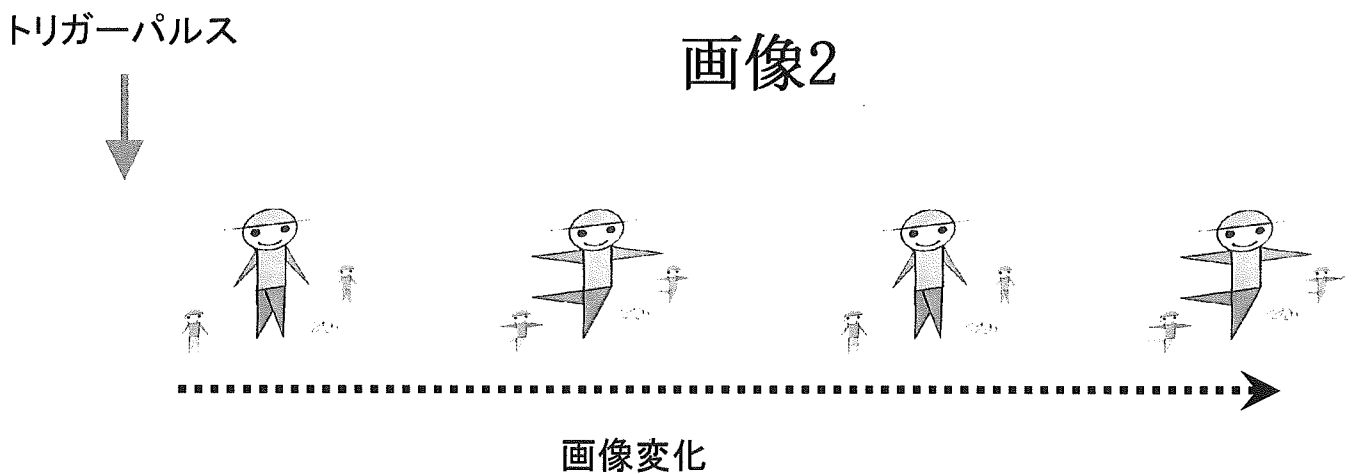
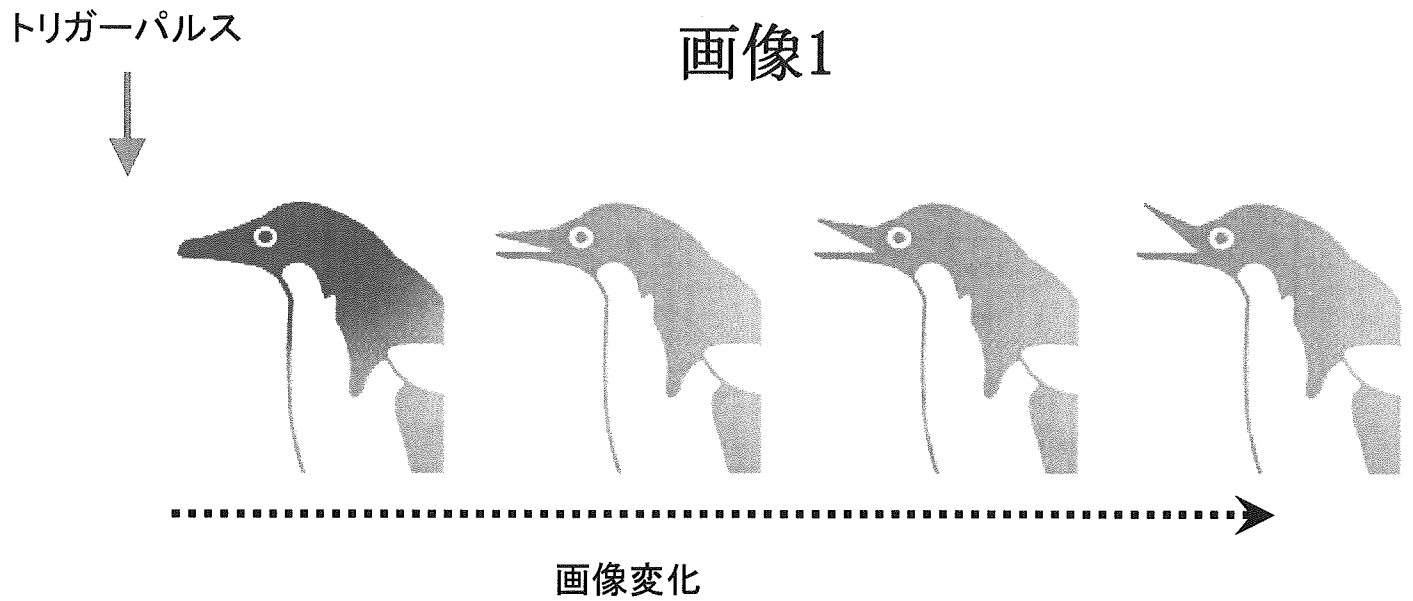


