7 by recruiting

either CD28 or CTLA-4 into immunological synapses (35). In contrast, it is unlikely that Treg cells have the ability to develop immunological synapse with effector T cells to select receptors. Moreover, it was demonstrated that CD86 expressed on T cells had a hypoglycosylated form and showed no detectable binding activity to CD28 with preserved binding to CTLA-4 (36). Thus, it is possible that CD86 on T cells preferentially give negative signaling through CTLA-4 (37).

CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells regulated CD8<sup>+</sup> T cell functions via mechanisms other than the CD86-CTLA-4 interaction, because the inhibitory mAb against CD86 or CTLA-4 did not abolish the suppression. Because neutralizing Ab against TGF- $\beta$  partially inhibited the induction of suppression, it is likely that membrane-bound TGF- $\beta$  participated in CD8<sup>+</sup> T cell suppression (data not shown).

These results indicate that Treg cells for tumor Ags are primed in draining LNs during tumor progression and that the balance between CD62L<sup>low</sup> effector T cell priming and CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cell priming in secondary lymphoid organs determines the outcome of antitumor immune reactions. Our data also indicate that promoting antitumor effector CD62L<sup>low</sup> T cell priming while eliminating CD62L<sup>high</sup>CD4<sup>+</sup>CD25<sup>+</sup> Treg cells or inhibiting regulatory mechanisms such as CD86-CTLA-4 interaction is critical to establishing effective antitumor immunotherapy. Furthermore, it might be possible to orchestrate adaptive immune reactions by manipulating the balance of effector and regulatory T cell priming against acquired Ags, such as infectious pathogens, alloantigens, and allergens.

## Disclosures

The authors have no financial conflict of interest.

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## A novel type of encephalopathy associated with mushroom Sugihiratake ingestion in patients with chronic kidney diseases

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### A novel type of encephalopathy associated with mushroom Sugihiratake ingestion in patients with chronic kidney diseases.

**Background.** The etiology of encephalopathy in uremic patients is multiple. We recently encountered a novel type of encephalopathy which occurred exclusively in patients with chronic kidney diseases after ingestion of a mushroom called Sugihiratake. While the exact etiology of this encephalopathy remained mysterious, we aimed to describe its clinical features.

**Methods.** A total of 32 patients with chronic kidney diseases who had presented with encephalopathy following ingestion of Sugihiratake were enrolled from seven prefectures in Japan, with 24 of the 32 patients undergoing regular hemodialysis. The patient's clinical data were from surveillance by The Japanese Society of Nephrology.

**Results.** There was a significant association between Sugihiratake ingestion and the occurrence of encephalopathy in 524 hemodialysis patients questioned for a recent ingestion of this mushroom ( $P = 0.0006$ ). The latent asymptomatic period before the onset of symptoms varied from 1 to 31 days (mean  $9.1 \pm 7.3$ ) days. The patient's symptoms consisted of disturbed consciousness in 30 patients (93.8%), convulsions in 25 (78.1%), myoclonus in 15 (46.9%), dysarthria in ten (31.3%), ataxia in eight (25.0%), paresis or paralysis in seven (21.9%), and skin parasthesia in two patients (6.3%). Nine (27.2%) patients died, mostly due to respiratory failure. The other patients were either discharged or still in hospitals with various degrees of clinical improvement.

**Conclusion.** Patients with chronic kidney diseases are at risk of having serious encephalopathy following Sugihiratake ingestion and must refrain from eating it. Physicians, in those parts of the world, where this mushroom harvesting is common, should be aware of this complication.

