

痴呆性疾患の介入予防に関する研究

（睡眠からの介入研究の理論指導と実践に関する研究）

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研究要旨：

高齢者では、睡眠障害が認知機能低下の引き起こし、認知症疾患発症の予防に睡眠健康改善が有用である。睡眠に係わる生活習慣のセルフマネジメント方式による改善で、睡眠健康が短期間で増進する。運動や昼寝による覚醒機能の改善が、特に睡眠改善に重要な役割を果たすことから、覚醒機能改善に係わる自律神経機能の睡眠におけるメカニズムについて検討した。

A. 研究目的

高齢者の認知症疾患の予防介入を、睡眠改善の面から遂行するための実践技術の開発及び確立とその科学的基盤解明を目的とする。

B. 研究方法

健康な成人男女 14 名（男性 11 人、女性 3 人）とし、サーカディアンリズムを統制するため、実験は各被験者の習慣的な就寝時刻に合わせ、就寝 3 時間前から行動の影響統制を行った。2 夜の終夜睡眠脳波記録を行い、心電図、直腸温を同時に連側測定し、2 夜目のデータを解析に使用した。心電図 R-R 間隔から、MEMCalc 法による心拍数変動周波数解析を行い、高周波成分(HF)、低周波成分(LF)と HF の比(LF/HF)、%LF(LF/(LF+HF))を求めた。

入眠期の睡眠段階による変化を検討するため、直腸温、HF、LF/HF、%LF の消灯 60

分前、消灯 30 分前、消灯時、入眠時（段階 2 が初めて出現した時点）、最初の段階 3 出現時、レム睡眠の直前の段階 2、レム睡眠出現時、レム睡眠終了時の夫々の値を抽出した。

また、入眠前 60 分～入眠後 120 分を抽出し、時間経過による変動も検討した。

（倫理面への配慮）

研究内容を書面で十分に説明し、自由意志での参加で、書面にて同意の得られた者のみを対象者とし、個人情報保護にも留意し、被験者情報はすべて ID のみの管理とした。

C. 研究結果

入眠潜時は平均 9.7 ± 2.4 分、睡眠率は平均 $96.4 \pm 3.2\%$ であり睡眠が良好な被験者であった。直腸温、HF、LF/HF、%LF のすべてにおいて、顕著な変化は入眠前に見られた。直腸温は消灯 60 分前から入眠時に向けて低下し、入眠後も徐々に低下した。LF/HF、%

LF は消灯 30 分前から入眠時に向けて低下し、入眠後は REM 睡眠の前の第 2 段階で有意に増加し、入眠、徐波睡眠、レム睡眠の出現に先行して変化していた。HF は消灯 60 分前から入眠時に向けて上昇し、特に消灯から入眠時に顕著な増加が見られたが、入眠後は睡眠段階による有意な変化は見られなかった。入眠前 60 分～入眠後 120 分の変化でも、直腸温、HF、LF/HF、%LF のすべてにおいて、有意な変化は入眠前に見られた。

D. 考察

入眠過程、入眠後の円滑な睡眠の進行には、心臓自律神経における交感神経活動が強く関係していることが判明した。入眠前の深部体温の急速な下降が円滑な入眠に重要であると多くの研究者により報告されているが、交感神経活動の低下が深部体温下降に先行あるいは同期し、交感神経活動が入眠を左右する支配的生理現象である可能性が示唆された。

また、交感神経活動の変化が入眠や各睡眠段階の出現に先行し睡眠段階の円滑な進行を調整している可能性が強く示唆された。

これらの結果は、睡眠においては迷走神経に代表される副交感神経活動よりも交感神経活動の変化が重要である可能性を示すものであった。

E. 結論

前頭連合野機能に強く影響を及ぼす睡眠の改善は、高齢者の意欲や記憶を含む認知機能の悪化を予防するための重要な要因である。良好な睡眠は、円滑な入眠と睡眠維持の安定および質的良好性を必要とする。入眠や円滑な睡眠の進行、ひいては睡眠の質的改善に交感神経活動の適切な動態が重要であることが、本研究より判明した。副交感神経活

動に比べ日中の状態を介入により変化させることで調整しやすい交感神経活動に注目し、より適切な睡眠改善のための生活介入技術を開発できる道が本研究により開かれた。

F. 研究発表

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G. 知的所有権の取得状況

1. 特許取得

なし

2. 実用新案登録

なし

3. その他

なし

Ⅲ. 研究成果の刊行に関する一覧表

研究成果の刊行に関する一覧表

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IV. 研究成果の刊行物・別刷

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Changes in hippocampal GABA_BR1 subunit expression in Alzheimer's patients: association with Braak staging

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Abstract Alterations in the γ -aminobutyric acid (GABA) neurotransmitter and receptor systems may contribute to vulnerability of hippocampal pyramidal neurons in Alzheimer's disease (AD). The present study examined the immunohistochemical localization and distribution of GABA_B receptor R1 protein (GBR1) in the hippocampus of 16 aged subjects with a range of neurofibrillary tangle (NFT) pathology as defined by Braak staging (I–VI). GBR1 immunoreactivity (IR) was localized to the soma and processes of hippocampal pyramidal cells and some non-pyramidal interneurons. In control subjects (Braak I/II), the intensity of neuronal GBR1 immunostaining differed among hippocampal fields, being most prominent in the CA4 and CA3/2 fields, moderate in the CA1 field, and very light in the dentate gyrus. AD cases with moderate NFT pathology (Braak III/IV) were characterized by increased GBR1-IR, particularly in the CA4 and CA3/2 fields. In the CA1 field of the majority of AD cases, the numbers of GBR1-IR neurons were significantly reduced, despite the presence of Nissl-labeled neurons in this region. These data indicate that GBR1 expression changes with the progression of NFT in AD hippocampus. At the onset of hippocampal pathology, increased or stable expression of GBR1 could contribute to neuronal resistance to the disease process. Advanced

hippocampal pathology appears to be associated with decreased neuronal GBR1 staining in the CA1 region, which precedes neuronal cell death. Thus, changes in hippocampal GBR1 may reflect alterations in the balance between excitatory and inhibitory neurotransmitter systems, which likely contributes to dysfunction of hippocampal circuitry in AD.

Keywords Alzheimer's disease · GABA_B receptor · Hippocampus · Neuronal degeneration · Neurofibrillary tangles

Introduction

Over-activation of excitatory amino acid (EAA) receptors leads to excitotoxic neuronal changes that can contribute to neuropathology in Alzheimer's disease (AD) [3, 4, 13, 30]. Excitotoxicity due to excessive EAA receptor stimulation could be countered by compensatory activation of inhibitory neurotransmission. γ -Aminobutyric acid (GABA) is the major inhibitory neurotransmitter in the central nervous system [29] and GABA signaling occurs through two major classes of receptors, the ionotropic GABA_A type, and the metabotropic GABA_B type [20]. Our previous studies demonstrated the relative stability of GABA_A receptor subunits in AD hippocampus [24, 25, 26]. The status of GABA_B receptors, however, remains to be examined.