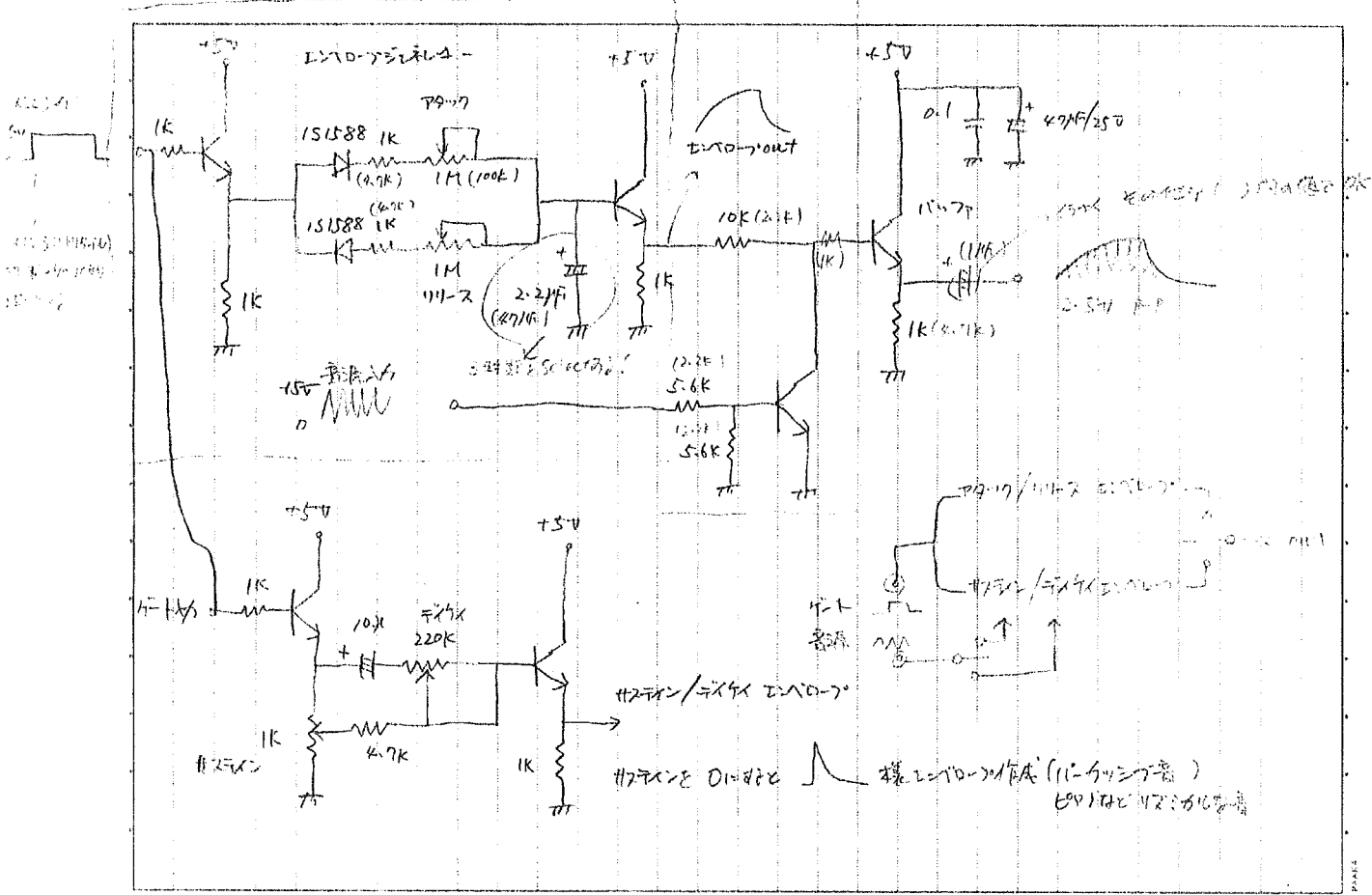
図19 トリガーパルスによる画像変化の例



咀嚼運動により発生したトリガーパルスを、アナログ-デジタル変換インターフェースを用いてデジタル信号化し、パソコンに入力する。
パソコン上で映像を変化させ、これをスクリーンに投影する。

図20 トリガーパルスにより音を発生させるための
アナログシンセサイザー回路図
(新規考案モデル)