**Key words:** mushroom intoxication, encephalopathy, chronic kidney disease, hemodialysis.

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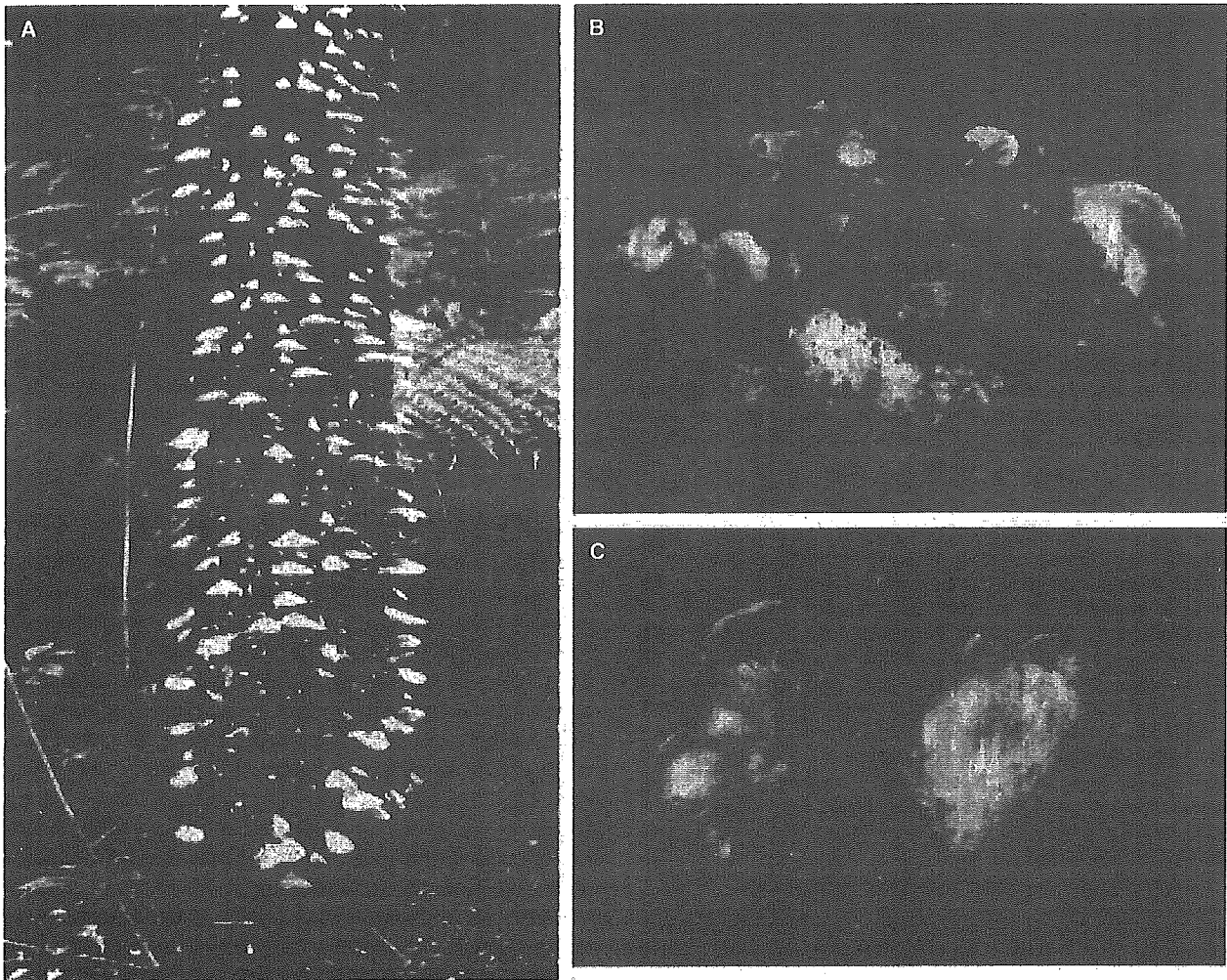
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Sugihiratake (Fig. 1) is the Japanese name of the fungus *Pleurocybella porrigens*, which is a small mushroom that grows in abundance during the fall season, not only in the forest of northern Japan, but is also widely distributed across the northern hemisphere [1]. It has an interesting flavor that many Japanese used to enjoy, usually consumed as a component of the highly popular miso (fermented bean paste) soup. Until now, there has been no report of significant adverse effect as a result of Sugihiratake ingestion. However, during the fall of 2004 in Japan, an outbreak of a serious encephalopathy exclusively occurred in patients with chronic kidney diseases after ingestion of this mushroom.

