

Fig. 2. Activity of HIV-1 *vif* genes derived from LTNPRs and PRs for AIDS. (A) Growth kinetics in H9 and A3.01 cells of various clones carrying *vif* sequence from the individuals. Cells were electrophorated, as previously described [23], with 10 μ g of proviral clones carrying the *vif* sequences indicated on the right, and RT production in the culture supernatants at intervals was determined, as previously described [24]. Selected samples are shown in the figure for clarity (all the data are presented in Table 1). The clones were constructed by insertion of the *vif* sequences derived from the individuals into *Sma*I and *Xba*I sites of pNL-SX (Fig. 1A). The *vif* sequences from the individuals are indicated by the individual no.-clone no. Sequences NLVif and NL Δ Vif (for details, see the legend to Fig. 1) were used as positive and negative controls, respectively. Mock, pUC19. (B) Ratio of active and inactive *vif* clones in LTNPRs and PRs for AIDS. The ratio of clones with active *vif* in each person versus viral RNA level in his/her plasma (10^4 copies per ml) [19] is plotted for all the clones examined (Table 1).

distinct growth phenotypes in H9 and A3.01 cells (Fig. 2A, Table 1).

The ratio of active *vif* clones (in both cell lines) to the total number of clones screened per individual is also shown in Table 1. When the ratio versus the viral load in plasma

(Table 1) was plotted, results as shown in Fig. 2B were obtained. LTNPRs enrolled in the present study had varied ratios of active *vif* clones, ranging from 0 to 1.0. The results from the majority of LTNPRs indicate no apparent relationship between presence or absence of functional *vif* and long-

term nonprogression of infection. Subject p5 was an exception and had inactive *vif* in all the clones screened. This finding suggested a possibility that a defect of *vif* function in p5 could be responsible for the long-term nonprogression. On the other hand, all PRs with high viral load (HIV RNA $>20 \times 10^4$ copies per ml) had high ratios (0.5 or >0.5) of active *vif* clones. We and others have reported that primary cultured PBMCs and macrophages are non or less permissive for HIV virions lacking *vif* [30–35]. Recently, molecular biological demonstration has shown that *vif* confers infectivity on virions in nonpermissive cells. Vif counteracts an endogenous cellular factor, APOBEC3G, that inhibits HIV-1 replication [36,37]. It is well anticipated that the activity of *vif* modulates HIV-1 replication in individuals infected with virus.

3.3. Conclusion

Table 1 summarizes the major data in our two papers (reference [7] and this article). In these two reports, *nef* and *vif* from the same individuals were systematically analyzed for their potential roles in the development of AIDS. The *nef* genes from LTNPRs were found to have lower activity to enhance viral infectivity than those from PRs. The results of our *nef* and *vif* studies have shown that all PRs with a high viral load have a high ratio of active genes, strongly suggesting active *vif* and *nef* are required for HIV-1 replication in individuals. The *vif* and *nef* genes, therefore, would influence the progression and outcome of the disease in infected individuals. This is consistent with the previous reports analyzing *vif* sequences in disease progression [11,12]. On the other hand, the active/inactive ratio of *vif* varied in the samples from LTNPRs (from 0 to 1.0), different from that of *nef*. Without active *nef*, infected individuals appeared not to develop ARC or AIDS. In tissue cultures, *nef* is critical for HIV replication in resting PBMC [38,39] but not in stimulated PBMC or macrophages [32,35]. The *vif* is known to be indispensable for virus replication in these cells [30–35]. Because *vif* is more essential in virus replication than *nef*, it is reasonable to assume that *vif*-inactive viruses cannot or hardly persist in individuals. Further study is necessary to draw a clear conclusion regarding the functional importance of *vif* for disease progression.

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References

- [1] O.J. Cohen, A.S. Fauci, Pathogenesis and medical aspects of HIV-1 infection, in: fourth ed, in: D.M. Knipe, P.M. Howley (Eds.), *Fields Virology*, 2, Lippincott Williams & Wilkins, Philadelphia, 2001, pp.2043–2094.
- [2] N.J. Deacon, A. Tsykin, A. Solomon, K. Smith, M. Ludford-Menting, D.J. Hooker, D.A. McPhee, A.L. Greenway, A. Ellett, C. Chatfield, V.A. Lawson, S. Crowe, A. Maerz, S. Sonza, J. Learmont, J.S. Sullivan, A. Cunningham, D. Dwyer, D. Dowton, J. Mills, Genomic structure of an attenuated quasi species of HIV-1 from a blood transfusion donor and recipients, *Science* 270 (1995) 988–991.
- [3] F. Kirchhoff, T.C. Greenough, D.B. Brettler, J.L. Sullivan, R.C. Desrosiers, Absence of intact *nef* sequences in a long-term survivor with nonprogressive HIV-1 infection, *New Engl. J. Med.* 332 (1995) 228–232.
- [4] J.C. Learmont, A.F. Gecey, J. Mills, L.J. Ashton, C.H. Raynes-Greenow, R.J. Garcia, W.B. Dyer, L. McIntyre, R.B. Oelrichs, D.I. Rhodes, N.J. Deacon, J.S. Sullivan, D.A. McPhee, S. Crowe, A.E. Solomon, C. Chatfield, S. Blasdall, H. Kuipers for the Sydney Blood Bank Cohort Research Group, Immunologic and virologic status after 14–18 years of infection with an attenuated strain of HIV-1—a report from the Sydney blood bank cohort, *New Engl. J. Med.* 340 (1999) 1715–1722.
- [5] D.I. Rhodes, L. Ashton, A. Solomon, A. Carr, D. Cooper, J. Kaldor, N. Deacon for the Australian Long-Term Nonprogressor Study Group, Characterization of three *nef*-defective human immunodeficiency virus type 1 strains associated with long-term nonprogression, *J. Virol.* 74 (2000) 10581–10588.
- [6] R. Salvi, A.R. Garbuglia, A.D. Caro, S. Pulciani, F. Montella, A. Benedetto, Grossly defective *nef* gene sequences in a human immunodeficiency virus type 1-seropositive long-term nonprogressor, *J. Virol.* 72 (1998) 3646–3657.
- [7] M. Tobiume, M. Takahoko, T. Yamada, M. Tatsumi, A. Iwamoto, M. Matsuda, Inefficient enhancement of viral infectivity and CD4 downregulation by human immunodeficiency virus type 1 Nef from Japanese long-term nonprogressors, *J. Virol.* 76 (2002) 5959–5965.
- [8] R.C. Desrosiers, J.D. Lifson, J.S. Gibbs, S.C. Czajak, A.Y.M. Howe, L.O. Arthur, R.P. Johnson, Identification of highly attenuated mutants of simian immunodeficiency virus, *J. Virol.* 72 (1998) 1431–1437.
- [9] H.W. Kestler III, D.J. Ringler, K. Mori, D.L. Panicali, P.K. Sehgal, M.D. Daniel, R.C. Desrosiers, Importance of the *nef* gene for maintenance of high virus loads and for the development of AIDS, *Cell* 65 (1991) 651–662.
- [10] J.S. Gibbs, D.A. Regier, R.C. Desrosiers, Construction and in vitro properties of SIVmac mutants with deletions in "nonessential" genes, *AIDS Res. Hum. Retroviruses* 10 (1994) 607–616.
- [11] G. Hassaine, I. Agostini, D. Candotti, G. Bessou, M. Caballero, H. Agut, B. Autran, Y. Barthalay, R. Vigne, the French ATL study group, Characterization of human immunodeficiency virus type 1 *vif* gene in long-term asymptomatic individuals, *Virology* 276 (2000) 169–180.
- [12] L. Alexander, M.J. Aquino-DeJesus, M. Chan, W.A. Andiman, Inhibition of human immunodeficiency virus type 1 (HIV-1) replication by a two-amino-acid insertion in HIV-1 Vif from a nonprogressing mother and child, *J. Virol.* 76 (2002) 10533–10539.
- [13] S. Bour, K. Strebel, HIV accessory proteins: multifunctional components of a complex system, *Adv. Pharmacol.* 48 (2000) 75–120.
- [14] B.R. Cullen, HIV-1 auxiliary proteins: making connections in a dying cell, *Cell* 93 (1998) 685–692.
- [15] M. Emerman, M.H. Malim, HIV-1 regulatory/accessory genes: keys to unraveling viral and host cell biology, *Science* 280 (1998) 1880–1884.
- [16] R. Inubushi, M. Tamaki, R. Shimano, A.H. Koyama, H. Akari, A. Adachi, Functional roles of HIV accessory proteins for viral replication, *Int. J. Mol. Med.* 2 (1998) 429–433.