GABA_B receptor activation increases K⁺ conductance, hyperpolarizing postsynaptic sites [19, 32] and inhibiting presynaptic Ca²⁺ conductance, thus suppressing neurotransmitter release and postsynaptic excitatory transmission [31, 37]. GABA_B receptors are composed of at least two heteromers, GABA_BR1 (GBR1) and GABA_BR2 (GBR2) [17, 18, 36]. Compared to the GBR2, GBR1 immunoreactivity (IR) is more prominent in neuronal soma and proximal dendrites [5],

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where pathological material is known to accumulate during the formation of neurofibrillary tangles (NFT) in AD. Therefore, immunohistochemical examination of the GBR1 protein in AD brain allows for the assessment of potential neuronal GABA_B receptor changes relative to the development of NFT pathology. The present study employed immunohistochemical techniques to examine the cellular localization and density of the GBR1 receptor subunit in the hippocampus of 16 elderly subjects at different stages of NFT pathology progression.

Materials and methods

Subjects

Postmortem brain tissue was obtained from 16 elderly subjects: 12 with a clinical diagnosis of AD (mean age \pm SD 77.8 ± 13.9 years) and 4 age-matched cognitively normal (CN) control subjects (mean age 73.3 ± 16.5 years). The mean postmortem interval and brain weight of the cases were 5.3 ± 1.9 h and $1,154 \pm 138$ g, respectively, with no significant difference between AD and CN groups (Table 1). Clinical diagnosis of CN subjects was based on the absence of dementia, determined through retrospective analysis of medical records as well as interviews with family physicians and immediate family members. All AD subjects were participants in a longitudinal research program maintained by the University of Pittsburgh's Alzheimer's Disease Research Center (ADRC). As participants in this program, patients underwent periodic neuropsychological and neurological evaluation. Clinical diagnosis of AD was based on a standardized ADRC evaluation at a Consensus Conference, utilizing DSM-IV [2] and NINCDS/ADRDA [21] criteria. Neuropathological diagnosis was determined by a certified neuropathologist, and was based in part on histological examination of brain tissue sections

stained with hematoxylin and eosin, thioflavin-S, and Bielschowsky silver stains. All AD subjects fulfilled CERAD criteria for the diagnosis of "definite" AD [22]. All brains (CN and AD) showed NFT, and dependent on the extent of NFT progression through the entorhinal, hippocampal, and neocortical areas, they were assigned a Braak score, according to neuropathological staging by Braak and Braak [7]. Of the 16 subjects, 4 CN controls were Braak stage I/II with only "mild" hippocampal pathology. Four of the AD patients were in Braak stage III/IV with "moderate" hippocampal pathology, and the remaining eight AD cases were Braak stage V/VI with "severe" hippocampal NFT pathology. Lewy bodies were detected in the cerebral cortex of one moderate case and three severe AD cases, but no Lewy bodies or neurites were detected in the hippocampus of any case. None of the patients included in this study had any confounding neurological or neuropathological disorder, except for isolated old infarcts in the cortex and thalamus of two severe cases (Table 1).

Tissue preparation

Brain tissue was processed according to previously described procedures [24, 26, 27]. The material for this study was obtained from a block of hippocampal tissue cut in the coronal plane at the level of the lateral geniculate body. Tissue was placed in 0.1 M phosphate buffer (PB, pH 7.4) containing 4% paraformaldehyde for 48 h at 4°C, and subsequently cryoprotected in 30% sucrose in PB for several days. Tissue sections for immunohistochemistry were cut at 40 μ m on a sliding, freezing microtome. For each case, adjacent sections were stained for Nissl substance to delineate the cytoarchitectural boundaries of hippocampal fields as defined by Duvernoy [10] and Amaral and Insausti [1].

Table 1 Case demographics [AD Alzheimer's disease, CN cognitively normal controls; Braak Braak score (0–VI), BW brain weight, PMI post mortem interval, Dx diagnosis]

Case	Dx	Age (years)	Gender	BW (g)	PMI (h)	Braak
1	CN	61	F	1,360	8	I/II
2	CN	57	F	1,400	8	I/II
3	CN	87	F	1,120	8	I/II
4	CN	88	F	990	5.5	I/II
5	AD	91	F	1,260	3	III/IV
6	AD	75	M	1,300	7	III/IV
7	AD	81	M	1,150	4	III/IV
8	AD	72	M	1,160	4	III/IV
9	AD	100	F	970	5	V/VI
10	AD	48	M	1,100	8	V/VI
11	AD	86	M	1,330	2	V/VI
12	AD	72	M	1,080	4	V/VI
13	AD	74	M	960	4	V/VI
14	AD	62	M	1,150	4	V/VI
15	AD	84	F	1,070	5	V/VI
16	AD	89	F	1,070	5	V/VI

Immunohistochemistry

Tissue sections were processed free-floating for immunohistochemistry of human GABA receptor subunits as described previously [27, 28], using a polyclonal antibody against the receptor subunit GABA_BR1 (GBR1, AB1531; Chemicon, Temecula, CA). The amino acid sequence is common to both the GABA_BR1a and GABA_BR1b receptor isoforms, but this antibody primarily recognizes the GABA_BR1a [38]. The primary antibody was diluted 1:5,000 in TRIS-saline containing 3% goat serum and 0.25% Triton X-100. At least three sections from each case were immunostained for this study. Sections from all cases were processed together to control for variability in the immunohistochemical procedure. As a control for nonspecific staining, sections were incubated with initial incubation media with the primary antibody omitted, and otherwise processed as described. No positive staining was detectable in these control sections. Control sections for each case were used for background correction in optical density (OD) analyses (described below). Double immunolabeling was performed on each case using GBR1 and MC1 antibodies to investigate alterations of GBR1 in neurons undergoing early NFT changes. MC1 (generously provided by Dr. Peter Davies; used at 1:1,000) is a monoclonal antibody that detects early cytoskeletal alterations involving changes in the conformation of the tau molecule [16]. The sequential double-immunolabeling procedure used diaminobenzidine (DAB) as a chromogen to visualize MC1 immunohistochemistry, with subsequent GBR1 immunohistochemistry using DAB and 2.5% nickel ammonium sulfate, yielding homogeneous light brown (DAB) and granular black reaction products (nickel-conjugated DAB). Additional tissue sections were stained using monoclonal 10D5 (1:3,000; Athena Neurosciences, San Francisco, CA) or polyclonal paired helical filament (PHF) tau (1:10,000; DAKO, Carpinteria, CA) antibodies, to visualize amyloid β (A β) plaques and NFT, respectively.

