

Fig. 5. Molecular hepatocarcinogenesis in HCV infection. Oxidative stress together with hepatic steatosis induced by the HCV core protein would play a pivotal role in the development of HCC. Alterations in cellular gene expressions, such as TNF- α or SOCS-1, and those in the intracellular signaling pathways including JNK would be co-accelerators to hepatocarcinogenesis in HCV infection. The latter pathway has not been found in NASH while the former may be common in the pathogenesis of hepatitis C and NASH. HCC, hepatocellular carcinoma; TNF- α , tumor necrosis factor- α ; SOCS-1, suppressor of cytokine signaling-1; NASH, non-alcoholic steatohepatitis.

established between non-alcoholic steatohepatitis (NASH) and HCC, it needs to be pursued energetically in view of histological resemblance of NASH to chronic hepatitis C.

When hepatitis C and NASH are compared, there are a number of similarities between these two medical conditions (Table 2). Steatosis, which is one of the definitions in NASH, is a characteristic trait of chronic hepatitis C [4–6,13]. Disturbances in the lipid metabolism are present in both conditions, although the phenotypes may show a distinction: hypo- β -lipoproteinemia in hepatitis C but hyperlipidemia in NASH. As described above, insulin resistance often arises in chronic hepatitis C, and is also a feature frequently observed in NASH [18]. Some cytokines, such as TNF- α , are considered to be critical in the pathogenesis of these conditions. TNF- α levels are increased in patients with hepatitis C and are implicated in insulin resistance. Single nucleotide polymorphism in TNF- α gene is significantly found in NASH patients [18]. Overproduction of oxidative stress or reactive oxygen species (ROS) plays a pivotal role in the progression of hepatitis and development of HCC in both the conditions: ROS is overproduced

in the liver of the core gene transgenic mice in the absence of inflammation, contributing, at least in part, to the development of HCC [13,19,20]. Functional abnormalities in the mitochondria are implicated, in both hepatitis C and NASH, in the pathogenesis of liver diseases including HCC. In HCV core gene transgenic mice, the malfunction of the electron transfer system of mitochondria has been suggested and is assumed to be an origin of ROS overproduction (Table 2).

Finally, HCC develops both in chronic hepatitis C and NASH. However, the association between NASH and HCC is not strong yet while there is a definite connection in the case of hepatitis C. Nevertheless, HCC develops in patients with NASH, regardless of the frequency. Hence the underlying mechanism of HCC development in NASH awaits further investigation. The analogy between chronic hepatitis C and NASH, as described above, may be a clue to solve a puzzle in the pathogenesis of NASH including hepatocarcinogenesis. In the pathogenesis of HCC in HCV infection, one of intracellular signaling MAPK systems, JNK, is activated in the liver. In the downstream of JNK, transcription factor AP1 and cell cycle machineries, CDK4 and cyclin D1, are subsequently activated, conferring advantage to cell proliferation [16,20]. However, such activations in cellular genes or signaling systems have not been identified yet for NASH. Overproduction of oxidative stress together with the presence of steatosis may be a common pathway to liver hepatocarcinogenesis in both hepatitis C and NASH (Fig. 5, upper half).

However, the alterations in cellular gene expressions and/or intracellular signaling systems are solely with hepatitis C in the presence of the viral protein(s), putting chronic hepatitis C onto the fast track for the development of HCC (Fig. 5, upper half). This aspect of NASH should be investigated. The

Table 2
Comparison of hepatitis C and NASH

Hepatitis C	NASH
Steatosis	Steatosis
Hypo- β -lipoproteinemia	Hyperlipidemia
Insulin resistance	Insulin resistance
Cytokines (TNF- α , etc.)	Cytokines (TNF- α , etc.)
Oxidative stress	Oxidative stress
Mitochondrial abnormality	Mitochondrial abnormality
Obesity?	Obesity
HCC	HCC?

analogy between hepatitis C and NASH would give a solution to problems in the pathogenesis of NASH.

7. Conclusion

Although HCV targets at the liver, it has become increasingly evident that HCV can induce diseases of many organs. Recently, much attention is drawn to metabolic disorders in HCV infection. First, hepatic steatosis and derangement in lipid metabolism have been found characteristic of HCV infection, and later on correlation was noted between HCV infection and diabetes as well as insulin resistance. We have demonstrated that HCV by itself can induce insulin resistance by means of disturbing the insulin signaling pathway by an HCV protein. The fact that HCV infection induces insulin resistance by the virus itself may influence the progression of chronic liver disease and open up novel therapeutic approaches. HCV infection would need to be viewed not only as liver disease but also a metabolic disease, which would be a clue to open up a novel way to the molecular understanding of pathogenesis of NASH.

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Hepatitis C Virus Infection Can Present with Metabolic Disease by Inducing Insulin Resistance

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Key Words

Diabetes · Hepatitis C virus · Insulin resistance · Insulin receptor substrate · Transgenic mouse

Abstract

Although hepatitis C virus (HCV) targets the liver, it has become increasingly evident that HCV can induce diseases of many organs. Recently, much attention is drawn to metabolic disorders in HCV infection. First, hepatic steatosis and derangement in lipid metabolism have been found characteristic of HCV infection, and later on, a correlation was noted between HCV infection and diabetes as well as insulin resistance. We have demonstrated that HCV by itself can induce insulin resistance through disturbing the insulin signaling pathway by HCV proteins. The fact that HCV infection induces insulin resistance by the virus itself may influence the progression of chronic liver disease and open up novel therapeutic approaches. In conclusion, towards the future, HCV infection needs to be viewed not only as a liver disease but also as a metabolic disease.

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Introduction

Hepatitis C virus (HCV) infects approximately 1.8 million people in Japan alone and as many as 200 million over the world and induces liver disease ranging from

chronic hepatitis through cirrhosis to hepatocellular carcinoma (HCC) [1, 2]. It has been noticed soon after the discovery that the infection with HCV does not exclusively involve the liver. In fact, type II cryoglobulinemia [3] and membranoproliferative glomerulonephritis [4] frequently occur in patients infected with HCV. Furthermore, strong associations of HCV infection with Sjogren's syndrome [5] and lichen planus [6] have been noted, which is verified in the animal model [7]. In addition, the relation between HCV infection and B cell lymphoma has attracted attention especially in Europe [8].

Recently, there have been increasing lines of evidence to indicate metabolic disturbances in HCV infection which, in turn, would influence the pathogenesis of chronic hepatitis C. The discovery of HCV in 1989 [9] enabled a comparison between chronic hepatitis C and other chronic liver diseases. As shown in the results, it has been repeatedly reported that steatosis is significantly associated with chronic hepatitis C [10, 11]. Steatosis in HCV infection is reproduced in animal models [12–14] to reinforce a pathologic role of HCV. Furthermore, patients infected with HCV have abnormalities in serum lipids, such as hypocholesterolemia and abnormal levels of apolipoproteins in serum [15, 16]; they are rectified in sustained virological responders to interferon (IFN) [16]. Thus, the association between HCV infection and a derangement in lipid metabolism has become increasingly strong, both in patients and experimental systems in animals. Finally, patients with chronic hepatitis C accompanied by severe steatosis develop hepatic fibrosis with an

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increased velocity [17]. All in all, we could say that abnormal lipid metabolism in HCV infection is deeply involved in the pathogenesis of hepatitis C.

HCV Infection and Diabetes

Diabetes is suggested as another metabolic disease in association with HCV infection. In 1994, Allison et al. [18] reported an epidemiological link between diabetes and HCV infection. However, doubts were cast on the association in view of a decreased glucose tolerance in advanced chronic hepatitis as well as an increased opportunity for HCV infection in diabetics who frequently receive determination of blood sugar. Several reports from the same group and others followed along this line. The trend to accept the solid association between diabetes and HCV infection seems to have been triggered in the United States by the population study by Metha et al. [19].

However, the association between diabetes and HCV infection is blemished by factors responsible for decreased glucose tolerance, such as advanced cirrhosis, obesity and ageing common in patients with hepatitis C; they make it difficult to prove this association. Hence, there is a need to evaluate the association by basic studies in experimental systems.