While the exact etiology of the encephalopathy remains mysterious and currently under investigation, we wish to report for the first time the neurotoxic effects of *P. porrigens* mushroom intoxication in a series of Japanese patients with chronic kidney disease.

### METHODS

Data used for this study were from surveillance by The Japanese Society of Nephrology. From September through October 2004, a total of 45 patients with chronic kidney diseases presented to several hospitals in eight prefectures in Japan (Fig. 2) because of acute neurologic disturbances. Extensive workup to find out a possible cause was negative other than a history of ingesting Sugihiratake in 44 of the 45 patients during its harvesting season from the end of August through October 2004. Among the 44 patients, 32 (19 females and 13 males) who had their clinical conditions reported to us by the physicians in charge were enrolled in the analysis. Twenty-four patients previously had received regular hemodialysis, whereas eight were not yet undergoing dialysis. The mean age of the patients was  $69.2 \pm 10.5$  years. The underlying kidney diseases were chronic glomerulonephritis in



**Fig. 1. Photographs of Sugihiratake (*Pleurocybella porrigens*).** Sugihiratake have a habitat rooting on woods, especially *Cryptomeria japonica* in Japan forming dense clusters (A). The mushroom, which the patient 1 (Table 2) ingested, was that of *P. porrigens* (B). The pileus of Sugihiratake, also known as “angel’s wings,” was 3 to 5 inches long, white in color, and had a wavy margin (C).

14 patients, diabetic nephropathy in eight, unknown etiology in five, hypertension in three, and polycystic kidney disease in two patients. All patients, whether dialyzed or not, were in stable clinical condition before this event. We also questioned a total of 524 hemodialysis patients at nine hospitals in the affected area for any recent history of Sugihiratake ingestion and the intoxication rate was calculated. Statistical analyses were conducted by Fisher exact probability test with StatView 5.0 software (Abacus Concepts, Inc., Berkeley, CA, USA).

## RESULTS

All cases of acute encephalopathy after Sugihiratake ingestion occurred in patients who had chronic kidney diseases, whereas none of the individuals with normal renal function who had eaten Sugihiratake were affected.

Out of 524 hemodialysis patients, 278 (53%) admitted recent ingestion of Sugihiratake, but only 12 patients (4.3%) manifested symptoms of intoxication. By Fisher’s exact probability test, there was a highly significant association between the mushroom ingestion by hemodialysis patients and the development of encephalopathy ( $P = 0.0006$ ).

The clinical profile, time of symptom onset, and the outcome are listed in Table 1 for the patients on hemodialysis, and in Table 2 for the patients with pre-end-stage renal disease (pre-ESRD). Nine (28.1%) patients had died so far, five had been uneventfully discharged, and the other 18 patients have remained hospitalized with various degrees of clinical improvement. While many patients continued to eat the mushroom for several days before the symptoms appeared, the time from the start of ingestion to onset of symptoms varied from 1 to 31



