

## Experimental Spinal Fusion With Recombinant Human Bone Morphogenetic Protein-2 Delivered by a Synthetic Polymer and $\beta$ -Tricalcium Phosphate in a Rabbit Model

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**Study Design.** An experimental animal study to achieve posterolateral intertransverse process spine fusion with recombinant bone morphogenetic protein in combination with a new delivery system.

**Objective.** To evaluate the efficacy of a new synthetic biodegradable bone-inducing material containing recombinant human bone morphogenetic protein-2 (rhBMP-2) as a bone-graft substitute for posterolateral intertransverse process fusion in a rabbit model.

**Summary of Background Data.** rhBMP-2, a powerful bone-inducing cytokine, has been used as a bone graft substitute in combination with animal-derived collagen to achieve spinal fusion in animal models. However, the minimum dose of rhBMP-2 required to obtain solid posterolateral intertransverse process fusion was high on the basis of previous reports ( $>100 \mu\text{g}$  in rabbit models). To improve the efficacy, performance of rhBMP-2, and the safety of the delivery system for this protein, a more sophisticated system is required.

**Methods.** To fabricate one implant for one-side L4-L5 intertransverse process fusion,  $\beta$ -tricalcium phosphate ( $\beta$ -TCP) powder (300  $\mu\text{g}$ ), a polymer gel (PLA-DX-PEG block copolymer; 300  $\mu\text{g}$ ) and rhBMP-2 (7.5, 15, or 30  $\mu\text{g}$ ) were mixed and manually shaped to resemble a rod. Through a posterolateral approach, two implants were placed on both sides (1 per side) by surgery so as to bridge the transverse processes of adult New Zealand white rabbits ( $n = 27$ ). In control animals, implants without rhBMP or autogenous cortico-cancellous bone chips from the iliac-

crest were placed in a similar location. The lumbar vertebrae were recovered 6 weeks after surgery. The posterolateral fusion was examined by manual palpation, radiography, biomechanical testing, and histology.

**Results.** Rabbits that received 15 or 30  $\mu\text{g}$  of rhBMP-2 showed consistent fusion. However, solid fusion was seen in 2 of 5 rabbits with autografting and rabbits that received 7.5  $\mu\text{g}$  of rhBMP-2. Fusion was not observed in the rabbits that did not receive rhBMP-2.

**Conclusions.** Consistent spinal fusion was obtained by implanting a biodegradable bone-inducing implant composed of  $\beta$ -TCP, PLA-DX-PEG, and rhBMP-2 within a period of 6 weeks. The rhBMP-2 doses required for the spinal fusion were significantly lower than those reported previously.

**Key words:** animal model, bone induction, posterolateral lumbar spine fusion, recombinant human bone morphogenetic protein-2. *Spine* 2005;30:1717-1722

Anterior or posterior fusion with autogenous bone grafting is a routine method for the treatment of spinal disorders associated with spinal instability resulting from degenerative changes, tumor resection, or trauma to the spine. To restore permanent stability of the spine, local new bone formation bridging the neighboring unstable vertebrae is essential. Autogenous iliac bone grafting is commonly used to promote bone formation. However, autogenous bone grafting is limited by some issues that remained unsolved. These are physical or cosmetic morbidities such as acute and chronic pain or dysesthesia, the potential risk for wound infection, extensive skin scarring, and deformity at the donor site.<sup>1,2</sup> In addition, the limited available mass of graft bone is also a disadvantage. To overcome these issues, new methods or materials that can substitute for the autogenous bone grafts have been desired. Allogeneic bone graft or banked bone is one of the alternatives that have been considered. However, banked bone has less osteogenic potential than autograft, and there is a potential risk for immunologic reaction from hosts and disease transfer to host with this material.<sup>3,4</sup> Biomaterials such as hydroxyapatite and bioactive ceramics also have been tested as bone-graft substitutes to avoid the potential risks arising from the use of allografts. Unfortunately, materials with osteoconductive potential but no osteoinductive capacity cannot substitute for autograft. Therefore, new materials

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with a potent osteoinductive capacity are required to avoid the disadvantages of autograft and to secure enhanced new bone formation for solid spinal fusion.

To manufacture an osteoinductive artificial bone graft substitute, cytokines retaining osteoinductive activity (bone morphogenetic proteins, BMPs) have been combined with biocompatible implant materials and used to obtain spinal fusion in experimental animals or in limited number of human cases.<sup>5-15</sup> To elicit the BMP-induced bone formation, a carrier material that delivers BMP slowly to the target cells is essential. As a carrier material, animal-derived collagen has been used routinely both in animal experiments and in clinical settings despite the potential risks for immunologic reaction in the host and transfer of diseases such as bovine spongiform encephalopathy (BSE).<sup>16,17</sup> To avoid those risks, we synthesized biodegradable polymers which work more effectively as the carrier for BMP-2 in *in vivo* conditions than bovine-derived collagen.<sup>18,19</sup> By use of this BMP delivery system, critical size defects in the long bones of rabbits and dogs were repaired successfully. New bone formation was achieved with these new porous solid biomaterials, which remained unresorbed in hosts.<sup>20,21</sup>

In this study, we attempted to achieve posterolateral intertransverse process spine fusion in the rabbit model by use of a biodegradable polymer and  $\beta$ -TCP composite as a delivery system for BMP. In this system, a successful outcome would be bone formation and the complete resorption of the carrier materials at the implanted sites.

## Materials and Methods

**rhBMP-2.** rhBMP-2 was produced at Genetics Institute (Cambridge, MA) and donated to us through Astellas Pharma Inc. (Ibaraki, Japan).

**PLA-DX-PEG Polymer.** Poly-D,L-lactic acid with a random insertion of *p*-dioxanone/polyethylene glycol block copolymer (PLA-DX-PEG, MW; 12,400, LA/DX/EO molar ratio; 42:14:44), was provided by Taki Chemical (Kakogawa, Japan). The chemical formula of the PLA-DX-PEG is shown in Figure 1. We have reported that this polymer worked effectively as a carrier for rhBMP in previous studies. Details of the physicochemical characteristics and efficacy as a carrier material for rhBMP-2 have been reported elsewhere.<sup>18,19</sup> The minimal efficacious content of rhBMP-2 in the synthetic polymer required to elicit new bone formation in rabbits was approximately 0.02%.<sup>20</sup>

**$\beta$ -TCP Powder.**  $\beta$ -TCP powder (less than 100  $\mu$ m in diameter of particles) was manufactured and provided to us by Olympus Biomaterial (Tokyo, Japan).

**Preparation of New Bone Graft Substitute Implants.** To prepare one implant (Figure 2A) to bridge L5 and L6 transverse

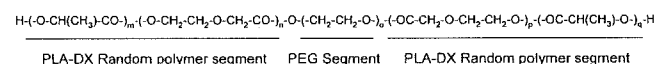


Figure 1. Structural formula of PLA-DX-PEG. The subscripts m, n, o, p, q represent variable number of units.

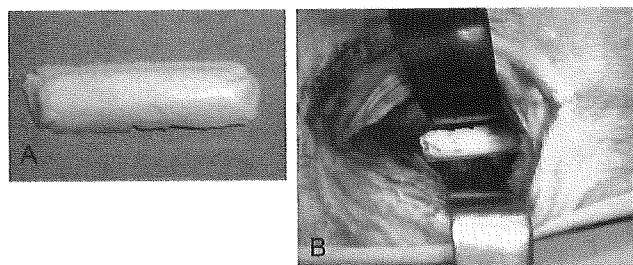


Figure 2. **A**, Prepared  $\beta$ -TCP dough implant. **B**,  $\beta$ -TCP dough was placed on the L5-L6 intertransverse region.

processes on one side, 300 mg of  $\beta$ -TCP powder, 300 mg of PLA-DX-PEG, and 3 dosages of rhBMP (7.5, 15, or 30  $\mu$ g) were mixed and stirred with a metal rod at 50°C for several minutes. The resultant dough was then cooled and fabricated by hand to resemble a rod. The hardened rods were stored at -30°C until implantation. As control implants, 300 mg of  $\beta$ -TCP powder and 300 mg of PLA-DX-PEG without rhBMP-2 was prepared in the same manner.

**Surgery and Experimental Protocols.** Twenty-seven New Zealand white rabbits (age, 1-2 years-old; weight, 3.5-4.5 kg) were divided randomly into five groups depending on the material to be implanted into the intertransverse process space. Before surgery, the animals were anesthetized with an intramuscular injection of ketamine (30 mg/kg) and xylazine (10 mg/kg). Cefazolin (100 mg) was administered subcutaneously as a prophylactic antibiotic. Each rabbit underwent surgery for a single level posterolateral intertransverse process fusion at L5-L6.<sup>7</sup> A dorsal midline skin incision was made, followed by two paramedian fascial incisions. The intermuscular plane between the multifidus and longissimus muscles was retracted to expose the transverse processes of L5 and L6 and the intertransverse membrane. An electric-driven burr (Stryker, Kalamazoo, MI) was used to decorticate posterior cortex of the respective transverse process, and one of the implant or transplant materials listed in Table 1 was implanted (Figure 2B). The wounds were then closed with 3-0 absorbable and 3-0 nylon sutures. Cefazolin (100-mg once daily) was administered to the respective animal subcutaneously for 3 days after surgery. The animals were killed by overdose of anesthetics at 6 weeks after surgery, and the L4-L7 lumbar spines were harvested and processed for further examination. This protocol was approved by the Institutional Committee for Animal Care and Experiments of Osaka City University Medical School.

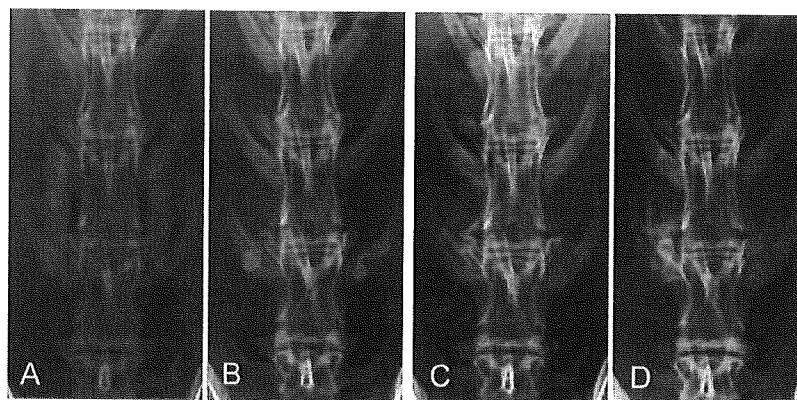
**Radiographic Evaluation.** The L5-L6 spines from each group animals were examined by posteroanterior plain radio-

Table 1. Implant Assignment

Group	rhBMP-2 ( $\mu$ g)	$\beta$ -TCP (mg)	PLA-DX-PEG (mg)	Concentration of rhBMP-2 (wt%)	n
BMP 30	30	300	300	0.005	5
BMP 15	15	300	300	0.0025	6*
BMP 7.5	7.5	300	300	0.00125	5
BMP 0	0	300	300	0	6*
Autogenous bone	Autogenous iliac bone graft (1-1.5 g)				5

\* Each one is for histological evaluation.

Figure 3. Representative posteroanterior radiographs of rabbit spines in the BMP 30 group, immediately after surgery (A), at 2 weeks (B), at 4 weeks (C), and at 6 weeks (D). In the radiograph, a remarkable fusion mass is visible between the L5–L6 transverse process at 6 weeks.



graphs and computed tomographic (CT) scans (GE Yokogawa Medical System, Tokyo, Japan) sequentially at 2, 4, and 6 weeks after surgery. CT images of 2-mm slice thickness were used to construct three-dimensional images. From the harvested lumbar spine samples, soft tissues were removed, and plain radiographs and CT images were taken again. A radiologic evaluation for intertransverse process fusion was made by three observers in a blinded fashion. Fusion was graded as solid when two of the three observers agreed that the presence and location of new bone formation were consistent with a successful fusion outcome.