OD measurements of neuronal GBR1-IR

Quantitative evaluation of the intensity of immunohistochemical reaction in individual pyramidal neurons was performed by OD measurements using an Olympus AHBT-3 light microscope equipped with a SPOT-2 digital video camera (Diagnostic Instruments, San Jose, CA) and public domain image analysis software (NIH Image, Scion, Frederick, MD). All images were obtained at $\times 40$ magnification under constant illumination and exposure conditions. Densitometric analysis was performed as described previously [23, 34]. Briefly, profiles of individual GBR1-immunoreactive neurons with visible nuclei were outlined with a free-hand marquee to obtain a morphometric mask. In three separate non-adjacent sections from each case, ten neurons were randomly selected for measurements in

the pyramidal layer of four hippocampal regions (CA1-4) and the subiculum (Sub). The dentate gyrus (DG) granule cell layer was evaluated by randomly choosing three microscopic fields as regions of interests (ROI), in which all neurons were measured collectively, due to their high packing density. The measurement of the relative concentration of GBR1-immunoreactive material in each ROI was obtained as the gray level (GL), related to the OD for the specimen using the following equation:

$$OD = GL_{\text{specimen}} - GL_{\text{background}}$$

where GL_{specimen} is the gray level of the image delineated by the morphometric mask (individual neurons) and $GL_{\text{background}}$ is the gray level of the background reference image (obtained by outlining neuronal layers in sections processed in the absence of primary antibody for each case). Specific staining was defined as the difference in immunostaining intensity between sections incubated with and without the primary antisera.

Values presented are means \pm standard deviation (SD). Where appropriate, analysis of variance (ANOVA) was used to make comparisons. If differences were detected by ANOVA, individual groups were compared using the Student-Newman-Keuls test. $P < 0.05$ was accepted as statistically significant for all comparisons.

Cell counts

To compare the relative loss of GBR1-immunoreactive compared to Nissl-labeled pyramidal neurons in CA1 hippocampus, cell counts were performed by randomly selecting ten $\times 40$ microscopic fields on three non-adjacent sections in all cases from each Braak group. All neurons on which the entire soma and initial primary dendrite could be identified were counted within each microscopic field, irrespective of neuronal size (i.e., all pyramidal neurons were counted). The total number of neurons within the entire CA1 field was not calculated; instead, numerical densities of GBR1-immunoreactive and Nissl-labeled neurons were obtained in each case and then calculated as means \pm SD for each Braak diagnostic group. Correction for laminar shrinkage was not performed, as shrinkage would affect equally GBR1-IR and Nissl-stained cells counted in adjacent tissue sections. However, it is possible that due to laminar shrinkage the reductions in both GBR1-IR and Nissl-positive neurons were underestimated in the Braak stage V/VI group (see discussion). Comparisons across Braak groups were made using ANOVA.

Results

GBR1-IR was detected as black, punctate chromogen precipitate that labeled the soma and proximal dendrites of hippocampal neurons and interneurons. GBR1-IR interneurons were infrequently observed, but were

present in every case; they were either bipolar or multipolar, with fusiform or round cell bodies and thin dendrites (not shown). Unlike pyramidal neurons, GBR1-IR interneurons were located primarily within the strata radiatum and oriens. Neuropil GBR1 immunostaining was not above background levels.

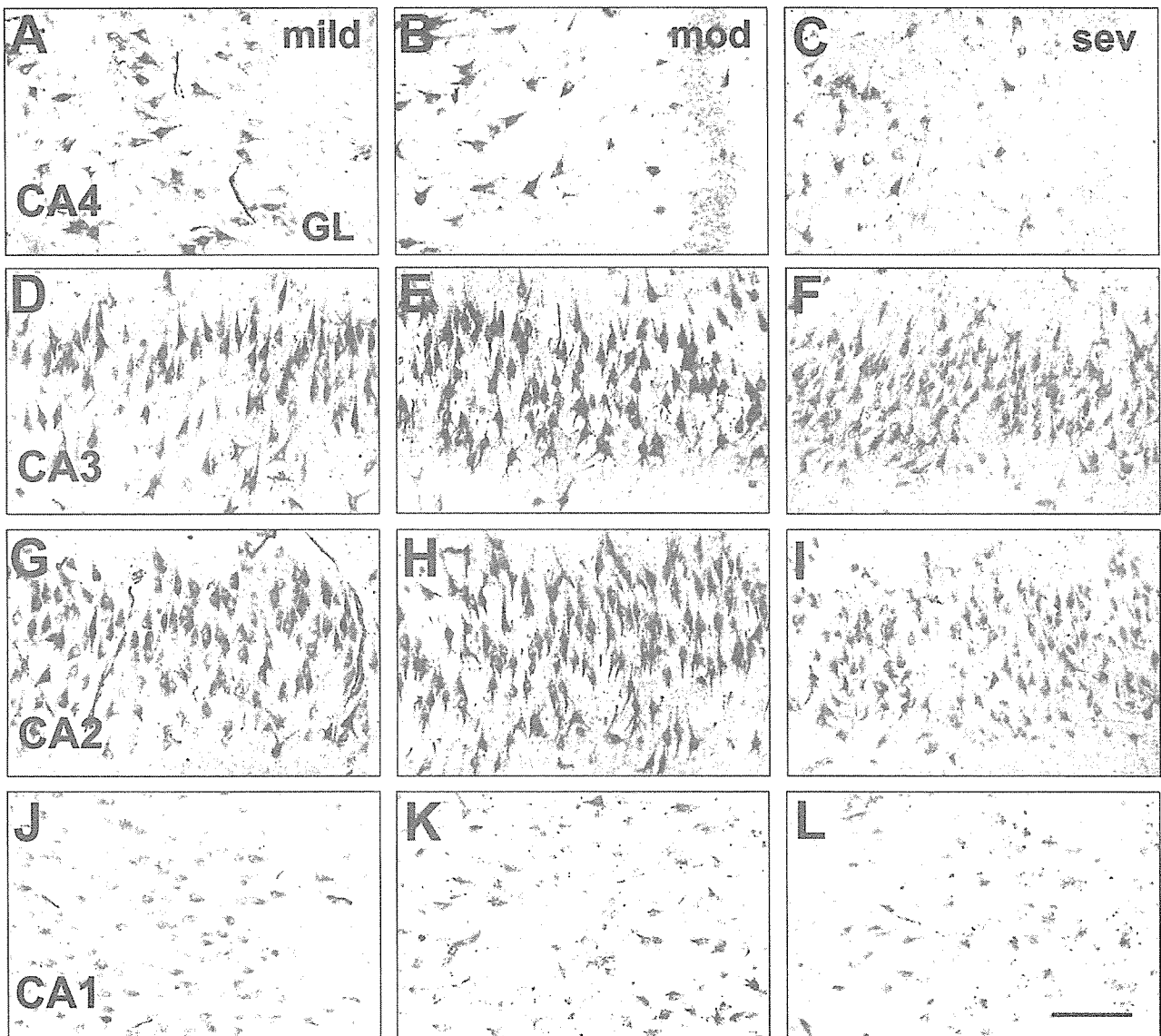
Fig. 1 Photomicrographs showing GBR1 immunohistochemistry in dentate granule cell layer (*GL*) and in CA4 (A–C), CA3 (D–F), CA2 (G–I) and CA1 (J–L) subfields of controls that are pathologically “mild” (Braak I/II; A, D, G and J), “moderate” AD (Braak III/IV; B, E, H, K), and “severe” AD (Braak V/VI; C, F, I, L) cases. Compared to mild and severe groups, the overall intensity of neuronal GBR1 immunoreactivity is increased in the moderate cases. In the CA1 subfield of moderate and severe cases, loss of pyramidal cell GBR1 staining is evident (K, L). This illustrates differences in neuronal GBR1 immunostaining intensity, and is not shown as a representative of differences in numbers of GBR1-immunoreactive neurons across Braak stage groups. The latter numbers are displayed in Table 2 (*GBR1* GABA_B receptor R1 protein, *AD* Alzheimer’s disease, *mod* moderate, *sev* severe). *Bar* 100 μm

