あると思われる。

F.健康危険情報
なし

G.研究発表
1.論文発表
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2.学会発表
なし

H.知的財産権の出願・登録状況（予定を含む）
1.特許取得　なし
2.実用新案登録　なし
3.その他　なし

別紙5
研究成果の刊行に関する一覧表
雑誌

発表者氏名	論文タイトル名	発表誌名	巻名	ページ	出版年
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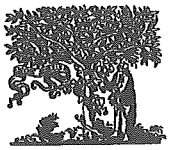
＊ 研究成果に関する書籍・論文一覧に記載された業績は別刷（ｺﾋﾟｰ可）を同封下さいますようお願い申し上げます。

Ⅳ．研究成果の刊行物･別刷

Lung Cancer (2006) 51, 363–368



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EKB-569, a new irreversible epidermal growth factor receptor tyrosine kinase inhibitor, with clinical activity in patients with non-small cell lung cancer with acquired resistance to gefitinib

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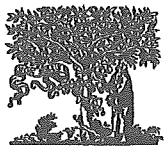

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EKB-569, a new irreversible epidermal growth factor receptor tyrosine kinase inhibitor, with clinical activity in patients with non-small cell lung cancer with acquired resistance to gefitinib

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KEYWORDS
EKB-569;
Non-small cell lung cancer;
EGFR mutation;
Resistance to gefitinib;
Irreversible inhibitor of EGFR

Summary EKB-569 is a potent, low molecular weight, selective, and irreversible inhibitor of epidermal growth factor receptor (EGFR) that is being developed as an anticancer agent. A phase 1, dose-escalation study was conducted in Japanese patients. EKB-569 was administered orally, once daily, in 28-day cycles, to patients with advanced-stage malignancies known to overexpress EGFR. Two patients with advanced non-small cell lung cancer with EGFR mutations and acquired gefitinib resistance from the phase 1 study are described in detail. *Case #1* is a 63-year-old man with smoking history. He received treatment from 4 March 2004. Because he had no severe adverse events, a total of 10 courses of therapy were completed through December 16. Grade 2 skin rash and ALT elevation, and grade 1 diarrhea and nail changes developed. A chest CT scan on 4 August 2003 revealed multiple pulmonary metastases that had decreased in size. *Case #2* is a 49-year-old woman with no smoking history. She received therapy from 9 February 2004. She received a total of five courses of the therapy until 22 June 2004. Grade 3 nausea and vomiting

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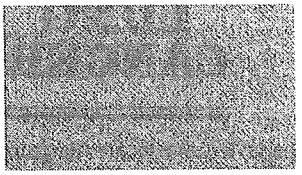

and grade 1 diarrhea and dry skin developed. A chest CT scan on March 3 revealed multiple pulmonary metastases that had decreased in size. A brain MRI on March 4 showed that multiple brain metastases also had decreased in size. Based on RECIST criteria, they had stable disease but radiographic tumor regression was observed.

1. Introduction

1.1. Efficacy of gefitinib

The epidermal growth factor receptor (EGFR) autocrine pathway contributes to a number of processes important to cancer development and progression, including cell proliferation, apotosis, angiogenesis, and metastatic spread [1]. EGFR-tyrosine kinase has become a particularly promising drug targeting for treating non-small cell lung cancer. Gefitinib is an orally active, selective EGFR tyrosine kinase inhibitor that blocks signal transduction pathways implicated in proliferation and survival of cancer cells [2]. Responsiveness characteristics include distinct subgroups of women, patients who have never smoked, patients with adenocarcinoma, and Asians [3–5]. Molecular predictive markers have also been investigated. It is suggested that MAPK is a predictive marker for survival after treatment with gefitinib in chemo-naive patients with bronchioloalveolar carcinoma [6]. Patients with P-Akt-positive tumors who received gefitinib had a better response rate, disease control rate, and time to progression than patients with P-Akt-negative tumors, suggesting that gefitinib may be most effective in patients with basal Akt activation [7]. However, it was not possible to predict gefitinib sensitivity by the level of EGFR overexpression as determined by immunohistochemistry [8] or immunoblotting [9]. Recently it has been reported that somatic mutations in the tyrosine kinase domain of the *EGFR* gene occur in a subset of patients with lung cancer who showed a dramatic response to the EGFR tyrosine kinase inhibitors gefitinib and erlotinib [10–12]. All of these mutations were within exons 18 through 21 of the kinase domain of the *EGFR* gene.

1.2. Drug summary

EKB-569 (Wyeth Research, Collegeville, PA) is a potent, low molecular weight, selective, and irreversible inhibitor of EGFR that is being developed as an anticancer agent. EGFR is a receptor tyrosine kinase that is activated by a variety of growth factors. Upon binding ligands, including epidermal growth factor (EGF) or transforming growth factor alpha (TGF-α), EGFR dimerizes and its intracellular kinase domain is activated, leading to the recruitment and phosphorylation of a number of proteins that ultimately lead to cell growth [13,14]. Several features of EKB-569 may provide certain advantages over other EGFR inhibitors. First, EKB-569 is an orally available, small-molecule EGFR inhibitor, whereas antibody-targeted EGFR inhibitors require intravenous (IV) administration. Second, EKB-569 is an irreversible inhibitor of EGFR, while other small-molecule EGFR inhibitors bind EGFR reversibly [15].