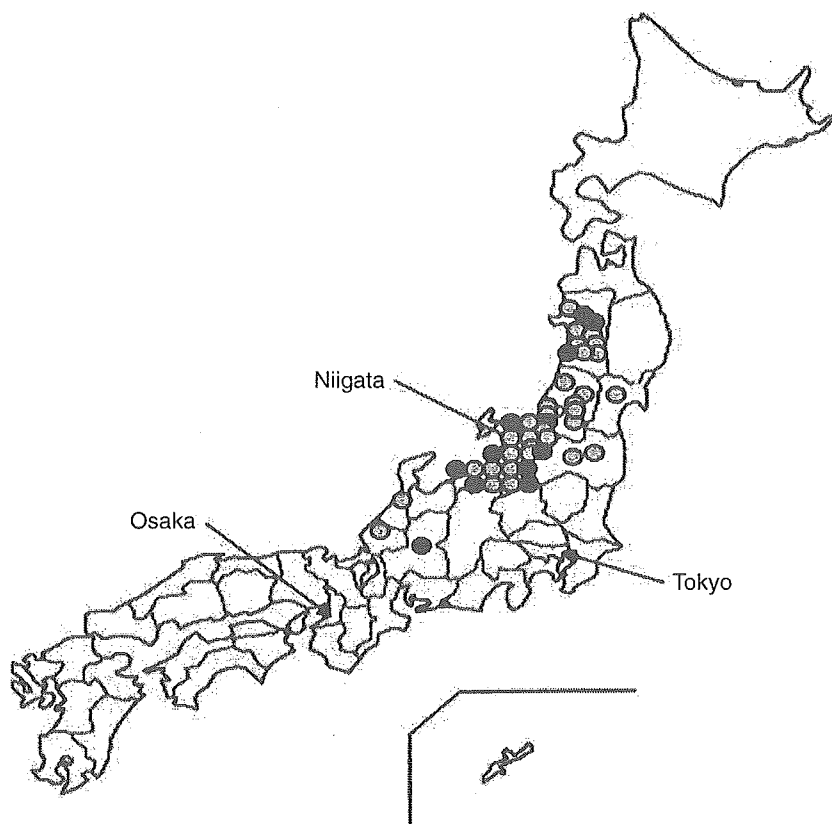


Fig. 2. The distribution of the cases with acute neurological disturbances. Cases included in the analysis (○) ( $N = 32$ ). Cases not included in the analysis because of insufficient clinical information (●) ( $N = 12$ ).

days ( $9.1 \pm 7.3$  days), with no difference between the patients who died or survived. The most common symptoms in all 32 patients included disturbed consciousness of various degrees in 30 patients (93.8%), convulsions in 25 (78.1%), myoclonus in 15 (46.9%), dysarthria in ten (31.3%), ataxia in eight (25.0%), paresis or paralysis in seven (21.9%), and skin parasthesia in two patients (6.3%). Typically, most patients presented initially with ataxia and/or myoclonus several days after Sughiratake ingestion, to be followed in a few days by generalized convulsions. The convulsive activities either stopped without neurologic sequelae or progressed to status epilepticus and death. The terminal event was profound central apnea and bradycardia. Noteworthy, all patients did not experience gastrointestinal symptoms, which are generally common features of mushroom poisoning. Fever was undetectable in almost all patients at the initial presentation; however, some patients had their temperature increased after admission secondary to a respiratory infection.

After being admitted, the patients were closely monitored either in a general ward or in an intensive care unit with the support of a neurologist. The differential diagnosis included metabolic encephalopathy, cerebrovascular accident, viral encephalitis, or possible drug intoxication. Nineteen patients underwent computerized tomographic (CT) brain scans to rule out stroke, but no organic lesion

could be identified, though in five patients, who ultimately died, brain edema was evident. Lumbar puncture was done in ten patients, and the cerebrospinal fluid analysis showed a mild increase of protein content, but normal cell count and sugar. There were no significant changes in the blood biochemistry, including blood sugar, liver function tests, serum aluminum levels, and blood gas analysis compared with the preintoxication levels in most patients.

Treatment depended on the severity of presenting symptoms. Fourteen patients on regular hemodialysis, and two other patients who are not yet on hemodialysis, were assigned to every-other-day dialysis. Ten patients with more severe mental confusion were managed in an intensive care unit with continuous venovenous hemodialysis (CVVHD) and mechanical ventilatory support. Nine patients died 4 to 15 days (mean  $8.2 \pm 4.1$  days) after admission. The principal characteristics of these patients were severe mental confusion in all and convulsive activities in eight.