**Manual Palpation.** The harvested lumbar spines were manually palpated by flexion and extension at the fusion level and comparison with the adjacent level. Each motion segment was graded as solid fusion or not solid.

**Biomechanical Testing.** Biomechanical testing to evaluate the solidity of the L5–L6 fusion site was performed by a three-point flexion-bending test using a materials testing machine (Instron 5882, Instron, Boston, MA). Three-point bending tests were performed with a 30-mm intersupport distance and a 1 mm/minute head speed. The bending moment at 1.5-mm middle-span deflection was determined from the moment-deflection curves.

**Histologic Examination.** The harvested specimens were fixed in 10% formalin in neutral buffer solution and decalcified in 10% formic acid solution, dehydrated in a gradient ethanol series, and embedded in paraffin. Sections of 4- $\mu$ m thickness at the intertransverse process region were cut in a sagittal plane, stained with hematoxylin and eosin, and observed under a light

microscope to examine for bony fusion between the newly formed bone and the original transverse processes.

**Statistical Analysis.** Comparisons of biomechanical testing of spines in each group were made using one-way analysis of variance. The post hoc Scedge test was performed to determine significant differences between groups. Significance for all tests was defined as  $P < 0.05$ .

## Results

### Radiographic Evaluation

An opaque shadow of  $\beta$ -TCP was noted at the operated site on radiographs immediately after surgery. Radiographs at 6 weeks showed homogeneous calcified shadows between the transverse processes in all animals of the BMP15, BMP30, and in part of the BMP7.5 groups. (Figures 3–5) Representative three-dimensional CT images of each group are shown in Figure 6. Fusion assessments in three-dimensional CT were difficult because the images tend to overestimate the fusion mass. Results of the evaluation from plain radiographs and three-dimensional CT are shown in Table 2.

### Manual Palpation

In all samples of the BMP 15 and BMP 30 groups, bony hard masses at the intertransverse process were palpable, and the passive motion between the vertebrae was significantly restricted when compared with that in control samples. Two of the five samples from the BMP 7.5 and autogenous bone groups, respectively, were evaluated as

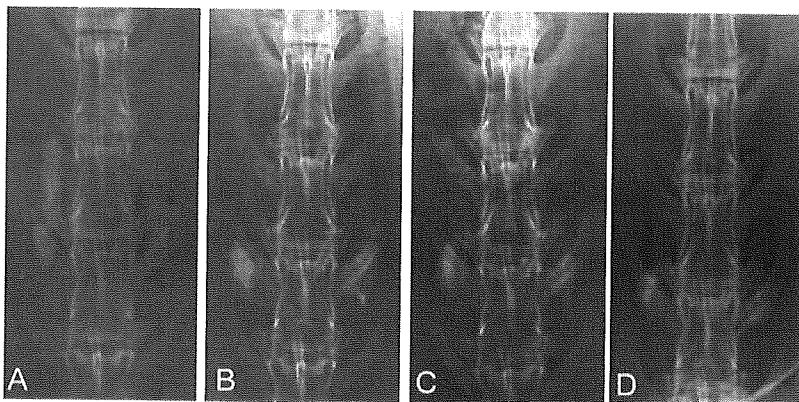


Figure 4. Representative posteroanterior radiographs of rabbit spines in the BMP 0 group, immediately after surgery (A), at 2 weeks (B), at 4 weeks (C), and at 6 weeks (D). In the radiograph, no fusion mass between the L5–L6 transverse process is visible.

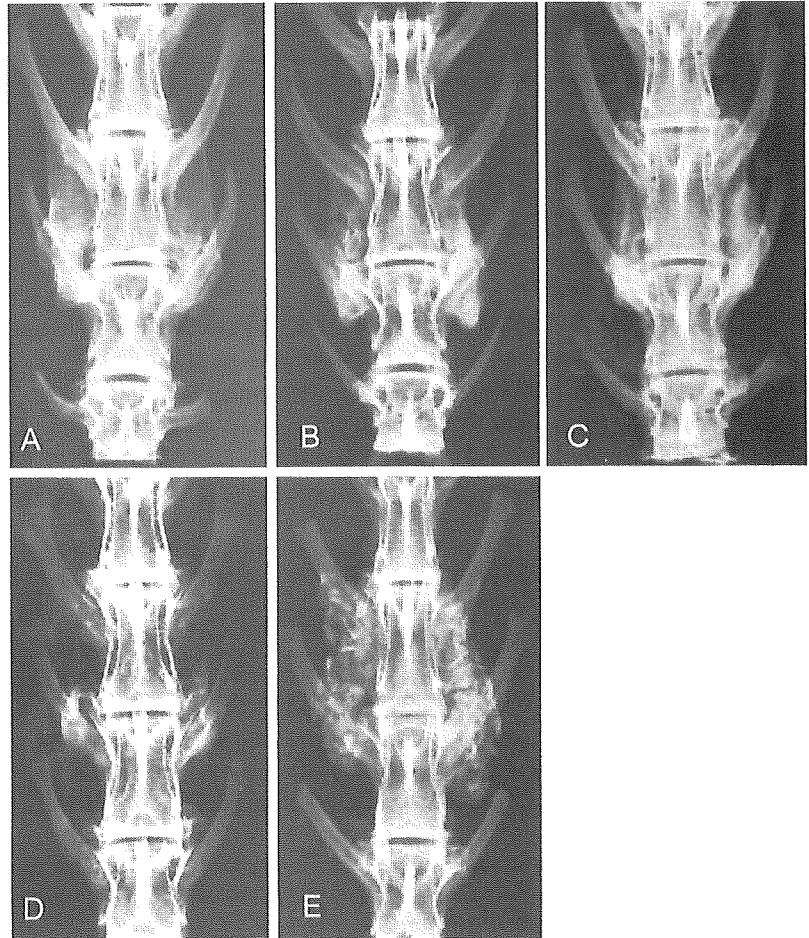


Figure 5. Harvested lumbar spine 6 weeks after surgery. BMP30 (A), BMP15 (B), BMP7.5 (C), BMP0 (D), autogenous bone (E).

solid fusion outcomes. In contrast, none of the samples from the BMP 0 groups achieved solid fusion (Table 3).

**Biomechanical Testing**

The results from biomechanical testing in each of the experimental groups are shown in Figure 7. The bending

moment at 1.5-mm middle-span deflection of the BMP 15 and BMP 30 groups were significantly larger than the BMP 0 group. The mean values in the BMP30 and BMP15 groups were higher than those from the autogenous bone group and the control BMP 0 group.

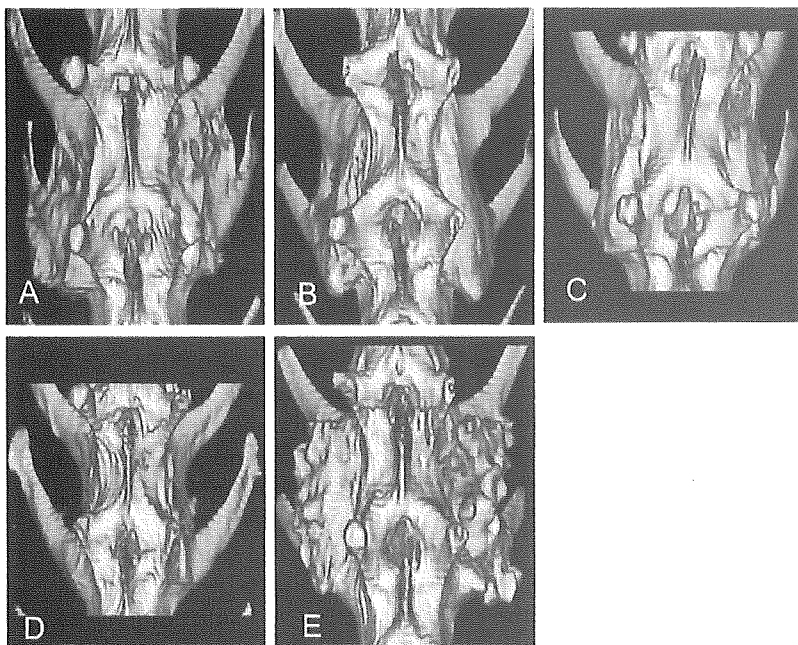


Figure 6. Three-dimensional-CT at 6 weeks after surgery. BMP30 (A), BMP15 (B), BMP7.5 (C), BMP0 (D), autogenous bone (E).

**Table 2. Fusion Rate in Radiological Assessments**

Group	Plain X-p	Three-Dimensional CT
BMP 30	5/5	5/5
BMP 15	5/5	5/5
BMP 7.5	4/5	4/5
BMP 0	0/5	0/5
Autogenous bone	5/5	5/5

### Histologic Examination

Lower magnification views of sagittal sections (hematoxylin and eosin staining) of fusion mass in animals that received rhBMP-2 showed bone mass with peripheral cortical bone bridging the transverse processes. Higher magnification of the bridging bone mass revealed woven bone and hematopoietic marrow. Tiny remnants of the  $\beta$ -TCP powder were also recognized. However, the intertransverse region sampled in a spine that did not receive rhBMP-2 revealed fibrous tissue and remnants of  $\beta$ -TCP with no evidence of new bone formation (Figure 8).

### Discussion

The present study was designed to test a synthetic and absorbable bone-graft substitute with osteoinductive ability equivalent or superior to the autogenous bone graft. The test was conducted in a critical bone defect model wherein the successful outcome was a solid posterolateral intertransverse process fusion. The results in this study were satisfactory, and in all animals with  $\beta$ -TCP (300 mg)/PLA-DX-PEG (300 mg) composite implants with 15 or 30  $\mu$ g of rhBMP-2, solid spinal fusion was obtained in 6 weeks. In the autogenous bone graft group, new bone formation was consistently recognized on radiographs, but in some specimens retrieved from the rabbits, failure of fusion was noted during biomechanical tests. It is interesting to note that in some clinical cases, the pseudarthrosis rate of posterolateral spine fusion has ranged from 5 to 35%.<sup>22,23</sup> These results are encouraging and point to the need for further clinical testing of this synthetic composite implant. The avoidance of additional surgery to harvest graft bone and thereby eliminate donor site morbidities is a potential key benefit of this approach.