1.3. Effects in humans (Japanese)

A phase 1, open-label, dose-escalation study to assess the safety, tolerability, and pharmacokinetics of EKB-569 was conducted in Japanese patients. EKB-569 was administered orally, once daily, in 28-day cycles, to patients (pts) with advanced-stage malignancies known to overexpress EGFR. Enrollment and treatment are completed; 15 pts (six men, nine women) were treated with 25 mg (3 pts), 35 mg (8 pts), or 50 mg (4 pts) of EKB-569. Their median age was 62 years (range 47–72); ECOG performance status varied: 0 = 4/15 (26.7%) or 1 = 11/15 (73.3%).

The most frequently occurring tumor types included non-small cell lung (10 pts) and breast (2 pts). The remaining tumors were renal, leiomyosarcoma, and malignant thymoma (1 pt each). The most frequently reported EKB-569-related adverse events were diarrhea (86.7%), rash (53.3%), anorexia (40.0%), and dry skin (40.0%). Dose-limiting toxicities were observed at the 50-mg dose level with grade 4 interstitial lung disease and grade 3 diarrhea, stomatitis, and increased blood calcium levels. Thus, the maximum tolerated dose was 35 mg EKB-569 per day.

1.4. Molecular analysis of lung cancer specimens

We obtained appropriate approval from the institution and written informed consent from the patients for the comprehensive use of tumor samples for molecular and pathologic analyses. Surgically resected tumor samples were obtained retrospectively before the patients received

any systemic treatment. All of these tumors were formalin fixed and paraffin embedded by the Department of Pathology. To minimize non-neoplastic tissue contamination, the tumor portion was first selected and marked on an H&E-stained tissue section slide by a pathologist. Only the tumor portion was dissected from the unstained tissue section and sent for DNA extraction.

DNA was extracted from the paraffin section containing a representative portion of each tumor, using the QIAamp DNA Mini kit (Qiagen, Hilden, Germany). For mutational analysis of the kinase domain of the *EGFR* coding sequence, exons 19, 20, and 21 were amplified with three pairs of primers (exon 19, F: 5′-TCACAATTGCCAGTTAACGTCT-3′-(this is the convention for writing a primer), R: 5#-cagcaaagcagaaactcacatc; exon 20, F: 5#-tgaaact-caagatcgcattcat, R: 5#-catggcaaactcttgctatcc; exon 21, F: 5#-gagcttcttcccatgatgatct, R: 5#-gaaaatgctggctgacctaaag). The PCR conditions were one cycle at 95 °C for 11 min, 46 cycles at 95 °C for 30 s, 60 °C for 30 s, 72 °C for 40 s, followed by one cycle at 72 °C for 7 min. PCR products were diluted and cycle-sequenced using the Big Dye Terminator v3.1/1.1 cycle sequencing kit (Applied Biosystems, Forster City, CA) according to the manufacturer's instructions. Sequencing products were electrophoresed on an ABI PRISM 3100 Genetic Analyzer (Applied Biosystems). All sequencing reactions were performed in both forward and reverse directions and chromatograms were reviewed manually and analyzed by BLAST (basic local alignment search tool). High-quality sequence variations found in both directions were scored as candidate mutations.

2. Clinical cases

Two patients from the Japanese phase 1 study are described in detail.

2.1. Case #1

A 63-year-old man with smoking history (BI: 720) who was treated for hyperlipidemia and hypertension showed an abnormal chest X-ray in February 1996. Further examinations including a chest computed tomography (CT) scan and bronchoscopy revealed an adenocarcinoma of the lung, c-T1N0M0, stage Ia, in the right upper lobe. He had undergone a right upper lobectomy with mediastinal lymph node dissection in July 1996 and was proven to have a well-differentiated adenocarcinoma, p-T1N0M0, stage Ia. After further follow-up, multiple pulmonary metastases in both lungs were found in January 2000. Then he was given first-line chemotherapy of cisplatin and docetaxel beginning in May 2000. After two courses of this regimen, multiple pulmonary metastases had not increased in size by CT scan; however skin metastases were found. He was started on oral gefitinib 250 mg/day on November 2000. After 4 weeks, a CT scan indicated a reduction of multiple pulmonary metastases. During this treatment, grade 2 rash and grade 1 nail changes, AST/ALT elevations, and diarrhea were observed. On June 2002, multiple pulmonary metastases had increased, and this treatment was discontinued. The patient entered a phase I study of a new EGFR tyrosine kinase inhibitor (TAK-165), starting treatment on October 2002. After 2 weeks of treatment, grade 3 anorexia was observed and the therapy was stopped. On February 2003, multiple pulmonary metastases had more increased, and on March 2003, he entered a phase I study of EKB-569, receiving treatment from 4 March 2004. EKB-569 (25 mg) was administered orally, once daily, in 28-day cycles. Because he had no severe adverse events, a total of 10 courses of therapy were completed through December 16. Grade 2 skin rash and ALT elevation, and grade 1 diarrhea and nail changes developed during this therapy. Based on RECIST criteria, the patient had stable disease (SD) but radiographic tumor regression was observed on 4 August 2003 (day 27 in the sixth course) (Fig. 1). The size of multiple pulmonary metastases increase by CT scan on 8 December 2003, and the treatment was stopped on 17 December 2003.

A lung cancer specimen was obtained at surgery and studied by immunohistochemistry. EGFR over-expression was detected. In addition, we found the heterozygous in-frame deletion E746-A750 in exon 19 of the *EGFR* gene by direct sequencing of the specimen.