## DISCUSSION

Acute encephalopathy specifically related to patients with uremia may result from any of the following causes; uremic encephalopathy, dialysis disequilibrium syndrome, cerebrovascular disease, electrolytes

**Table 1.** Clinical characteristics and outcome of the patients undergoing hemodialysis

| Patients | Age | Gender | Duration of hemodialysis years | Onset of symptoms days <sup>a</sup> | Symptoms <sup>b</sup> | Outcome <sup>c</sup> |
|----------|-----|--------|--------------------------------|-------------------------------------|-----------------------|----------------------|
| 1        | 69  | F      | 5                              | 1                                   | 1, 2                  | Died                 |
| 2        | 60  | F      | 8                              | 10                                  | 1, 3                  | In hospital          |
| 3        | 62  | F      | 8                              | 11                                  | 1, 2, 4, 7            | In hospital          |
| 4        | 65  | F      | 8                              | 3                                   | 1, 2, 3               | In hospital          |
| 5        | 70  | F      | 15                             | 14                                  | 1, 2, 3, 6            | In hospital          |
| 6        | 77  | F      | 5                              | ND                                  | 1, 3, 5               | Discharged           |
| 7        | 57  | F      | 5                              | 14                                  | 1, 2, 4, 5            | In hospital          |
| 8        | 66  | F      | 10                             | ND                                  | 1, 2, 3               | In hospital          |
| 9        | 58  | F      | 4                              | 14                                  | 1, 2, 6               | In hospital          |
| 10       | 60s | F      | 0.3                            | 12                                  | 1, 2, 3               | In hospital          |
| 11       | 72  | F      | 2                              | 13                                  | 1, 2, 6               | In hospital          |
| 12       | 71  | F      | 11                             | ND                                  | 1, 4, 6, 7            | In hospital          |
| 13       | 72  | F      | 3                              | 4                                   | 1, 2, 4, 6            | Died                 |
| 14       | 48  | F      | 20                             | ND                                  | 1, 2                  | Died                 |
| 15       | 85  | M      | 1                              | 18                                  | 1, 2, 3               | In hospital          |
| 16       | 68  | M      | 6                              | 12                                  | 1, 2, 4, 5            | In hospital          |
| 17       | 78  | M      | 1                              | 31                                  | 1, 2, 3               | In hospital          |
| 18       | 66  | M      | 3                              | 14                                  | 1, 2, 3, 5, 6         | Died                 |
| 19       | 73  | M      | 1.8                            | 14                                  | 1, 2, 3, 4            | Died                 |
| 20       | 64  | M      | 7                              | ND                                  | 4, 6                  | Discharged           |
| 21       | 50  | M      | 0.3                            | 1                                   | 1, 2, 3               | Discharged           |
| 22       | 53  | M      | 16                             | 3                                   | 1, 2                  | Died                 |
| 23       | 83  | M      | 1.5                            | ND                                  | 1, 2                  | Died                 |
| 24       | 60  | M      | 2                              | 7                                   | 1, 2, 3, 4            | In hospital          |

Abbreviations are: F, female; M, male; ND, not determined.

<sup>a</sup>Time of symptom onset after Sugihiratake ingestion.

<sup>b</sup>1, disturbed consciousness; 2, convulsions; 3, myoclonus; 4, dysarthria; 5, ataxia; 6, paresis/paralysis; 7, parasthesias.

<sup>c</sup>As confirmed by the end of October 2004.

**Table 2.** Clinical characteristics of the patients with pre-end-stage renal disease (pre-ESRD)

| Patients | Age | Gender | Serum creatinine mg/dL | Onset of symptoms days <sup>a</sup> | Symptoms <sup>b</sup> | Outcome <sup>c</sup> |
|----------|-----|--------|------------------------|-------------------------------------|-----------------------|----------------------|
| 1        | 87  | F      | 4.8                    | 3                                   | 1, 2, 3, 4, 5         | Discharged           |
| 2        | 68  | F      | 5.6                    | ND                                  | 1, 5                  | Died                 |
| 3        | 71  | F      | 5.5                    | 7                                   | 1, 2, 5               | Died                 |
| 4        | 84  | F      | 6.2                    | 2                                   | 1, 4                  | In hospital          |
| 5        | 80  | F      | 1.5                    | 1                                   | 1, 2, 3               | In hospital          |
| 6        | 89  | M      | 2.0                    | ND                                  | 1, 2, 5               | In hospital          |
| 7        | 71  | M      | 4.0                    | 8                                   | 1, 2                  | In hospital          |
| 8        | 67  | M      | 8.0                    | 20                                  | 3                     | Discharged           |

Abbreviations are: F, female; M, male; ND, not determined.