CN subjects with mild (Braak stage I/II) NFT pathology

In CN subjects, neuronal GBR1-IR was detected in all hippocampal fields, although the intensity of immunostaining differed between fields (Figs. 1A, D, G, J; 2). Within the DG, light GBR1-IR was observed in granule cells (Figs. 1A, 2). In the CA4 field, moderate GBR1-IR was observed in the soma and proximal dendrites of mossy cells (Fig. 1A). In CA3 and CA2, pyramidal cells showed more intense GBR1-IR, particularly in comparison to CA1 field and DG (Figs. 1, 2). In contrast, GBR1-IR of CA1 and Sub pyramidal cells was considerably less intense (Figs. 1J, 2).

AD subjects with moderate (Braak stage III/IV) and severe (Braak stage V–VI) NFT pathology

In hippocampus of moderate AD cases, GBR1-IR increased markedly (Figs. 1, 2) in DG granule cells and



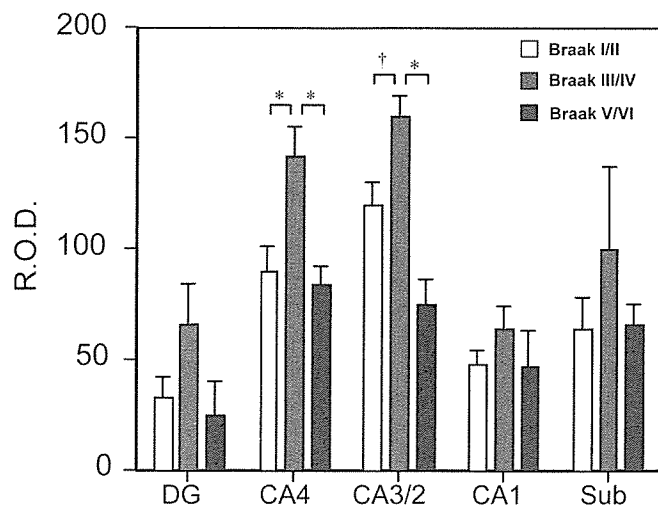


Fig. 2 Bar graph showing R.O.D. measurements of neuronal GBR1 immunostaining intensity in five hippocampal fields in control cases with Braak I/II (mild), and AD patients with Braak III/IV (moderate) or Braak V/VI (severe) pathology. Different patterns can be observed in regions more resistant (DG, CA4, CA3/2) versus those more vulnerable (CA1, Sub) to AD. In cases with an onset of hippocampal NFT pathology (Braak III/IV), significant increases of GBR1 staining intensity are observed in CA4 and CA3/2 regions, while a trend for an increase is seen in DG. Asterisks: $P < 0.01$; dagger: $P < 0.05$ (R.O.D. relative optical density. DG dentate gyrus, CA4, CA3/2, CA1 hippocampal CA fields, Sub subiculum, NFT neurofibrillary tangles)

in CA4 and CA3/2 pyramidal cells (Figs. 1B, E, H; 2). In contrast, severe AD cases showed a comparable to decreased cellular GBR1-IR in all hippocampal fields when compared to the mild CN group (Figs. 1C, F, I, L; 2). We next used relative OD (R.O.D.) measurements of neuronal GBR1 immunostaining in individual hippocampal fields to compare GBR1 immunostaining intensity in the three NFT severity groups, and detected significant differences in the CA4 and CA3/2 regions. Moderate AD cases had higher R.O.D. values (Fig. 2) than either mild CN or severe AD groups in CA4 (both $P < 0.01$) or in CA3/2 ($P < 0.05$ and $P < 0.01$). There was a considerable variability in numbers of GBR1-IR and Nissl-stained neurons in the CA1 field of AD cases. Moderate and severe AD cases showed marked loss of GBR1-IR CA1 pyramidal cells compared to the mild CN group (Table 2). Nissl staining of adjacent tissue sections revealed only moderate CA1 pyramidal cell loss across both AD groups, but this is likely an underestimate due to hippocampal atrophy and laminar shrinkage (Table 2; see discussion).

Table 2 Cell counts in the CA1 region (GBR1 GABA_B receptor R1 protein)

Braak stage	GBR1 positive	Nissl positive
I/II	37.5 ± 14.5	55.8 ± 8.7
III/IV	13.6 ± 9.0*	49.7 ± 12
V/VI	11.3 ± 9.2*	34.2 ± 17.6

* $P < 0.05$ (compared to I/II)

In addition to cellular IR, plaque-like clusters of GBR1-immunoreactive dystrophic neurites were detected in the CA1/Sub (Fig. 3B) and DG molecular layer (not shown) in both moderate and severe AD cases. These clusters were entirely composed of neuritic elements, with no associated neuropil-IR (Fig. 4).

GBR1 and MC1 co-localization in AD hippocampus

Immunostaining with MC1, a marker of early neurofibrillary changes, revealed a differential distribution of MC1-positive neurons in various hippocampal fields of the three groups of cases, consistent with their pathological classification by Braak stages. CN cases showed infrequent MC1-IR cells in the CA1 region, and no labeled neurons in DG, CA3/2 and CA4 (not shown). These CA1 MC1-IR cells did not co-localize GBR1.

In moderate AD cases, MC1-positive neurons were of substantial numbers in CA1/Sub (Fig. 3C), and only infrequent in other fields. In severe AD cases, there was an abundance of MC1-positive cells in CA1/Sub, and substantial numbers in other hippocampal fields. In both AD groups, only a small percentage of CA1/Sub neurons showed co-localization of MC1 and GBR1. In contrast, the CA4 and CA3/2 regions in these cases showed numerous GBR1/MC1 dual-labeled neurons (Fig. 5).

GBR1-positive neuritic plaque-like deposits in AD cases also contained MC1-IR neurites (Fig. 3C). There were no correlations between intensity of neuronal GBR1 immunostaining and distribution of A β plaques in hippocampal fields.

Discussion

The present study demonstrates alterations in GBR1-IR during the progression of NFT pathology in the hippocampus of AD subjects. Detection of neuronal GBR1-IR in all hippocampal fields in non-demented cases with only mild neurodegenerative changes is consistent with the results of previous immunohistochemical studies of GABA_B receptors in hippocampus of aged humans [5, 27]. We detected GABA_B-IR exclusively in neuronal soma and proximal dendrites. In contrast, Billinton et al. [5] reported that GBR1-IR is also present in the neuropil of the dentate molecular layer and stratum lacunosum-moleculare of the CA fields. Differences in specificity of the antibodies employed in the two studies could account for the discrepancies between the two studies.