In considering the practical application of the rhBMP retaining implants, one of the remaining issues is the extremely high dose of rhBMP required to elicit new bone, especially in humans. Typically, several milligrams of rhBMP are necessary for one level spinal fusion, and this fact results in a high cost of the BMP-retaining im-

**Table 3. Fusion Rate in Manual Palpation**

Group	Solid Fusion
BMP 30	5/5
BMP 15	5/5
BMP 7.5	2/5
BMP 0	0/5
Autogenous bone	2/5

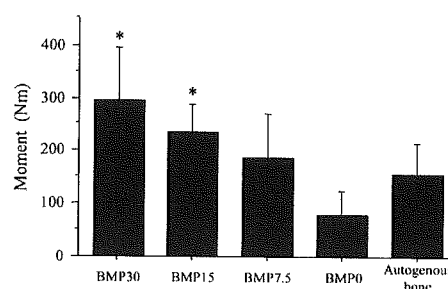
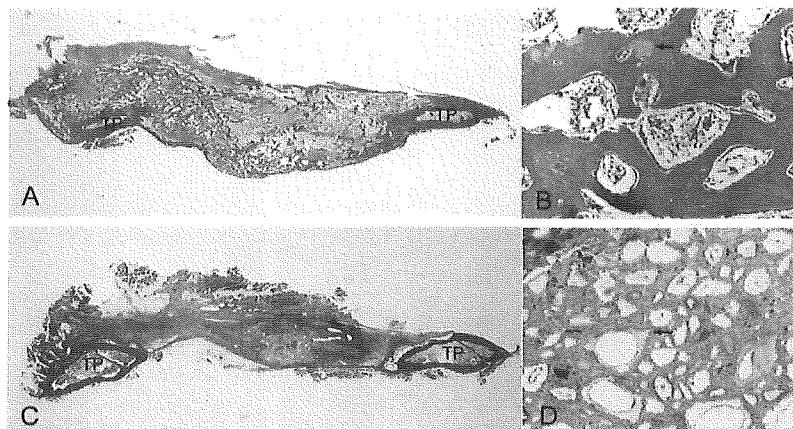


Figure 7. Results of three-point flexion bending tests for each specimen. These results indicate the fusion sites of BMP 30 and BMP 15 groups are stiffer than BMP 0 group. \*, statistically significant difference from BMP 0 group.

plant.<sup>6</sup> To reduce the cost, a more effective method to deliver efficacious but lower doses of rhBMP-2 would be desired. In previous experimental studies of the rabbit posterolateral intertransverse process spine fusion model, >100  $\mu$ g of rhBMP-2 were required to achieve the solid one-level spine fusion in 5 or 6 weeks. This was accomplished by the use of either animal-derived collagen sheets, hydroxyapatite with collagen, an open cell polylactic acid polymer, or sintered bovine bone as the delivery system for rhBMP-2.<sup>5,10,12-15</sup> Interestingly, in this study, the minimal effective dose of rhBMP-2 was significantly lower (15–30  $\mu$ g) than the amounts used in previous studies. Those prior experiments were not reproduced in this study; therefore, a direct comparison of the minimal effective dose of rhBMP is difficult. However, our results suggest an advantage of this new delivery system to reduce the effective dose of rhBMP for spinal fusion. The use of  $\beta$ -TCP powder to construct the bone-inducing implants provides three advantages; 1) the addition of the  $\beta$ -TCP powder to the “sticky” PLA-DX-PEG resulted in a dough-like material with easy handling and molding characteristics, 2) a volume expansion of the implant by addition of  $\beta$ -TCP powder, and 3) resorption of the  $\beta$ -TCP powder by osteoclasts recruited during the BMP-induced bone formation phase and disappears after establishment of spinal fusion. In our previous study in mice, it was noted that the  $\beta$ -TCP granules coated with the rhBMP-2-retaining polymer (PLA-DX-PEG) elicited ectopic new bone in situ at 3 weeks.  $\beta$ -TCP encased within the ectopic ossicles was resorbed by a large population of osteoclasts attached on the surface.<sup>24</sup> As expected in the current study, the  $\beta$ -TCP powder was almost completely resorbed and replaced by new bone with marrow on histologic findings at 6 weeks after surgery.

In summary, a new bone-inducing and biodegradable implant was produced by combining three synthetic materials (PLA-DX-PEG, rhBMP-2, and  $\beta$ -TCP). Posterolateral intertransverse process fusion was successfully achieved in a rabbit model with this implant in 6 weeks. The fusion rate appeared to exceed that obtained by autogenous bone grafting. These data provide support for the use of this new biomaterial as a substitute for autogenous bone grafting. The avoidance of the need for and

Figure 8. Hematoxylin and eosin stained sagittal section of L5–L6 intertransverse region in BMP 15 (A,B) and BMP 0 (C,D) group. New bone formation between the L5–L6 transverse process (TP), the cortical rim around fusion mass, and tiny residual  $\beta$ -TCP (arrow) was seen in BMP 15 group. The specimen from the group that did not receive rhBMP-2 showed no new bone formation between the L5–L6 transverse process (A and C,  $\times 0.5$ , B and D,  $\times 10$ ).



risks associated with surgery for graft material procurement is an additional benefit. Additional preclinical study involving nonhuman primates will be required to evaluate the utility and safety of this implant for spine fusion.

### Key Points

- The efficacy of the  $\beta$ -TCP/PLA-DX-PEG composite that contained low doses of rhBMP-2 was evaluated in a New Zealand White rabbit posterolateral lumbar intertransverse process fusion model.
- Rabbits that received 15 or 30  $\mu$ g of rhBMP-2 on each side, which is a significantly lower efficacious dose than previously reported, achieved solid fusion within 6 weeks in all experimental animals.
- This new synthetic biodegradable bone-inducing material could reduce the dose of rhBMP-2 required to achieve solid fusion with no residual trace of the implant.
- The current study demonstrated the potential of the new material as a substitute for autogenous bone graft material in spine fusion. This material obviates the need for bone graft procurement and thereby reduces the risk of morbidity often associated with this surgery.

### References

1. Arrington ED, Smith WJ, Chambers HG, et al. Complications of iliac crest bone graft harvesting. *Clin Orthop* 1996;300–9.
2. Kurz LT, Garfin SR, Booth RE Jr. Harvesting autogenous iliac bone grafts. A review of complications and techniques. *Spine* 1989;14:1324–31.
3. Buck BE, Malinin TI, Brown MD. Bone transplantation and human immunodeficiency virus. An estimate of risk of acquired immunodeficiency syndrome (AIDS). *Clin Orthop* 1989;129–36.
4. Leads from the MMWR. Transmission of HIV through bone transplantation: case report and public health recommendations. *JAMA* 1988;260:2487–8.
5. Akamaru T, Suh D, Boden SD, et al. Simple carrier matrix modifications can enhance delivery of recombinant human bone morphogenetic protein-2 for posterolateral spine fusion. *Spine* 2003;28:429–34.
6. Boden SD, Kang J, Sandhu H, et al. Use of recombinant human bone morphogenetic protein-2 to achieve posterolateral lumbar spine fusion in humans: a prospective, randomized clinical pilot trial: 2002 Volvo Award in clinical studies. *Spine* 2002;27:2662–73.
7. Boden SD, Schimandle JH, Hutton WC. An experimental lumbar intertransverse process spinal fusion model. Radiographic, histologic, and biomechanical healing characteristics. *Spine* 1995;20:412–20.
8. David SM, Gruber HE, Meyer RA Jr, et al. Lumbar spinal fusion using recombinant human bone morphogenetic protein in the canine. A comparison of three dosages and two carriers. *Spine* 1999;24:1973–9.
9. Grauer JN, Patel TC, Erulkar JS, et al. 2000 Young Investigator Research Award winner. Evaluation of OP-1 as a graft substitute for intertransverse process lumbar fusion. *Spine* 2001;26:127–33.
10. Itoh H, Ebara S, Kamimura M, et al. Experimental spinal fusion with use of recombinant human bone morphogenetic protein 2. *Spine* 1999;24:1402–5.
11. Johnsson R, Stromqvist B, Aspenberg P. Randomized radiostereometric study comparing osteogenic protein-1 (BMP-7) and autograft bone in human noninstrumented posterolateral lumbar fusion: 2002 Volvo award in clinical studies. *Spine* 2002;27:2654–61.
12. Konishi S, Nakamura H, Seki M, et al. Hydroxyapatite granule graft combined with recombinant human bone morphogenetic protein-2 for solid lumbar fusion. *J Spinal Disord Tech* 2002;15:237–44.
13. Minamide A, Kawakami M, Hashizume H, et al. Evaluation of carriers of bone morphogenetic protein for spinal fusion. *Spine* 2001;26:933–9.
14. Sandhu HS, Kanim LE, Kabo JM, et al. Effective doses of recombinant human bone morphogenetic protein-2 in experimental spinal fusion. *Spine* 1996;21:2115–22.
15. Schimandle JH, Boden SD, Hutton WC. Experimental spinal fusion with recombinant human bone morphogenetic protein-2. *Spine* 1995;20:1326–37.
16. Butler D. Last chance to stop and think on risks of xenotransplants. *Nature* 1998;391:320–4.
17. DeLustro F, Dasch J, Keefe J, et al. Immune responses to allogenic and xenogenic implants of collagen and collagen derivatives. *Clin Orthop* 1990; 263–79.
18. Saito N, Okada T, Horiuchi H, et al. A biodegradable polymer as a cytokine delivery system for inducing bone formation. *Nat Biotechnol* 2001;19: 332–5.
19. Saito N, Takaoka K. New synthetic biodegradable polymers as BMP carriers for bone tissue engineering. *Biomaterials* 2003;24:2287–93.
20. Murakami N, Saito N, Horiuchi H, et al. Repair of segmental defects in rabbit humeri with titanium fiber mesh cylinders containing recombinant human bone morphogenetic protein-2 (rhBMP-2) and a synthetic polymer. *J Biomed Mater Res* 2002;62:169–74.
21. Murakami N, Saito N, Takahashi J, et al. Repair of a proximal femoral bone defect in dogs using a porous surfaced prosthesis in combination with recombinant BMP-2 and a synthetic polymer carrier. *Biomaterials* 2003;24: 2153–9.
22. DePalma AF, Rothman RH. The nature of pseudarthrosis. *Clin Orthop* 1968;59:113–8.
23. Steinmann JC, Herkowitz HN. Pseudarthrosis of the spine. *Clin Orthop* 1992;80–90.
24. Matsushita N, Terai H, Okada T, et al. A new bone-inducing biodegradable porous beta-tricalcium phosphate. *J Biomed Mater Res* 2004;70A:450–8.

## Repair of an intercalated long bone defect with a synthetic biodegradable bone-inducing implant

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### Abstract

Recombinant human bone morphogenetic protein (rhBMP)-2 in a block copolymer composed of poly-D,L-lactic acid with randomly inserted *p*-dioxanone and polyethylene glycol (PLA-DX-PEG) as a carrier and porous  $\beta$ -tricalcium phosphate ( $\beta$ -TCP) blocks were used to generate a new fully absorbable osteogenic biomaterial. The bone regenerability of the rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP composite was studied in a critical-sized rabbit bone defect model. In an initial study, a composite of PLA-DX-PEG (250 mg) and  $\beta$ -TCP (300 mg) loaded with or without rhBMP2 (50  $\mu$ g) was implanted into a 1.5 cm intercalated bone defect created in a rabbit femur. Defects were assessed by biweekly radiography until 8 weeks postoperatively. The bony union of the defect was recognized only in the BMP-loaded group. To obtain further data on biomechanical and remodeling properties, another BMP-loaded composites group was made and observed up to 24 weeks. All defects were completely repaired without residual traces of implants. Anatomical and mechanical properties of the repaired bone examined by histology, 3-dimensional CT (3D-CT) and mechanical testing were essentially equivalent to the nonoperated-on femur at 24 weeks. These experimental results indicate that fully absorbable rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP is a promising composite having osteogenicity efficient enough for repairing large bone defects.

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**Keywords:** BMP (bone morphogenetic protein); Bone tissue engineering; Biodegradation; Osteoconduction; Drug delivery; Calcium phosphate

### 1. Introduction

The long history of orthopedic practice has confirmed the efficacy of autogenous bone grafting, but significant problems such as donor site morbidity and the limited supply associated with autogenous bone harvesting remain. To overcome these problems, a variety of osteoconductive biomaterials, e.g., ceramics and titanium alloys, have been considered as bone graft substitutes [1,2]. Experience to date with osteoconductive

biomaterials suggests that these can be made in greater quantities, would improve biomechanical strength, and present no concerns with immunogenicity [3]. These osteoconductive biomaterials are already efficacious in repairing unloaded bone cavities or small bone defects where an efficient osteogenic cell supply can be anticipated. Repairing large loaded bone defects using osteoconductive biomaterials alone remains a challenge, however, because osteogenic cells are not likely to be recruited without osteoinductivity in the center of large defects, and the lack of absorbability presents an obstacle to remodeling necessary to withstand repetitive mechanical loading. These considerations led us to conclude that biomaterials used to repair large bone

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defects should be completely biodegradable and have good osteoconduction and osteoinduction.