HCV Infection Induces Insulin Resistance

We set out to demonstrate the association between HCV infection and diabetes using the animal model. Mice transgenic for the HCV core gene were employed to this end [12, 13]. These mice are engineered to have the HCV core gene of genotype 1b in the absence of other viral genes. They express HCV core protein of the expected size in the liver, in levels comparable with those of patients with chronic hepatitis C (fig. 1). Half of them develop HCC later during their lives [13]. These transgenic mice were fed with their normal littermates, and the glucose metabolism was compared between them [20].

Although mice transgenic for the core gene did not develop overt diabetes, they had markedly elevated serum levels of insulin. Plasma glucose levels were somewhat higher in transgenic mice than in their normal littermates, both in the fast and after ample feeding, with no significant differences between them. In remarkable contrast, serum insulin levels were significantly higher in transgenic than in normal mice in both conditions (fig. 2). Since such a combination of normal glucose levels and

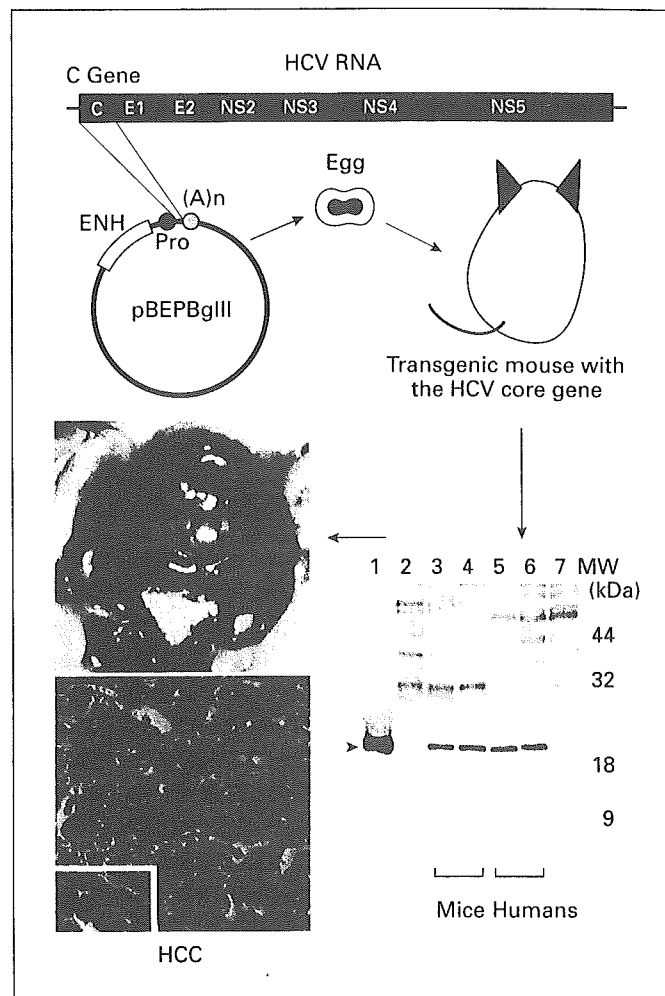


Fig. 1. Expression of HCV core gene in transgenic mouse. It carries the core gene of HCV genotype 1b alone and expresses the core protein of expected size in the liver, at levels similar to those in human patients. Mice eventually develop HCC later in their lives.

hyperinsulinemia points to insulin resistance, glucose and insulin tolerance tests were conducted.

Mice transgenic for the HCV core gene exhibited glucose levels a little higher than those in normal littermates, without any significant differences between them. In insulin tolerance tests, glucose levels were significantly higher in transgenic than in normal mice, both 40 and 60 min after they were injected with insulin intraperitoneally (fig. 3). These results indicate suppression of the activity of insulin to decrease blood glucose levels for inducing insulin resistance in core-transgenic mice. Since only the HCV core gene had been incorporated into these transgenic mice, HCV core protein was able to induce insulin resistance *in vivo*.

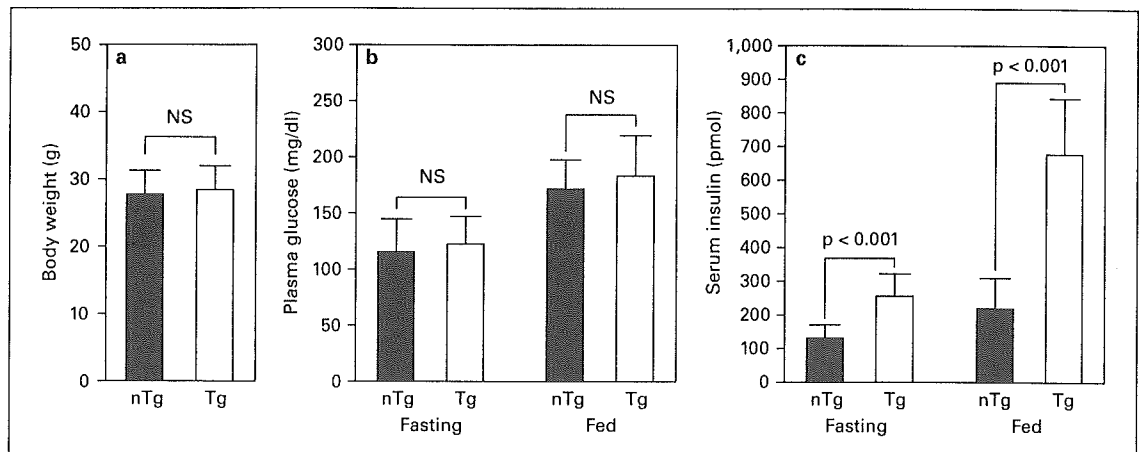


Fig. 2. Altered homeostasis of glucose in mice transgenic for the HCV core gene. Body weight of 2-month-old mice (a), plasma glucose levels in fasting or fed mice (b) and serum insulin levels in fasting or fed mice (c) are shown. Values represent means \pm SE. NS = Not significant statistically; nTg = nontransgenic mice; Tg = transgenic mice.

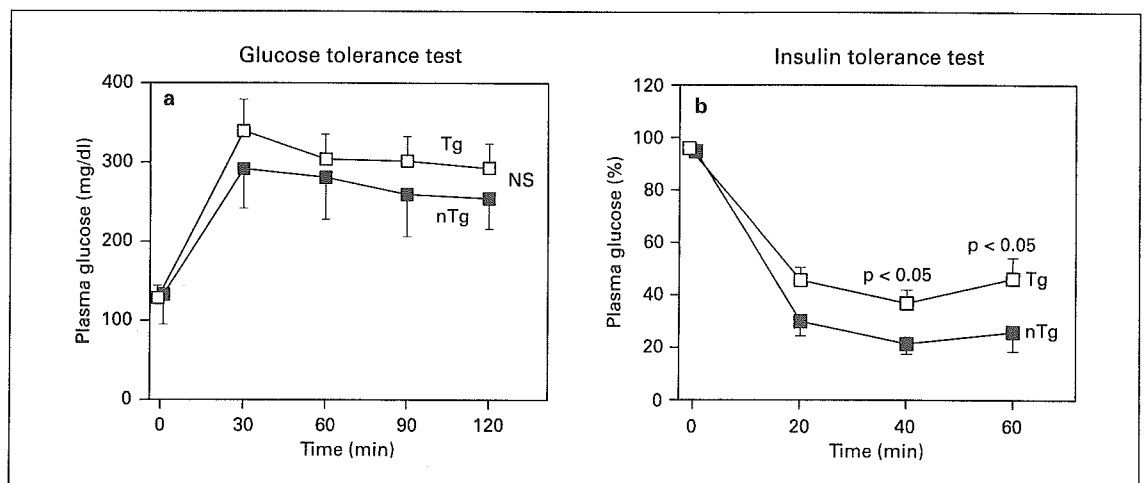


Fig. 3. Insulin resistance in transgenic mice. Glucose tolerance in mice after overnight fasting (a). *D*-Glucose (1 g/kg body weight) was given intraperitoneally to conscious mice, and plasma glucose levels were determined at time points indicated. **b** Insulin tolerance in mice fasted overnight. Human insulin (1 U/kg body weight) was injected intraperitoneally, and glucose concentrations were determined sequentially. Values were normalized to the baseline glucose concentration at the time of insulin administration. NS = Not significant statistically; nTg = nontransgenic mice; Tg = transgenic mice.