- [17] M. Fujita, A. Sakurai, N. Doi, M. Miyaura, A. Yoshida, K. Sakai, A. Adachi, Analysis of the cell-dependent replication potentials of human immunodeficiency virus type 1 *vif* mutants, *Microbes Infect.* 3 (2001) 1093–1099.
- [18] M. Fujita, A. Sakurai, A. Yoshida, S. Matsumoto, M. Miyaura, A. Adachi, Subtle mutations in the cysteine region of HIV-1 Vif drastically alter the viral replication phenotype, *Microbes Infect.* 4 (2002) 621–624.
- [19] T. Yamada, A. Iwamoto, Expression of a novel Nef epitope on the surface of HIV type 1-infected cells, *AIDS Res. Hum. Retroviruses* 15 (1999) 1001–1009.
- [20] T. Yamada, A. Iwamoto, Comparison of proviral accessory genes between long-term nonprogressors and progressors of human immunodeficiency virus type 1 infection, *Arch. Virol.* 145 (2000) 1021–1027.
- [21] T. Folks, S. Benn, A. Rabson, T. Theodore, M.D. Hoggan, M. Martin, M. Lightfoote, K. Sell, Characterization of a continuous T-cell line susceptible to the cytopathic effects of the acquired immune deficiency syndrome (AIDS)-associated retrovirus, *Proc. Natl. Acad. Sci. USA* 82 (1985) 4539–4543.
- [22] J.S. Lebkowski, S. Clancy, M.P. Calos, Simian virus 40 replication in adenovirus-transformed human cells antagonizes gene expression, *Nature* 317 (1985) 169–171.
- [23] A. Adachi, H.E. Gendelman, S. Koenig, T. Folks, R. Willey, A. Rabson, M.A. Martin, Production of acquired immunodeficiency syndrome-associated retrovirus in human and nonhuman cells transfected with an infectious molecular clone, *J. Virol.* 59 (1986) 284–291.
- [24] R.L. Willey, D.H. Smith, L.A. Lasky, T.S. Theodore, P.L. Earl, B. Moss, D.J. Capon, M.A. Martin, In vitro mutagenesis identifies a region within the envelope gene of the human immunodeficiency virus that is critical for infectivity, *J. Virol.* 62 (1988) 139–147.
- [25] H. Akari, T. Fukumori, A. Adachi, Cell-dependent requirement of human immunodeficiency virus type 1 gp41 cytoplasmic tail for Env incorporation into virions, *J. Virol.* 74 (2000) 4891–4893.
- [26] A. Adachi, N. Ono, H. Sakai, K. Ogawa, R. Shibata, T. Kiyomasu, H. Masuike, S. Ueda, Generation and characterization of the human immunodeficiency virus type 1 mutants, *Arch. Virol.* 117 (1991) 45–58.
- [27] H. Sakai, R. Shibata, J.I. Sakuragi, S. Sakuragi, M. Kawamura, A. Adachi, Cell-dependent requirement of human immunodeficiency virus type 1 Vif protein for maturation of virus particles, *J. Virol.* 67 (1993) 1663–1666.
- [28] M. Fujita, S. Matsumoto, A. Sakurai, N. Doi, M. Miyaura, A. Yoshida, A. Adachi, Apparent lack of *trans*-dominant negative effects of various *vif* mutants on the replication of HIV-1, *Microbes Infect.* 4 (2002) 1203–1207.
- [29] C. Ochsenbauer, V. Bosch, I. Oelze, U. Wieland, Unimpaired function of a naturally occurring C terminally truncated *vif* gene product of human immunodeficiency virus type 1, *J. Gen. Virol.* 77 (1996) 1389–1395.
- [30] M. Fujita, A. Sakurai, A. Yoshida, M. Miyaura, A.H. Koyama, K. Sakai, A. Adachi, Amino acid residues 88 and 89 in the central hydrophilic region of human immunodeficiency virus type 1 Vif are critical for viral infectivity by enhancing the steady-state expression of Vif, *J. Virol.* 77 (2003) 1626–1632.
- [31] D.H. Gabuzda, K. Lawrence, E. Langhoff, E. Terwilliger, T. Dorfman, W.A. Haseltine, J. Sodroski, Role of *vif* in replication of human immunodeficiency virus type 1 in CD4⁺ T lymphocytes, *J. Virol.* 66 (1992) 6489–6495.
- [32] M. Kawamura, T. Ishizaki, A. Ishimoto, T. Shioda, T. Kitamura, A. Adachi, Growth ability of human immunodeficiency virus type 1 auxiliary gene mutants in primary blood macrophage cultures, *J. Gen. Virol.* 75 (1994) 2427–2431.
- [33] U. Schwedler, J. Song, C. Aiken, D. Trono, *vif* is crucial for human immunodeficiency virus type 1 proviral DNA synthesis in infected cells, *J. Virol.* 67 (1993) 4945–4955.
- [34] J.H.M. Simon, M.H. Malim, The human immunodeficiency virus type 1 Vif protein modulates the postpenetration stability of viral nucleoprotein complexes, *J. Virol.* 70 (1996) 5297–5305.
- [35] K. Tokunaga, A. Ishimoto, K. Ikuta, A. Adachi, Growth ability of auxiliary gene mutants of human immunodeficiency virus types 1 and 2 in unstimulated peripheral blood mononuclear cells, *Arch. Virol.* 142 (1997) 177–181.
- [36] B.R. Cullen, HIV-1 Vif: counteracting innate antiretroviral defenses, *Mol. Ther.* 8 (2003) 525–527.
- [37] A.M. Sheehy, N.C. Gaddis, J.D. Choi, M.H. Malim, Isolation of a human gene that inhibits HIV-1 infection and is suppressed by the viral Vif protein, *Nature* 418 (2002) 646–650.
- [38] M.D. Miller, M.T. Warmerdam, I. Gaston, W.C. Greene, M.B. Feinberg, The human immunodeficiency virus-1 *nef* gene product: a positive factor for viral infection and replication in primary lymphocytes and macrophages, *J. Exp. Med.* 179 (1994) 101–113.
- [39] C.A. Spina, T.J. Kwok, M.Y. Chow, J.C. Guatelli, D.D. Richman, The importance of *nef* in the induction of human immunodeficiency virus type 1 replication from primary quiescent CD4 lymphocytes, *J. Exp. Med.* 179 (1994) 115–123.