Among biodegradable and osteoconductive biomaterials such as synthetic porous polymers (poly[L-lactide-co-glycolide] copolymer (PLGA)) and tricalcium phosphates, porous beta-tricalcium phosphate ( $\beta$ -TCP) has proved most popular in current orthopedic surgery. Porous  $\beta$ -TCP has interconnected pores, which aid in infiltrating osteogenic cells, and is strong enough to maintain implant shape during bone formation [4].  $\beta$ -TCP, as well as natural bone matrix, is mainly absorbed by cells positive for tartrate-resistant acid phosphatase (TRAP) and is replaced by newly formed bone. Reports have also confirmed its osteoconductivity and degradability at orthotopic sites [5].

Osteoinductivity is currently added to biomaterials three main ways: (1) applying cultured osteogenic cells from autologous bone marrow (cell-based), (2) applying osteoinductive cytokines (cytokine-based), and (3) applying osteoinductive genes [6–8]. We used the cytokine-based approach because of its promising clinical application since several human osteoinductive cytokines have been produced by recombinant techniques [9].

Bone morphogenetic proteins (BMPs) are biologically active osteoinductive cytokines with significant clinical potential, but the lack of a delivery system enabling full osteoinduction has precluded their wider implementation in clinical therapeutics [10]. Ideal delivery should be synthetic to avoid the disease transmission possible with allogenic materials. Current carrier materials for BMPs, such as collagen sponges and hydroxyapatite, show successful bone formation in vivo, but require a high BMP dose because of their inability to retain BMPs [11,12]. Another requirement for ideal delivery is thus controlled BMP release enabling the amount of BMPs required for bone repair to be reduced by ensuring effective BMP retention in reactive cells. For this, we developed a synthetic block copolymer composed of poly-D,L-lactic acid with randomly inserted *p*-dioxanone and polyethylene glycol (PLA-DX-PEG), reported to deliver BMPs effectively [13–15].

To this end, we used porous  $\beta$ -TCP and recombinant human BMP-2 (rhBMP-2) with PLA-DX-PEG to generate a new, fully absorbable osteogenic biomaterial for repairing loaded large bone defects. We evaluated the efficacy of rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP composite in bone induction and degradability using a critical-sized intercalated rabbit bone defect model.

## 2. Materials and methods

### 2.1. Materials

Porous  $\beta$ -TCP cylinders of OSferion<sup>®</sup> 6 mm in diameter, 5 mm high, and weighing approximately

100 mg were manufactured and donated (Olympus Biomaterials Co., Ltd., Tokyo, Japan) for the purpose of these studies. Pores were 100–400  $\mu$ m in size, porosity was 75%, and the sintering was at 1050° [4]. rhBMP-2 produced at the Genetics Institute (Cambridge, MA) and donated through Yamanouchi Pharmaceutical Co., Ltd. (Ibaraki, Japan), was dissolved in a buffer of 5 mM glutamic acid, 2.5% glycine, 0.5% sucrose, and 0.01% Tween 80 at a concentration of 1  $\mu$ g/ $\mu$ l. As a rhBMP-2 delivery system, a block copolymer of poly-D,L-lactic acid with randomly inserted *p*-dioxanone and polyethylene glycol (PLA-DX-PEG; MW 9,600; PLA/DX/PEG molar ratio 43/14/43) was synthesized and provided by Taki Chemical Co., Ltd. (Kakogawa, Japan). The physicochemical properties and efficacy of the rhBMP-2 carrier material are detailed elsewhere [13–15]. We determined the minimal optimal rhBMP-2 content in the synthetic polymer required to elicit new bone formation to be approximately 0.02% (wt/wt) in rabbits from previous experiments [16–18].

### 2.2. Preparation of implants

To combine polymer and rhBMP-2, 250 mg of polymer was dissolved in 3 ml of distilled acetone and 50  $\mu$ g of rhBMP in 0.01N HCl was mixed in a glass vial. Three porous  $\beta$ -TCP cylinders were then submerged in the mixed solution and placed in a vacuum for a few seconds to replace air in  $\beta$ -TCP cylinder pores with solvent. Acetone was then removed with a centrifuge evaporator. The glass vials were shaken several times during evaporation to thoroughly impregnate cylinders with the rhBMP-2 delivery material. Treated porous  $\beta$ -TCP cylinders were coated with the rhBMP-2/PLA-DX-PEG composite (50  $\mu$ g of rhBMP2, 250 mg of PLA-DX-PEG, and 300 mg of  $\beta$ -TCP (3 cylinders) for each defect). Implants were kept in a freezer at –30 °C until use. Control implants were  $\beta$ -TCP cylinders coated with PLA-DX-PEG without rhBMP-2 prepared the same way as above.

### 2.3. Scanning electron microscopy (SEM)

The surface of porous  $\beta$ -TCP blocks was observed by scanning electron microscopy (SEM; Hitachi 4700SI) to ensure that the pore surface was coated by rhBMP-2/PLA-DX-PEG homogeneously (Fig. 1).

### 2.4. Animals and operative procedures

Of 24 Japanese white rabbits 3 months old purchased from Japan SLC Co., Ltd. (Shizuoka, Japan), 15 were used in Study 1 and 9 in Study 2. All experiments were conducted strictly observing institutional guidelines for the care and use of laboratory animals.



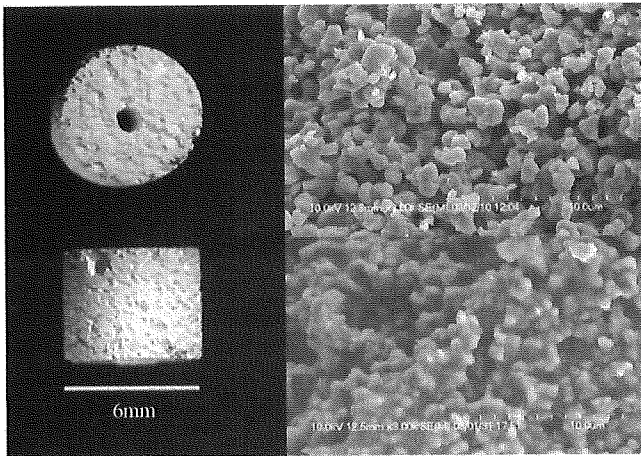


Fig. 1. Macroscopic aspect of  $\beta$ -TCP cylinders: images at left are axial and side views of the  $\beta$ -TCP cylinder, and at right are SEM micrographs of the  $\beta$ -TCP cylinder coated with PLA-DX-PEG  $\times 3000$  (above) and the noncoated control ( $\times 3000$ , below). The  $\beta$ -TCP cylinder consists of  $1\mu\text{m}$  sintered  $\beta$ -TCP granules with  $100\text{--}400\mu\text{m}$  interconnected pores. The surface of  $\beta$ -TCP granules in the experimental group is uniformly covered by a thin polymer layer.

Rabbits were anesthetized using an intramuscular injection of ketamine (60 mg/kg body weight) before surgery. Under sterile conditions, the mid-shaft of the femur was exposed through a lateral longitudinal skin incision and thigh muscles divided. A 1.5 cm long section of the diaphysis was removed together with the periosteum using a cutting saw and the bone defect either filled with prepared implants or left as is [19]. Femurs were then fixed with external fixators for up to 8 weeks postoperatively.

## 2.5. Experimental design

### 2.5.1. Effects of rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP on bone defect repair (Study 1)

Fifteen rabbits were divided into 3 groups of 5 each based on implants received, i.e., (1) 50  $\mu\text{g}$  of rhBMP2, 250 mg of PLA-DX-PEG, and 300 mg of  $\beta$ -TCP; (2) no rhBMP2 but the same amounts PLA-DX-PEG and  $\beta$ -TCP; and (3) no implant (defect controls). Femurs were assessed every 2 weeks using X-ray radiography, and bone formation was evaluated as shown in X-ray photographs below. All animals were euthanized by anesthesia overdose at 8 weeks and femurs in the first group collected and stored at  $-30^\circ\text{C}$  until mechanical testing in Study 2.  $\beta$ -TCP alone was not assessed in this study because PLA-DX-PEG alone only negligibly affected  $\beta$ -TCP osteoconductivity and degradability in previous studies [20].

### 2.5.2. Mechanical and remodeling properties of the repaired bone by rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP with long-term observation (Study 2)

Five rabbits underwent the same implants as group (1) in Study 1, i.e., 50  $\mu\text{g}$  of rhBMP2, 250 mg of PLA-

DX-PEG, and 300 mg of  $\beta$ -TCP, and external fixators were removed at 8 weeks postoperatively and observed for 24 weeks until animals were euthanized as described above. Femurs were collected to examine bone mineral density, then mechanically tested together with the 5 specimens in Study 1. Four other rabbits undergoing implants of 50  $\mu\text{g}$  of rhBMP2, 250 mg of PLA-DX-PEG, and 300 mg of  $\beta$ -TCP were euthanized (1 each) at 2, 4, 8, and 24 weeks postoperatively for histological examination.

## 2.6. X-ray analysis

New bone formation was evaluated by measuring the ratio of recognizable occupying callus to the defect in length because it is difficult to distinguish newly formed bone from  $\beta$ -TCP due to its radiopacity in the central portion of the defect [21].

## 2.7. Dual-energy X-ray absorptiometry analysis

Femurs harvested 24 weeks after surgery in Study 2 were examined via dual-energy X-ray absorptiometry (DXA) (DCS-600EX-III; Aloka Co., Ltd., Tokyo, Japan) using an analysis program designed for small animals. Bone mineral content (BMC) and area of new bone were measured and bone mineral density (BMD) calculated as BMC/bone area. Nonoperated-on femurs harvested from animals euthanized at 24 weeks were used as controls.

## 2.8. Mechanical testing

Five regenerated femurs each, harvested at 8 weeks in Study 1 and at 24 weeks, were used in 3-point bending mechanical testing with an apparatus designed for this purpose (Maruto Testing Machine Co., Tokyo, Japan). Nonoperated-on femurs harvested from animals euthanized at 24 weeks were used as controls. Other femurs implanted without rhBMP-2 in Study 1 showed no union, and hence were not mechanically tested. Maximum bending strength was measured.

## 2.9. Histological examination

For histological examination, samples were decalcified in 10% formic acid, dehydrated in a gradient ethanol series, mounted in paraffin, sectioned  $4\mu\text{m}$  thick, and stained with hematoxylin-eosin. To observe bone resorption, specimens were stained with tartrate-resistant acid phosphatase (TRAP). Briefly, deparaffinized sections were placed in a TRAP-staining solution consisting of acetate buffer (pH 5.0) 50 mM sodium tartrate, 25 mg/ml Naphthol-AS MX phosphate (Sigma Chemical Co., MO, USA), and 0.5 mg/ml

fast red violet salt (Sigma Chemical Co.) and incubated at 37 °C for 120 min. After the solution was removed by washing, specimens were counterstained with hematoxylin and observed under light microscopy.

### 2.10. Computed tomography (CT) scanning images

Computed tomography (CT) data on harvested distal femurs at 24 weeks was collected with a helical CT (GE Yokogawa, Tokyo, Japan) and images were reconstructed using 3-dimensional image reconstruction software (Aze, Tokyo, Japan).

### 2.11. Statistical analysis

Student's *t*-test was used to determine statistical significance, with  $P < 0.05$  considered significant.