By what mechanism does insulin resistance arise in this animal model? The insulin resistance is considered to involve two factors, namely central and peripheral insulin resistances (table 1) [21]. The hyperinsulinemic-euglycemic clamp method was employed to differentiate between them. In this method, hepatic glucose production (HGP) is calculated on the basis of amounts of glu-

cose required to keep plasma glucose levels within a certain range at serum insulin levels higher than physiological ones. In normal control mice, HGP was suppressed by 60% by the administration of insulin, in contrast to core-transgenic mice in which there was no appreciable suppression of HGP by insulin (fig. 4). These results indicate a hepatic (central) origin of the insulin resistance

Table 1. Two types of insulin resistance

Type	Mechanism
Peripheral	A shortage of insulin action in the muscle due to deficit in the insulin-induced uptake of glucose into muscles
Central	A shortage of insulin action in the liver due to deficit in the insulin-induced suppression of glucose production in hepatocytes

in transgenic mice. For further confirmation, an uptake of glucose into muscle was determined. There were no differences in the uptake in response to administration of insulin between normal and transgenic mice. Therefore, the insulin resistance in mice transgenic for the HCV core gene is central and hepatic.

HCV Core Protein Suppresses the Transduction of Insulin Signaling in Hepatocytes

Next, we evaluated how insulin resistance elicits in mice transgenic for the HCV core gene. For this purpose, liver homogenate was immunoblotted with antiphosphotyrosine and antiphosphoserine antibodies after insulin receptor substrate (IRS)-1 and IRS-2 had been immunoprecipitated. Tyrosines in IRS-1 were weakly phosphorylated both in normal and transgenic mice before they received insulin, with no differences between them. However, after the administration of insulin, the phosphorylation of tyrosines in IRS-1 increased in normal but not in transgenic mice (fig. 5). Obtained results suggested disturbance in tyrosine phosphorylation as one of the factors responsible for insulin resistance in the liver. There were no differences in phosphorylation of serines in IRS-1 or tyrosines in IRS-2 between normal and transgenic mice. Combined, they provided experimental evidence for the development of insulin resistance by the presence of HCV in the liver that would occur by disturbing the transduction of insulin signaling in hepatocytes (fig. 6).

There remains a possibility for the HCV core protein to directly prohibit phosphorylation of tyrosines, or else, it might inhibit tyrosine phosphorylation via certain cytokines. In our extensive searches for the expression of cytokines in the liver of transgenic mice, only TNF- α and IL-1 β have been found with an increased expression [22]. Therefore, for the purpose of evaluating the role of

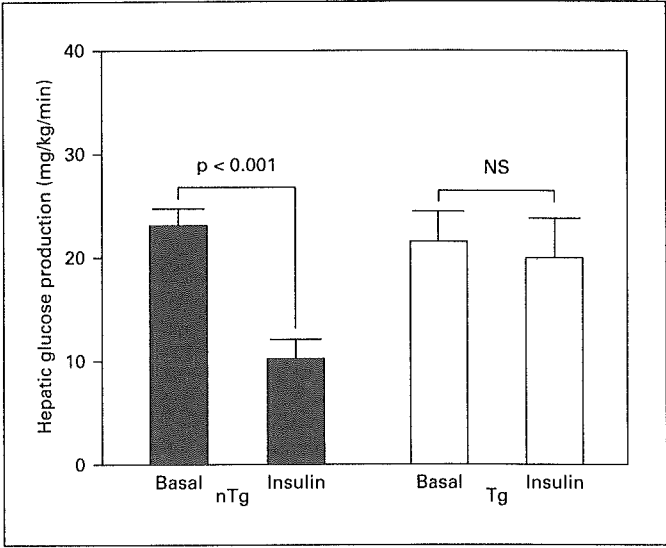


Fig. 4. Characterization of glucose metabolism in transgenic mice. Glucose production in the liver was calculated using the hyperinsulinemic-euglycemic clamp method. NS = Not significant statistically; nTg = nontransgenic mice; Tg = transgenic mice.

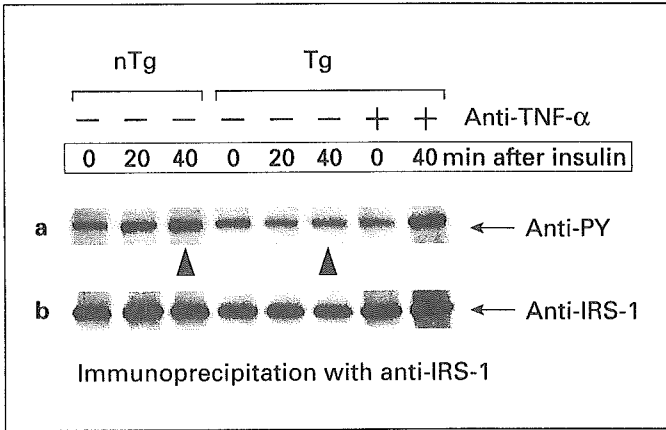


Fig. 5. Phosphorylation of tyrosine in IRS-1 in response to insulin stimulation. Liver tissues from control mice, transgenic mice with or without anti-TNF- α antibody treatment, were analyzed before and 20 as well as 40 min after administration of insulin. Samples were subjected to immunoprecipitation with anti-IRS-1 antibody and then immunoblotted with indicated antibodies. Experiments were performed in triplicate, and a representative picture is exhibited. Immunoblotting with antiphosphotyrosine (PY) antibody (lane a) did not augment phosphorylation of tyrosine in IRS-1 after stimulation with insulin in the core gene transgenic mice (Tg), in contrast to tyrosine phosphorylation markedly enhanced in control mice (nTg). Insulin-stimulated tyrosine phosphorylation was restored 40 min after treatment with anti-TNF- α antibody. Note differences in the intensity of bands 40 min after the administration of insulin (arrowheads). Immunoblotting with anti-IRS-1 antibody (lane b) served as control for the IRS-1 load.

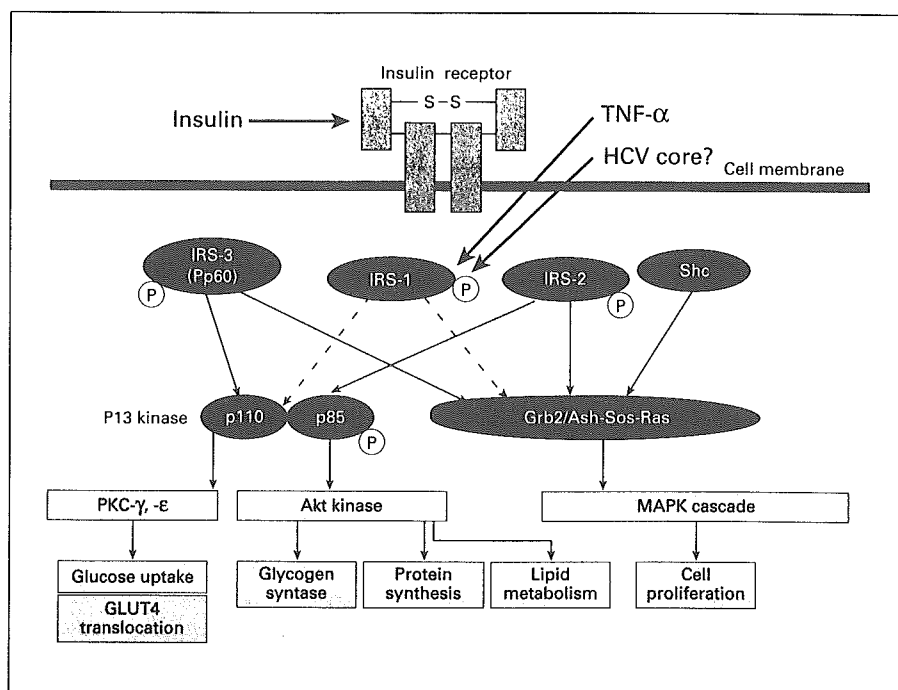


Fig. 6. A proposed mechanism for insulin resistance in HCV infection. HCV itself or elevated levels of cytokines such as TNF- α may inhibit tyrosine phosphorylation of IRS-1 in the liver, suppress intracellular transduction of insulin signal and lead to insulin resistance. PKC = Protein kinase C; MAPK = mitogen-activated protein kinase.