Our results indicate that changes in the expression of hippocampal GBR1 subunit correlate with the progression of neuronal degeneration defined by a general assessment of NFT pathology by Braak and Braak [7], as well as by a marker of early NFT changes (MC1). The GBR1 immunostaining was most robust in CA3/2 regions, where it increased markedly in moderate AD

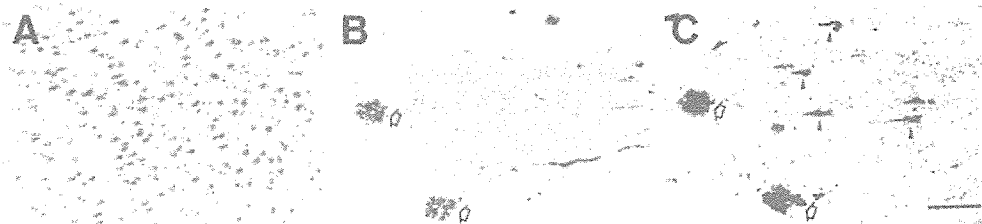


Fig. 3 Photomicrographs of the CA1 field in serial tissue sections, processed for Nissl staining (A), GBR1 immunohistochemistry (B) and MCI immunohistochemistry (C), from an AD case with moderate (Braak III/IV) NFT pathology. Despite unremarkable neuronal loss (A), there is a loss of GBR1 immunoreactivity on pyramidal neurons (B) that coincides with appearance of NFTs (C, arrowheads). Neuritic plaques (arrowheads) are both GBR1 positive (B) and MCI positive (C). Bar 100 μ m

(Braak III/IV) cases. In contrast, numbers of GBR1-immunoreactive neurons decreased in the CA1 region in both moderate and severe AD, and no GBR1 up-regulation was observed in these cells. The more pronounced loss of GBR1-immunoreactive compared to Nissl-labeled pyramidal neurons in the CA1 region suggests that GBR1-expressing neurons may be more sensitive to degeneration than CA1 neurons collectively. Notably, the lack of significant loss of Nissl-labeled pyramidal neurons in CA1 in the severe (Braak V/VI) group in the present study differs from that shown in studies employing unbiased stereological techniques [33, 35]. We recognize potential limitations of our non-stereological cell counting method; the latter inconsistency could be due to laminar shrinkage that likely occurs in end-stage AD hippocampus. However, correcting for laminar shrinkage would further reduce the numbers of both GBR1-IR and Nissl-positive CA1 neurons in severe AD

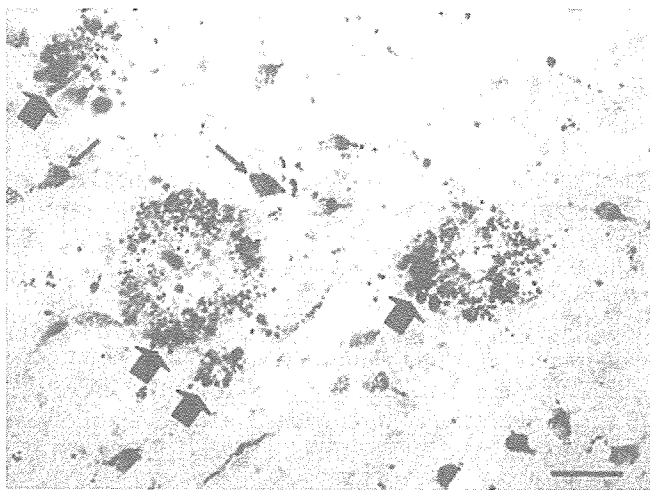


Fig. 4 High-power photomicrograph of three GBR1-immunoreactive plaque-like structures in the CA4. GBR1 immunostaining is present in numerous dystrophic neurites, many of which appear as large bulbous swellings (large arrows). Note the punctate GBR1-immunoreactive labeling in nearby neurons (small arrows), contrasting the dark uniform labeling seen in neuritic processes. Bar 75 μ m

cases. While such correction would bring our Nissl cell counting data in closer agreement with previous stereological investigations, we would still observe relatively lower numerical density of GBR1-immunoreactive neurons compared to Nissl-labeled neurons in AD. The CA1 region is particularly vulnerable to AD pathology, while CA3/2 is relatively resistant [11, 12]; thus, elevations in GBR1-IR in CA3/2 pyramidal neurons could reflect a compensatory response to increased excitatory neurotransmission, and render these cells more resistant to excitotoxic insults. The GABA_B receptor is known to modulate postsynaptic excitatory transmission [31, 37], and when excitation increases, GABA_B-mediated slow inhibition is recruited to offset excessive neuronal excitation [6]. Thus, up-regulation of GBR1 in pyramidal cells could serve to reduce excessive (toxic) stimulation of these cells by increasing inhibitory tone. Thus, transient up-regulation of the GBR1 in pyramidal cells, which occurs at the onset of hippocampal NFT pathology (i.e., Braak stage III/IV), could function to reduce excitotoxic neuronal damage in this region. Previous studies reported that excitotoxicity mediated via calcium-permeable α -amino-3 hydroxy-5-methyl-4-isoxazolepropionate (AMPA) and *N*-methyl-D-aspartate (NMDA) types of glutamate receptors may contribute, at least in part, to the development of neurodegenerative changes in the hippocampus [13, 14] as well as in other vulnerable regions [3, 15] of AD brains. In accordance with these studies, we have shown previously that loss of the GluR2 AMPA receptor subunit precedes hippocampal NFT formation [13]. However, it is possible that not only increased excitatory signal, but also reduced GABA compensatory (inhibitory) mechanisms contribute to NFT formation and cell death during the progression of AD. Alternatively, metabolic alterations due to neurofibrillary tangle pathology may impede synthesis of the receptor. Previous studies suggest that GABA receptor subunits can be substantially and selectively affected in AD. For example, radiolabeled ligand binding experiments in the hippocampus [8] and frontal cortex [9] from AD subjects have demonstrated significant reductions in GABA_B receptors, while GABA_A receptors were relatively preserved. Furthermore, GABA_A receptor density was reduced only in stratum pyramidale of CA1 subfield, while reductions in GABA_B receptor density were more extensive [8]. In agreement with these studies, our previous work demonstrated that GABA_A receptor alpha and beta 2/3 protein and beta 2 mRNA signals were well preserved in the DG, even in cases with severe AD pathology, [24, 25, 26]. Collec-

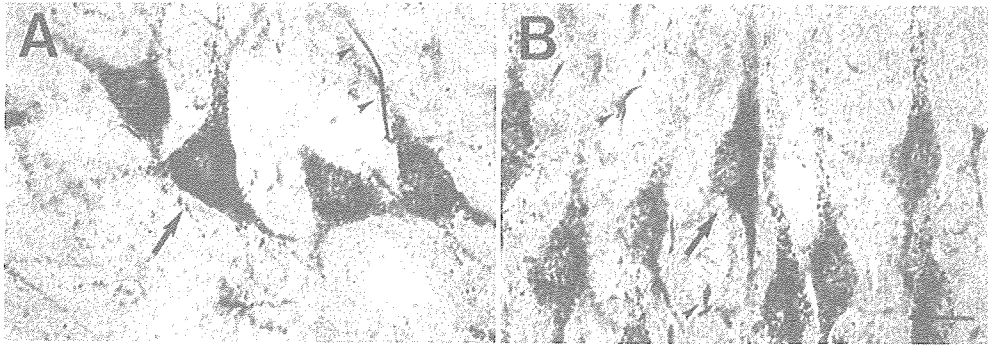


Fig. 5 Photomicrographs showing neuronal double immunolabeling with MC1 (*homogeneous brown*) and GBR1 (*granular purple*) in CA4 (**A**) and CA3 (**B**) subfields of a severe AD case. Colocalization of MC1 and GBR1 is observed in several pyramidal cells (*arrows*). In contrast, MC1-stained neuronal processes and threads (*arrowheads*) are GBR1 negative. Bar 20 μ m

tively, these results suggest that pathological processes might differentially affect the stability of GABA receptor subunits, with GABA_B being more vulnerable to AD.