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Unrelated cord blood transplantation for a human immunodeficiency virus-1-seropositive patient with acute lymphoblastic leukemia

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The concurrent use of highly active antiretroviral therapy (HAART) improves results of high-dose chemotherapy with autologous stem cell transplantation (SCT) for human immunodeficiency virus-1 (HIV)-associated lymphomas.¹ Recently, successful allogeneic SCT from HLA-matched sibling donors was reported in HIV-infected patients.^{2–4} Here, we describe the first case of an HIV-infected patient with acute lymphoblastic leukemia (ALL) who underwent umbilical cord blood transplantation (CBT).

In July 1996, a 23-year-old Japanese woman presented with fever and genital herpes. She was confirmed as seropositive for HIV, probably transmitted from her boyfriend. In March 2001, a real-time quantitative polymerase chain reaction (PCR) analysis showed that the HIV-RNA level was elevated to 25 000 copies/ml (lower limit of detection, 50). The CD4 count decreased to 28/μl.

Therefore, HAART consisting of 60 mg stavudine, 300 mg lamivudine, and 600 mg efavirenz was initiated. In July 2001, the HIV-RNA level decreased to 220 copies/ml, and the CD4 count increased to 129/μl. In May 2003, her complete blood count tests showed a white blood cell count (WBC) of 3990/μl with 29% lymphoblasts. Bone marrow (BM) examination showed hypercellularity with 96% lymphoblasts, which were positive for CD4, CD10, CD13, CD19, CD33, CD34, and HLA-DR. Cytogenetic analysis disclosed the presence of t(9;22)(q34;q11) in 12 of 20 metaphases. The p190^{BCR-ABL} transcript was shown by a reverse transcriptase (RT)-PCR analysis. She was diagnosed as Philadelphia chromosome-positive ALL. She achieved hematological complete remission after two courses of chemotherapy. She has been taking HAART during and after the chemotherapy and her HIV-RNA level continued to be below detectable levels. She was negative for hepatitis B virus surface antigen and anti-hepatitis C virus antibody, and positive for anti-cytomegalovirus antibody. As she had no HLA-matched related or unrelated BM donors, the patient underwent CBT from an unrelated donor with mismatches at two loci (HLA-B and DR) in September 2003 (Figure 1). The numbers of total nucleated cells and CD34-positive cells in the cord

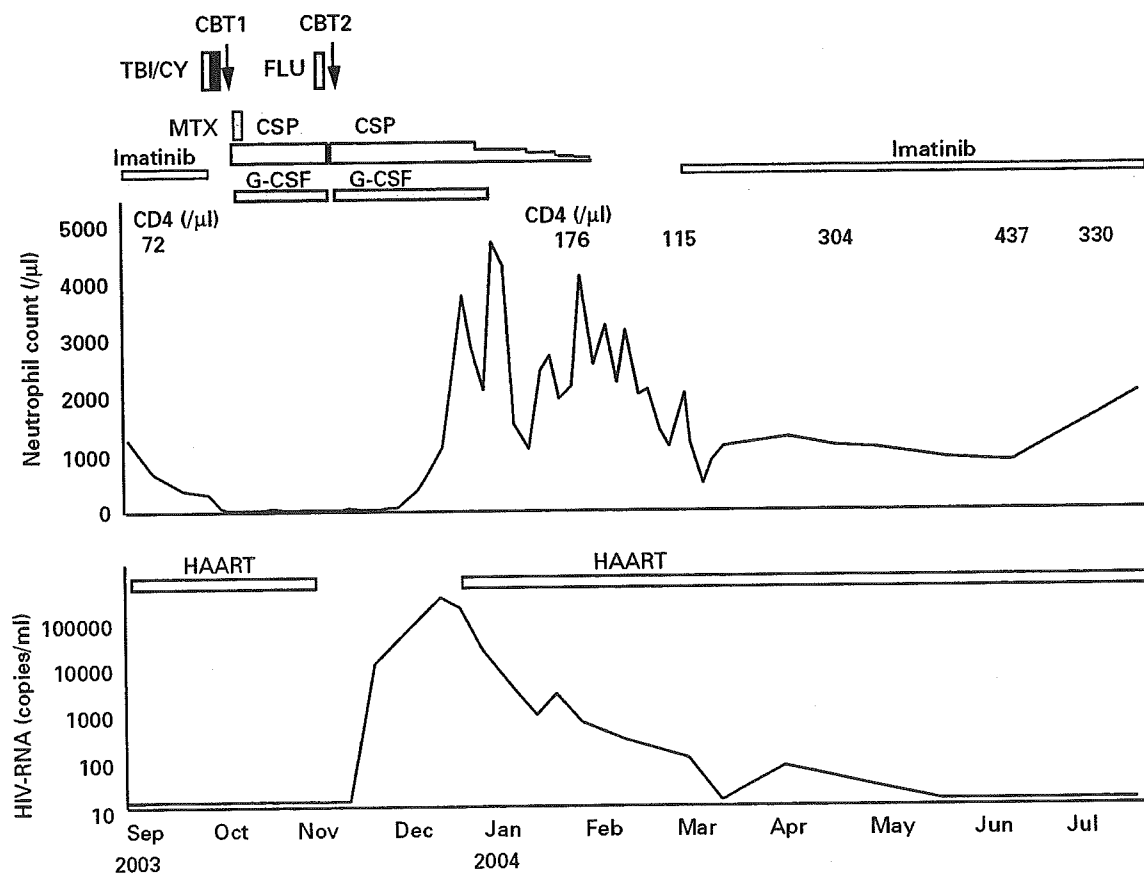


Figure 1 Clinical course of the patient.