TNF- α in insulin resistance in transgenic mice, serum insulin was determined and an insulin tolerance test performed after they had received anti-TNF- α intraperitoneally. Pretreatment with anti-TNF- α partially improved insulin resistance in mice transgenic for the HCV core gene. Albeit a direct anti-insulin activity of core protein and direct or indirect factors for insulin resistance are not to be excluded, high levels of TNF- α in the liver would be one of the factors for expression of insulin resistance in this mouse model.

Insulin Resistance in Patients with Chronic Hepatitis C

Concurrently with our report in experimental systems, Aytug et al. [23] investigated insulin signaling in biopsied liver specimens from patients with chronic hepatitis C. Specifically, they evaluated changes in IRS-1, IRS-2 and phosphatidylinositol (PI)3 kinase levels in the liver of patients. With insulin stimulation of biopsied liver samples, insulin receptor proteins and IRS-1 increased, while phosphorylation of tyrosines in IRS-1 decreased to one half of the baseline value, along with a diminished activity for PI3 kinase associated with IRS-1, in patients with chronic hepatitis C. The authors went on to propose a possibility for disturbed transduction of the insulin sig-

naling pathway in the liver to induce insulin resistance in patients with chronic hepatitis C [23]. Their report is quite intriguing in that it opens up the way for evaluating an association between HCV infection and insulin resistance in clinical samples at the molecular level.

The results of Aytug et al. [23] inadvertently coincide with ours in analyzing the mechanism of insulin resistance with the experimental system in mice (*vide supra*). They unanimously incriminate impaired tyrosine phosphorylation in IRS-1 in the induction of insulin resistance by HCV infection. It struck us as a surprise that the mechanism of insulin resistance induced by HCV infection has been in agreement between clinical samples and experimental animals, in spite of hepatic IRS-2 that was preferred to IRS-1 for its role in development of insulin resistance in former studies [24]. HCV infection is peculiar in that IRS-1 weighs heavier than IRS-2 in the induction of hepatic insulin resistance.

Although our data strongly indicate a hepatic character of insulin resistance in HCV infection, they by no means exclude roles of other factors in the induction of this resistance. There is little expression of the HCV core gene in muscles of our animal model; it is not known if HCV infects muscular cells in patients with chronic hepatitis C. Factors not intrinsic to the liver would have to be evaluated to sort this out, including dysfunction of mitochondria for induction of insulin resistance [25].

Insulin Resistance for Advanced Hepatic Fibrosis

Insulin resistance in HCV infection may have an additional significant clinical implication. In 260 patients with chronic hepatitis C, Hui et al. [26] have tried to establish the relationship between liver histology and indicators of glucose metabolism, as well as insulin resistance represented by the homeostasis model assessment of insulin resistance. They have found that insulin resistance already exists in hepatitis C patients with stage 0 or 1 fibrosis in the liver. This indicates that insulin resistance in HCV infection is not attributable to advanced liver disease. In their study, independent predictors of insulin resistance in HCV infection were body mass index, non-response to antiviral treatment, intensity of portal inflammation and infection with HCV genotype 3 [26]. Furthermore, the homeostasis model assessment of insulin resistance was a significant and independent predictor of the stage and velocity of hepatic fibrosis. The results of the study are of much importance, because they implicate a role of insulin resistance and hyperinsulinemia by inference, in promoting the progression of hepatic fibrosis. Insulin has been proven as an aggravating factor not only in atherosclerosis, but also in systemic inflammation and fibrosis. The liver is no exception to this.

Conclusions: Hepatitis C Viewed as a Metabolic Disease and Outlook for Therapeutic Strategies in the Future

We have demonstrated that HCV per se induces insulin resistance in the animal model. Superimposed high-fat diet and obesity may lead to overt diabetes. Since insulin resistance accelerates the progression of chronic hepatitis C, it would naturally influence the development of HCC. Although the association has not been established between nonalcoholic steatohepatitis and HCC, it needs to be energetically pursued in view of the histological homology of nonalcoholic steatohepatitis to chronic hepatitis C. Drugs for improving glucose metabolism and insulin resistance need to be kept in store in the treatment of hepatitis C patients who have failed to respond to antivirals, because they may well prevent progression of fibrosis and development of HCC in such patients. Traditional 'high-protein and high-calorie' diet, especially advocated in Japan after World War II, is obviously detrimental, except in some patients with advanced cirrhosis. Consultation on the dietary habit with hepatitis C patients should include iron restriction [27] as well as weight control, because high-calorie intakes are likely to accelerate hepatic fibrosis by aggravating insulin resistance.

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Review

Oxidative stress and hepatitis C viral infection

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Available online 20 December 2005**Abstract**

The involvement of oxidative stress in the pathogenesis of hepatitis and hepatocellular carcinoma has been strongly suggested. Oxidative stress is produced by inflammatory processes that occur in hepatitis via immunological mechanisms. In addition, in hepatitis C virus (HCV) infectious disease, some role has been assigned to viral proteins in the induction of oxidative stress. In the presence of hepatic steatosis, insulin resistance and increased levels of some cytokines, all of which are also induced by viral protein expression, oxidative stress is enhanced in HCV infection. In this sense, the role of oxidative stress in the progression of chronic hepatitis and hepatocarcinogenesis is greater in hepatitis C than in other types of hepatitis such as hepatitis B or autoimmune hepatitis. The additive effects of oxidative stress caused by the inflammatory process and that induced by HCV proteins may, furthermore, exert synergistic effects with alterations in intracellular signaling systems such as mitogen-activated protein kinases (MAPK), which are also induced by HCV proteins. These synergistic effects may be responsible for rare characteristics, that is, the high incidence and multicentric nature of hepatocarcinogenesis in HCV infection.

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Keywords: Oxidative stress; Hepatitis C virus; Hepatocarcinogenesis; Lipid peroxidation; Steatosis; Insulin resistance**Contents**

1. Introduction	66
2. Oxidative stress, reactive oxygen, and the liver	66
2.1. Oxidative stress and reactive oxygen	66
2.2. Antioxidation system and oxidative stress markers	66
3. Viral infection and oxidative stress	67
3.1. ROS production associated with viral infection	67
3.2. Nitric oxide production associated with viral infection	67
4. Oxidative stress caused by viral protein	68
5. Relationship of HCV infection with insulin resistance	68
6. Relationship of HCV infection with hepatic steatosis	69
7. Iron and reactive oxygen	69
8. Interactions with alcohol	69
9. Hepatocarcinogenesis and oxidative stress	69
10. Conclusions	70
References	71

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1. Introduction

There are approximately 200 million people infected with hepatitis C virus (HCV) worldwide, of which about 1.8 million are in Japan. It is one of the most serious causes of liver disease. It was reported that approximately 70% of those with HCV infection suffer from persistent infection, causing active or inactive chronic hepatitis and that about 30% of patients with chronic hepatitis are assumed to develop cirrhosis within their lifetime. Once HCV infection develops into cirrhosis, hepatocellular carcinoma (HCC) develops at an annual rate of 5–7% [1]. The strong association of oxidative stress with HCV infection has been demonstrated recently and it has become possible to explain at least part of the clinical progression of the disease. The pathogenesis of chronic hepatitis C is not merely ascribed to inflammation caused by viral infection, but the role of viral proteins in the pathogenesis was also reported [2–4]. Of proteins constituting HCV, the core protein, in particular, has various functions with respect to host cells [5] and is closely related to oxidative stress. In this overview, the relationship between HCV infection and oxidative stress is reviewed focusing on the pathological effect of the core protein of HCV, and the significance of oxidative stress in the pathogenesis of liver disease will be discussed.

2. Oxidative stress, reactive oxygen, and the liver

2.1. Oxidative stress and reactive oxygen

The main source of reactive oxygen species (ROS) in hepatocytes is the mitochondria. Outside of hepatocytes, ROS also originate from nicotinamide adenine dinucleotide phosphate

(NADPH) oxidase and xanthine oxidase in Kupffer cells and inflammatory cells. Several percent of consumed oxygen is constantly converted into ROS in the mitochondria accompanied by oxygen consumption in the electron transport system (ETS, Fig. 1). Hepatocytes contain many mitochondria and therefore have a high ROS production. Generated ROS are very unstable and highly reactive, and attack biomolecules such as DNA, lipids, and proteins. The liver not only produces much ROS but is also the center of the anti-oxidative effect in the form of protein synthesis. Oxidative stress refers to the oxidation-reaction-dominant state of the living body induced by an imbalance between the oxidation reaction caused by ROS and the anti-oxidation reaction. Main ROS include superoxide ($^{\circ}\text{O}_2^-$), hydrogen peroxide (H_2O_2) and the hydroxyl radical (HO°). ROS are mainly produced from $^{\circ}\text{O}_2^-$ and converted into stable H_2O_2 through dismutation reaction. H_2O_2 is converted into highly reactive HO° in the presence of a transition metal.