ボリオン型 B-type
TRは 2SC1815

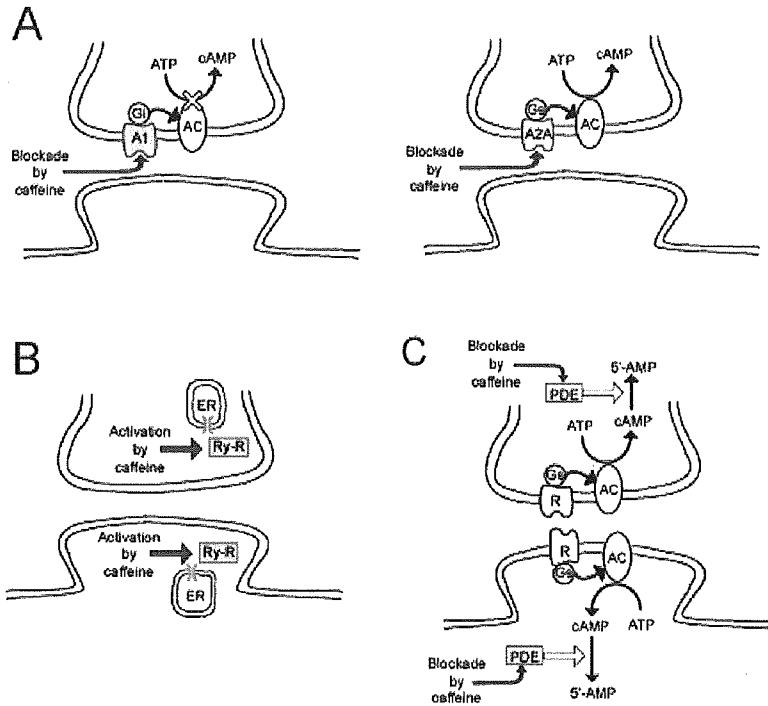


7907/10-2 と #270x/4.5x と 5.5x
 2.2nF/2.2k と 2.2nF/2.2k と 2.2k/2.2k (1.1k) ... LM13700: 2.2k/2.2k

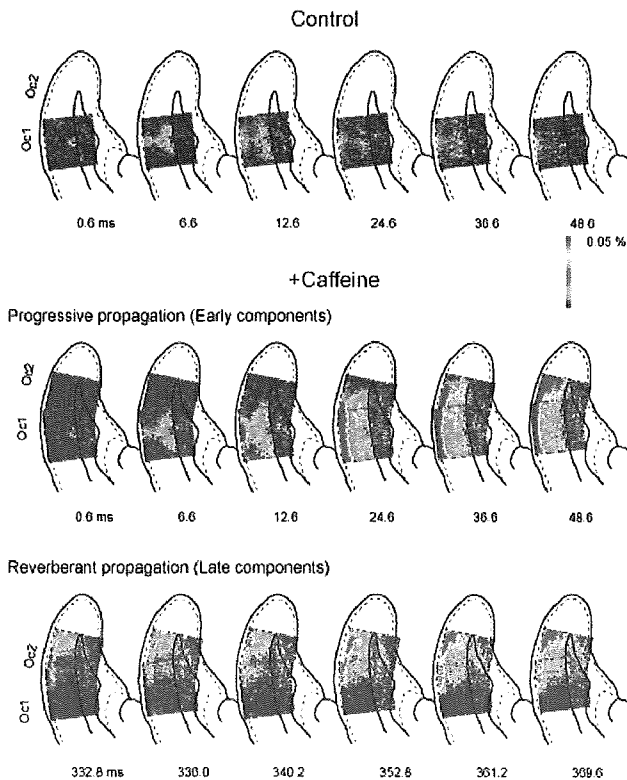
咀嚼運動により発生したトリガーパルスを、アナログ入力し、簡単なVCO-VCF-VCA回路を経て音を作製し、増幅する。低音の聴覚刺激として用いると同時に、ボディーソニックを駆動する周波数としても利用する。

図21

ニューロンのシナプス部位における カフェインの薬理的作用



カフェイン投与による 1次-2次視覚野間の 水平的相互神経連絡の強化



カフェイン投与による 振動性神経活動の誘発

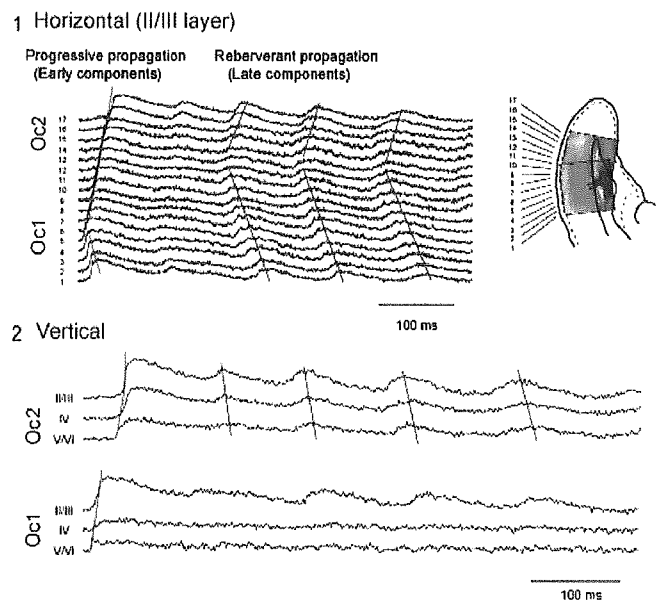
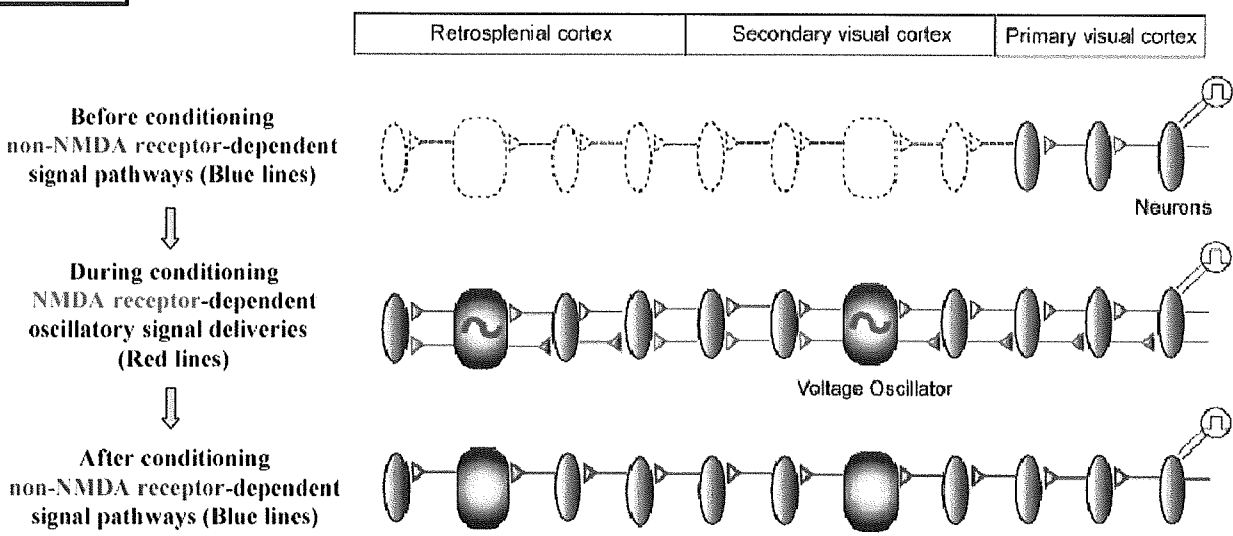