2.2. Case #2

A 49-year-old woman with no smoking history, who was treated for Basedow's disease, insomnia, and bronchial asthma, had an abnormal chest X-ray in October 2000. Further examinations including a chest CT scan and bronchoscopy revealed lung cancer in the left upper lobe. She was diagnosed with adenocarcinoma, c-T1N0M0, stage Ia. She had a left-upper lobectomy with mediastinal lymph node dissection, which revealed a well-differentiated adenocarcinoma, p-T4N2M1, stage IV. She was then given first-line chemotherapy of carboplatin and paclitaxel beginning in January 2001. After two courses of therapy, she discontinued treatment because of adverse events. Right supraclavicular lymph node metastases were found on August

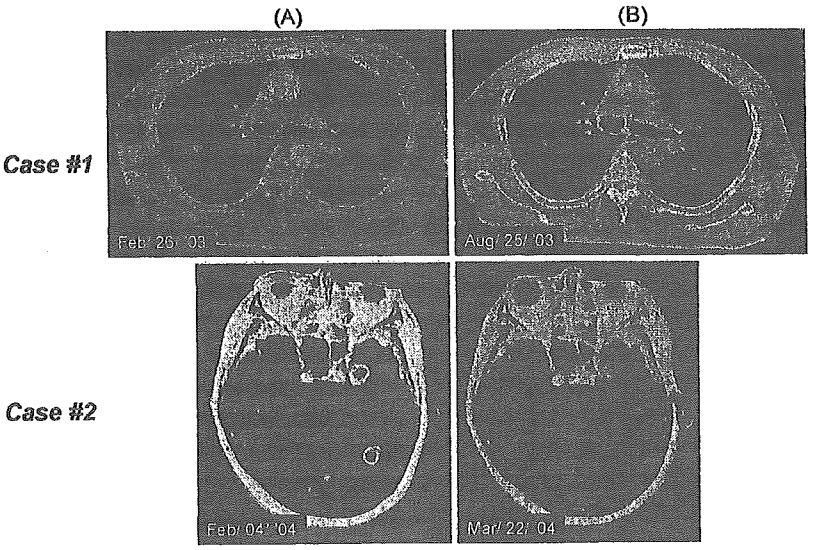

Fig. 1 *Clinical case* #1: a 63-year-old man with adenocarcinoma of lung. CT scan before treatment (A) and after initiation of EKB-569 (B). *Clinical case* #2: a 49-year-old woman with adenocarcinoma of brain metastasis. MRI scan before treatment (A) and after initiation of EKB-569 (B).

2001. Radiotherapy for the metastases (60 Gy/30 fractions) was done, and they decreased in size. On March 2002, right supraclavicular lymph node metastases increased and left clavicular lymph node metastases were found. On April 2002, the patient enrolled in a phase II trial of cisplatin, gemcitabine, and irinotecan for non-small-cell lung cancer. After two courses of therapy, bone metastases were found and pulmonary metastases had grown slowly so the treatment was stopped. She entered a phase I study of a new EGFR tyrosine kinase inhibitor (TAK-165) and started treatment on July 2002. The treatment was stopped after a week later due to grade 3 fatigue. In September 2002, the patient was started on oral gefitinib 250 mg/day. While she was taking 250 mg gefitinib daily for 15 months, the size of multiple pulmonary and bone metastases did not increase by CT scan and she had SD. On December 2003, the patient developed grade 3 oral mucositis and discontinued treatment. On January 2004, the size of multiple pulmonary and bone metastases increase by CT scan. She then entered a phase I study of EKB-569 and received therapy from 9 February 2004. EKB-569 (35 mg) was administered orally, once daily, in 28-day cycles. She received a total of five courses of the therapy until 22 June 2004. Grade 3 nausea and vomiting and grade 1 diarrhea and dry skin developed during the therapy. A chest CT scan on March 3 (day 24 in the first course) revealed multiple pulmonary metastases that had decreased in size. A brain MRI on March 4 (day 25 in the first course) showed that multiple brain metastases also had decreased in size (Fig. 1). The response was SD by RECIST criteria, although tumor regression was observed. The size of bone metastases increase by CT scan on 18 June 2004, and the treatment was stopped on 22 June 2004.

A lung cancer specimen was obtained by surgery and studied by immunohistochemistry. EGFR overexpression was detected. This lung cancer specimen had a heterozygous point mutation in exon 21 (L858R, CTG to CGG) of the *EGFR* gene.

3. Discussion

This is the first case report to describe the effects of EKB-569 on patients with adenocarcinoma of the lung. Case 1 is a 63-year-old man with a smoking history (BI: 720), and case 2 is a 49-year-old woman with no smoking history. Case 1 had an exon 19 deletion of E746-A750, and case 2 had an exon 21-point mutation. These patients underwent surgery and were treated with platinum-based chemotherapy and EGFR tyrosine kinase inhibitors. The treatment with EKB-569 was effective in these two patients after resistance to gefitinib and cytotoxic chemotherapy. These cases suggest that EKB-569 is effective in patients with *EGFR* mutations as has been reported for gefitinib and erlotinib. Despite initial responses to these EGFR inhibitors, patients eventually progress by unknown mechanisms of "acquired" resistance.

Recently, a second mutation in the *EGFR* kinase domain, which is associated with acquired resistance of non-small cell lung cancer to gefitinib or erlotinib, was reported [16,17]. Pao et al. showed that in two of five patients with acquired resistance

to gefitinib or erlotinib, progressing tumors contained, in addition to a primary drug-sensitive mutation in EGFR, a secondary mutation in exon 20. This mutation leads to a substitution of methionine for threonine at position 790 (T790M) in the kinase domain [16]. Kobayashi et al. reported the case of a patient with EGFR-mutant, gefitinib-responsive, advanced non-small cell lung cancer who relapsed after two years of complete remission during treatment with gefitinib. The DNA sequence of the EGFR gene in his tumor biopsy specimen at relapse also revealed the presence of the secondary point mutation, T790M [17]. Kurata et al. reported an interesting case in which acquired resistance to gefitinib could be overcome [18]. In this case, the patient received gefitinib, then a combination of nedaplatin and gemcitabine, and then gefitinib again. The cytotoxic agents may have altered the *EGFR* gene or associated genes to produce acquired sensitivity to gefitinib.