2.2. Antioxidation system and oxidative stress markers

Antioxidants include glutathione (GSH), thioredoxin (TRX), vitamin E, vitamin C, and β -carotene. Reactive oxygen elimination enzymes include superoxide dismutase (SOD), glutathione peroxidase (GPx), and catalase. SOD is induced by oxidative stress and dismutates $^{\circ}\text{O}_2^-$ to H_2O_2 and oxygen. GSH is a compound belonging to the SH group and is highly abundant in the living body, and the SH group provides electrons to free radicals to stabilize the radicals. GSH exists in a reduced form in cells. Because it is converted into dimeric oxidized glutathione (GSSG) and becomes stable after donating electrons, GSSG prevents free radicals from continuously scrambling for electrons. GPx decomposes H_2O_2 into water

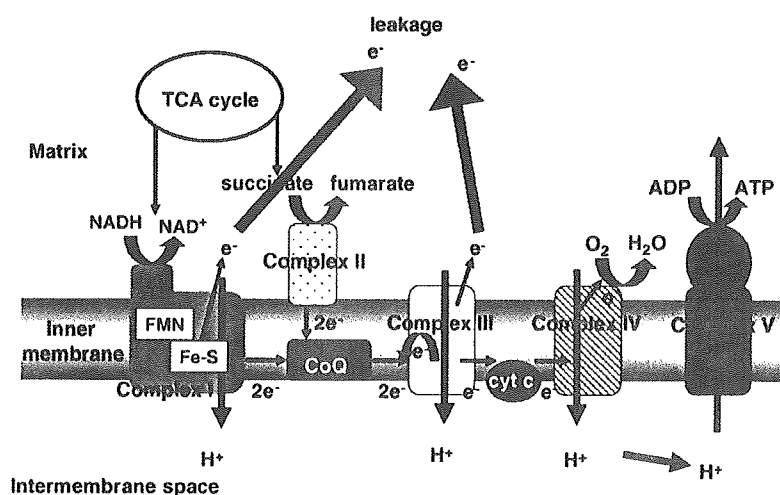


Fig. 1. The electron transfer system (ETS) of the mitochondrion. Most of the oxygen, consumed by mammalian cells, is converted to water via the mitochondrial ETS. However, up to 5% of the electrons entering the mitochondrial ETS can become uncoupled and singly leak out onto oxygen to form superoxide. Therefore, if there is impairment in the mitochondrial ETS function, it can be a cause of the overproduction of reactive oxygen species (ROS). TCA, tricarboxylic acid; NADH, nicotinamide adenine dinucleotide phosphate; FMN, flavine mononucleotide; CoQ, coenzyme Q; cyt c, cytochrome c.

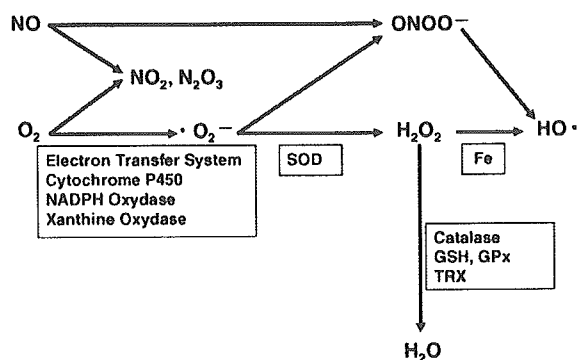


Fig. 2. Generation and scavenging of oxidative stress. SOD, super-oxide dismutase; GSH, reduced glutathione; GPx, glutathione peroxidase; TRX, thioredoxin.

and oxygen with GSH as an electron donor and reduces lipid peroxide to become neutralized. GSSG is converted back to GSH when glutathione reductase transfers an electron from NADPH to GSSG. Catalase in peroxisomes also decomposes H_2O_2 to water and oxygen. TRX is also a protein induced by oxidative stress, and is reduced via the S–S binding of the substrate protein by two SH groups in TRX and acts on the H_2O_2 elimination system via peroxiredoxins (Fig. 2).

ROS cause various forms of cellular damage. 4-Hydroxy-2-nonenal (HNE) and malondialdehyde (MDA) are the peroxidation reaction products of lipids, and 8-hydroxydeoxyguanosine (8-OHdG) is the product of DNA base modification (Fig. 3). These products serve as oxidative stress markers.

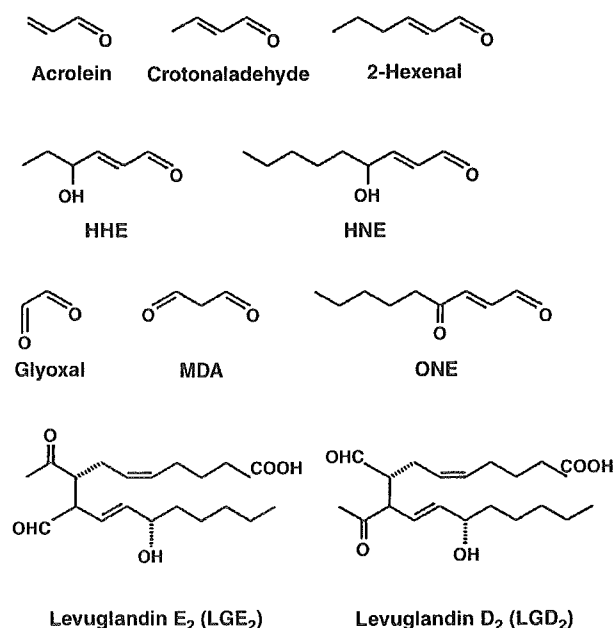


Fig. 3. Representative aldehyde species generated via lipid peroxidation reaction.

3. Viral infection and oxidative stress

3.1. ROS production associated with viral infection

Upon viral infection, ROS are produced by NADPH oxidase and xanthine oxidase in neutrophils and macrophages. In particular, NS3, one of the non-structural proteins of HCV, was reported to induce ROS production by NADPH oxidase in neutrophils [6]. Furthermore, in viral hepatitis, ROS are also produced in hepatocytes through the release of inflammatory cytokines such as $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ from inflammatory cells. Increased hepatic or serum 8-OHdG, HNE and MDA levels are observed in chronic hepatitis C, indicating an increase in ROS production [7–13]. Findings that indicate an increase in the activity of the ROS elimination system including decreased hepatic and blood GSH levels, an increased GSSG/GSH + GSSG ratio, and an increased serum TRX level have been reported [13–16]. The findings of markedly decreased HNE level following viral eradication with interferon [12] and decreased serum ALT and TRX levels following the administration of vitamin E, an antioxidant [17], also demonstrated that oxidative stress plays an important role in chronic hepatitis C.

3.2. Nitric oxide production associated with viral infection

In the presence of an inflammation, inducible nitric oxide synthase (iNOS) is induced in macrophages and hepatocytes by $\text{TNF-}\alpha$ and $\text{IFN-}\gamma$ [18–20]. Other investigators reported that protein kinase (PKR) activated by double-stranded RNA formed during virus reproduction in turn activates the transcripts of $\text{NF-}\kappa\text{B}$ and IRF-1 to induce iNOS [21]. In the case of HCV, it was reported that its constituent proteins (E2 and non-structural (NS) protein 5A) inhibit PKR activity [22,23], but iNOS induction by viral RNA via PKR is also suspected. Indeed, iNOS synthesis correlates with intrahepatic viral load in chronic hepatitis C [24].