図22

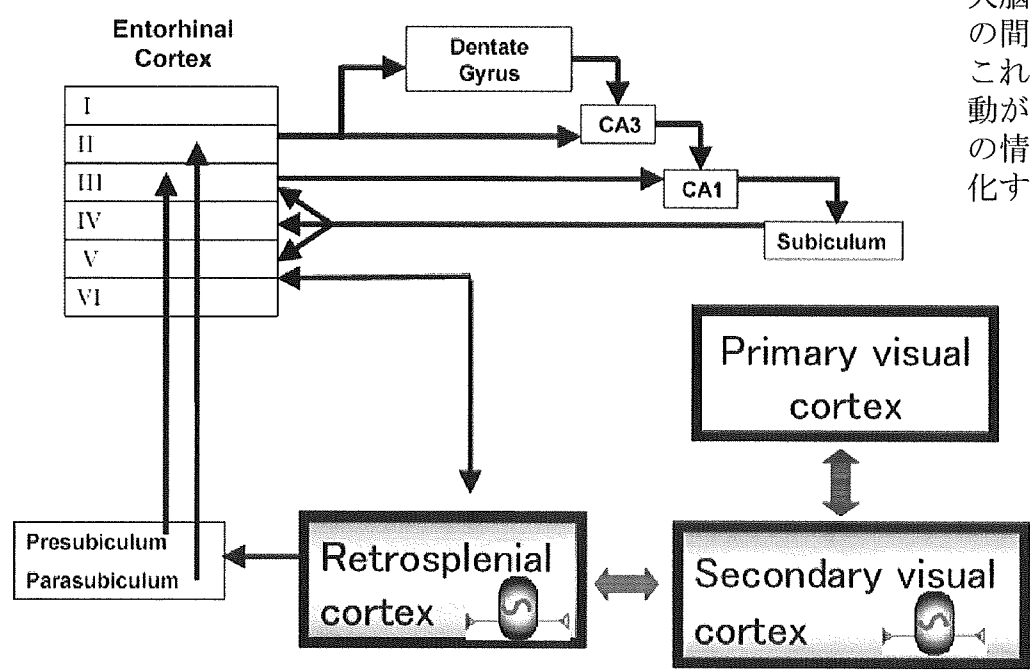
振動装置依存性可塑性仮説



膜電位振動を引き起こす振動源がある間隔で存在し、NMDA受容体依存性の信号を周囲へと配信し、皮質間結合が強化される。

振動装置の配置

Signal flows between visual cortex and hippocampal formation and voltage oscillators



大脳皮質視覚野と海馬の間で振動源が存在し、これらによる振動性活動が大脳皮質-海馬間の情報のやりとりを強化するという仮説。

分担研究報告書

老人性痴呆症・アルツハイマー病の予防および治療を目的とした
中枢機能賦活口腔スプリントの開発

分担研究者 瀬上 夏樹 金沢医科大学・顎口腔機能病態学・教授

当該研究課題のうち、「口腔機能低下と老人性痴呆（認知症）・アルツハイマー病の進行度の評価及びこれらの因果関係の解明」を担当し、調査研究を遂行した。詳細については、総括研究報告書中、上記項目として記載した。

分担研究報告書

老人性痴呆症・アルツハイマー病の予防および治療を目的とした
中枢機能賦活口腔スプリントの開発

分担研究者 長尾 壽和 金沢医科大学・顎口腔機能病態学・助手

当該研究課題のうち、「脳機能評価のための電気生理学的脳活動計測」を担当し、調査研究を遂行した。詳細については、総括研究報告書中、上記項目として記載した。

研究成果の刊行に関する一覧表

雑 誌

発表者氏名	論文タイトル名	発表誌名	巻号	ページ	出版年
Yoshimura, H.	The potential of caffeine for functional modification from cortical synapses to neuron networks in the brain.	Current Neuropharmacology	3	309-316	2005
Yoshimura, H., Honjo, M., Segami, N., Kaneyama, K., Sugai, T., Mashiyama, Y., Onoda, N.	Cyclic AMP-dependent attenuation of oscillatory activity-induced intercortical strengthening of horizontal pathways between insular and parietal cortices.	Brain Research	1069	86-95	2006