Kobayashi et al. also found that CL-387,785, a specific and irreversible, anilinoquinoline EGFR inhibitor [19], strongly inhibited the EGFR kinase in cells transfected with DNA containing the L747-S752 deletion in the *EGFR* gene or a double mutation with the L747-S753 deletion and the T790M point mutation. They speculated that CL-387,785 inhibited the EGFR kinase of the double mutant because of its altered binding to the kinase domain or its covalent binding to EGFR [17]. Kwak et al. used a bronchoalveolar cancer cell line with an L746-A750 deletion in the *EGFR* gene to isolate gefitinib-resistant clones. These clones had not acquired secondary EGFR mutations but were sensitive to the irreversible, anilinoquinoline EGFR inhibitor EKB-569 [20].

We have shown that EKB-569 had clinical activity in two patients with advanced non-small cell lung cancer with EGFR mutations and acquired gefitinib resistance. Thus, irreversible EGFR inhibitors may be an effective therapy for patients with EGFR-mutant advanced non-small cell lung cancer who have relapsed after treatment with gefitinib.

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Enhancement of Sensitivity to Tumor Necrosis Factor α in Non–Small Cell Lung Cancer Cells with Acquired Resistance to Gefitinib

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Abstract Tumor cells that have acquired resistance to gefitinib through continuous drug administration may complicate future treatment. To investigate the mechanisms of acquired resistance, we established PC-9/ZD2001, a non-small-cell lung cancer cell line resistant to gefitinib, by continuous exposure of the parental cell line PC-9 to gefitinib. After 6 months of culture in gefitinib-free conditions, PC-9/ZD2001 cells reacquired sensitivity to gefitinib and were established as a revertant cell line, PC-9/ZD2001R. PC-9/ZD2001 cells showed collateral sensitivity to several anticancer drugs (vinorelbine, paclitaxel, camptothecin, and 5-fluorouracil) and to tumor necrosis factor α (TNF-α). Compared with PC-9 cells, PC-9/ZD2001 cells were 67-fold more sensitive to TNF-α and PC-9/ZD2001R cells were 1.3-fold more sensitive. Therefore, collateral sensitivity to TNF-α was correlated with gefitinib resistance. PC-9/ZD2001 cells expressed a lower level of epidermal growth factor receptor (EGFR) than did PC-9 cells; this down-regulation was partially reversed in PC-9/ZD2001R cells. TNF-α-induced autophosphorylation of EGFR (cross-talk signaling) was detected in all three cell lines. However, TNF-α-induced Akt phosphorylation and IκB degradation were observed much less often in PC-9/ZD2001 cells than in PC-9 cells or PC-9/ZD2001R cells. Expression of the inhibitor of apoptosis proteins c-IAP1 and c-IAP2 was induced by TNF-α in PC-9 and PC-9/ZD2001R cells but not in PC-9/ZD2001 cells. This weak effect of EGFR on Akt pathway might contribute to the TNF-α sensitivity of PC-9/ZD2001 cells. These results suggest that therapy with TNF-α would be effective in some cases of non-small-cell lung cancer that have acquired resistance to gefitinib.

Gefitinib (Iressa, ZD1839), a small-molecule epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor, has been approved for the treatment of refractory and relapsed non-small-cell lung cancer (NSCLC) patients in a number of countries around the world. This drug, which is given continuously as a once-daily oral dose, showed antitumor activity in patients with relapsed or recurrent NSCLC; however, tumor responses were observed in 12% to 18% of patients with chemotherapy-refractory advanced NSCLC (1, 2). Even in cases sensitive to gefitinib, resistance might be acquired through continuous drug administration. Additional treatments for cases of NSCLC relapsing during treatment with gefitinib are urgently needed.

To investigate the mechanism of acquired resistance to gefitinib, we previously established gefitinib-acquired resistant cells, PC-9/ZD2001, from a NSCLC, PC-9, which is hypersensitive to gefitinib and has a 15-del mutation in exon 19 of EGFR (data not shown). After >6 months of culture in gefitinib-free conditions, the sensitivity of PC-9/ZD2001 cells to gefitinib was restored, and the cells were subsequently established as a revertant cell line, PC-9/ZD2001R. The active mutation of EGFR was sustained in both the resistant and the revertant cell lines and the existence of revertant cell line suggests the additional mutation of EGFR, such as a secondary mutation of T790M in EGFR that causes resistance to gefitinib (3, 4), is unlikely to be contribute to this gefitinib resistance. In the gefitinib-resistant cells, the expression levels of EGFR and mRNA decreased to 30% to 50% of those in parental cells. A ligand-induced EGFR activation minimally activated mitogen-activated protein kinase signaling pathways and the inhibitory effect of gefitinib on this

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pathway was significantly decreased in the resistant cells.[5] To elucidate the cross-resistance to other anticancer agents, we examined the sensitivity to the conventional anticancer agents and tumor necrosis factor α (TNF-α). PC-9/ZD2001 showed cross-resistance to another EGFR inhibitor, AG1478. Interestingly, gefitinib-resistant cells were ~3-fold more sensitive than PC-9 cells to the cytotoxic effects of vinorelbine, paclitaxel, camptothecin, 5-fluorouracil, and a cytokine, TNF-α.[5] The same tendency was confirmed in the other gefitinib-resistant clones established along with PC-9/ZD2001. The restoration of these collateral sensitivities (except 5-fluorouracil) in revertant PC-9/ZD2001R cells suggests that such sensitivities are correlated with the mechanism of gefitinib resistance.