NO is generally synthesized as a non-specific defense reaction to infectious diseases; however, in viral infection, antiviral activity may be present or absent in various viral types [20]. NO is reported to exhibit no antiviral activity against a tick-borne encephalitis virus (TBE-V), flavivirus [25], and NO may also have no antiviral activity against HCV. On the contrary, NO causes cellular damage upon its reaction to O_2 or simultaneously produced $\text{O}_2^{\cdot-}$ (reactive nitrogen species, RNS). Upon reaction to $\text{O}_2^{\cdot-}$, in particular, NO acts as a strong oxidant with the generation of peroxynitrous acid (ONOO^-), and ONOO^- also produces nitrotyrosine through the nitration of aromatic amino acid residues in the presence of a transition metal. Nitrotyrosine accumulation was observed in correlation to inflammation severity in chronic hepatitis C tissue [26]; suggesting that the production of both NO and ROS increased. ROS and RNS are produced as defense factors for biological viral clearance, but these factors also have cytotoxic effects that

are assumed to contribute to the exacerbation of the disease state.

4. Oxidative stress caused by viral protein

The HCV genome comprises the genes of four structural proteins and six non-structural proteins (Fig. 4), and it has been reported that at least two viral proteins cause oxidative stress in cells. The core protein, a structural protein, was found to have various actions, including the induction of oxidative stress and accumulation of lipids, in experimental studies using cultured cells and transgenic mice [2,27]. Experiments using mice transgenic for the core gene showed an increased ROS production, an increased intrahepatic catalase activity, a decreased intrahepatic GSH level and a decreased GSH/GSH – GSSG ratio indicating an anti-oxidation effect inhibition, although there was no increase in serum ALT level nor a histological finding of hepatitis. Increased levels of intrahepatic peroxide lipids in the core gene transgenic mice with aging as compared with those in the control mice also indicate increased oxidative stress. As a mechanism underlying oxidative stress induction by the core protein, mitochondrial damage is considered. Morphological abnormalities of the mitochondria were observed in the core gene transgenic mouse liver [2], and an increased ROS production caused by damage of the mitochondrial electron transport system was noted in core-protein-expressing cells [27]. Mitochondrial DNA, which has no protective proteins such as histone, is susceptible to damage by ROS [28,29]. Mitochondrial DNA in the core gene transgenic mice showed damage as early as 3-months old. This mitochondrial damage disrupts the synthesis of proteins constituting the electron transport system complex and could also increase oxidative stress caused by damage of the electron transport system.

A study using a cell culture system demonstrated that non-structural protein 5A (NS5A) also causes oxidative stress. NS5A induces endoplasmic reticulum calcium release via

endoplasmic reticulum stress, and this leads to an increased ROS production in the mitochondria [4]. Although the effect of NS5A has not been confirmed yet by other study groups, HCV has the direct action of increasing intracellular ROS production via its proteins, separate from oxidative stress induction by inflammation caused by viral infection. A report that oxidative stress was also observed in HCV carriers with a normal ALT level [13] indicates that it is caused by a direct oxidative stress induction without being mediating inflammatory reactions.

5. Relationship of HCV infection with insulin resistance

The relationship of HCV infection with insulin resistance and type 2 diabetes has been suggested epidemiologically [30–32]. Insulin resistance was also observed in core gene transgenic mice before the onset of hepatic steatosis [33]. A disrupted tyrosine phosphorylation of the insulin receptor substrate (IRS-1) was observed in the liver of these transgenic mice. The analysis of hepatic tissues in patients with chronic hepatitis C not complicated by diabetes showed that insulin receptor and IRS-1 expression levels are elevated in patients with HCV infection, whereas the tyrosine phosphorylation of IRS-1 induced by insulin is inhibited. An excessive oxidative stress may be another potential cause of this insulin resistance. Oxidative stress indirectly blocks the phosphorylation of tyrosine residues of insulin receptors and IRS-1 and inhibits insulin signaling [34].

These reported results thus indicate an insulin signaling disorder in the liver infected with HCV [35]. There has been no report to date directly proving that hepatic insulin signaling disorder in patients with HCV infection is attributable to oxidative stress. However, because diabetes, which is the state of having abnormally high blood sugar levels that cannot be self-regulated by individual organisms, also induces oxidative stress [34], the close relationship between insulin resistance or diabetes and oxidative stress as the cause and the

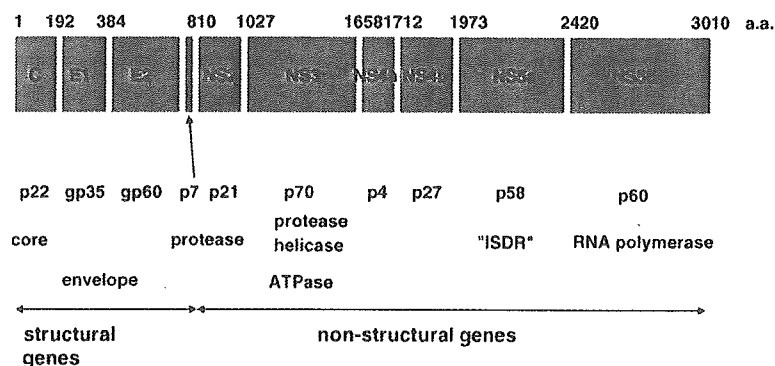


Fig. 4. Structure of hepatitis C virus genome. The genome of HCV consists of two parts, structural and non-structural regions. The former comprises the core and envelope regions, and the latter consists of NS2 to NS5a, which regions chiefly code enzymes necessary for viral replication. NS, non-structural; ISDR, interferon sensitivity-determining region.

result, respectively, is a very interesting issue to investigate in the future.

6. Relationship of HCV infection with hepatic steatosis

Hepatic steatosis is frequently observed in patients with HCV infection. The relationships of HCV infection with intrahepatic viral loads and core protein levels, different prevalence of hepatic steatosis by viral genotype [higher incidence for genotype 3a], and improved steatosis following viral eradication were reported [36–38]. It is presumed from these reports that HCV itself causes hepatic steatosis. A similar hepatic steatosis caused even by the core protein alone was observed in a study using an expression system in cultured cells and transgenic mice, and it was thus suggested that the core protein plays a significant role in hepatic steatosis as the direct action of HCV [39,40]. Hyperinsulinemia induced by insulin resistance mentioned above causes the overloading of the liver with fatty acids from fat cells, and mitochondrial damage inhibits the β -oxidation of fatty acids [41]. Furthermore, the core protein was reported to inhibit microsomal triglyceride transfer protein (MTP) activity that is required when neutral fat is released as very low-density lipoproteins (VLDLs) [42]. All these actions could cause hepatic steatosis. In the liver of non-alcoholic steatohepatitis (NASH) patients, it was reported that β -oxidation in the mitochondria and peroxisomes or the metabolism of fatty acids by cytochrome P450 2E1 (CYP2E1) in microsomes is promoted under an excessive load of fatty acids, resulting in ROS production [43,44]. In HCV infection as well, intrahepatic fat accumulation possibly increases ROS production as in NASH. Because hepatic steatosis in chronic hepatitis C was reported to be a factor for disease progression [45–47], increased oxidative stress associated with hepatic steatosis is presumably involved in disease progression.

7. Iron and reactive oxygen

The iron content in the liver and spleen is high, and transition metals facilitate electron transfer and play an important role in the production of free radicals. Iron in combination with transferrin and ferritin is stable, but an unstable iron ion is freed when ferritin is decomposed by lysosomes [48]. ROS additionally promote iron release from ferritin [49]. A free iron ion catalyzes changes from relatively poor reactive O_2^- and H_2O_2 to a highly reactive HO^\bullet (Fenton reaction) [50,51]. HO^\bullet oxidizes membrane phospholipids, which compose cells and intracellular organelles, and iron forms radicals from produced peroxide lipids, thereby enhancing lipid peroxidation. Iron site-specifically combines with DNA and promotes DNA damage caused by ROS. Iron also increases ROS production by CYP2E1 [52]. A report that an enhanced peroxidation of intrahepatic lipids is attenuated by exsan-

guination in hemochromatosis also supports the involvement of iron in oxidative stress [53].

An excessively high iron content in the liver was observed in chronic hepatitis C [8,54]. Other investigators reported that iron removal therapy by exsanguination of chronic hepatitis C patients significantly improves serum ALT level without affecting viral load [55–57]. Another study showed that hepatic impairment is exacerbated following the administration of iron to chimpanzees with chronic hepatitis C [58]. Furthermore, oxidative stress is decreased by the iron removal therapy for chronic hepatitis C using intrahepatic 8-OHdG level as an index [57]. The above-mentioned reports show the close relationships of chronic hepatitis C with iron metabolism and oxidative stress.