The Potential of Caffeine for Functional Modification from Cortical Synapses to Neuron Networks in the Brain

Hiroshi Yoshimura*

Departments of Oral and Maxillofacial Surgery, Kanazawa Medical University, Uchinada-cho 920-0293, Japan

Abstract: Structure and function of the brain are use-dependent variables based on "synapse plasticity". Since synapses are driven by chemical transmitters, synaptic functions are liable to be modified by extrinsic chemicals displaying affinities for synaptic receptors or modulators. Caffeine is a widely used chemical substance that can invade synapses, and has several biochemical and metabolic actions on synaptic activities. This review focuses on the actions of caffeine on changes in structure and function in the region of the hippocampal formation and neocortex, which exhibit high synapse plasticity. At the synapse level, various synaptic receptors and channel activities are modulated by caffeine *via* mobilization of intracellular calcium, inhibition of phosphodiesterase, antagonism of adenosine receptors and GABA receptors. These actions of caffeine enable neurons to induce plastic changes in the properties of synaptic activities, such as synaptic transmission efficiency and morphology. At the network level, caffeine has the ability to activate cortical neural oscillators that deliver repetitive *N*-methyl-D-aspartate receptor-dependent signals to surrounding areas, causing strengthening of long-range inter-cortical communications. Caffeine might thus allow reorganization of cortical network functions *via* synaptic mobilizations.

1. INTRODUCTION

The brain is a complex system for information processing. The intellectual device requires harmonic and coherent action of the component neuron network units, resulting in consistent and intensive operation of the network systems [23,76]. One prominent property of the brain is that structure and function, such as neural wiring and signal communicating efficiency, remain use-dependent and developmentally variable, allowing the brain to acquire the ability to process various modes of information in accordance with changing circumstances [3,9,13,78]. Mechanisms at the synapse level in local dimensions provide this brain variability. Use-dependent induction of synaptic changes is called "synapse plasticity" [11,19,43,52,53,54,59,75]. In general, induction of the synapse plasticity requires repetitive synaptic experiences. Ionotropic or metabotropic receptor activities elicited by synaptic transmission play important roles in the generation of use-dependent synapse plasticity. Production of the electro-motive forces that drive the network systems is triggered at the synapse level. Synapto-motive forces are generated by presynaptic chemical-transmitter release and postsynaptic receptor activities. Interestingly, various natural and synthetic chemicals in the external environment display affinities for synapse receptors and modulators. When these chemicals invade the synaptic cleft and chemical actions are exerted, synaptic functions are liable to be modified.

Among the natural chemicals in the external environment, caffeine is one of the most well-known chemicals able to invade the synaptic cleft. Caffeine displays affinities for several kinds of receptors embedded in the synaptic membranes and internal calcium store, and also has an

affinity for cytoplasmic phosphodiesterases (PDEs), enabling caffeine to modify synaptic activities [31,32,66]. Caffeine thus displays various biochemical and metabolic actions at the synapse level. In general, plastic changes in synaptic transmission efficiency and synaptic architecture are induced according to synaptic activities *via* various kinds of modulation system [6,16,48]. If local synaptic changes are induced systematically and extensively, local changes may develop into network changes. The chemical activity of caffeine might therefore provide the potential for reorganization of brain function from synapse to wide-ranging networks.

Among the various areas of the brain, the hippocampal formation and neocortex exhibit a high susceptibility to the induction of synapse plasticity [11,13,53,75]. The present review focuses attention on these cortical regions, and explores the action of caffeine on plastic changes in structure and function from synapse to cortical network levels.

2. BASIC NEUROPHARMACOLOGICAL EFFECTS OF CAFFEINE

2.1. Effects of Caffeine on Adenosine A1 and A2A Receptors

Purines such as adenosine triphosphate (ATP) and adenosine play central roles in energy metabolism for all cells, and purinergic receptors are located on the cell surface and hence bind purines in the extracellular space [14,31,34]. Interestingly, xanthines such as caffeine block adenosine receptors, but not ATP receptors [20,31]. Adenosine receptors are coupled with G-protein, and can be divided into subtypes A1, A2A, A2B and A3 [20,22,24,31,32,34,69]. Among these subtypes, caffeine blocks A1 receptors that inhibit adenylyl cyclase (AC), in addition to A2A receptors that activate AC [22,24,31,32,34]. In neurons, A1 and A2A receptors are expressed at presynaptic terminals. A1 receptors negatively influence transmitter release from presynaptic terminals, whereas A2A receptors positively influence transmitter release [32]. While A1 receptors are widely distributed

*Address correspondence to this author at Department of Oral and Maxillofacial Surgery, Kanazawa Medical University, Uchinada-cho 920-0293, Japan; Tel: +81-76-286-2211; Fax: +81-076-286-2010; E-mail: hyoshimu@kanazawa-med.ac.jp

throughout the brain, A1 receptors are expressed at the highest level in the hippocampus and neocortex, where glutamate is used as an excitatory transmitter. Conversely, A2A receptors are not distributed widely, but are distributed locally at the highest level in the striatum and nucleus accumbens [22,24,32]. A2A receptors are expressed in dopamine-rich regions, and are co-expressed with dopamine D2 receptors [28,29,35,49,80]. A2A receptors are thus dominantly linked with the dopaminergic system, whereas A1 receptors in the hippocampus and neocortex are dominantly linked with the glutamatergic system. In addition, in hippocampal CA3 neurons, A1 receptor-selective blockade induces bursting activities, but not A2A receptor-selective blockade [82]. Caffeine is therefore considered to act predominantly on A1 receptors in the cortical regions, and positively influence presynaptic transmitter release *via* blockade of A1 receptors.