## 3. Results

### 3.1. Defect repair effect of rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP (Study 1)

Representative radiology of bone defects at 2, 4, and 8 weeks after surgery (Fig. 2) demonstrated opaque calcified shadows bridging both ends of defects as early as 2 weeks in the group implanted with  $\beta$ -TCP combined with rhBMP-2 and polymer. Calcification became more evident at 4 weeks, and newly formed bone connecting both ends of defects appeared to have been remodeled into cortical bone with a bone marrow cavity. The BMP-loaded group showed a time-dependent increase in callus of nearly 100% at 6 weeks, but the group with  $\beta$ -TCP and polymer without rhBMP-2 showed only small amounts of newly formed bone formation—less than 20%. No bone formation was recognized in the control group (Fig. 3).

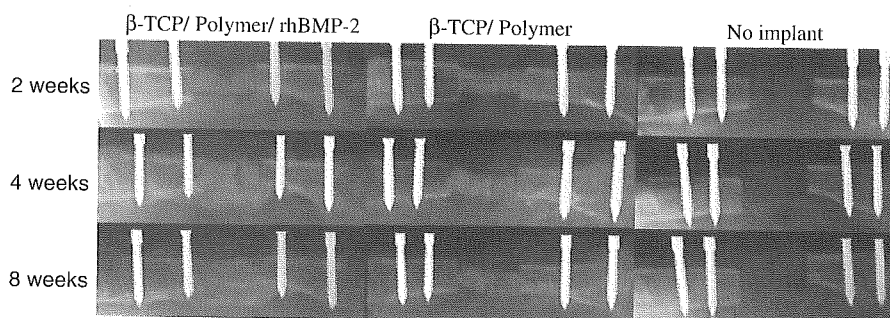


Fig. 2. Representative femur radiographs. From left, implanted with  $\beta$ -TCP with PLA-DX-PEG and rhBMP-2,  $\beta$ -TCP with PLA-DX-PEG without rhBMP-2, and critical size bone defect without implantation (sham surgery). Sequential radiographs show bone repair at 2, 4, and 8 weeks after implantation in the experimental group.

### 3.2. Mechanical and remodeling properties of the repaired bone by rhBMP-2/PLA-DX-PEG/ $\beta$ -TCP with long-term observation (Study 2)

Femurs in this group were stable and enabled free movement in caged animals until their sacrifice at 24 weeks. Radiology of femurs harvested 24 weeks after surgery showed complete regeneration of the intercalated defect. The 3D CT image and frontal tomographic image of regenerated femurs showed that normal femur anatomy had been restored with cortical bone with no residual evidence of implanted  $\beta$ -TCP cylinder blocks (Fig. 4).

#### 3.2.1. Dual-energy X-ray absorptiometry analysis

Bone mineral density of femurs regenerated by biodegradable bone-inducing implants at 24 weeks was 350 mg/cm<sup>2</sup>, essentially equal to control (normal) levels. No significance was seen between experimental and control groups.

#### 3.2.2. Biomechanical properties of repaired bone

The 3-point bending test of femurs regenerated by biodegradable bone-inducing implants at 8 weeks showed significantly lower stiffness (160 N/m) than

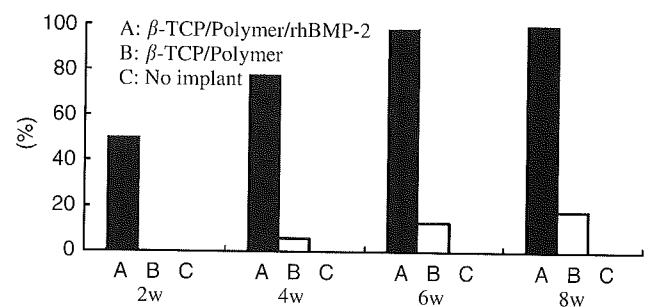


Fig. 3. BMP-loaded group promoting a time-dependent increase in callus (nearly 100% at 6 weeks). The group of  $\beta$ -TCP and polymer without rhBMP-2 promoted only negligible new bone formation—less than 20%. No bone formed in the control group.

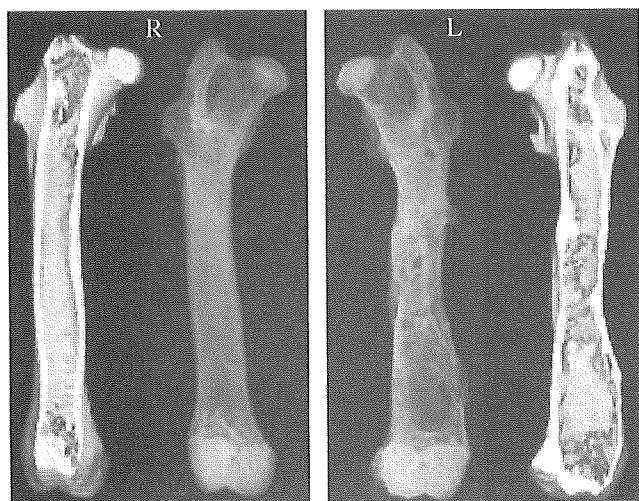


Fig. 4. Soft X-ray and 3D CT images of femurs 24 weeks after surgery. The repaired defect with biodegradable bone-inducing implants is shown at right (L). Images at left are of the counterpart femur in the control rabbit (R). The external fixator was removed 8 weeks after surgery. Note that  $\beta$ -TCP was absorbed and cortical walls remodeled anatomically with the marrow cavity.

controls (400 N/m, nonsurgical femurs 24 weeks after surgery). Stiffness increased at 24 weeks (370 N/m) and was essentially equal to control (normal) levels (Fig. 5).

### 3.2.3. Histological findings

In histological sections from defect sites 2 weeks postoperatively, fibrous tissue and a thin layer of bone running parallel to the long axis and encasing implants were seen in the experimental group. A femur from the experimental group at 4 weeks with increased bone mass connected to  $\beta$ -TCP implants and fibrous tissue was observed. Regenerated cortical bone had united with ends of the original cortex of the femur. In sections from the experimental group 8 weeks postoperatively, bone occupying defects had remodeled to where cortical bone and hematopoietic marrow-like tissue were clearly visible (Fig. 6). On the  $\beta$ -TCP surface at 4 weeks, large numbers of TRAP-positive multinucleated cells (osteoclasts) appeared (Fig. 7). In sections of regenerated defects at 24 weeks, cortical bone was remodeled to lamellar bone connected to original ends of the femur. Marrow tissue was also completely restored and no remnants of  $\beta$ -TCP implants were visible (Fig. 8).

## 4. Discussion

Our experimental results indicated successful regeneration of a critical intercalated bone defect in femur implantation using porous  $\beta$ -TCP with rhBMP-2 and a synthetic PLA-DX-PEG block copolymer as its delivery system. This approach clearly demonstrated that combining these synthetic materials and recombinant protein repaired large defects. The osteogenic potential of composite implants has yet to be compared critically, however.

The rhBMP-2 dose and rhBMP-2 concentration in polymer we used was determined based on previous study of critical bone defect repair in rabbits in which 1.5 cm of an intercalated bone defect in the humerus was

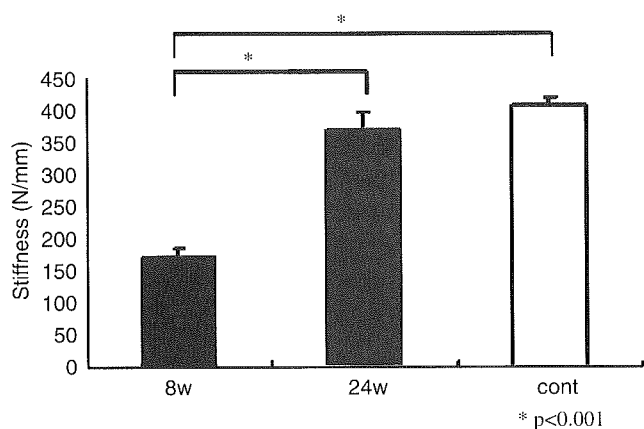


Fig. 5. Results of 3-point bending tests at 8 and 24 weeks after surgery (bar chart). (\*: significant difference,  $p < 0.001$ ).

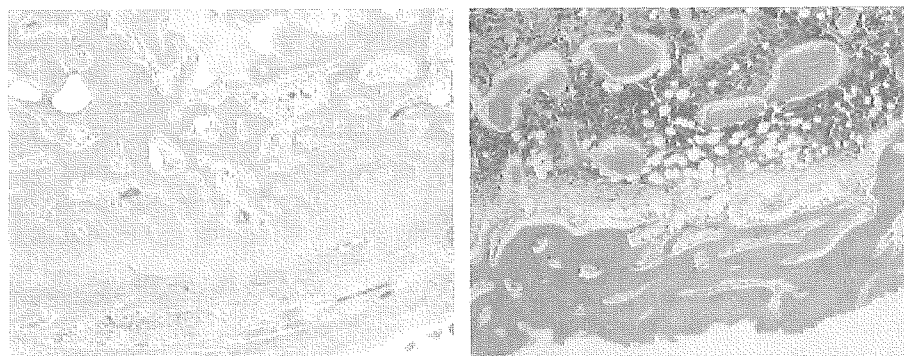


Fig. 6. Typical histological sections at 4 weeks (left, HE staining  $\times 40$ ) and 8 weeks (right, HE staining  $\times 40$ ). Abundant bone formed around  $\beta$ -TCP but not the outside of the femur in sections of specimens at 4 weeks. Cortical and bone marrow-like tissue clearly identified at 8 weeks.

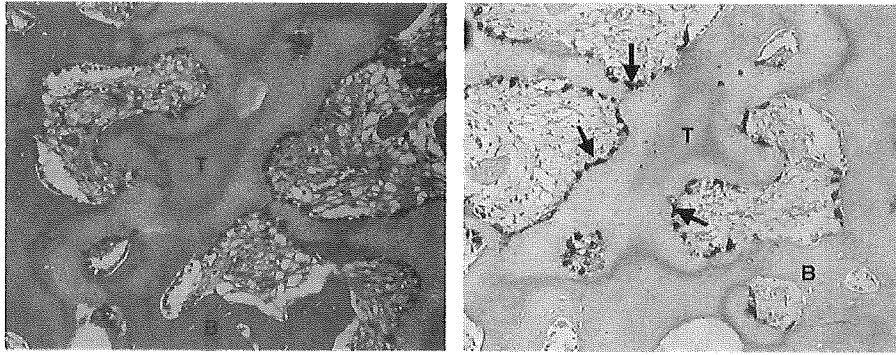


Fig. 7. Histological section of decalcified specimens harvested at 4 weeks and stained with HE (left,  $\times 40$ ) and tartrate-resistant acid phosphatase (TRAP) (right,  $\times 40$ ).  $\beta$ -TCP blocks coated with PLA-DX-PEG and rhBMP-2 are surrounded by abundant TRAP-positive multinucleated osteoclasts (arrows, stained red). (B = bone, T =  $\beta$ -TCP).