TNF-α is the prototype of ~20 related cytokines that act through specific members of the TNF receptor (TNFR) super family (5–7). Several cancer therapies exploiting the cytotoxic effect of TNF-α on solid tumors and soft-tissue sarcomas have recently been examined in clinical trials (8, 9). The TNF-α stimulates inflammation by turning on gene transcription through signaling cascades such as the Akt/nuclear factor κB (NF-κB) pathway. This signaling subsequently serves as the primary mechanism to protect cells against apoptotic stimuli through several transcriptional genes, such as inhibitor of apoptosis proteins (IAP), the specific inhibitor of caspases (10, 11). In contrast, TNF-α-mediated signaling also triggers apoptosis through the activation of caspase-8 and the downstream caspase-3 or caspase-7 in a wide variety of cells (12). From these observations, it is possible to say that TNF-α has two different signaling pathways that contradict each other. The cytotoxic effect of TNF-α might be determined by ratios between the apoptosis-inducing and the apoptosis-inhibiting effects.

Akt/NF-κB signaling also occurs downstream of EGFR and this signaling mediates cell proliferation and antiapoptotic signaling through this pathway (13). In the case of the antiapoptotic signaling of TNF-α, TNFR is known to activate Akt/NF-κB in three ways: directly through phosphatidylinositol 3-kinase activation, or indirectly through cross-talk signaling to EGFR, or both together (5–7, 12, 14, 15). Moreover, several recent articles report that the TNFR-mediated cross-talk signaling to EGFR occurs in a ligand-dependent and -independent manner (16–21). Therefore, to investigate the mechanisms of the collateral sensitivity to TNF-α in gefitinib-acquired resistant cells, we focused on TNF-α-induced cross-talk signaling to EGFR and analyzed the Akt/NF-κB signaling pathway in response to TNF-α.

In this article, we show that a weakness of Akt/NF-κB signaling from TNF-α-mediated cross-talk signaling via EGFR causes the collateral sensitivity to TNF-α in the gefitinib-acquired resistant cell line. Moreover, this cross-talk signaling is thought to be a dominant pathway of TNF-α-mediated Akt activation.

Materials and Methods

Chemicals and antibodies. Gefitinib was donated by AstraZeneca Pharmaceuticals (Wilmington, DE). An anti-phospho-EGFR antibody (Tyr1068) was purchased from Cell Signaling Technology (Beverly, MA). Other antibodies and chemicals were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA) and Sigma-Aldrich Co. (St. Louis, MO), respectively, unless otherwise specified.

Cell lines and cultures. The PC-9 human NSCLC cell line, established from a previously untreated patient, was kindly donated by Prof. K. Hayata (Tokyo Medical College, Tokyo, Japan.). The PC-9 cells were cultured with RPMI 1640 supplemented with 10% FCS and maintained in a 5% CO_2 incubator at 37°C under humidified conditions.

Establishment of gefitinib-resistant cell lines. To establish gefitinib-resistant cell lines, PC-9 cells were continuously exposed to increasing dosages of gefitinib for >1 year. The surviving cells were cloned and three gefitinib-resistant cell lines, designated as PC-9/ZD2001, PC-9/ZD2002, and PC-9/ZD2003, were established. These cell lines can survive exposure to 200 nmol/L gefitinib. Sensitivity to gefitinib was restored by culture of PC-9/ZD2001 in gefitinib-free conditions for >6 months. The restored cells were cloned and subsequently established as a revertant cell line, PC-9/ZD2001R.

Established resistant cell lines were maintained by culture in a medium containing 200 nmol/L gefitinib. To eliminate the effects of gefitinib, the resistant cells were cultured in a drug-free medium for at least 2 weeks before all experiments. As the relative resistance values of these cell lines were stable for at least 3 months after culture under drug-free conditions (data not shown), we used the cells for experiments during this period.

Growth inhibition assay. To measure sensitivity to gefitinib, a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay was done (Cell Titer 96 assay kit, Promega Corp., Madison, WI). In brief, PC-9, PC-9/ZD2001, and PC-9/ZD2001R cells were seeded onto 96-well plates and preincubated overnight. The cells were continuously exposed to the indicated concentrations of gefitinib for 4 or 5 days. Absorbance was measured at 570 nm with a microplate reader (Model 550, Bio-Rad Laboratories, Hercules, CA).

Analysis of tumor necrosis factor α–induced apoptotic cell death. The PC-9, PC-9/ZD2001, and PC-9/ZD2001R cells were treated with 100 ng/mL TNF-α for the indicated time periods. They were then fixed with 4% paraformaldehyde at 4°C for 30 minutes. After 100 μL of 70% ethanol were added, the cells were permeabilized by incubation overnight at −20°C. Apoptotic DNA fragments were probed with the terminal deoxyribonucleotidyl transferase-mediated dUTP nick end labeling method (MEBSTAIN Apoptosis TUNEL Kit Direct, Medical & Biological Laboratories, Nagoya, Japan) and subpopulations of apoptotic cells were measured with a flow cytometer (FACSCalibur, BD Biosciences Immunocytometry Systems, San Jose, CA).