blood (CB) unit were $2.9 \times 10^7/\text{kg}$ and $0.76 \times 10^5/\text{kg}$, respectively. The conditioning regimen included 12 Gy total body irradiation and 120 mg/kg cyclophosphamide. Graft-versus-host disease (GVHD) prophylaxis consisted of cyclosporine and methotrexate. The patient tolerated the procedure well with minimal regimen-related toxicity. Owing to possible myelosuppression, HAART was discontinued on day +28. On day +33, her WBC remained below $100/\mu\text{l}$ and all of the BM cells were shown to be derived from the recipient. At 40 days after the first CBT, second CBT was performed from an unrelated donor with a one-locus mismatch at HLA-DR. The numbers of total nucleated cells and CD34-positive cells in the CB unit were $2.1 \times 10^7/\text{kg}$ and $0.46 \times 10^5/\text{kg}$, respectively. The conditioning regimen included $40 \text{ mg}/\text{m}^2$ fludarabine for 3 days. Cyclosporine was administered for GVHD prophylaxis. A neutrophil count consistently greater than $500/\mu\text{l}$ was achieved on day +27. Full donor chimerism of BM cells was shown on day +28. The HIV-RNA level increased to 3×10^6 copies/ml on day +31. After the administration of HAART from day +38, the HIV-RNA levels returned to below detectable levels from day +195, and the CD4 count increased to above $300/\mu\text{l}$ from day +170. No bacterial or fungal infections were documented during the first and second CBT processes and cytomegalovirus reactivation was successfully treated with ganciclovir and foscarnet. Grade I acute GVHD occurred, but resolved without any additional immunosuppressants. No chronic GVHD was observed. An RT-PCR analysis showed continuous negative test results for the p190^{BCR-ABL} transcript until the last follow-up evaluation at 15 months post-CBT.

CBT for adults has been associated with a high rate of early transplantation-related mortality (TRM).^{5,6} However, our single-institution experience showed a 1-year TRM of 9% and 2-year disease-free survival of 74% in 68 adults after CBT.⁷ Both CB donors and the patient in the present study were Japanese. The lesser genetic diversity in a single ethnic population in our studies might be associated with the favorable outcomes of CBT, such as the lower rates of severe acute GVHD. Although our results suggest that CBT is feasible for HIV-infected patients on HAART, the safety and efficacy should be further examined by prospective studies.

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References

- 1 Krishnan A, Zaia J, Forman SJ. Should HIV-positive patients with lymphoma be offered stem cell transplants? *Bone Marrow Transplant* 2003; **32**: 741-748.
- 2 Schlegel P, Beatty P, Halvorsen R, McCune J. Successful allogeneic bone marrow transplant in an HIV-1-positive man with chronic myelogenous leukemia. *J Acquir Immune Defic Syndr* 2000; **24**: 289-290.
- 3 Sora F, Antinori A, Piccirillo N *et al.* Highly active antiretroviral therapy and allogeneic CD34(+) peripheral blood progenitor cells transplantation in an HIV/HCV coinfecting patient with acute myeloid leukemia. *Exp Hematol* 2002; **30**: 279-284.
- 4 Kang EM, de Witte M, Malech H *et al.* Nonmyeloablative conditioning followed by transplantation of genetically modified HLA-matched peripheral blood progenitor cells for hematologic malignancies in patients with acquired immunodeficiency syndrome. *Blood* 2002; **99**: 698-701.
- 5 Laughlin MJ, Eapen M, Rubinstein P *et al.* Outcomes after transplantation of cord blood or bone marrow from unrelated donors in adults with leukemia. *N Engl J Med* 2004; **351**: 2265-2275.
- 6 Rocha V, Labopin M, Sanz G *et al.* Transplants of umbilical-cord blood or bone marrow from unrelated donors in adults with acute leukemia. *N Engl J Med* 2004; **351**: 2276-2285.
- 7 Takahashi S, Iseki T, Ooi J *et al.* Single institute comparative analysis of unrelated bone marrow transplantation and cord blood transplantation for adult patients with hematological malignancies. *Blood* 2004; **104**: 3813-3820.



CASE REPORT

AIDS-related cerebral toxoplasmosis with hyperintense foci on T1-weighted MR images: A case report

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KEYWORDS

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Summary The neuroradiological findings are helpful for the diagnosis of toxoplasmic encephalitis. The T1 hypersignal intensity foci on brain magnetic resonance (MR) images without contrast enhancement are presented and can be a pathognomonic sign of this disease.

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Introduction

Most toxoplasmic encephalitis is opportunistic infection complicated with the acquired immunodeficiency syndrome (AIDS) and immunosuppressive conditions. The diagnosis of this disease is difficult

because of the incompetence of the serological examination for the immunocompromised patients.¹ Although the direct detection method for the pathogen by polymerase chain reaction (PCR) using the cerebrospinal fluid (CSF) has high specificity, the sensitivity of this method is insufficient for definitive diagnosis.² We, therefore, have to synthetically diagnose with clinical symptoms, signs, laboratory data, neuroradiological images and the response to anti-toxoplasmosis therapy.

We report here our experience of a unique MR imaging finding of toxoplasmic encephalitis in an

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AIDS patient and emphasize the hyperintense foci on T1-weighted MR images that can be one of the pathognomonic MR images of this disease.

Case report

A 44-year-old man with disturbance of consciousness and respiratory insufficiency was admitted to our hospital in April 2005. His consciousness had been rapidly deteriorated and he developed coma 2–3 days before hospitalization. Serological tests of HIV antibodies and *Toxoplasma gondii* IgG antibody were positive, but the *T. gondii* IgM antibody was not detected. The concentration of HIV RNA in plasma was 120,000 copies/ml and the CD 4 cell count was 8 mm³. The chest X-ray showed bilateral ground glass shadow and *Pneumocystis jiroveci* (carinii) was detected from bronchoalveolar lavage (BAL) fluid. CSF showed mild elevated protein level of 65 mg/dl and pleocytosis, and the opening pressure was over 300 mmH₂O. No malignant cells or microorganisms were detected. *T. gondii* B1-gene fragment was detected by PCR using CSF, therefore, the diagnosis of an AIDS case with toxoplasmic encephalitis was made.³

MRI of the brain showed multiple high intensity lesions on T2-weighted image (Fig. 1a) and the corresponding T1-weighted image showed low intensity lesions. Contrast enhanced T1-weighted images showed multiple nodular and ring enhancement lesions.

The chemotherapy with trimethoprim/sulfamethoxazole (TMP/SMX) was very effective and the patient's consciousness level was improved gradually. *P. jiroveci* pneumonia was also cured. MR imaging after 4 weeks of treatment demonstrated that the multiple nodular lesions on T1 and T2-weighted images had significantly been reduced. After 8 weeks of treatment, the contrast enhanced T1-weighted images showed only residual small lesions without contrast enhancement. Interestingly, the hypersignal intensity foci appeared at bilateral basal ganglia obviously after 2 weeks of treatment on the non-enhanced T1-weighted images (Fig. 1b). Corresponding computed tomography (CT) image did not show hemorrhagic or calcified densities (Fig. 1c). These T1 hypersignal intensity foci regressed gradually along with anti-toxoplasmic chemotherapy in proportion to other mass lesions. The T2* (star)-weighted image, which can detect the hemosiderin deposition as hypointensity lesion, operated after 12 weeks of treatment showed no hypointensity at corresponding T1 hypersignal intensity foci on basal ganglia (Fig. 1d).⁴ We concluded that the toxoplasmic

encephalitis showed the hypersignal intensity foci on T1-weighted MR imaging without hemorrhage or calcification.