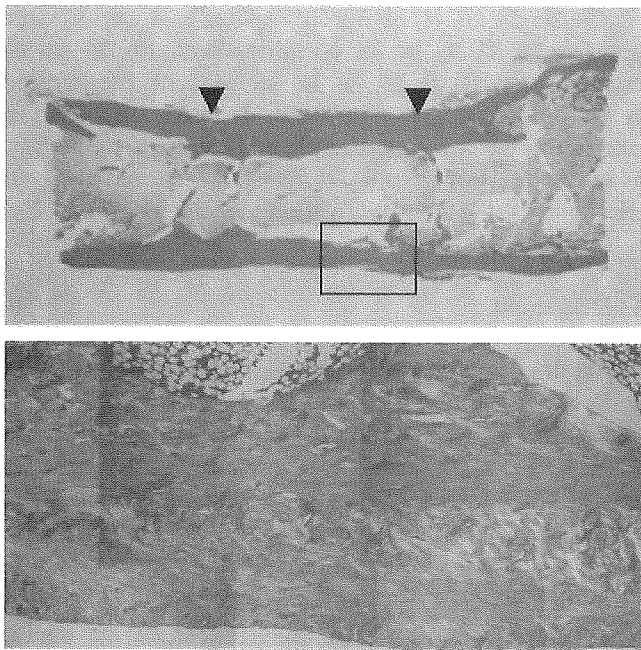


Fig. 8. Images of sagittal histological section at 24 weeks. Cortical walls were repaired completely. Arrowheads indicate fixator pin insertion locations. Polarized magnified image of junction between original and repaired bone (rectangle) (below). Randomly arrayed collagen in the repaired cortical wall contrasts with the more ordered layers of collagen in the original cortical wall.

successfully repaired by filling the defect with titanium mesh cylinders impregnated with a delivery system containing rhBMP-2 [17]. Titanium cylinders were not resorbed in repaired bone. Sustained permanent release of metal ions from the implant could increase the potential risk of allergic reactions or carcinogenesis, especially in infants. For these reasons, we replaced titanium in the present study with biodegradable material. As expected,  $\beta$ -TCP was completely resorbed and replaced by host bone within 24 weeks with no apparent adverse events from resorbed  $\beta$ -TCP.

The successful regeneration of the critical bone defect may be due in part to the porosity of  $\beta$ -TCP cylinders having 100–400  $\mu\text{m}$  pores, which may enable rhBMP to accumulate locally within pores to where osteogenesis is initiated [22–25]. An appropriate local rhBMP concentration in pores would in turn enable successful invasion and ingrowth of mesenchymal cells in the implant and subsequent differentiation into osteoblasts. Additionally, the porosity of  $\beta$ -TCP appeared to be crucial to its rapid degradation in host animals, facilitating contact with host cells and resulting clearly in large numbers of osteoclasts contacting  $\beta$ -TCP [26]. The ability of these cells to permeate and resorb the  $\beta$ -TCP mass likely contributed to the relatively rapid replacement of implants by bone and marrow in our study. Although the location of BMP receptors on the surface of osteoclasts was reported previously [27], the effect of rhBMP-2 on osteoclastic differentiation remains to be clarified, requiring further study to determine the potential action of BMP on osteoclasts to explain the increased recruitment of osteoclasts in new bone induced by rhBMP2. Biodegradable osteoconductive  $\beta$ -TCP combined with a BMP delivery system is replaced by fully integrated biomechanically competent bone, eliminating one of the major limitations of other osteoconductive biomaterials.

The newly formed bone repairing the defect was remodeled to restore the normal anatomy of the original bone with concurrent resorption of  $\beta$ -TCP in 8 weeks. This means that BMP-induced new bone could remodel to adapt to the local biomechanical environment. Osteoconductive material must disappear to generate the physiological biomechanical environment and restore the original anatomy. Our results suggest that absorbable  $\beta$ -TCP is suitable as an ideal bone graft substitute.

The physicochemical properties and degradation profiles of polymer used in this study have been reported elsewhere [13–15]. A detailed safety check for clinical use of the polymer is currently in process. No systemic or local adverse effects have been noted to date.

The optimal content of rhBMP-2 in the carrier required to elicit new bone formation depends on the animal host [16–18], and a higher dose of rhBMP-2 is required in highly evolved animal species. Determining the optimal clinical dose thus requires additional experiments in primates.

## 5. Conclusion

A new absorbable bone graft substitute with osteogenic capacity was made by combining 3 artificial materials—porous  $\beta$ -TCP, rhBMP-2, and a PLA-DX-PEG delivery system. The capacity of this composite implant to regenerate bone is satisfactory. The composite implant was completely absorbed and replaced by newly formed bone, then remodeled into the femur to restore the natural anatomy. Although further safety checks and clinical trials are required, the practical use of this implant to promote bone regeneration is without doubt a realistic possibility.

## Acknowledgements

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## References

- [1] Bauer TW, Muschler GF. Bone graft materials. An overview of the basic science. *Clin Orthop* 2000;371:10–27.
- [2] Greenwald AS, Boden SD, Goldberg VM, Khan Y, Laurencin CT, Rosier RN. Bone-graft substitutes: fact, fictions & applications. *J Bone Joint Surg Am* 2001;83(Suppl 2):98–103.
- [3] Hoogendoorn HA, Renooij W, Akkermans LMA, Visser DDS, Wittebol P. Long-term study of large ceramic implants in dog femora. *Clin Orthop* 1984;187:281–8.
- [4] Ozawa M. Experimental study on bone conductivity and absorbability of the pure  $\beta$ -TCP. *J Jpn Soc Biomater* 1995;13:167–75.
- [5] Debruijn JD, Bovell YP, Davies JE, Vanblitterswijk CA. Osteoclastic resorption of calcium phosphate is potentiated in postosteogenic culture conditions. *J Biomed Mater Res* 1994;28-1:105–12.
- [6] Dong J, Kojima H, Uemura T, Kikuchi M, Tateishi T, Tanaka J. In vivo evaluation of a novel porous hydroxyapatite to sustain osteogenesis of transplanted bone marrow-derived osteoblastic cells. *J Biomed Mater Res* 2001;57-2:208–16.
- [7] Lieberman JR, Daluiski A, Einhorn TA. The role of growth factors in the repair of bone. Biology and clinical application. *J Bone Jt Surg Am* 2002;84-A6:1032–44.
- [8] Baltzer AWA, Lattermann C, Whalen JD, Wooley P, Weiss K, Grimm M, Ghivizzani SC, Robbins PD, Evans CH. Genetic enhancement of fracture repair: healing of an experimental segmental defect by adenoviral transfer of the BMP-2 gene. *Gene Therapy* 2000;7-9:734–9.
- [9] Wang EA, Rosen V, D'Alessandro JS, Bauduy M, Cordes P, Harada T, Israel DI. Recombinant human bone morphogenetic protein induces bone formation. *Proc Natl Acad Sci USA* 1990;87:2220–4.
- [10] Takaoka K, Nakahara H, Yoshikawa H, Masuhara K, Tsuda T, Ono K. Ectopic Bone Induction on and in porous hydroxyapatite combined with collagen and bone morphogenetic protein. *Clin Orthop* 1988;234:250–4.
- [11] Zabka AG, Pluhar GE, Edwards RB, Manley PA, Hayashi K, Heiner JP, Kalscheur VL, Seeherman HJ, Markel MD. Histomorphometric description of allograft bone remodeling and union in a canine segmental femoral defect model: a comparison of rhBMP-2, cancellous bone graft, and absorbable collagen sponge. *J Orthop Res* 2001;19-2:318–27.
- [12] Jeppsson C, Bostrom M, Aspenberg P. Intraosseous BMP implants in rabbits—Inhibitory effect on bone formation. *Acta Orthop Scand* 1999;70-1:77–83.
- [13] Saito N, Okada T, Toba S, Miyamoto S, Takaoka K. New synthetic absorbable polymers as BMP carriers: plastic properties of poly-D,L-lactic acid-polyethylene glycol block copolymers. *J Biomed Mater Res* 1999;47-1:104–10.
- [14] Saito N, Okada T, Horiuchi H, Murakami N, Takahashi J, Nawata M, Ota H, Nozaki K, Takaoka K. A biodegradable polymer as a cytokine delivery system for inducing bone formation. *Nat Biotechnol* 2001;19-4:332–5.
- [15] Saito N, Okada T, Horiuchi H, Murakami N, Takahashi J, Nawata M, Ota H, Miyamoto S, Nozaki K, Takaoka K. Biodegradable poly-D,L-lactic acid-polyethylene glycol block copolymers as a BMP delivery system for inducing bone. *J Bone Jt Surg Am* 2001;83-A(Suppl 1-Pt 2):S92–8.
- [16] Saito N, Okada T, Horiuchi H, Ota H, Takahashi J, Murakami N, Nawata M, Kojima S, Nozaki K, Takaoka K. Local bone formation by injection of recombinant human bone morphogenetic protein-2 contained in polymer carriers. *Bone* 2003;32-4:381–6.
- [17] Murakami N, Saito N, Horiuchi H, Okada T, Nozaki K, Takaoka K. Repair of segmental defects in rabbit humeri with titanium fiber mesh cylinders containing recombinant human bone morphogenetic protein-2 (rhBMP-2) and a synthetic polymer. *J Biomed Mater Res* 2002;62-2:169–74.
- [18] Murakami N, Saito N, Takahashi J, Ota H, Horiuchi H, Nawata M, Okada T, Nozaki K, Takaoka K. Repair of a proximal femoral bone defect in dogs using a porous surfaced prosthesis in combination with recombinant BMP-2 and a synthetic polymer carrier. *Biomaterials* 2003;24-13:2153–9.
- [19] Crigel MH, Balligand M. Critical size defect model on the femur in rabbits. *Vet Comp Orthop Traumatol* 2002;3:158–63.
- [20] Matsushita N, Terai H, Okada T, Nozaki K, Inoue H, Miyamoto S, Takaoka K. A new bone-inducing biodegradable porous beta-tricalcium phosphate. *J Biomed Mater Res* 2004;70A-3:450–8.
- [21] Kirkerhead CA, Gerhart TN, Schelling SH, Hennig GE, Wang E, Holtrop ME. Long-term healing of bone using recombinant human bone morphogenetic protein 2. *Clin Orthop* 1995;318:222–30.
- [22] Urist MR, Lietze A, Dawson E.  $\beta$ -tricalcium phosphate delivery system for bone morphogenetic protein. *Clin Orthop* 1983;187:277–80.
- [23] Urist MR, Nilsson O, Rasmussen J, Hirota W, Lovell T, Schmalzreid T, Finerman GA. Bone regeneration under the influence of a bone morphogenetic protein (BMP) beta tricalcium phosphate (TCP) composite in skull trephine defects in dogs. *Clin Orthop* 1987;214:295–304.
- [24] Laffargue P, Hildebrand HF, Rtaimate M, Frayssinet P, Amoureux JP, Marchandise X. Evaluation of human recombinant bone morphogenetic protein-2-loaded tricalcium phosphate implants in rabbits' bone defects. *Bone* 1999;25-2(Suppl):55S–8S.

- [25] Jingushi S, Urabe K, Okazaki K, Hirata G, Sakai A, Ikenoue T, Iwamoto Y. Intramuscular bone induction by human recombinant bone morphogenetic protein-2 with beta-tricalcium phosphate as a carrier: in vivo bone banking for muscle-pedicle autograft. *J Orthop Sci* 2002;7-4:490–4.
- [26] Yamada S, Heymann D, Bouler JM, Daculsi G. Osteoclast resorption of calcium phosphate ceramics with different hydroxyapatite/ $\beta$ -tricalcium phosphate ratios. *Biomaterials* 1997;18:1037–41.
- [27] Kaneko H, Arakawa T, Mano H, Kaneda T, Ogasawara A, Nakagawa M, Toyama Y, Yabe Y, Kumegawa M, Hakeda Y. Direct stimulation of osteoclastic bone resorption by bone morphogenetic protein (BMP)-2 and expression of BMP receptors in mature osteoclasts. *Bone* 2000;27:479–86.