Activity assays for CPP32/caspase-3 and FLICE/caspase-8. Activities of CPP32/caspase-3 and FLICE/caspase-8 were measured with caspase-3 and caspase-8 colorimetric assay kits (MRL Diagnostics, Cypress, CA) according to the instructions of the manufacturer. The PC-9, PC-9/ZD2001, and PC-9/ZD2001R cells were incubated for 12 hours with 10 ng/mL TNF-α and then resuspended in 50 μL of chilled cell lyses buffer. The cells were incubated on ice for 10 minutes and the protein concentration of the supernatant was assayed with a bicinchoninic acid protein assay kit (Sigma-Aldrich). A certain amount of each sample was added to 50 μL of 2× reaction buffer containing the respective substrates DEVD-pNA and IETD-pNA, then incubated at 37°C for 1 hour. After incubation, absorbance was measured at 400 and 405 nm with a microtiter plate reader (Model 550, Bio-Rad Laboratories).

Immunoblot analysis. Cells were treated with 10 ng/mL of TNF-α for 30 minutes, then washed twice with ice-cold PBS and lysed in EBC buffer [50 mmol/L Tris-HCl (pH 8.0), 120 mmol/L NaCl, 0.5% NP40, 100 μmol/L NaF, 200 μmol/L Na orthovanadate, and 10 μg/mL of leupeptin, aprotinin, and phenylmethylsulfonyl fluoride] with an ultrasonic disrupter (Tomy Digital Biology Co., Ltd., Tokyo, Japan). The cell lysate was precleared by centrifugation, resolved by 10% SDS-PAGE, transferred to nitrocellulose membrane, and probed with antibodies against EGFR, phospho-EGFR (Tyr1045), phosphatase and tensin homologue, Akt, phospho-Akt, IκB, c-IAP1, and c-IAP2. Bound antibodies were detected with horseradish peroxidase-linked immunoglobulin (Amersham Biosciences, Buckinghamshire, United Kingdom)

[5] T. Yamaoka, T. Ohmori, F. Inoue, et al. Characteristics of gefitinib-acquired resistance in non–small cell lung cancer cell lines, submitted for publication.

and enhanced chemiluminescence reagents (Perkin-Elmer Life and Analytical Sciences, Boston, MA).

Real-time reverse transcription-PCR method. Total RNA was isolated with the guanidium isothiocyanate method using an RNA purification kit (RNeasy Mini Kit, Qiagen, Venlo, the Netherlands) according to the instructions of the manufacturer. After RNA isolation, cDNA was prepared in the presence of random 9-mers with a reverse transcription-PCR (RT-PCR) kit (Takara Shuzo Co., Ltd., Kyoto, Japan). Expression levels of EGFR, c-IAP1, and c-IAP2 mRNA were quantified with a fluorescence-based real-time detection method (GeneAmp 5700 Sequence Detection System, Applied Biosystems, Foster City, CA). Cycling conditions were 40 cycles at 94°C for 20 seconds, 55°C (EGFR) and 64°C (c-IAPs) for 20 seconds, and 72°C for 30 seconds. Expression of the mRNA was measured with the following primer sets: EGFR, 5′-ACGAATGGGCCTAAGATC-3′ and 5′-TGCTTACCCGGATTCTAGG-3′; c-IAP1, 5′-ATGTGGGTAACAGTGATGATGTCA-3′ and 5-AAACCAC-TTGGCATGTTGAAC-3′; and c-IAP2, 5′-CTAGTGTTCATGTTGAAC-3′ and 5′-CCTCAAGCCACCATCACAAC-3′. The expression of β-actin mRNA was used as an internal control.

Statistical analysis. Statistical analysis was done with the StatView II software program (Abacus Concepts, Berkeley, CA). Activities of CPP32/caspase-3 and FLICE/caspase-8 were analyzed with paired Student's t test. $P < 0.05$ was considered significant.

Results

Establishment of acquired gefitinib-resistant cell lines. To elucidate the mechanism of acquired resistance against gefitinib, we established gefitinib-resistant NSCLC cell lines through continuous exposure of this drug. Resistance against gefitinib developed quite slowly; the relative resistant values of 3- to 4-fold were reached after >1-year exposure to gefitinib. We picked the clones of gefitinib-resistant cell lines named PC-9/ZD2001, PC-9/ZD2002, and PC-9/ZD2003. These cell lines can survive in 200 nmol/L gefitinib-contained medium. Sensitivities to gefitinib were measured by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay. In the case of PC-9/ZD2001 cells, the cell line was able to survive by >50% at the concentration of >500 nmol/L gefitinib. This concentration caused maximum inhibition in PC-9. The IC_{40} value of gefitinib in PC-9 cells was 53.0 ± 8.1 nmol/L. The gefitinib-resistant cell line PC-9/ZD2001 showed a 4-fold higher resistance to gefitinib than PC-9 cells (IC_{40} = 211.1 ± 32.4 nmol/L; Fig. 1). Culture of the cells in gefitinib-free conditions for 6 months restored sensitivity to gefitinib in PC-9/ZD2001 and subsequently established a revertant cell line, PC-9/ZD2001R, in which sensitivity to gefitinib was completely restored (IC_{40} = 46.3 ± 10.2 nmol/L).

Analysis for tumor necrosis factor α–induced apoptotic cell death. TNF-α-induced cytotoxic effect was measured by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay. The IC_{40} values of TNF-α in PC-9, PC-9/ZD2001, and PC-9/ZD2001R cell lines were 815.0 ± 44.8, 12.2 ± 1.4, and 626.2 ± 18.5 ng/mL, respectively. PC-9/ZD2001 cells acquired new sensitivity to TNF-α. PC-9/ZD2001 was ~67-fold more sensitive to TNF-α as compared with PC-9, but this sensitization was restored to 1.3-fold in PC-9/ZD2001R (Fig. 2A). This collateral sensitivity to TNF-α was confirmed in the other gefitinib-resistant cell lines, PC-9/ZD2002 and PC-9/ZD2003 (data not shown).