Discussion

Toxoplasmic encephalitis progresses rapidly and is life threatening to immunocompromised patients. Therefore, we often have to start the anti-toxoplasmosis therapy when this encephalitis is suspected on the neuroradiologic images and laboratory data. Typically, the toxoplasmic encephalitis lesions on MRI studies appear as T2 hypersignal intensity foci and T1 hypo-isosignal intensity foci, and reveal a rim of enhancement surrounding the edema on contrast enhanced T1-weighted images. Nevertheless, even characteristic foci on these MR imagings are not pathognomonic. Since the differential diagnosis of toxoplasmic encephalitis from other infections or CNS lymphoma is difficult, improvement in the diagnostic methods is an urgent necessity.

In our case, the toxoplasmic encephalitis was diagnosed with the highly specific PCR and confirmed by the response to anti-toxoplasmosis therapy. Brain MRI revealed unusual findings, T1 hypersignal intensity foci, accompanied by typical multiple high intense lesions on T2-weighted image during the treatment. These unique MR findings have been reported on only a few cases of non-HIV/AIDS-related toxoplasmic encephalitis. Terada et al.⁵ reported a case of toxoplasmic encephalitis after stem cell transplantation with T1 hypersignal intensity foci. Autopsy revealed the disseminated toxoplasmosis, and coagulative necrosis without hemorrhage or calcification was revealed at corresponding T1 hypersignal intensity foci by neuropathological study. In another post-bone marrow transplantation case, inflammatory and vascular changes without hemorrhage appeared to be the cause of iso or hypersignal intensity rings by the stereotactic biopsy of T1 hypersignal intensity foci.⁶ On the other hand, Navia et al.⁷ demonstrated that the T1 hypersignal intensity foci were caused by coagulative necrosis with lipid-laden macrophages. The pathophysiological and neuroradiological mechanisms to create these MRI findings are far from clear yet. The reason why the T1 hypersignal intensity foci tend to localize in the basal ganglia is not clear either.^{5,6}

CNS lymphoma, which is important for the distinction from toxoplasmic encephalitis, shows T1 hypo-isosignal intensity foci and never shows T1 hypersignal intensity foci except subacute

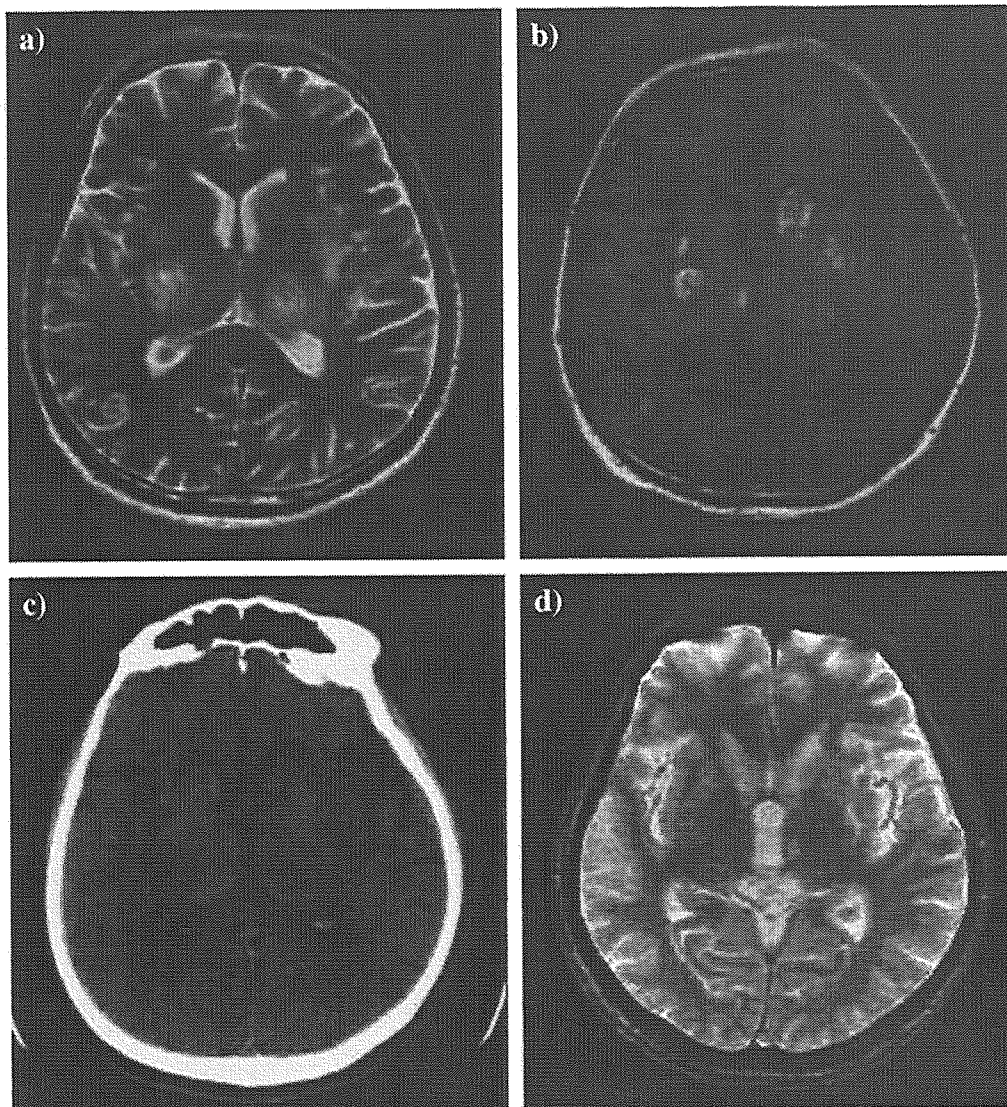


Figure 1 (a) The T2-weighted magnetic resonance image presented multiple high intense lesions. (b) Non-enhanced T1-weighted image showed hypersignal intensity foci at bilateral basal ganglia. (c) The corresponding CT image showed non-hemorrhagic or non-calcified density. (d) The T2* (star)-weighted image showed non-hemorrhagic observations at basal ganglia.

hemorrhage with hypervascular CNS lymphoma.^{8,9} However, the CT imaging and T2* (star)-weighted MR imaging can simply distinguish it from the toxoplasmic T1 hypersignal intensity foci without hemorrhage or calcification.

We reported here the unique MRI findings, T1 hypersignal intensity foci, without hemorrhage or calcification on HIV/AIDS-related toxoplasmic encephalitis. It will be helpful for the diagnosis of toxoplasmic encephalitis and may be a pathognomonic finding. Unfortunately, we have not experienced another case of toxoplasmic encephalitis after this case, but we would like to continue to explore this unique MRI finding of this disease.

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References

1. Luft BJ, Brooks RG, Conley FK, McCabe RE, Remington JS. Toxoplasmic encephalitis in patients with acquired immune deficiency syndrome. *JAMA* 1984;252:913-7.