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## 骨組織再生

*Regeneration of the bone tissue*

### Keywords

骨組織再生

骨形成因子

生体吸収性→用語解説 78 頁

ポリマー

$\beta$ -TCP→用語解説 78 頁

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大阪市立大学大学院医学研究科 整形外科学教室

### Summary

Regeneration of the bone tissue has been required in various clinical scenes, especially in orthopaedic surgery, dental and oral surgeries. Regeneration of tissues requires cells, cytokines (growth factors) and scaffold. In the field of bone regeneration, bone morphogenetic proteins (BMPs) have utilized as main cytokines with osteoinductive capacity. We introduce the efficacy of implants composed of recombinant BMP and biodegradable polymers and  $\beta$ -tricalcium phosphate ( $\beta$ -TCP) as new scaffold or drug delivery system for the BMP to regenerate various types of bone defects with potential use in clinical practice as bone graft substitute.

Although various lines of researches for the bone regeneration have been progressed all over the world with variety of technologies including tissue engineering and gene therapy, these have problems remained to be solved and appear to be fur from practical use so far.

### 背景

近年、さまざまな組織における再生医学技術が進歩し、一部が先端医学として実際に臨床応用されつつある。骨組織再生は、必要な部位に必要な量・形態の骨を形成することを目的としている。臨床的には、脊椎疾患に対する脊椎固定術・外傷や腫瘍摘出に伴う巨大骨欠損の再建・顔面骨再建などで必要とされている。従来は骨再生の目的には自家骨移植が用いられてきた。しかし、自家骨移植による骨再生にはさまざまな問題があることが指摘されてきた。採取部の外科的侵襲や慢性的疼痛、知覚障害、変形などである。さらに移植骨の採取量の限界、力学的強度の不足など多くの問題があった。それらの欠点を補う方法として同種骨移植や生体親和性生体材料が用いられることもあったが、それらには骨形成促進(骨誘導)能力に欠けるという決

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定的な問題点があること、また同種骨移植ではウイルス感染などの危険性や、保存、運搬などのシステム(骨銀行)が必要であるなどの理由で、わが国では汎用には至っていないのが現状である。したがって、臨床的な実用という面からみても、さらに優れた骨再生技術が求められている。理想的な骨再生技術または骨再生材料に求められる仕様としては、①移植材作成に生体への外科侵襲を伴わないこと、②生物由来でなく合成材料であること、③骨形成促進活性を有すること、④生体吸収性があり、骨再生完成後消失すること、⑤形成される骨の量・形態の制御が可能であること、⑥力学的強度に優れること、⑦可塑性があり、容易に成型できること、⑧生体に対する安全性に優れること、⑨安価で経済性に優れていること、⑩保存・搬送が容易であること、などが求められる。このような特性を満足する実用的な再生医療技術を可能にする材料を開発することは必ずしも容易ではない。

組織再生という生物現象には3要素が必須であるとされる。①分化能を有する未分化細胞、②細胞の増殖・分化を刺激する成長因子/サイトカイン、③その細胞が増殖分化する足場(Scaffold)である。骨組織の再生では、未分化間葉系細胞とそれらを軟骨細胞や骨芽細胞に分化誘導するサイトカインである一群の骨形成蛋白(Bone morphogenetic proteins: BMPs)が大きな役割を担っている。ScaffoldはBMPの担体であり、動物由来のコ

ラーゲンが主に用いられてきた。

BMPsは、1965年にDr. Uristが、塩酸処理骨基質による異所性新生骨の誘導を報告し、その存在を提唱した<sup>1)</sup>。その後、長期間にわたりさまざまな研究がなされたが、抽出は困難を極めた<sup>2)</sup>。1980年代後半、アメリカGenetic Institute社により遺伝子情報がcloningされ、リコンビナント蛋白として分離・精製された<sup>3)</sup>。現在では、骨再生医療の基本的な材料としてヒト型リコンビナントBMP(Recombinant human: rhBMP)が用いられている。すでに限られた用途(脊椎前方固定)で臨床利用が始まっている<sup>4)</sup>。しかし、使用については多くの問題があり、その解決が迫られている。

本稿ではBMPを用いた骨再生技術の現状について記述する。

### BMPの生物化学的特性

BMPsは、正常骨芽細胞系細胞によって産生分泌されるホモ2量体の活性蛋白(サイトカイン)である。その成熟領域の分子構造の特徴として、分子量は30kDa前後で疎水性の強い中性蛋白分子である。また、C末端からみたアミノ酸配列では7個のシステイン残基を有し、その位置がTransforming growth factor- $\beta$  (TGF- $\beta$ )と同一である。したがって、BMPsはTGF- $\beta$  super-familyに属する一群のfamilyをなしている(図1)<sup>5)</sup>。

現在ではBMP-2~BMP-15の14種類の分子が同定されている。このうち

骨形成促進活性を有して異所性に骨形成を誘導する活性が確認されているのはBMP-2, BMP-4, BMP-6, BMP-7(OP-1), BMP-9である<sup>6)</sup>。それぞれのBMPの詳細な生理学的役割の違いは明らかではないが、発生過程<sup>7)</sup>や骨折治癒過程<sup>8)</sup>でみられる骨形成にはBMP発現が必須である。

BMPの分子構造にはcystine knotと称する安定化構造があり、極めて安定した蛋白である。したがって、変性しにくく、90℃30分程度の熱処理でも生物活性は保たれる。すなわち、保存・搬送に関する問題はほとんどないとされる。

### BMPによる新生骨誘導

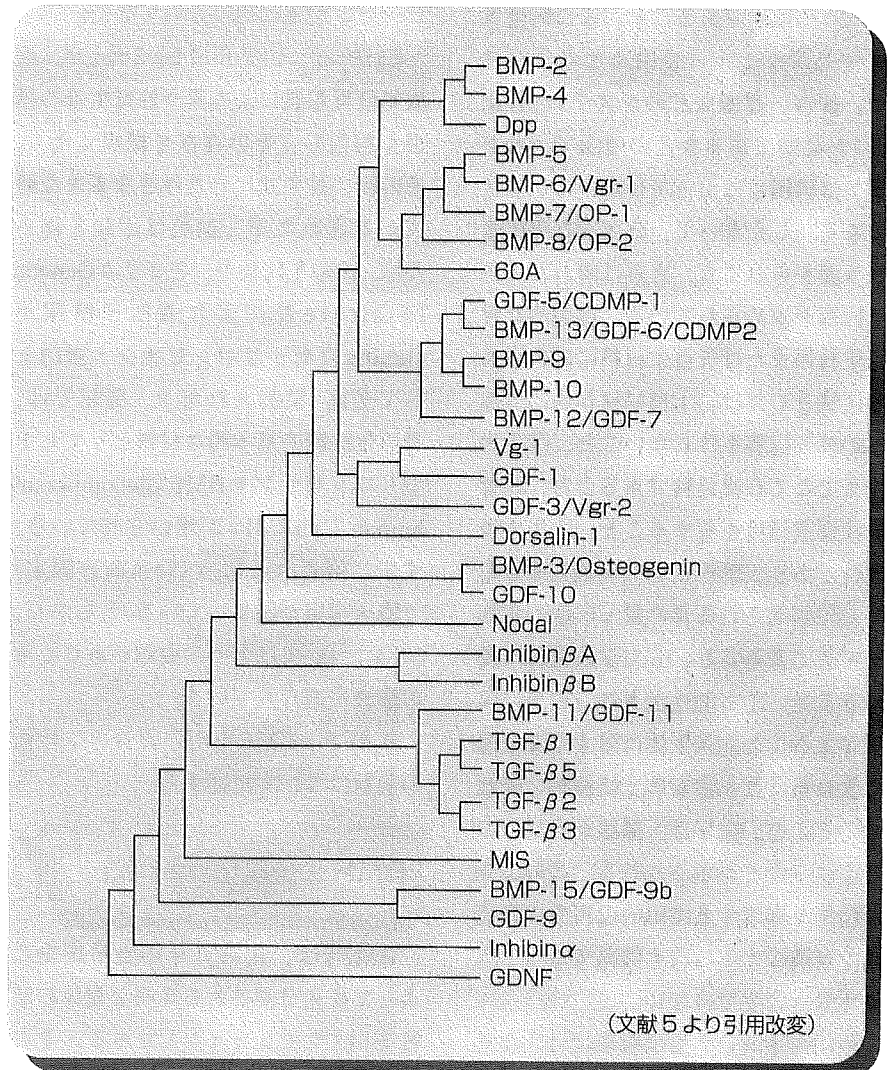
rhBMPの生体内埋植による局所的骨形成促進効果は、これまで多くの実験で示されている。大きな骨欠損修復や脊椎固定がrhBMPによって可能であることも実験動物での結果が報告されている<sup>9)</sup>。実際の臨床治療試験もすでに一部行われており、欧米では脊椎固定、骨折後偽関節治療などでの限定的使用が許可されている<sup>10)</sup>。しかし、BMPの臨床応用のためには、まだ改良を加え解決しなければならないいくつかの問題が残されている。現時点での主な問題点は、BMPの担体としてウシのコラーゲンが使用されていることからくる狂牛病などのリスク、ヒトでのBMPに対する低い応答性に由来する起因した高用量のBMPの必要性、その結果として高価となっており一般

的普及が困難な点である。この問題解決には新しい安全な担体の開発、低用量BMPで骨再生を可能とする新しい担体 (drug delivery system : DDS) の開発が望まれている。リコンビナントBMPとして実際に臨床応用されているBMPは、BMP-2およびBMP-7(OP-1)であるが、いずれについてもこのような欠点が解決されていない。

脊椎固定術に関して欧米では、リコンビナントBMPと担体(コラーゲン)を一体化して生体材料として用いている。rhBMP-2についてはMedotronic Sofamor Danec社から、またBMP-7(OP-1)についてはStryker社から発売されている。わが国でのBMPの臨床応用ははまだ認可されていない。

### BMPの効果的担体

重要な問題はBMPをいかに優れたDDSによって生体内の必要な限局した部位に作用させ、より安全かつ効果的に骨形成を促進するかである。現在使われている担体であるコラーゲンの使用についてはBSEなどの感染症問題が指摘されており、さらに力学的強度に欠ける点などの問題があり、今後コラーゲンに代替し得る担体の開発は骨組織の再生医療を拡大していく上で急を要する。すなわち、BMPの骨形成促進活性を有効に利用するためには、安全で安価な生体吸収性で成型しやすい合成担体が必要である。我々はこのような特性を有する担体として数種類の合成ポリマーを開発した(poly-

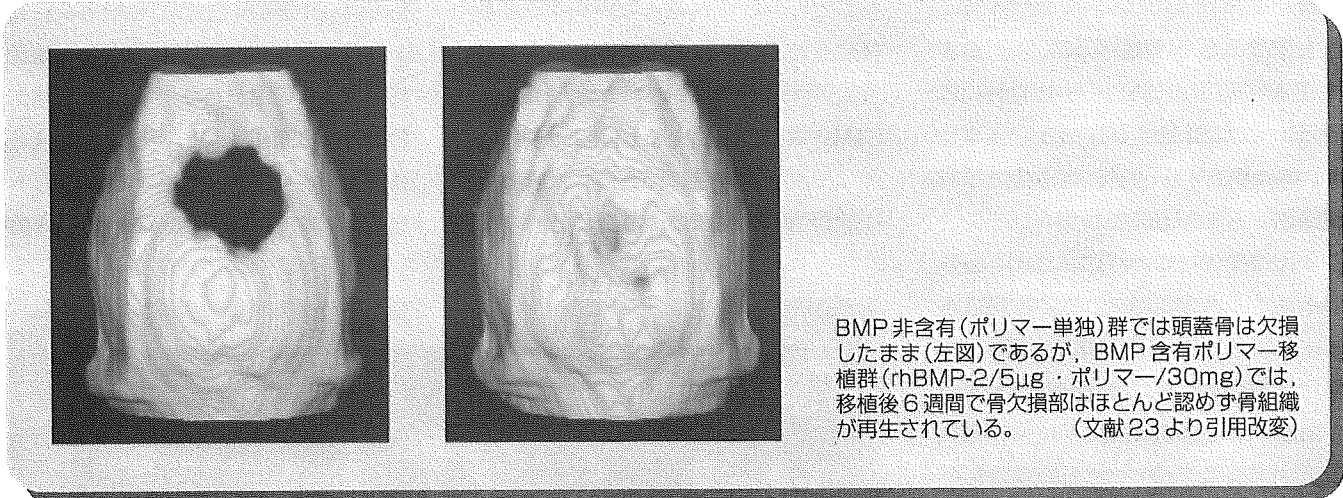


(文献5より引用改変)