We observed that in CA1, an area particularly affected by the development of mature NFT and cell death in AD, neurons expressing an early marker of neurofibrillary changes (MC1) did not co-localize GBR1. In contrast, in the CA3/2 and CA4 regions many of the MC1-immunoreactive neurons also expressed GBR1. Because neurons in the CA1/Sub region are known to develop NFT sooner in the progression of AD than those in any other field in the hippocampus, loss of GBR1 subunits from this cell population might contribute to their propensity to transform into NFT. In contrast, pyramidal neurons from CA3/2 and CA4 regions are resistant to the conversion into NFT, and this might be due to their ability to up-regulate, or sustain, functional GBR1. Thus, early in the course of neurofibrillary change, up-regulation of GBR1 in the CA2/3–4 regions may render these neurons more resistant to the progression of neurofibrillary pathology, whereas in CA1, this compensatory change is either not occurring, or is ineffective. Alternatively, neurons in CA2/3–4 may be more resistant than those in CA1 to the effects of NFT formation, and thereby retain a greater level of GBR1-IR synthesis.

In conclusion, our results indicate that there is a transient increase in GBR1-IR in the hippocampus during the progression of neurofibrillary pathology in the course AD. This potentially compensatory change is not sustainable, as GBR1-IR in severe AD cases is comparable to subjects with only mild NFT pathology. Additionally, there is considerable intersubject variability in AD, with loss of CA1 GBR1-IR in several cases of moderate and severe AD. Thus, changes in GABA_B receptor subunit expression in AD hippocampus could mark a compensatory response to the onset of regional neurodegenerative changes, while loss of such a response might facilitate or be due to further progression of NFT pathology.

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EEG filtering based on blind source separation (BSS) for early detection of Alzheimer's disease

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Abstract

Objective: Development of an EEG preprocessing technique for improvement of detection of Alzheimer's disease (AD). The technique is based on filtering of EEG data using blind source separation (BSS) and projection of components which are possibly sensitive to cortical neuronal impairment found in early stages of AD.

Methods: Artifact-free 20 s intervals of raw resting EEG recordings from 22 patients with Mild Cognitive Impairment (MCI) who later proceeded to AD and 38 age-matched normal controls were decomposed into spatio-temporally decorrelated components using BSS algorithm 'AMUSE'. Filtered EEG was obtained by back projection of components with the highest linear predictability. Relative power of filtered data in delta, theta, alpha 1, alpha 2, beta 1, and beta 2 bands were processed with Linear Discriminant Analysis (LDA).

Results: Preprocessing improved the percentage of correctly classified patients and controls computed with jack-knifing cross-validation from 59 to 73% and from 76 to 84%, correspondingly.

Conclusions: The proposed approach can significantly improve the sensitivity and specificity of EEG based diagnosis.

Significance: Filtering based on BSS can improve the performance of the existing EEG approaches to early diagnosis of Alzheimer's disease. It may also have potential for improvement of EEG classification in other clinical areas or fundamental research. The developed method is quite general and flexible, allowing for various extensions and improvements.

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Keywords: Alzheimer's disease; Diagnosis; EEG; Blind Source Separation; AMUSE; Filtering

1. Introduction

Alzheimer's disease (AD) is one of the most frequent disorders among the elderly population (Jeong, 2004; Petersen, 2003). Recent studies have demonstrated that AD has a presymptomatic phase, likely lasting years, during which neuronal degeneration is occurring but clinical symptoms do not yet appear. This makes preclinical discrimination between people who will and will not

ultimately develop AD critical for early treatment of the disease which could prevent or at least slow down the onset of clinical manifestations of disease (Blennow and Hampel, 2003; DeKosky and Marek, 2003; Rapoport, 2000; Wagner, 2000). Moreover, early diagnostic tools could significantly facilitate the development of drugs for the treatment at the early stage of AD: without preclinical diagnosis, many times more subjects (potential patients with huge percentage of those who actually would never develop AD) should be involved for testing of these drugs (DeKosky and Marek, 2003). A diagnostic method should be relatively inexpensive to make possible screening of many individuals who are at risk of developing this dangerous disease

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(DeKosky and Marek, 2003). The electroencephalogram (EEG) is one of the most promising candidates to become such a method.

To date, many signal processing techniques were applied for revealing pathological changes in EEG associated with AD (see Jeong, 2004, for review). For example, combination of linear and nonlinear measures improved the classification accuracy of AD versus normal subjects up to 92% (Pritchard et al., 1994). Using principal component analysis (PCA) as a postprocessing tool for compressing linear and nonlinear EEG features over channels and age as a moderator variable in a study with rigorous validation procedure (jack-knifing), Besthorn et al. (1997) obtained 89% correct classification. However, high classification accuracy was obtained for patients who already developed serious cognitive impairment (e.g. Mini Mental State Examination (MMSE) score was 11.5 ± 7.9 in the study of Besthorn et al. (1997)).

Finding a method for identification of patients who have no clinical signs of AD at the moment of EEG registration but later progress to AD is the main challenge in this field. The studies of this kind are very rare. Huang et al. (2000) obtained 87% classification accuracy for discrimination between patients with mild cognitive impairment (MCI) who later progressed and not progressed to AD, however, without reporting the use of cross-validation. Musha and co-authors demonstrated, in a computer simulation, that local cortical neuronal impairment should lead to lower dipolarity (goodness-of-fit for dipole localizations) of alpha EEG frequency components (Hara et al., 1999), and then, based on these results, developed a technique for estimation of cortical impairment in AD using a single index of dipolarity (Musha et al., 2002). Alpha dipolarity was able to differentiate MCI patients who showed no clinical signs of AD at the time when EEG was recorded but developed AD later, as diagnosed in the follow-up, from normal controls with high probability; it also correlated with the degree of cortical neuronal impairment, estimated by SPECT (Musha et al., 2002).

However, in spite of all of the achievements made in the above cited studies, the problem of preclinical diagnosis of AD using EEG is not yet solved and further improvement of the methodology is necessary.