2. Cingolani A, De Luca A, Ammassari A, Murri R, Linzalone A, Grillo R, et al. PCR detection of *Toxoplasma gondii* DNA in CSF for the differential diagnosis of AIDS-related focal brain lesions. *J Med Microbiol* 1996;45:472–6.
3. Castro KG, Ward JW, Slutsker L, Buehler JW, Jaffe HW, Ruth L, et al. 1993 revised classification system for HIV infection and expanded surveillance case definition for aids among adolescents and adults. *Morb Mortal Wkly Rep* 1992; 41:1–19.
4. Bulte JW, Kraitchman DL. Iron oxide MR contrast agents for molecular and cellular imaging. *NMR Biomed* 2004;17: 484–99.
5. Terada H, Kamata N, Yokoyama Y, Ohashi K, Akiyama H, Sakamaki H. T1-hypersignal foci in cerebral toxoplasmosis. *Riv Neuroradiol* 2001;14:665–7 [Case report].
6. Dietrich U, Maschke M, Dorfler A, Prumbaum M, Forsting M. MRI of intracranial toxoplasmosis after bone marrow transplantation. *Neuroradiology* 2000;42:14–8.
7. Navia BA, Petito CK, Gold JW, Cho ES, Jordan BD, Price RW. Cerebral toxoplasmosis complicating the acquired immune deficiency syndrome: clinical and neuropathological findings in 27 patients. *Ann Neurol* 1986;19:224–38.
8. Jenkins CN, Colquhoun IR. Characterization of primary intracranial lymphoma by computed tomography: an analysis of 36 cases and a review of the literature with particular reference to calcification haemorrhage and cyst formation. *Clin Radiol* 1998;53:428–34 [Review].
9. Rubenstein J, Fischbein N, Aldape K, Burton E, Shuman M. Hemorrhage and VEGF expression in a case of primary CNS lymphoma. *J Neurooncol* 2002;58:53–6.

Effects of human interleukin 7 on HIV-1 replication in monocyte-derived human macrophages

Running head: human interleukin 7 and HIV-1 replication in monocyte derived human macrophages.

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Summary

Interleukin 7 (IL-7) contributes to development and proliferation of T cells. We investigated the effect of IL-7 on HIV-1 infected monocyte-derived human macrophages. IL-7 treatment for macrophages at a concentration of 10ng/ml reduced replication of the R5 HIV-1 strain by approximately 50%. Meanwhile, HIV-1-infected macrophages themselves could excrete IL-7 approximately 20% more than uninfected macrophages. These results suggested the advantage of IL-7 as a therapeutic modality to recover CD4+ T cells.

Text

Macrophages and CD4+ T cells are the predominant cell types for human immunodeficiency virus-1 (HIV-1) infection [1,14]. Macrophages can be infected by HIV-1 in many tissues, including the brain, lung and lymph nodes [16]. Unlike infected CD4+ T cells, which have a short half-life of 1-1.5 days, macrophages are quite resistant to the cytopathic effect of the virus, and may thus provide a reservoir for persistent infection and virus dissemination. It is known that cytokines play an important role in HIV-1 infection. Interleukin 7 is a cytokine produced by stromal cells of the thymus and bone marrow [2-4]. It has capacity to induce growth of immature B lymphocytes [5], and contributes to development and proliferation of T cells [6-9]. With respect to HIV-1 infection, there is a reverse correlation between CD4+ T cell number and IL-7 serum levels in HIV-1-infected patients [17-19]. After patients started to receive antiretroviral drugs, the elevated IL-7 in the serum decreased to normal levels [17]. On the other hand, IL-7 increases HIV-1 replication in thymic organ cultures [10-12] and to induce latent HIV-1 in resting CD4+

T cells [13]. However, effects of IL-7 on HIV-1 replication in macrophages remain unclear. In the present study, we evaluated levels of HIV-1 proliferation in monocyte-derived human macrophages treated with or without exogenous IL-7.

Peripheral blood mononuclear cells (PBMC) from blood buffy coats of healthy donors were isolated by centrifugation through Ficoll-Hypaque and plated on a 24 well MULTIWELL™ PRIMARIA™ plate (Becton Dickinson, Franklin Lakes, New Jersey, USA) with RPMI 1640 supplemented with 10% fetal calf serum (FCS). After incubation at 37°C for one day, the floating cells were removed by washing the plate with phosphate-buffered saline four times and the adherent cells were incubated at 37°C for 11 days with 0.5ml of RPMI 1640 supplemented with 10% FCS plus 100ng/ml of granulocyte macrophage colony stimulating factor (GM-CSF).

To determine whether or not differentiated macrophages express the IL-7 receptor, we analyzed total RNA extracted from macrophages. PBMC stimulated with PHA and 10ng/ml of IL-2 for three days and monkey kidney CV-1 cells served as positive and negative controls, respectively. RNA was reverse-transcribed into cDNA, and levels of IL-7 receptor cDNA were measured by an ABI 7500 Real-time PCR System. As shown in Fig. 1a, IL-7 receptor mRNA in macrophages was clearly detected. It was also detected in PHA and IL-2-stimulated PBMC. Levels of IL-7 receptor mRNA in macrophages were approximately 1/10 of those in PHA and IL-2-stimulated PBMC, while CV-1 cells were totally negative for expression of the IL-7 receptor.

Macrophages were infected with 6.5ng of p24 of HIV-1 SF162 strain for two hours on day 12. Cells were then incubated with 1ml of RPMI 1640 supplemented with 10% FCS plus 100ng/ml of GM-CSF

and 0, 3 or 10ng/ml of IL-7 (Techne, Minneapolis, Minnesota, USA). Infected macrophages were fed on days 3, 6, 9 and 12 (after infection) by exchanging one half of the culture supernatant with fresh media containing the same amount of cytokines. Levels of HIV-1 p24 antigen in culture supernatants on days 1, 3, 6, 9 and 12 were measured by using an HIV-1 P24 Antigen ELISA Kit (Fig. 1b). The p24 antigen level in the culture supernatant of macrophages treated with 3ng/ml of IL-7 was apparently less than that of macrophages without IL-7 treatment (Fig. 1b). This difference was evident on day 3 and became greater with time. On day 12, the p24 level of macrophages treated with 3ng/ml of IL-7 was 2220 pg/ml while that of untreated macrophages was 3542 pg/ml. That is, the suppression with 3ng/ml of IL-7 on HIV-1 replication was 37.4% on day 12. Treatment of macrophages with 10ng/ml of IL-7 showed a greater suppressive effect on HIV-1 replication (Fig. 1b). On day 12, the p24 level of macrophages treated with 10ng/ml of IL-7 was 1760 pg/ml. That is, suppression with 10ng/ml of IL-7 on HIV-1 replication was 50.3% on day 12.

Monocyte-derived human macrophages were also infected with the Z strain of the Sendai virus. Hemagglutination assay (HA) titers were measured on days 1, 3 and 6 after infection. As shown in Fig. 1c, there was no difference in HA titers between IL-7-treated and untreated macrophages, indicating no apparent effect of IL-7 on Sendai virus replication in macrophages.

Levels of IL-7 in supernatants of macrophages were measured on days 1 and 3 after HIV-1 infection (Fig. 1d). On day 1, there was no difference in levels of IL-7 between HIV-1 infected and uninfected macrophages. On day 3, IL-7 levels in HIV-1-infected macrophages slightly increased, while those in uninfected macrophages did not.