Additionally, we measured TNF-α-induced apoptotic cell death by flow cytometry. The apoptotic cells were stained by the terminal deoxyribonucleotidyl transferase-mediated dUTP nick end labeling method. No significant apoptosis was observed in these three cell lines until 24 hours of exposure to TNF-α (10 ng/mL). Forty-eight hours of TNF-α exposure induced a 6-fold higher apoptotic cell death in PC-9/ZD2001 cells (70.3%) as compared with the parental PC-9 cells (11.8%). This enhancement was completely recovered in PC-9/ZD2001R cells (16.6%; Fig. 2B; Table 1). These results suggest that the collateral sensitivity to TNF-α might be correlated with the resistance to gefitinib in these cell lines.

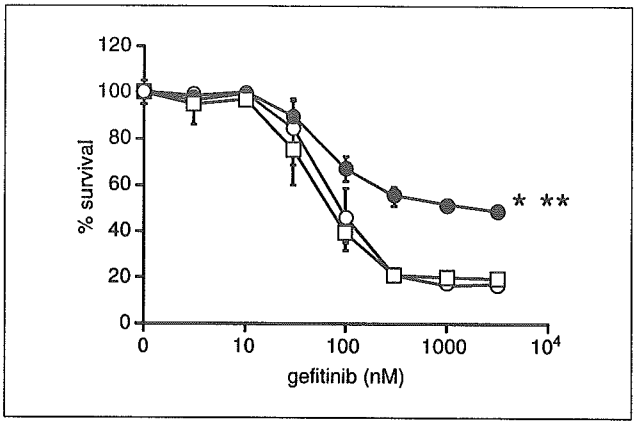

Fig. 1. Cytotoxic effects of gefitinib in a gefitinib-resistant NSCLC cell line. The cells (2×10^3 per well) were seeded onto a 96-well plate and preincubated overnight, then continuously exposed to the indicated concentrations of gefitinib for 4 or 5 days. The growth inhibition rate was analyzed with 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay as described in Materials and Methods. ○, PC-9; ●, PC-9/ZD2001; □, PC-9/ZD2001R. Points, mean of three different experiments; bars, SD. *, $P < 0.001$, PC-9 versus PC-9/ZD2001; **, $P < 0.001$, PC-9/ZD2001R versus PC-9/ZD2001.

Analysis of tumor necrosis factor α–mediated activations of CPP/caspase-3 and FLICE/caspase-8. To clarify the difference of TNF-α-induced apoptotic cell death in these cell lines, we analyzed TNF-α-mediated CPP32/caspase-3 and its upstream FLICE/caspase-8 activations by caspase-8 and caspase-3 colorimetric protease assay kits (Medical and Biological Laboratories), respectively. PC-9, PC-9/ZD2001, and its revertant PC-9/ZD2001R cells were incubated with the indicated concentrations of TNF-α for 12 hours. In the case of caspase-3, TNF-α did not cause any increases in the activity in PC-9 and PC-9/ZD2001R cells even at the highest concentration of 100 ng/mL. In contrast, TNF-α significantly enhanced caspase-3 activity in PC-9/ZD2001 cells even at the concentration of 1 ng/mL within this time course (Fig. 3A). In the case of caspase-8, TNF-α enhanced the activities in all three cell lines from 10 ng/mL (Fig. 3B). TNF-α at 100 ng/mL activated caspase-8 ~1.6-, 2.9-, and 1.9-fold higher in PC-9, PC-9/ZD2001, and PC-9/ZD2001R, as compared with the respective untreated cells. In PC-9/ZD2001 cells, TNF-α caused the highest relative induction of caspase-8 (Fig. 3B).

Immunoblot analysis for the tumor necrosis factor α–induced cross-talk signaling to epidermal growth factor receptor and Akt/nuclear factor κB pathway activation. EGFR expression was significantly lower in PC-9/ZD2001 than in PC-9 cells (Fig. 4A). When measuring the expression of EGFR protein by a densitometer (calculated by the NIH image software), the expression was decreased to 52.4 ± 2.6% of that in parental cell line. Moreover, we measured the expression levels of EGFR mRNA by a real-time RT-PCR method. The expression level in PC-9/ZD2001 was decreased to 37.0 ± 3.2% of that in parental