<sup>a</sup>Time of symptom onset after Sugihiratake ingestion.

<sup>b</sup>1, disturbed consciousness; 2, convulsions; 3, myoclonus; 4, dysarthria; 5, ataxia; 6, paresis/paralysis; 7, parasthesias.

<sup>c</sup>As confirmed by the end of October 2004.

disorders, and aluminum intoxication [2]. Because of the impaired excretory function, uremic patients are also especially vulnerable to drugs and toxins from many different sources, including food. There have been reports of encephalopathy in uremic patients caused by star fruit (*Averrhoa carambola*) intoxication [3, 4]. However, acute encephalopathy related to mushroom poisoning in uremic patients have not been previously reported.

Two theories for the etiology of this encephalopathy have been speculated. The first theory suggested that an aberrant viral infection, to which uremic patients particularly susceptible, is to be the culprit. However, the absence of family history of similar symptoms, fever, or

other markers of acute inflammatory response, in addition to the lack of cerebrospinal fluid pleocytosis stood against this theory. Moreover, the examination for common viral infections was negative. The second theory accused a toxin that is normally metabolized through the kidney. Reviewing the history for a possible intoxication disclosed that almost all of the patients had eaten Sugihiratake. Statistical analysis of hemodialysis patients in the affected area indicated a significant association between Sugihiratake ingestion and the intoxication episodes. Although these findings suggest a causal connection, several questions remained to be answered. If the mushroom is to blame, then, why intoxication occurred this year only,



and why only a small percentage of hemodialysis patients who took the mushroom turned to be symptomatic? What was the nature of this toxin, and how did it cause the encephalopathy?

Despite the fact that wild mushrooms are collected and consumed, becoming poisoned is still a probability. It is well-known that even some experts have difficulty in discriminating between the mushrooms. The extremely wet and hot regional weather in 2004 may have boosted the growth of some poisonous species that are morphologically indistinguishable from the benign mushrooms. In fact, an exceptionally good harvest of Sugihiratake was reported in the affected areas, suggesting that somewhat change in property of the mushroom occurred in this season. In addition, this may support that the amount of ingested mushroom was higher in 2004 than previous years. The toxic characteristics of a mushroom may vary from region to another depending on the soil type [5], and the individual response to certain fungal toxins is also very variable [6]. Taken together, these factors may explain why only limited number of hemodialysis patients were affected.

The connection of the encephalopathy to mushroom poisoning in our series was particularly difficult because of the extraordinary long latent period, and the lack of digestive and hepatic abnormalities, nonetheless, this possibility was worth considered after excluding the other common causes of encephalopathy in uremic patients. In orellanine-containing mushrooms, a latent period of 36 hours to 17 days postingestion is usual before the onset of symptoms, although the clinical symptoms are incompatible [7]. The presentation of mushroom poisoning generally depends on the species and the amount ingested. Encephalopathy was the hallmark of Sugihiratake mushroom poisoning. The fact that hemodialysis was inefficient in preventing the encephalopathy in our series is probably because the elimination capacity was overwhelmed by the amount ingested. Alternatively, it may suggest that the molecular size of the toxin was not small enough to dialyze or that the toxin could have passed the blood-brain barriers, and hence became inaccessible for dialysis. This may be an important contributing factor to its neurotoxic effect. The permeability of the

blood brain barrier may have been increased by chemical mediators and cytokines [8]. Experiments to identify and isolate the specific toxin in the mushroom are still undergoing.