The main idea of this paper can be formulated as ‘filtering based on Blind Source Separation (BSS)’, that is, filtering of EEG by selection of most relevant components followed by reconstruction of the relevant part (subspace) of EEG signal using back projection of only these components. We propose a preprocessing technique based on this idea for improving EEG-based AD diagnosis (possibly useful also in other fields of EEG analysis). Its usefulness was evaluated in combination with standard procedures, namely the linear discriminant analysis (LDA) applied to spectral power in several frequency bands. To make comparison clear and fair, we used only most reliable but simple procedures. However, more sophisticated analysis based on recent

advances in techniques for EEG processing and data classification may provide, in combination with proposed preprocessing, further significant improvement of early AD diagnosis, and some relevant emerging techniques will be mentioned in Discussion.

2. Methods

2.1. Blind source separation filtering for EEG classification

Intuitively, one can expect that some hidden components of such a complex signal like EEG can be more sensitive to Alzheimer’s disease and the related disorders than others. These more sensitive components can be considered as useful ‘signal’, and the other components of EEG as ‘noise’ or ‘unwanted signals’. Improving the ‘signal-to-noise ratio’ by filtering off the ‘noise’ could enhance the performance of subsequent feature extraction and data classification. Blind Source Separation (BSS) algorithms (see Cichocki and Amari, 2003, for extensive review) can be used for the purpose of such filtering.

BSS, in its application to EEG analysis, assume that EEG signal is composed of a finite number of components (signals from the brain and other sources), $s(t) = [s_1(t), \dots, s_n(t)]^T$. Here t is a discrete time index, n is the number of components and $[\dots]^T$ means transpose of row vector. Components are mixed through unknown linear mixing process (described by $n \times n$ mixing matrix A), and n sensors (EEG electrodes) record the mixed signals $x(t) = As(t)$. Each of the components changes in time, but has a fixed weight for each channel. BSS algorithm finds an unmixing (separating) $n \times n$ matrix W consisting of coefficients with which the electrode signals should be taken to form, by summation, the estimated components: $y(t) = Wx(t)$. (In more general case, the number of components can be not equal to the number of sensors.) The entries of the estimated mixing matrix $\hat{A} = W^{-1}$ are components’ weights in the mixing process; in other words, they indicate how strongly each electrode picks up each of individual components. *Back projection* of some selected components $x_r(t) = W^{-1}y_r(t)$ (where $x_r(t)$ is a vector of reconstructed sensor signals and $y_r(t)$ is the vector obtained from the vector $y(t)$ after removal of all the undesirable components (i.e. by replacing them with zeros)) allows us to filter the EEG data.

In strict sense, BSS means estimation of true (original) sources, though exactly the same procedure can be used for separation of two or more subspaces of the signal without estimation of true sources. One procedure currently becoming popular in EEG analysis is removing artifact-related BSS components and back projection of components originating from brain (e.g. Jung et al., 2000; Joyce et al., 2004; Vorobyov and Cichocki, 2002). In this procedure, components of brain origin are not required to be separated from each other exactly, because they are mixed again by

back projection after removing artifact-related components. But by the same procedure we can filter off the ‘noise’ also in wider sense, improving the relative amount of any types of useful information in the signal. Specifically, we can try to increase the relative amount of signal content related to AD (i.e. to improve signal to noise ratio — SNR).

Finding the rules or fundamental principles for identification of *relevant and irrelevant components* is critical for the proposed approach and, in general, may require extensive studies. In the case of removing artifact-related components, such components typically can be easily identified by visual inspection, but in more general case exact discrimination of relevant and non-relevant components is more difficult. In this paper we attempt to differentiate clusters or subspaces of components with similar properties or features. For the purposes of EEG classification the estimation of individual components corresponding to separate and meaningful brain sources is not required, unlike in other applications of BSS to EEG processing (including its most popular variant, Independent Component Analysis (ICA)). The use of clusters of components is especially beneficial when the data from different subjects are compared: similarity between individual components in different subjects is usually low, while subspaces formed by similar components are more likely to be sufficiently overlapped. Differentiation of subspaces with high and low amount of diagnostically useful information can be made easier if components are separated and sorted according to some criteria which, at least to some extent, correlate with the diagnostic value of components. BSS algorithm ‘AMUSE’, in our opinion, can be relevant for this task.

2.2. AMUSE algorithm and its properties

AMUSE (Cichocki and Amari, 2003; Szupiluk and Cichocki, 2001; Tong et al., 1991, 1993) is a BSS algorithm which arranges components not only in the order of decreasing variance (that is typical for the use of singular value decomposition (SVD) which is implemented within the algorithm), but also in the order of their decreased linear predictability. Low values for both characteristics can be specific for many of EEG components related to high frequency artifacts, especially electromyographic signal (which cannot be sufficiently removed by usual filtering in frequency domain, see Goncharova et al., 2003). Thus, a first attempt of selection of diagnostically important components can be made by removing a range of components separated with AMUSE (below referred to as ‘AMUSE components’) with the lowest linear predictability. Automatic sorting of components by this algorithm makes it possible to do this simply by removing components with indices higher than some chosen value.

AMUSE algorithm belongs to the group of second-order-statistics spatio-temporal decorrelation (SOS-STD) BSS algorithms. It provides similar decomposition as the well

known and popular SOBI algorithms (Belouchrani et al., 1997; Tang et al., 2002). AMUSE algorithm uses simple principles that the estimated components should be spatio-temporally decorrelated and be less complex (i.e. have better linear predictability) than any mixture of those sources. The components are ordered according to decreasing values of singular values of a time-delayed covariance matrix. As in Principal Component Analysis (PCA) and unlike in many ICA algorithms, all components estimated by AMUSE are uniquely defined (i.e. any run of algorithms on the same data will always produce the same components) and consistently ranked. Fig. 1 illustrates typical components obtained by decomposing EEG using AMUSE algorithm.

AMUSE algorithm can be considered as two consecutive PCAs: first, PCA is applied to input data; second, PCA (SVD) is applied to the time-delayed covariance matrix of the output of previous stage. In the first step standard or robust prewhitening (sphering) is applied as a linear transformation $\mathbf{z}(t) = \mathbf{Q}\mathbf{x}(t)$, where $\mathbf{Q} = \mathbf{R}_x^{-1/2}$ of the standard covariance matrix $\mathbf{R}_x = E\{\mathbf{x}(t)\mathbf{x}^T(t)\}$ and $\mathbf{x}(t)$ is a vector of observed data for time instant t . Next, SVD is applied to a time-delayed covariance matrix of pre-whitened data: $\mathbf{R}_z = E\{\mathbf{z}(t)\mathbf{z}^T(t-1)\} = \mathbf{U}\mathbf{S}\mathbf{V}^T$, where \mathbf{S} is a diagonal matrix with decreasing singular values and \mathbf{U} , \mathbf{V} are matrices of eigenvectors. Then, an unmixing matrix is estimated as $\mathbf{W} = \hat{\mathbf{A}}^{-1} = \mathbf{U}^T\mathbf{Q}$ or $\hat{\mathbf{A}} = \mathbf{Q}^T\mathbf{U}$.