8. Interactions with alcohol

Alcohol metabolism plays an important role in ROS production. Mainly alcohol dehydrogenase (ADH) in the cytosol and CYP2E1 (microsomal ethanol-oxidizing system) in microsomes are responsible for alcohol metabolism in the liver. When alcohol dehydrogenase oxidizes ethanol to acetaldehyde, the reduction from NAD^+ to NADH simultaneously occurs. NADH accumulation causes stress on the mitochondrial electron transfer system, leading to an increased production of ROS [59]. NADH also inhibits xanthine dehydrogenase activity, and xanthine is thereby oxidized by xanthine oxidase with the production of ROS [60]. CYP2E1 is induced by chronic alcohol intake and ROS are produced when CYP2E1 oxidizes ethanol to acetaldehyde [52,61].

There is no significant difference in hepatic peroxide level between core gene transgenic mice at 3–6-months old and control transgenic mice, but hepatic peroxide level significantly increases following the administration of a low dose of alcohol in the core gene transgenic mice [2]. ROS production increases upon glutathione reduction in HepG2 cells, with the co-expression of the core protein and CYP2D1, the latter of which is induced by alcohol [62]. These findings show that the core protein and alcohol in combination increase oxidative stress. Indeed, it was reported that alcohol intake plays a role in promoting the progression of chronic hepatitis C [63,64] and that increased levels of oxidative stress markers such as HNE and lipid hydroperoxide also support these findings [65]. From the viewpoint of oxidative stress also, HCV infection and alcohol intake are both considered to promote hepatic impairment.

9. Hepatocarcinogenesis and oxidative stress

It has been demonstrated that oxidative stress plays a key role in carcinogenesis [66,67]. Animal experiments using hepatocarcinogenesis models with the administration of a chemical substance (diethyl-nitrosamine, peroxisome proliferators) and with the administration of a choline-deficient

amino acid diet also indicates the involvement of oxidative stress [68–72]. In Long Evans Cinnamon (LEC) rats, an animal model that spontaneously develops heritable hepatitis and HCC caused by an abnormal copper accumulation, a congenitally decreased glutathione peroxidase expression level was reported, and the close relationship between oxidative stress and hepatocarcinogenesis was indicated [73].

The epidemiological relationship between HCV infection and HCC is evident [74,75], but the mechanism underlying this relationship has not been fully elucidated yet. Among postulated hypotheses on the mechanism of HCV-associated hepatocarcinogenesis, that of the involvement of the viral protein, in particular, the core protein of HCV is attractive: HCC develops in core gene transgenic mice, and carcinogenesis starts with well-differentiated carcinoma with an excessively high fat content, similar to hepatocarcinogenesis in human chronic hepatitis C, and poorly differentiated carcinoma with a low fat content develops in the form of “nodules in nodules” [76]. Because oxidative stress is increased in the core gene transgenic mice as mentioned above, it is assumed that oxidative stress plays an important role in hepatocarcinogenesis in chronic hepatitis C. Because the development of HCC is also observed in transgenic mice carrying the full-length HCV protein gene, the non-structural protein may have an additive effect to the effect of the structural proteins including the core protein, contributing to hepatocarcinogenesis [77]. NS5A, which was also reported to induce ROS production [4], may also contribute to hepatocarcinogenesis, although ROS induction by NS5A is not unequivocally confirmed yet.

Mitochondrial DNA has no potent protective proteins such as histone and is near the electron transport system, the major ROS production site. Hence, it is 10 to 15 times more susceptible to mutation caused by ROS than nuclear DNA [28,29]. In an investigation of mitochondrial DNA mutation in the human normal liver, both cancerous and non-cancerous liver tissues in patients with HCC showed very high incidences of DNA mutations [78]; thus, a relationship between oxidative stress persistence and hepatocarcinogenesis is suggested.

In the core protein expression system in the hepatic tissue and cultured cells of core gene transgenic mice, the activation of transcription factor AP-1 via mitogen-activated protein (MAP) kinase was observed [79–83]. The activation of the transcription factors AP-1, NF- κ B, and signal transducer and activator of transcription (STAT) 3 by NS5A were also reported [4,84]. The activation of these transcription factors may facilitate cell proliferation, contributing to tumorigenic transformation.

It was also reported that ROS facilitate apoptosis via c-Jun N-terminal kinase (JNK)/p38 MAP kinase or by directly attacking the mitochondria. Apoptosis is a protective mechanism of the host against viral infection and carcinogenesis. Some reports stated that the core protein facilitates apoptosis [85–88], whereas other reports stated that the core protein inhibits apoptosis [89–92]; thus, no fixed view has yet been established. If it indeed inhibits apoptosis, it is assumed that this inhibition proceeds by maintaining oxidative stress and

that the core protein has a beneficial effect against carcinogenesis and persistent viral infection.

In HCV infection, viral proteins such as the core protein and, possibly, NS5A protein induce oxidative stress, intracellular signaling, and transcription factors, which are not reflected in blood ALT level, contributing to the progression of carcinogenesis. Carcinogenesis, however, is slow as is observed in humans and core gene transgenic mice, the latter of which developed HCC in the latter half of their life. Recently, Okanoue et al. reported a long-term follow-up study of subjects with persistent HCV infection who had persistently normal ALT levels (PNAL) [93]. In their study, serum thioredoxin levels were not elevated in those with PNAL compared to those with chronic hepatitis. This may apparently seem contradictory to the results of our above-mentioned animal model studies. However, we should realize that anti-oxidant system is also instrumental in the liver. In these relatively younger people with PNAL than those with CH [93], active anti-oxidant system may erase the apparent elevation of ROS. Such a phenomenon was described in a mouse model by Moriya et al. [2], in which ROS was apparently normal in young core gene transgenic mice with the activation of catalase and reduction of GSH. Clinically, the presence of inflammation is thought to facilitate the process of hepatocarcinogenesis.

10. Conclusions

A very close pathological relationship between oxidative stress and HCV infection is observed, as shown by the above overview of relevant publications and discussion. The causes of oxidative stress in HCV infection are considered to include various factors such as mitochondrial damage, endoplasmic reticulum stress, iron accumulation, and lipid accumulation in the liver. Various study results demonstrated that even only viral proteins, mainly the HCV core protein, cause oxidative stress. When inflammation via immunoreactions to viral infection is added to oxidative stress, ROS production is expected to further increase, leading to a state in which the anti-oxidation system cannot cope with. In this sense, inflammation in chronic hepatitis C is considered to be qualitatively different from inflammation observed in other types of hepatitis such as autoimmune hepatitis or hepatitis B [94] (Fig. 5). As a treatment of chronic hepatitis C, the eradication of the virus is ideal. If it is not possible, however, the control of factors that exacerbate oxidative stress, such as inflammation via immune reaction and alcohol, and the relief of oxidative stress by the iron removal therapy and the administration of an anti-oxidation agent are considered to delay the progression of chronic hepatitis.

The development of such new anti-oxidation agents is being awaited. In further studies on the development of new therapies for hepatitis C and control methods for hepatocarcinogenesis in the future, the importance of those focusing on oxidative stress is expected to markedly increase.

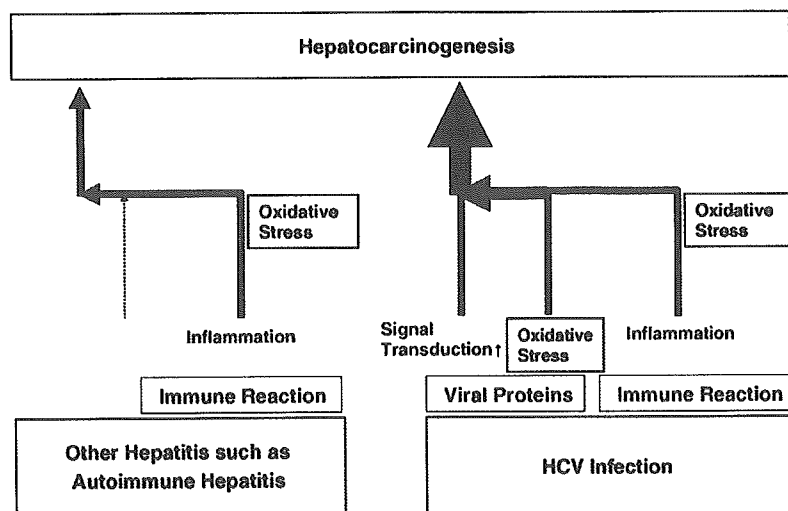


Fig. 5. Oxidative stress and hepatocarcinogenesis in various types of hepatitis (hypothesis). Oxidative stress is generated in all types of hepatitis via inflammation accompanied by continual cell death and regeneration. In HCV infection, HCV itself causes the production of oxidative stress in a synergy with inflammation. In this sense, the quality of "inflammation" in HCV infection may be different from that in other types of hepatitis. Additional impact of HCV proteins on the intracellular signal transduction would provoke the development of HCC. These may explain the conspicuous properties of HCC development.