The mechanisms of the suppressive effect of IL-7 on R5 HIV-1 replication in macrophages are still not clear. A previous study showed that IL-7 treatment for PBMC enhanced excretion of β chemokines including MIP-1 β , which can suppress R5 HIV-1 replication [15]. We pretreated macrophages with 10ng/ml of IL-7 for one day, and then inoculated them with HIV-1. However, we failed to detect any further suppression of HIV-1 replication compared with macrophages treated with IL-7 only after HIV-1 infection (data not shown). Therefore, it is unlikely that IL-7 could suppress HIV-1 replication by up regulation of MIP-1 β . It is possible that IL-7 could affect HIV-1 replication after HIV-1 enters macrophages.

We have shown that HIV-1-infected macrophages could excrete more IL-7 than uninfected ones. Therefore, it is likely that IL-7 is a self-defense system for macrophages against HIV-1 infection. Furthermore, our finding that IL-7 could moderately suppress HIV-1 replication in macrophages suggested the advantage of IL-7 as an immune modulator which could be used to recover CD4+ T cell numbers in HIV-1-infected individuals.

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References

1. Collman R, Hassan NF, Walker R, Godfrey B, Cutilli J, Hastings JC, *et al.* Infection of monocyte-derived macrophages with human immunodeficiency virus type 1 (HIV-1). Monocyte-tropic and lymphocyte-tropic strains of HIV-1 show distinctive patterns of replication in a panel of cell types. *J Exp Med* 1989; 170:1149-1163.
2. Heufler C, Topar G, Grasseger A, Stanzl U, Koch F, Romani N, *et al.* Interleukin 7 is produced by murine and human keratinocytes. *J Exp Med* 1993; 178:1109-1114.
3. Sudo T, Nishikawa S, Ohno N, Akiyama N, Tamakoshi M, Yoshida H. Expression and function of the interleukin 7 receptor in murine lymphocytes. *Proc Natl Acad Sci USA* 1993; 90:9125-9129.
4. Wolf SS, Cohen A. Expression of cytokines and their receptors by human thymocytes and thymic stromal cells. *Immunology* 1992; 77:362-368.
5. Namen AE, Lupton S, Hjerrild K, Wignall J, Mochizuki DY, Schmierer A, *et al.* Stimulation of B-cell progenitors by cloned murine interleukin-7. *Nature* 1988; 333:571-573.
6. Fry TJ, Connick E, Falloon J, Lederman MM, Liewehr DJ, Spritzler J, *et al.* A potential role for interleukin-7 in T-cell homeostasis. *Blood* 2001; 97:2983-2990.
7. Grabstein KH, Namen AE, Shanebeck K, Voice RF, Reed SG, Widmer MB. Regulation of T cell proliferation by IL-7. *J Immunol* 1990; 144:3015-3020.
8. Plum J, De Smedt M, Leclercq G, Verhasselt B, Vandekerckhove B. Interleukin-7 is a critical growth factor in early human T-cell development. *Blood* 1996; 88:4239-4245.
9. Schluns KS, Kieper WC, Jameson SC, Lefrancois L. Interleukin-7 mediates the homeostasis of naive and memory

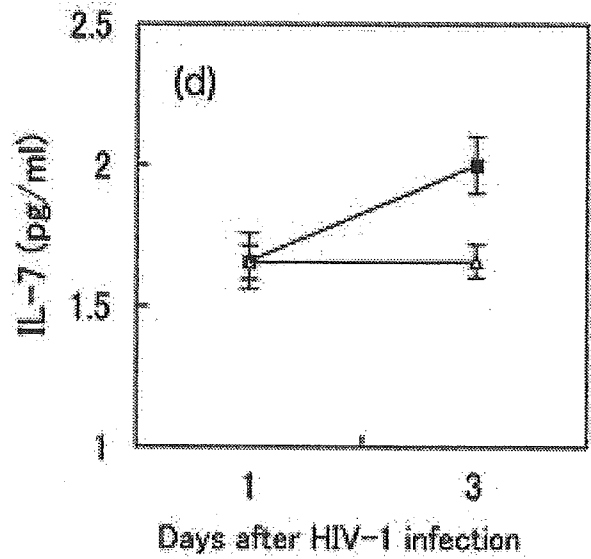
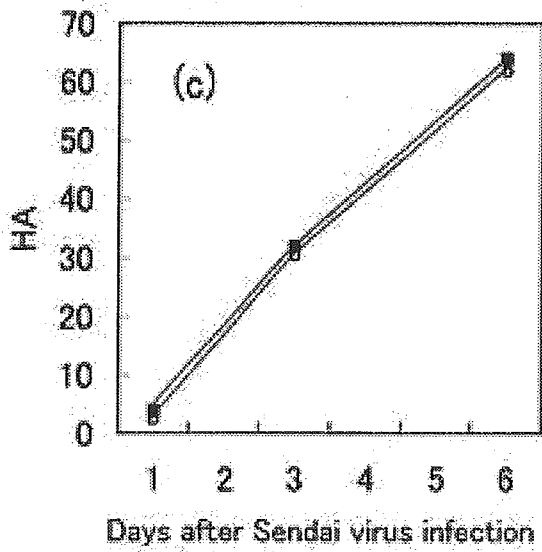
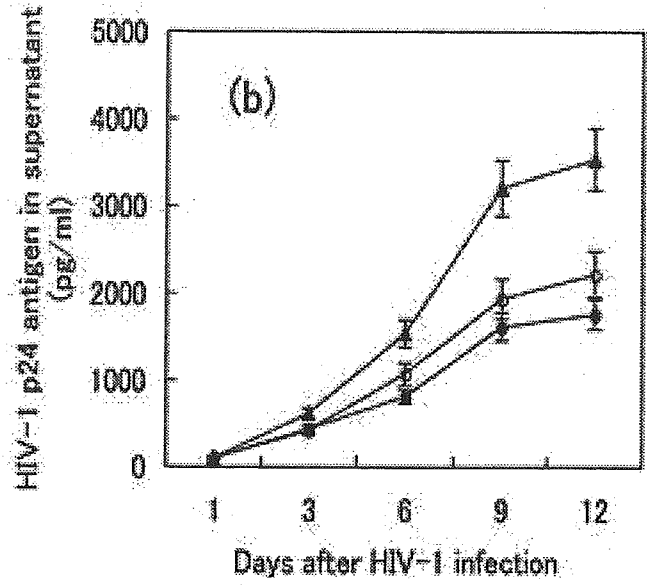
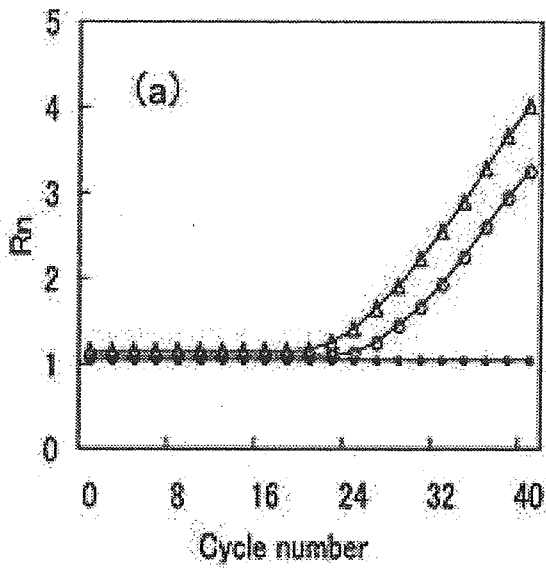
- CD8 T cells in vivo. *Nat Immunol* 2000; 1:426-432.
10. Uittenbogaart CH, Boscardin WJ, Anisman-Posner DJ, Koka PS, Bristol G, Zack JA. Effect of cytokines on HIV-induced depletion of thymocytes in vivo. *AIDS* 2000; 14:1317-1325.
 11. Chene L, Nugeyre MT, Barre-Sinoussi F, Israel N. High-level replication of human immunodeficiency virus in thymocytes requires NF-kappaB activation through interaction with thymic epithelial cells. *J Virol* 1999; 73:2064-2073.
 12. Schmitt N, Chene L, Boutolleau D, Nugeyre MT, Guillemard E, Versmisse P, *et al.* Positive regulation of CXCR4 expression and signaling by interleukin-7 in CD4+ mature thymocytes correlates with their capacity to favor human immunodeficiency X4 virus replication. *J Virol* 2003; 77:5784-5793.
 13. Wang FX, Xu Y, Sullivan J, Souder E, Argyris EG, Acheampong EA, *et al.* IL-7 is a potent and proviral strain-specific inducer of latent HIV-1 cellular reservoirs of infected individuals on virally suppressive HAART. *J Clin Invest* 2005; 115:128-137.
 14. Ho DD, Neumann AU, Perelson AS, Chen W, Leonard JM, Markowitz M. Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection. *Nature* 1995; 373:123-126.
 15. Llano A, Barretina J, Gutierrez A, Clotet B, Este JA. Interleukin-7-dependent production of RANTES that correlates with human immunodeficiency virus disease progression. *J Virol* 2003; 77:4389-4395.
 16. Koenig S, Gendelman HE, Orenstein JM, Dal Canto MC, Pezeshkpour GH, Yungbluth M *et al.* Detection of AIDS virus in macrophages in brain tissue from AIDS patients with encephalopathy. *Science* 1986; 233:1089-1093.
 17. Llano A, Barretina J, Gutierrez A, Blanco J, Cabrera C,