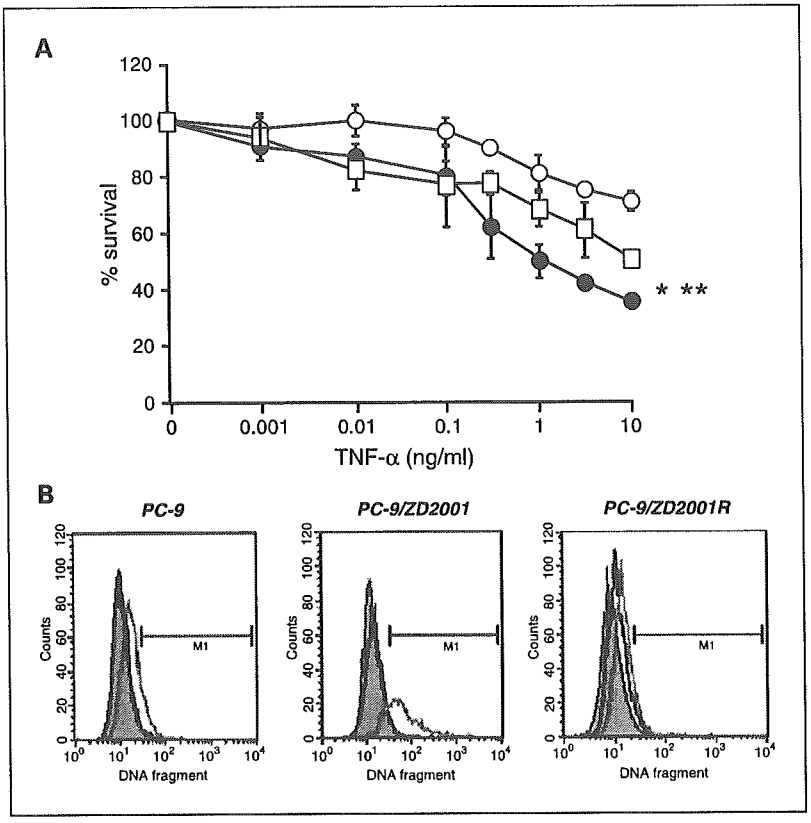

Fig. 2. Gefitinib-resistant cells acquired sensitivity to TNF-α. *A*, the cells were continuously treated with the indicated concentrations of TNF-α for 4 or 5 days. The growth inhibition rate was analyzed with 3-(4,5-dimethylthiazol-2-yl) -2,5-diphenyltetrazolium bromide assay as described in Materials and Methods. ○, PC-9; ●, PC-9/ZD2001; □, PC-9/ZD2001R. PC-9/ZD2001 cells were ~67-fold more sensitive to TNF-α than were PC-9 cells but the sensitivity of revertant PC-9/ZD2001R cells decreased to 1.3-fold that in PC-9 cells. Points, mean of three different experiments; bars, SD. *, $P < 0.001$, PC-9 versus PC-9/ZD2001, **, $P < 0.001$, PC-9/ZD2001R versus PC-9/ZD2001. *B*, the cells were treated with 10 ng/mL TNF-α for the indicated time periods. After treatment, the cells were fixed with 4% paraformaldehyde at 4°C and permeabilized with 70% ethanol. Fragments of apoptotic DNA were stained with the terminal deoxyribonucleotidyl transferase-mediated dUTP nick end labeling method and measured with flow cytometry as described in Materials and Methods.

cells. The same down-regulation of EGFR was seen in the other resistant cell lines (data not shown). In the case of PC-9/ZD2001R, expression levels of EGFR protein and mRNA were also decreased to 69.3 ± 1.1% and 56.8 ± 2.2%, respectively, as compared with PC-9. The expression of EGFR was restored, but not completely, in the revertant cell line.

In PC-9 cells, cross-talk signaling from TNFR to EGFR was observed and treatment with 10 ng/mL TNF-α for 30 minutes induced significant autophosphorylation of EGFR (Fig. 4A). According to the autophosphorylation of EGFR, definite phosphorylation of Akt and a decrease in IκB content were observed. The activation of Akt and down-regulation of IκB were inhibited by gefitinib at concentrations <10 nmol/L. Because gefitinib (100 nmol/L) mostly inhibited this signaling, we concluded that the cross-talk signaling from TNFR to EGFR might be the dominant pathway of TNF-α-mediated Akt/NF-κB activation in this cell line rather than the direct signaling from TNFR to Akt. In contrast, although EGFR autophosphorylation was observed, only partial phosphorylation of Akt and down-regulation of IκB, compared with those in PC-9, were observed after TNF-α exposure in PC-9/ZD2001 cells (Fig. 4A and B). Treatment with gefitinib inhibited this cross-talk signaling to EGFR but had no effect on downstream Akt phosphorylation.

These observations suggest that TNF-α-mediated EGFR signaling has less effect on the Akt/NF-κB pathway in the gefitinib-resistant PC-9/ZD2001 cell line. Other stimuli might activate Akt in an EGFR-independent manner. In the revertant PC-9/ZD2001R cell line, this weak effect of EGFR was largely reversed and TNF-α exposure induced autophosphorylation of EGFR and subsequent activation of the Akt/NF-κB pathway. The expression levels of phosphatase and tensin homologue, a suppressor of Akt signaling, did not differ significantly among PC-9, PC-9/ZD2001, and PC-9/ZD2001R cells. This decreased effect of EGFR might be partially caused by the down-regulation of EGFR expression in PC-9/ZD2001. However, although the EGFR-mediated signaling and the resistance to gefitinib were mostly restored, EGFR expression remained only partially restored in PC-9/ZD2001R. For this reason, we speculated that the down-regulation of EGFR expression might not fully explain the weak EGFR signaling to Akt pathway in PC-9/ZD2001 cells.

To clarify the decreased EGFR signaling in PC-9/ZD2001, we examined the inhibitory effect of a phosphatidylinositol 3-kinase inhibitor, wortmannin, on the TNF-α-induced activation of this pathway (Fig. 4B). Interestingly, wortmannin inhibited the TNF-α-mediated phosphorylation of Akt in PC-9/ZD2001 cells at the same level as it did in PC-9 and PC-9/ZD2001R cells.

Expresstion of c-IAP1 and c-IAP2 on treatment with tumor necrosis factor α. After treatment with TNF-α (10 ng/mL) for 30 minutes, expression of c-IAP1 and c-IAP2 proteins was

Table 1. Percentage of apoptotic subpopulations

%Apoptosis	PC-9	PC-9/2001	PC-9/2001R
Control	1.1	1.2	1.1
24 h	1.4	2.3	3.2
48 h	11.8	70.3	6.6

NOTE: After 72 hours of exposure to TNF-α, significant apoptotic cell death was observed in PC-9/ZD2001 cells but not in PC-9 or PC-9/ZD2001R cells.