2.2. Effects of Caffeine on PDEs

The cyclic AMP (cAMP) cascade is one of the most important intracellular signaling pathways, playing a key role in the expression and modulation of neural function in the central nervous system (CNS) [8]. Activation of membrane receptors coupled to a specific G protein, Gs, such as β -adrenergic receptors or specific metabotropic glutamate (mGlu) receptors, initiates the operation of membrane-bound AC and production of cAMP as a second messenger. Protein phosphorylation or gene expression is finally induced by way of cAMP-dependent protein kinase (PKA) or cAMP response element-binding (CREB) proteins [18,55,85]. These cAMP cascades are negatively controlled by PDEs that breakdown cAMP and turn off the cAMP signaling pathways [4,30,79]. Caffeine depresses PDE activity, and intracellular cAMP is accumulated, resulting in the enhancement of cAMP signaling pathways [5,15,32].

2.3. Effects of Caffeine on Ryanodine Receptors

Calcium signaling pathways play an important role in regulating various brain functions [7]. In particular, increases in cytoplasmic calcium triggers down-stream of the intracellular calcium-dependent cascades. There are extra- and intracellular sources of calcium. Neurons include endoplasmic reticulum (ER) to store high concentrations of calcium. Calcium-release channels called ryanodine receptors are expressed in the membrane of the ER. When extracellular calcium enters the endoplasm through voltage- or receptor-operated calcium channels, ryanodine receptor channels are opened by the binding of calcium with the ryanodine receptors, and calcium is then released from the calcium store into the cytoplasm as a calcium-induced calcium release (CICR) [7,26]. Concentrations of endoplasmic calcium are thus amplified, and intracellular calcium signaling pathways are activated in a feed-forward manner. Caffeine permeating into the cell through the cell membrane combines with ryanodine receptors. This results in activation of the ryanodine receptors, reducing the threshold of the CICR and resulting in intense facilitation of CICR [25,32,36,61]. In rat hippocampal CA3 neurons, caffeine promotes epileptic discharges *via* enhancement of the CICR [56], and caffeine enhances action potential-triggered CICR in rat hippocampal CA1 neurons [71]. Amplification of intracellular calcium is

thus positively controlled by caffeine through ryanodine receptors.

2.4. Effects of Caffeine on GABA Receptors

Neuron network activities are based on excitatory and inhibitory synaptic activities. The GABAergic network plays important roles in the stabilization of overall network activities. A recent study revealed that caffeine can modulate the GABAergic system. In ganglion cells of the turtle retina, caffeine depresses the activities of GABA-A receptors. This depression is mediated by caffeine facilitating CICR [1]. Similarly, in dentate gyrus cells of the hippocampus, CICR elicited by caffeine depresses the activities of GABA-A receptors [21]. In neonatal hippocampal neurons, mobilization of Ca^{2+} from caffeine-ryanodine-sensitive stores facilitates GABA release, while caffeine simultaneously depresses the activities of postsynaptic GABA-A receptors [72]. Caffeine thus affects GABA-A receptor activities by way of facilitating CICR. Conversely, although the mechanisms remain unclear, Ca^{2+} -independent inhibition of GABA-A receptor activities by caffeine occurs in hippocampal neurons [81].

3. NEUROPHARMACOLOGICAL EFFECTS OF CAFFEINE AT CORTICAL SYNAPSES

At the synapses, local synaptic potentials are generated by synaptic inputs. As local synaptic potentials are summated spatio-temporally, neurons have the ability to integrate various input signals. In addition, synapses can display changes in the efficiency of synaptic transmissions and induce morphological changes according to activity. The basic targets of caffeine mentioned above are concentrated at the synapses (Fig. 1). Synapses are therefore considered to represent the dominant targets of caffeine.

3.1. Effects of Caffeine on Presynaptic Sites

Release of excitatory transmitter is more strongly inhibited by adenosine than release of inhibitory neurotransmitters [33]. Blockade of adenosine receptors by caffeine can thus occasionally generate overactivity at excitatory synapses [2,47]. In hippocampal CA3 in guinea pigs, blockade of A1 receptors by caffeine generates paroxysmal depolarizing shifts, and the underlying mechanisms may be increased by intracellular cAMP and Ca^{2+} influxes [62,63]. In hippocampal CA1 neurons, caffeine enhances excitatory postsynaptic potentials (EPSPs), which are mediated by antagonism of presynaptic adenosine receptors [37].