AMUSE algorithm is much faster than the vast majority of BSS algorithms (its processing speed is mainly defined by the PCA processing within it) and is very easy to use, because no parameters are required. It is implemented as a part of package ‘ICALAB for signal processing’ (Cichocki et al., online) freely available online and can be called also from the current version of EEGLAB toolbox (Delorme and Makeig, 2004) (which is freely available online at <http://www.sccn.ucsd.edu/eeqlab/>) if both toolboxes are installed.

2.3. Subjects and EEG recording

We used EEG recordings collected in the previous study (Musha et al., 2002). In that study, patients who complained only for memory impairment, but had no apparent loss in general cognitive, behavioral, or functional status, were recruited. Fifty-three patients of this group met the following criteria for Mild Cognitive Impairment (MCI): MMSE score 24 or higher, Clinical Dementia Rating (CDR) scale score of 0.5 with memory performance less than one standard deviation below the normal reference (Wechsler Logical Memory Scale and Paired Associates Learning subtests, IV and VII, ≤ 9 (Wechsler, 1987), and/or ≤ 5 on the 30 min delayed recall of the Rey-Osterreith figure test (Hodges, 1993)). These patients were followed clinically for 12–18 months. Twenty-five of them developed probable or possible AD according to NINDS-ADRDA criteria (McKhann et al., 1984). Normal age-matched controls were recruited from family members of the patients

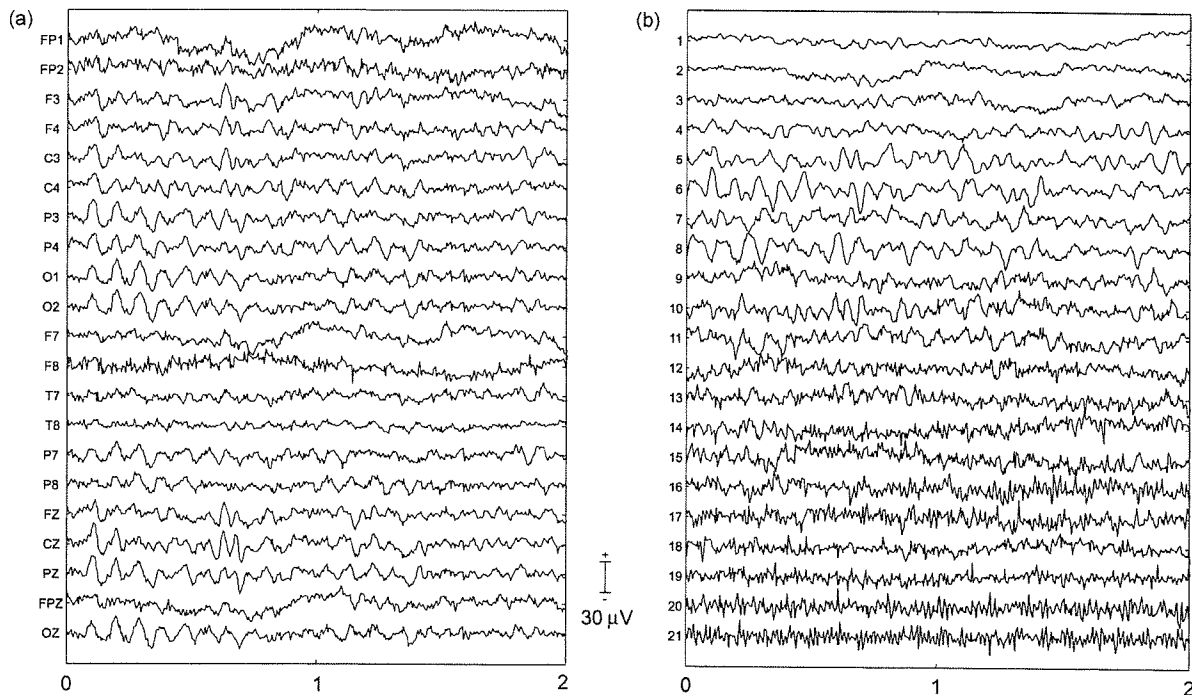


Fig. 1. Example of raw EEG (a) and its components separated with AMUSE algorithm (b) for a patient with MCI who later progressed to AD (MildAD002). AMUSE was applied to 20 s artifact-free interval of EEG, but only 2 s are shown. The scale for the components is arbitrary but linear. Note that the components are automatically ordered according to decreasing linear predictability (increasing complexity).

(mainly spouses) participated in the study as control group. Both patients and controls underwent general medical, neurological, psychiatric, and neuroimaging (SPECT, CT and MRI) investigation for making the diagnosis more precise.

EEG was recorded within 1 month after entering the study from all patients and controls, but only EEG recorded from the patients who progressed to AD ($n=25$; below: MCI group) and age-matched controls ($n=56$) was used for the analysis. No patient or control subject received psychotropic medication at the period when EEG was recorded. Mean MMSE score was 26 ± 1.8 in MCI group and 28.5 ± 1.6 in control group; age 71.9 ± 10.2 and 71.7 ± 8.3 , respectively. EEG recording was done in an awake resting state with eyes closed, under vigilance control. Ag/AgCl electrodes (disks of diameter 8 mm) were placed on 21 sites according to 10–20 international system, with the reference electrode on the right ear-lobe. EEG was recorded with Biotop 6R12 (NEC San-ei, Tokyo, Japan) using analog filtering bandpass 0.5–250 Hz and sampling rate 200 Hz.

2.4. EEG data analysis

All computations were done using MATLAB (The MathWorks, Inc.). EEGLAB (Delorme and Makeig, 2004) was used for visual analysis of EEG recordings, and AMUSE algorithm implemented in ICALAB (Cichocki et al., online) was used for BSS processing.

Out of the EEG database described above (from the study of Musha et al., 2002), we selected 25 MCI patients (later progressed to AD) and 47 age-matched controls who had relatively little artifacts. Their EEGs were visually inspected by an experienced EEG researcher and the first continuous artifact-free 20 s interval of each recording was chosen for the analysis. Due to the lack of such interval in some recordings, the number of patients and controls were reduced to 22 and 38, correspondingly. The reason for selecting artifact-free intervals was that most of the artifacts produced amplifier blocking (saturation) due to its low amplitude range, which lead to strongly nonlinear distortion of the signal. AMUSE, as most of BSS methods, assumes a linear model of summation of source signals, and amplifier blocking should be excluded from the data.

Each EEG was decomposed into 21 decorrelated components by BSS algorithm AMUSE (see above). Some of the components (see Results) were selected for back projection, which formed preprocessed ('AMUSE filtered') EEG data. Spectral analysis based on Fast Fourier Transform (Welch method, Hanning 1 s window, 2 s epochs overlapped by 0.5 s) was applied to raw data, to the components and to the projections of selected components. Relative spectral powers were computed by dividing the power in delta (1.5–3.5 Hz), theta (3.5–7.5 Hz), alpha 1 (7.5–9.5 Hz), alpha 2 (9.5–12.5 Hz), beta 1 (12.5–17.5 Hz) and beta 2 (17.5–25 Hz) bands by the power in 1.5–25 Hz band. These values were normalized for better fitting