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Hepatic Arterial Anatomy for Right Liver Procurement From Living Donors

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Living donor liver transplantation (LDLT) using right liver grafts is now widely performed. Anatomic classifications of the hepatic artery for right liver procurement, however, are limited. In this study, celiac and mesenteric angiograms of 223 consecutive living donors in a single institution were evaluated. Details of the arterial anastomosis and results were reviewed in 72 patients who underwent primary LDLT using right liver grafts. There was a 6% incidence of hepatic arterial bifurcations that might provide multiple orifices in a right liver graft. Only one right liver graft (1%) had multiple arterial orifices. Single arterial anastomosis without interposition was possible in all patients with right liver grafts and none of them were complicated with hepatic arterial thrombosis. Single arterial anastomosis, therefore, has a high probability of success in right liver graft implantation. (*Liver Transpl* 2004;10:129–133.)

Living donor liver transplantation (LDLT) is a preferable treatment for adults with end-stage liver disease due to the limited number of available cadaveric donors.¹ Fundamental to the application of this technique is an understanding of hepatic vascular anatomy.² Michels first reported 10 basic types of hepatic arterial supply.³ Since then, common and rare hepatic artery variants have been reported. Most of these studies, however, focused only on replaced or accessory arterial branches that are helpful for whole-liver harvesting and transplantation. Without information regarding bifurcation of the right hepatic artery (RHA), the classification is of little help for right liver harvesting.

Recently, Marcos et al proposed the use of interposition arterial grafts in right liver graft because double hepatic arteries were common in their series.⁷ Their report conflicted with our experience because, in our series, no patients underwent double hepatic artery reconstruction in right liver LDLT. To clarify this inconsistency, we evaluated celiac and mesenteric angiograms of 223 consecutive living donors in a single institution. The aim of the study was to determine a useful anatomic classification of the hepatic arteries for LDLT using right liver grafts.

Materials and Methods

Donors

From January 1996 until May 2003, 223 consecutive living donors underwent hepatectomy at the University of Tokyo Hos-

pital. They comprised 126 men and 97 women with a median age of 34 years (range, 18–63 years). Details regarding selection criteria and evaluation are described elsewhere.⁸ Only one case was rejected due to arterial anatomy.⁹ All of the donors were related to the recipients. The relation of the donors to the patients was 84 parents, 65 children, 37 siblings, 22 spouses, 9 nephews, and 4 uncles and two cousins. The type of graft was determined by volumetric analysis and not by vascular anatomy. The graft estimate was determined by computed tomography (CT). A graft-volume-to-recipient-standard-liver-volume ratio¹⁰ of 40% was the lower limit. Candidates in whom the right liver comprised more than 70% of the whole liver were rejected as prospective donors. The most common procedure was left liver with or without caudate lobe resection (n = 85), followed by right liver resection (n = 72), left lateral segmentectomy (n = 51), and right lateral resection of right lateral sector (n = 15). All donors provided written informed consent.

Angiography of celiac and mesenteric arteries was performed in each donor to evaluate the anatomy of the donor's hepatic artery. First, the anatomy was reviewed according to Michels's classification.³ Thereafter, the anatomy was classified from the point of view of whether single or multiple anastomoses were needed in LDLT using the right liver. In

Abbreviations: A6, accessory branch from segment VI; CT, computed tomography; Ce, celiac axis; GDA, gastroduodenal artery; LDLT, living donor liver transplantation; LHA, left hepatic artery; LGA, left gastric artery; MHA, middle hepatic artery; PSPDA, superior pancreaticoduodenal artery; RHA, right hepatic artery; RL, lateral branch of right hepatic artery; RPM, paramedian branch of right hepatic artery; SA, splenic artery; SMA, superior mesenteric artery.

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Table 1. Anatomy Classification Stratified by Michels

Type	Description	Michels's Series (n = 200)	Present Series (n = 223)
1	Normal	55%	61%
2	Replaced LHA from LGA	10%	14%
3	Replaced RHA and MHA from SMA	11%	4%
4	Replaced LHA from LGA, and replaced RHA from SMA	1%	0
5	Accessory LHA	8%	12%
6	Accessory RHA	7%	3%
7	Accessory LHA and accessory RHA	1%	2%
8	Accessory LHA and replaced RHA, or replaced LHA and accessory RHA	2%	0
9	PHA from SMA	4.5%	6%
10	PHA from LGA	0.5%	0

Abbreviations: LGA, left gastric artery; LHA, left hepatic artery; MHA, middle hepatic artery; RHA, right hepatic artery; SMA, superior mesenteric artery.

this classification, the anatomy of the left hepatic artery (LHA) was not always considered.

Patients Receiving Right Liver Grafts

The actual arterial anastomosis was reviewed in 72 patients who underwent primary LDLT using right liver grafts. The patients were 53 males and 19 females with a mean age of 50 years. The indications for LDLT in these patients included hepatitis C virus with cirrhosis (n = 25), hepatitis B virus with cirrhosis (n = 13), primary biliary cirrhosis (n = 12), fulminant hepatic failure (n = 8), cryptogenic cirrhosis (n = 5), metabolic disorders (n = 3), biliary atresia (n = 3), primary sclerosing cholangitis (n = 2), and autoimmune hepatitis (n = 1).

The surgical details of the recipients were described previously.¹¹ In brief, hepatic arterial reconstruction was performed under a surgical microscope by a microsurgeon (HA). The donor and recipient arterial branches were anastomosed in an end-to-end manner with interrupted sutures using 9-0 monofilament nylon. When the donor's arterial branch was long enough to turn over, the anterior suture was performed first. Otherwise, the posterior wall was sutured with an inside-outward procedure using double needles; thereafter, the anterior wall was sutured without turning the anastomotic site over.¹² After reconstruction, the intrahepatic arterial signals in each segment were examined using Doppler ultrasonography; the other branches were ligated after confirming pulsatile back-bleeding from the nonanastomosed cut stumps¹³ (k). When these criteria were not satisfied, the remaining arteries were anastomosed to the recipient hepatic arteries.

Results

Angiographic Classification

The frequency of each type proposed by Michels is shown in Table 1. There was a similar distribution between types in Michels's series and ours. There were no Michels types 4, 8, and 10 in our series, however.

To predict the number of hepatic artery stumps in right liver LDLT, the anatomy was classified into four types (Fig. 1). The frequency of each type is shown in Table 2. Type I secures a single arterial orifice in the right liver graft. This type is divided into six subcategories. Type IA, normal anatomy in which RHA originates from the common hepatic artery and the middle hepatic artery (MHA) originates from the LHA; Type IB, same variation as Type IA, except that the MHA originated from the RHA; Type IC, replaced RHA from superior mesenteric artery (SMA); Type ID, replaced RHA and MHA from the SMA; Type IE, entire common hepatic artery from the SMA and the MHA from the LHA; Type IF, same as with Type IE except for the MHA originated from the RHA.

The hepatic arterial bifurcations that might provide multiple orifices in the right liver graft were divided into three types. A total of 14 donors (6%) were classified into these types. In Type II, the MHA originated from the paramedian (Type IIA) or lateral branch (Type IIB) of the RHA. In Type III, the right paramedian and lateral branch of the RHA had separated origins. This type was divided into two subtypes with a right lateral branch from the LHA (Type IIIA) or from the SMA (Type IIIB). In Type IV, there was an accessory branch from segment VI (A6). This type was divided into three subtypes according to the root of A6 as follows: Type IVA, from the hepatic artery proper; Type IVB, from the celiac trunk; and Type IVC from the superior pancreaticoduodenal artery. The relation between Michels's classification and ours is shown in Table 3.

Donors of Right Liver Resection

Among the 72 donors who underwent right liver resection, angiographic analysis predicted one Type IVA