図1 TGF-βスーパーファミリー

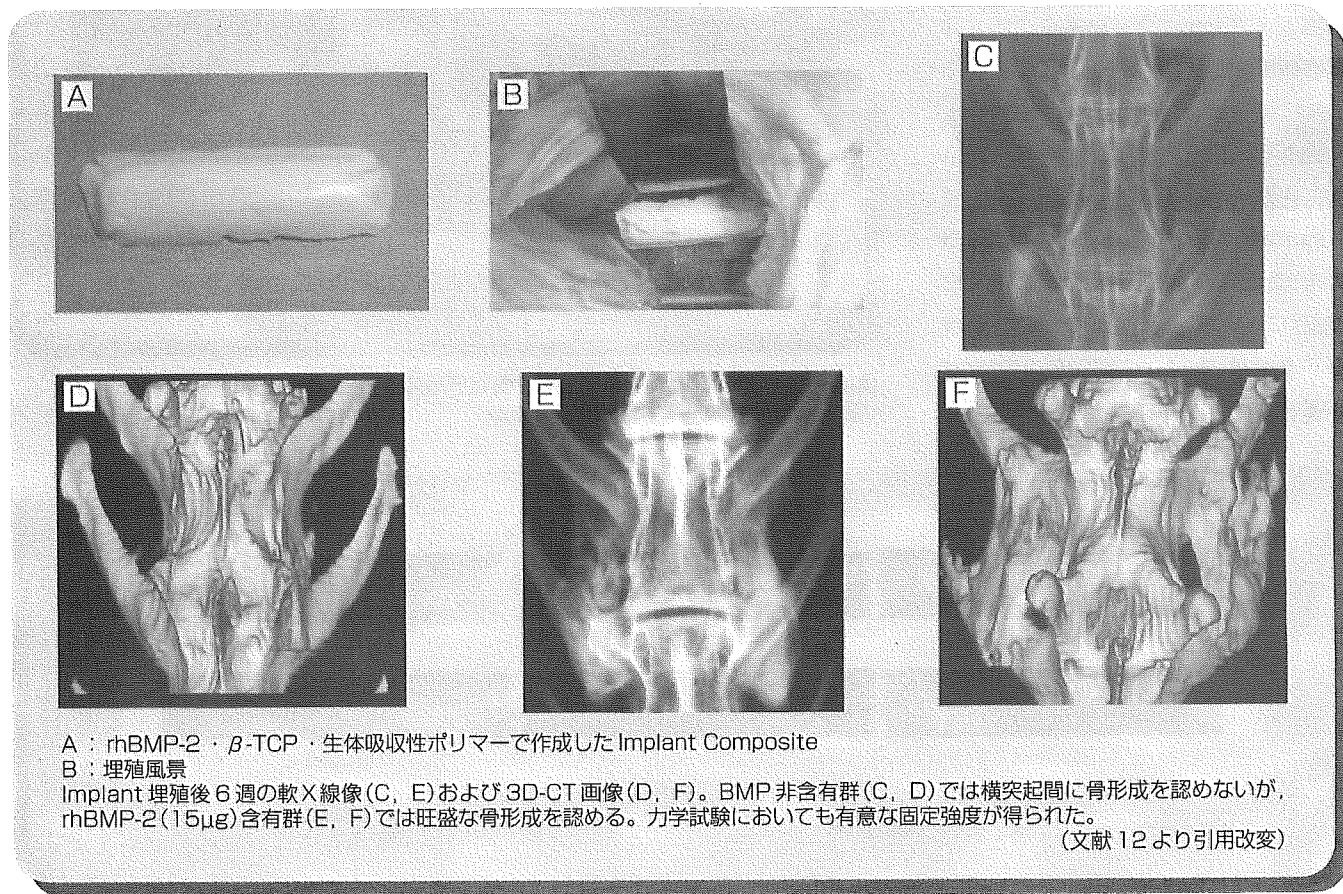
lactic acid-polyethyleneglycol block copolymer : PLA-PEG) <sup>11)</sup>。この polymer に少量のBMPを混和して生体に埋植すると異所性にも同所性にも骨新生が起り、埋植した polymer と置き換わる。この polymer の詳細についてはすでに報告してあるので、ここでは省略する。このBMP/polymer複合材料を用いると比較的大きな骨欠損を修復で

きる。図2はラットの頭蓋骨欠損の例を示す。しかしこの polymer の欠点として、常温では粘着性が強いために扱い難い点である。この点を改良するために、生体吸収性β-tricalcium phosphate (β-TCP) 粉末を polymer と等量混和することで粘土状にした。これによってインプラント材料の容積がほぼ倍加し、形成される骨量もほぼ倍加す



BMP非含有(ポリマー単独)群では頭蓋骨は欠損したまま(左図)であるが、BMP含有ポリマー移植群(rhBMP-2/5 $\mu$ g・ポリマー/30mg)では、移植後6週間で骨欠損部はほとんど認めず骨組織が再生されている。(文献23より引用改変)

図2 ラット頭蓋骨欠損モデル3D-CT画像



A : rhBMP-2・ $\beta$ -TCP・生体吸収性ポリマーで作成した Implant Composite  
 B : 埋植風景  
 Implant 埋植後6週の軟X線像(C, E)および3D-CT画像(D, F)。BMP非含有群(C, D)では横突起間に骨形成を認めないが、rhBMP-2(15 $\mu$ g)含有群(E, F)では旺盛な骨形成を認める。力学試験においても有意な固定強度が得られた。(文献12より引用改変)

図3 rhBMP-2・ $\beta$ -TCP・生体吸収性ポリマーによる日本白色家兎脊椎固定(→巻頭Color Gravure参照)

るため結果としてBMPの骨形成効率を上げることも可能となる。また粘着性を減ずることで扱いやすい材料にできる。このBMP/polymer/ $\beta$ -TCP powder複合材料を用いた動物での脊椎後側方固定を図3に示す<sup>12)</sup>。

多孔性生体材料の孔内にBMP/polymerを封入することで、立体的構造をもったインプラントを構成し骨欠損再生も可能である。ウサギ大腿骨の1.5cmのcritical defectの再生例を図4に示す<sup>9)</sup>。この例のように多孔性 $\beta$ -TCPを用いれば生体材料を残さずに骨再生が可能である。この材料を使えばイヌの8cmに及ぶ欠損の再生も比較的短時間で可能である(図5)。

### BMPに対する反応性の種差

rhBMPを普遍的に臨床応用するための大きな障害として、ヒトでのBMPに対する応答性が低いことがある。その結果として高用量のBMPを必要とし、必然的に高価な治療法とな

ることがあげられる。コラーゲンを担体とした場合、ヒトでは1cm<sup>3</sup>の新生骨を形成するためには約1mgのBMPが必要である。上述の新規担体では約半分の用量にできる。できるだけ低用量のBMPで効果的に骨再生を

達成するための工夫も必要である。そのための工夫として①DDSを至適化すること、②BMPの生物活性を増幅する方法の開発、があげられる。またBMPの生物活性を増幅する方法として、我々はprostaglandin E<sub>2</sub> (PGE<sub>2</sub>)の

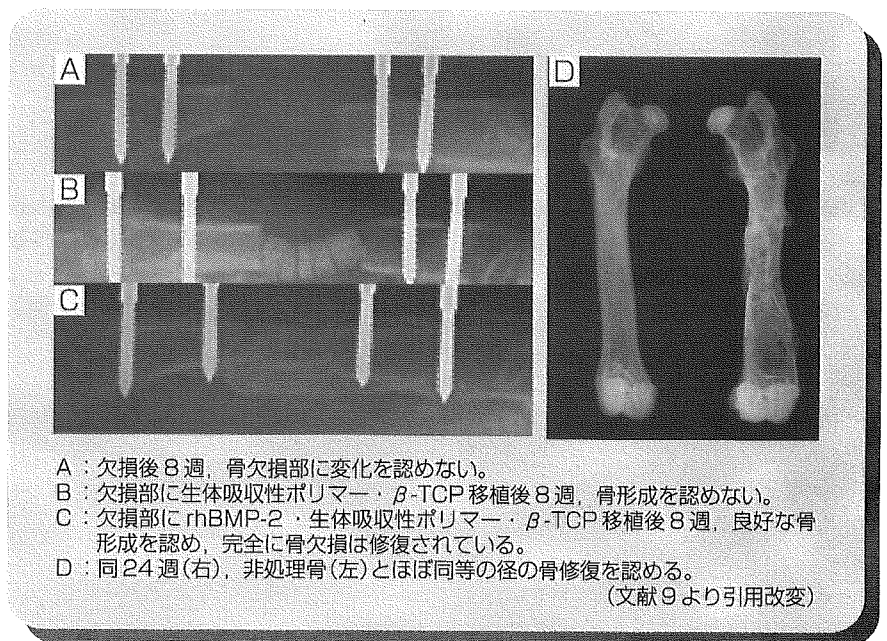


図4 日本白色家兎大腿骨巨大骨欠損モデルにおけるrhBMP-2・ $\beta$ -TCP・生体吸収性ポリマーによる骨欠損修復

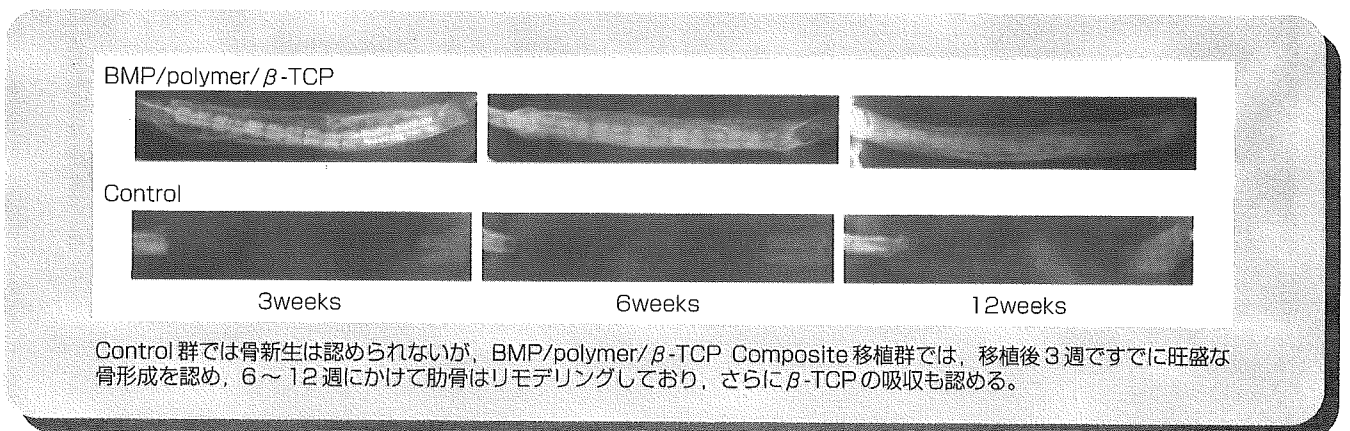


図5 肋骨の再生(イヌ)