- Clotet B. Interleukin-7 in plasma correlates with CD4 T-cell depletion and may be associated with emergence of syncytium-inducing variants in human immunodeficiency virus type 1-positive individuals. *J Virol* 2001; 75:10319-10325.
18. Beq S, Rannou MT, Fontanet A, Delfraissy JF, Theze J, Colle JH. HIV infection: pre-highly active antiretroviral therapy IL-7 plasma levels correlate with long-term CD4 cell count increase after treatment. *AIDS* 2004; 18:563-565.
19. Kopka J, Mecikovsky D, Aulicino PC, Mangano AM, Rocco CA, Bologna R. High IL-7 plasma levels may induce and predict the emergence of HIV-1 virulent strains in pediatric infection. *J Clin Virol* 2005; 33:237-242.

FIG. 1. (a) Expression of IL-7 receptor mRNA in macrophages measured by real-time monitoring of fluorescence signals. IL-7 receptor mRNA in macrophages (open circles), PHA-stimulated PBMC (open triangles) as the positive control and CV-1 cells (closed squares) as the negative control are shown. Rn is the ratio of the fluorescence intensity of the target cDNA to the fluorescence intensity of water instead of cDNA. Data shown are representative of two independent experiments with similar results. (b) The HIV-1 p24 antigen in the supernatant of HIV-1 infected macrophages treated with 0 (closed triangles), 3 (open circles) or 10ng/ml (closed circles) of IL-7 was measured on days 1, 3, 6, 9 and 12 after HIV-1 infection. Data show the mean of duplicate samples from the same blood donor with bars indicating the actual fluctuation between two individual measurements. Data shown are the representative of three independent experiments with similar results. (c) Growth of the Sendai virus in monocyte-derived

macrophages treated with 10ng/ml of IL-7 (open circles) or un-treated with IL-7 (closed squares). HA titers in culture supernatants were measured on days 1, 3 and 6 after infection. Data shown are representative of two independent experiments with similar results. (d) Levels of IL-7 in the supernatants of HIV-1 infected macrophages (closed squares) or uninfected macrophages (open triangles) were measured on days 1 and 3 after HIV-1 infection. Data show the mean of duplicate samples from the same blood donor with bars indicating the actual fluctuation between two individual measurements. Data shown are representative of two independent experiments with similar results.

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Protective Effects of *IL4-589T* and *RANTES-28G* on HIV-1 disease progression in infected Thai females

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Objective: To evaluate the effect of polymorphisms in interleukin-4 (*IL4*) and *RANTES* promoters on disease progression in HIV-1 infected Thais.

Design: Antiretroviral (ARV) drug-free HIV-1 infected females from the prospective cohort.

Methods: A total of 246 DNA samples were genotyped for *IL4* and *RANTES* promoter polymorphisms by PCR-RFLP. Associations of genotype with HIV-1 disease progression were assessed with respect to baseline clinical data including plasma HIV-1 load, CD4 cell counts, and proportion of symptomatic/AIDS, and survival status during 3 years of follow-up.

Results: Patients with homozygous *IL4-589T* allele showed a significantly lower HIV-1 viral load ($P = 0.005$) and a higher CD4 cell count ($P = 0.003$) than the other patients with heterozygous *IL4-589C/T* or homozygous *IL4-589C* allele. Kaplan–Meier analysis demonstrated an apparent but insignificant trend towards better survival in homozygous *IL4-589T* patients. On the other hand, patients with *RANTES-28G* allele showed a significantly better survival while those with *RANTES In1.1C* allele without *RANTES-28G* showed a significantly poorer survival compared with those who did not possess either *RANTES In1.1C* or *RANTES-28G* ($P = 0.02$), although those polymorphisms only weakly associated with baseline viral load and CD4 cell counts.

Conclusions: Our results implicate the significant protective effect of *IL4-589T* and *RANTES-28G* on HIV disease progression in Thais. In contrast, *RANTES In1.1C* without *RANTES-28G* had an accelerating effect on HIV disease progression.

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Keywords: *IL4-589T*, *RANTES* promoter polymorphisms, HIV disease progression, survival, Thailand, viral load, CD4 cells

Introduction

HIV-1 infected individuals have widely different rates of disease progression. Some infected individuals become symptomatic within 2–3 years while others remain asymptomatic for more than 10–15 years [1]. It is

important to investigate factors modulating rates of disease progression for designing novel therapies and vaccines.

RANTES is a natural CCR5 ligand and potently inhibits cell entry of HIV-1 that uses CCR5 as a coreceptor

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