In both glutamatergic and cholinergic neurons, caffeine affects presynaptic sites. In rat hippocampal neurons, caffeine enhances acetylcholine (ACh) release from presynaptic terminals *via* blockade of A1 receptors [17].

Changes in the probability of transmitter release induced by caffeine have been investigated by focusing miniature excitatory postsynaptic currents (EPSCs) [74]. The study proposed that in rat barrel cortex, caffeine enhances glutamate release from presynaptic terminals *via* calcium release from ryanodine-sensitive internal Ca^{2+} stores.

3.2. Effects of Caffeine on Postsynaptic Sites

Postsynaptic activities can be divided into two categories: direct synaptic transmission, and indirect synaptic trans-

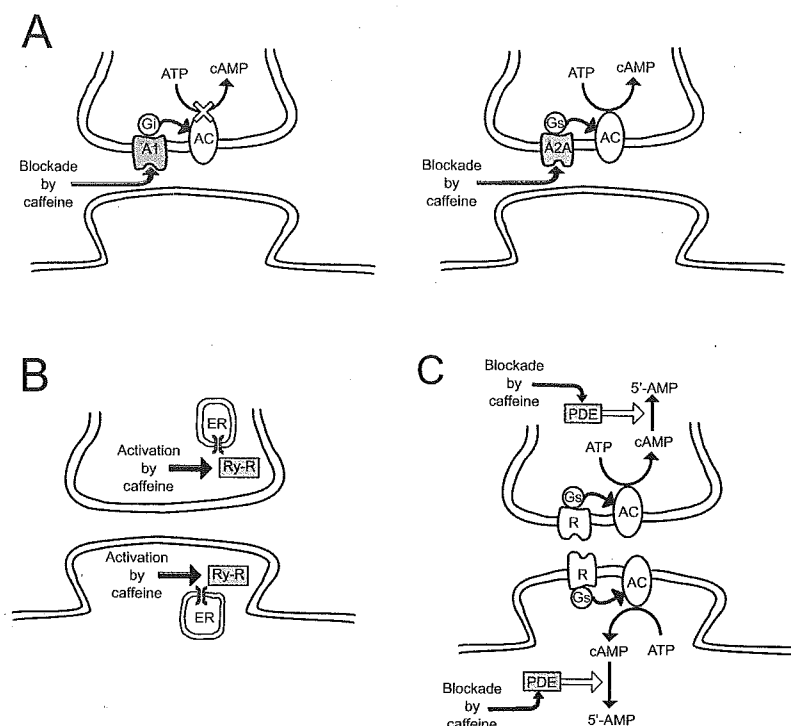


Fig. (1). Basic actions of caffeine at synapses.

(A) Presynaptic A1 receptors inhibit adenylyl cyclase (AC), resulting in decreased levels of intracellular cAMP, whereas presynaptic A2A receptors stimulate AC, increasing intracellular cAMP levels. Caffeine blocks both A1 and A2 receptors, resulting in increased and decreased intracellular cAMP levels, respectively. (B) Intracellular calcium stores are present for both pre- and postsynaptic sites. Ryanodine-sensitive calcium release channels are expressed on the calcium stores. Caffeine activates ryanodine receptor channels, reducing the threshold for calcium-induced calcium-release (CICR). (C) Intracellular phosphodiesterases (PDEs) breakdown intracellular cAMP at both pre- and postsynaptic sites. Caffeine blocks PDEs, increasing intracellular cAMP levels.

mission. Direct synaptic transmission is mediated by ligand-gated ionic channel-coupled receptors. Electro-motive force at the synapse is produced by ligand-gated ionic channel-coupled receptors. In general, two types of ionotropic glutamate receptors produce EPSPs: *N*-methyl-D-aspartate (NMDA) receptors, and non-NMDA receptors. NMDA receptors are postsynaptic activity-dependent calcium permeable channels, and play a central role in the induction of synapse plasticity, as mentioned below.

In rat hippocampal CA3 neurons, caffeine enhances both NMDA and non-NMDA receptor activities, inducing high-frequency oscillations [39]. In rat visual and parietal cortices, caffeine also enhances both NMDA and non-NMDA receptor activities, inducing α -range oscillations [89,90,92,93]. Caffeine-enhanced synaptic activities are triggered by activation of both receptors, in turn causing enhancement of the receptor activities themselves. Since adequate repetitive synaptic inputs are required for caffeine-dependent enhancement of synaptic activities, this enhancement is considered use-dependent.

Indirect synaptic transmission is mediated by G-protein-coupled metabotropic receptors. Expressed at both pre- and postsynaptic sites, mGlu receptors are involved in modulation of synaptic activities by activation of a second messenger system. Generation of a transient increase in intracellular

Ca^{2+} to switch on the Ca^{2+} -signaling second messenger pathway is one of the principal roles of mGlu receptors [68]. In hippocampal CA1 neurons, the Ca^{2+} transient induced by postsynaptic mGlu receptor activation is blocked by caffeine [10]. Caffeine thus acts on mGlu receptor activities at postsynaptic sites.