In this report, we warn practicing nephrologists that mushroom poisoning must be considered when patients with chronic renal diseases present with conscious disturbances and unexplained neurologic symptoms.

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## 2 針刺し・切創発生時の 倫理的な対応

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### ▶ Summary and Keywords

- ①針刺し・切創防止のために必要な情報を、医療従事者の目線に立って伝達する。
- ②当事者に適切な対処が行えるよう、針刺し・切創直後のサポート体制を整える。
- ③曝露後の対応を行う者は、当事者が自己決定できるよう、情報提供とサポートを行う。
- ④感染性の有無を確認する検査を行う場合は、患者さんへのインフォームドコンセントを行う。
- ⑤当事者のプライバシーには十分に配慮する。
- ⑥報告システムにおける個人情報の取り扱いについて確認する。

■針刺し・切創防止の情報提供   ■サポート体制   ■プライバシーの保護  
■個人情報の取り扱い   ■インフォームドコンセント

### ▶▶ はじめに

2005年4月、個人情報保護法が施行されたことにより、医療中の「倫理的な配慮」について再認識する機会となっている。感染管理担当者も、医療関連感染を取り巻く倫理的な側面について認識し、業務を遂行することが重要である。

私たち医療従事者は「人々の健康の維持・増進を図ること」を使命としており、感染管理担当者は、「患者さんを感染から守る」という視点での使命を果たすばかりでなく、「医療従事者を職業感染から守る」という使命もある。その意味で、針刺し・切創発生時の対応は重要である。

表1に、私見ではあるが、針刺し・切創時の倫理的配慮についてのチェックポイントを示した。

自施設の現状を評価してみしてほしい。

今回は、「医療従事者を職業感染から守る」という側面から、針刺し・切創発生時の倫理的な対応について考えてみたい。

### ▶▶ 針刺し・切創防止のための 情報提供

針刺し・切創を防止するために、各施設でさまざまな取り組みがなされていると思われる。当施設では、EPINet™ (Exposure Prevention Information Network) 日本版の、集計・解析ソフトEpisys<sup>1)</sup>を活用し分析した結果、「リキャップ」「携帯用針廃棄容器の活用の不徹底」「安全機能付針の不適切な使用」が問題点であることが判

表1 針刺し・切創時の倫理的配慮についての  
チェックポイント

- ①自施設の医療従事者に対して、針刺し・切創を防止するための方法について、適切に情報提供しているか。
- ②針刺し・切創直後、当事者が曝露後の対応に専念できるサポート体制ができているか。
- ③曝露後の対応では自己決定が尊重されているか。
- ④感染症検査を依頼する際、患者さんへのインフォームドコンセントは十分に行われているか。
- ⑤プライバシーへの配慮がなされているか。
- ⑥報告システムにおける個人情報の取り扱いは適切か。

明し、それらを解決すべく取り組みを行ってきた。しかし、上記3つによる針刺し・切創は減らない現状にある。

特に、2004年に導入された安全機能付留置針は、操作方法を誤ったことによる医師の針刺し件数が増加した。導入時に各部署の責任者に説明が行われたが、個々の医師の理解を得るまでには至らなかったことが原因の一つと考えられた。そのため、各部署単位で、医師が多く集まる検討会などを利用し、使用方法の説明会を再度実施した。その説明会に参加してみると、実際に使用している医師が正しく使用方法を理解していないという事実が明らかになり、導入時の細やかな情報提供の重要性を認識した。

感染管理担当者は、医療従事者の針刺し・切創による感染リスクや防止方法について、種々の情報を得て学習することで認識している。しかし、学習する前はどうかであったらうか？筆者自身、それほど針刺し・切創に意識を向けていなかったように記憶している。その当時の自分と同じ状況

にある人々に理解してもらうには、針刺し・切創による感染リスクを正しく認識できるように、また、防止方法を適切に実施できるように、細やかな情報提供と訓練を行うことが重要である。感染管理担当者は、常に、実際に針などを扱う医療従事者の目線に立って対応していくことが求められる。

## ▶▶▶ 針刺し・切創直後のサポート

筆者は以前、針刺し・切創を起こした時、精神的に動揺しながらも業務を中断することができずに、そのまま仕事を続けていたという経験がある。しかしこの時、同僚が、筆者の仕事をほかのスタッフに分担し、曝露後の対応に専念できるよう配慮してくれた。それ以来、自分の周囲で針刺し・切創が発生した場合には、当事者がその対応に専念できるように配慮することを心がけるようになった。このような針刺し・切創発生直後のサポート体制は重要である<sup>2)</sup>。

サポートを行う者は、プライバシーに配慮しながら、適切に当事者が対処できるようアドバイスできる身近な存在であることが望ましく、その部署の感染対策担当者（リンクドクターやリンクナースなど）あるいは責任者が適任である。また、担当者が不在の場合や不明な点がある場合には、ICNやICDなどに相談できることも必要で、当事者が「いつでも対応してもらえる」という安心感を得られるような体制が望ましい。曝露後の対応は、その病原体によって対処方法が異なったり、公務災害補償や労務災害補償の申請を含めた院内

の届け出の方法に戸惑ったりするケースが多いため、専門的な知識を持った者が相談窓口となることが適切な対処につながる。

また、当事者を責めるのではなく、支援的態度で接することも重要である。責められることを恐れ報告しないでいることは、当事者に適切な曝露後の対処が行われないばかりでなく、報告を生かした防止方法の検討も行われないことから、施設全体にも不利益が生じる<sup>3)</sup>。

## ▶▶▶ 自己決定の尊重

針刺し・切創を起こした場合、当事者は自己決定を迫られる場面がある。HBVに曝露した場合は、HBIGという人血液由来の特定生物由来製品を接種するかどうか、HIVの場合は、予防内服を行うかどうか、などである。当事者の自己決定にあたり、対応者は、使用する薬剤の効果や安全性、副作用、相互作用、日常生活上の注意点などについて十分な情報提供を行う。当事者が納得したうえで、対処方法を自己決定できるように支援を行うことが重要である。

日ごろから、針刺し・切創が起こった場合に「自分だったらどうするか」を考えてもらったり、シミュレーションを行ったりする機会を持っておくと、自己決定がスムーズに行える。

## ▶▶▶ 患者さんへの インフォームドコンセント

汚染源の感染性が不明な場合は、当該患者さん

に依頼し、感染症の有無を明らかにするための抗体検査を実施する。その際、針刺し・切創が起こった事実、検査の必要性、検査の結果の通知方法、得られた情報（検査結果）の取り扱い、費用などについて十分な説明を行い、同意を得る。書式を用意しておく、患者さんに伝えるべき情報が不足しないであろう。

また、「検査は、陰性であることを確認し当事者が安心するための検査ではない」ということを認識しておかなくてはならない。陰性確認のための検査と認識していたために「感染性がある」と判明して慌てる場合が少なくない。抗体検査の結果が陰性でも患者さんに事実を説明するのはもちろんであるが、陽性であった場合は、新たな疾患の事実を伝えられる患者さんの精神面に配慮しながら説明し、その感染症に対する適切な医療が受けられるよう準備を行う。感染管理担当者は、検査を受ける患者さんへの対応について、あらかじめ確認を行っておくことが必要である。

## ▶▶▶ プライバシーの保護

針刺し・切創に対処する場合、プライバシーの保護については常に配慮する必要がある。私たち医療従事者は患者さんのプライバシーについては十分に配慮しているが、同じ医療従事者、自身のプライバシーに無頓着な場合がある。

相談を受ける場合に、スタッフの多く存在する場所に対応し、針刺し・切創が起こった事実が、必要以上のスタッフにまで知られてしまう場合がある。対応者は、この点に十分配慮し、相談には