GABA receptors are concerned with generation of inhibitory postsynaptic potentials, so GABA receptors negatively influence synaptic activities. In hippocampal dentate gyrus neurons, elevation of intracellular calcium levels by caffeine depresses postsynaptic ionotropic GABA-A receptor activities, showing that depression of GABA-A receptors by caffeine is dependent on intracellular calcium elevation [21, 72]. In contrast, caffeine depresses GABA-A receptor activities in hippocampal CA3 neurons, which are independent of intracellular calcium elevations [81].

3.3. Effects of Caffeine on Long-Term Potentiation and Long-Term Depression

At many cortical synapses, repetitive synaptic activities can produce long-term changes in synaptic efficiency [11,12, 59]. According to the patterns of temporal coincidence, location and intensity of pre- and postsynaptic activities, synaptic efficiency is potentiated or depressed over the long term, and termed long-term potentiation (LTP) or long-

term depression (LTD) respectively. Various kinds of LTP, LTD and associated mechanisms have been investigated and summarized in many reviews [43,48,52,53,54,59,75]. Considering the mechanisms of LTP and LTD, whether LTP and LTD are NMDA receptor-dependent represents a central issue, since induction of activity-dependent synapse plasticity is deeply affected by NMDA receptor activities [16,43,50,73,75]. Activities of postsynaptic NMDA receptors are blocked by extracellular Mg^{2+} , and reduction of this Mg^{2+} block requires postsynaptic depolarization, allowing NMDA receptors to function as important detectors of coincident pre- and postsynaptic activities [60,67]. The coincidence between pre- and postsynaptic activities is deeply involved in the induction of synapse plasticity [40].

In general, LTP in hippocampal CA1 neurons requires both postsynaptic NMDA receptors and increased levels of intracellular Ca^{2+} by way of NMDA receptors. In contrast, caffeine induces another form of LTP in hippocampal CA1 neurons. Caffeine-dependent LTP requires neither postsynaptic NMDA receptors nor increased intracellular Ca^{2+} by way of NMDA receptors, but does require the interaction of caffeine with P1 adenosine receptors, P2 purinergic receptors and ryanodine receptors, indicating that caffeine-dependent CA1 LTP is caused by increases in presynaptic transmitter release [57,58]. Another presynaptically induced caffeine-dependent LTP in hippocampal CA1 has been reported. In rat hippocampal CA1 neurons, caffeine promotes forskolin-induced LTP, where adenosine A1 receptor antagonism underlies the effects of caffeine [51]. Caffeine thus increases susceptibility to the induction of cAMP-dependent LTP, *via* enhancement of presynaptic cAMP accumulation. Actions of caffeine at presynaptic sites may be sufficient to induce such cAMP-dependent NMDA receptor-independent LTP.

As for LTD, postsynaptically induced caffeine-dependent LTD has been reported. Caffeine-dependent LTD in hippocampal CA neurons is postsynaptically induced in a stimulation frequency-dependent manner. LTD requires both NMDA receptor activities and calcium release from internal calcium store [65].

3.4. Effects of Caffeine on Morphological Changes in Synapses

The morphology of dendritic spines exerts a substantial effect on important aspects of synaptic activities, such as synaptic transmission and integration of synaptic information [41,73,77,87,95,96]. In dendritic spines, calcium dynamics play an important role in the expression of those synaptic functions, by way of various calcium-dependent biochemical processes. Particularly in hippocampal CA1 neurons, individual spines play an important role in detecting temporal coincidence between pre- and postsynaptic activities *via* NMDA receptors [96].

In cultured hippocampal neurons, application of caffeine causes a transient rise in intracellular calcium levels *via* ryanodine receptors in dendrites and spines, resulting in increased size of excitatory dendritic spines and changes in spine shapes [44,45]. Calcium from internal stores elicited by caffeine can thus modify dendritic spine shape [38]. Dynamics of intracellular calcium increases differently between short- and long-neck dendritic spines, suggesting

that control of spatio-temporal calcium increases is provided by the shape of dendritic spines [84]. Since changes in spine structure contribute to changes in brain function [64,95], caffeine might modulate brain function *via* increases in intracellular Ca^{2+} level [38].

4. NEUROPHARMACOLOGICAL EFFECTS OF CAFFEINE ON CORTICO-CORTICAL SIGNAL INTERACTIONS

Local synaptic changes may in turn induce reorganization of cortical network function. However, this might require strong and long-range synchronization of synaptic activities or firing between neuron clusters [27,46,70,76]. In this respect, a strong relationship may exist between neural oscillations and synapse plasticity [27,83]. To understand the mechanisms of neural oscillation, several network oscillation models have been proposed [27]. Theoretically, synchronization as a non-local event is convenient for induction of synapse plasticity between long-range discrete cortical areas. Even if synchronization is a local event, however, synapse plasticity is inducible between long-range discrete areas, on the condition that the propagating system is strong and stable.

Recently, a protocol for inducing synchronized membrane potential oscillation at a frequency of 8-10 Hz in the visual cortex has been developed, by applying caffeine to rat brain slices [90,92,93]. The start of oscillation requires a trigger input, and oscillation comprises several propagating wavelets. Oscillation induction requires low-frequency activation of input fibers in conjunction with caffeine application, suggesting that use-dependent mechanisms underlie oscillation induction. Notably, induction of oscillation requires both NMDA receptor activation and the release of intracellular calcium from the internal calcium store, suggesting that functional coupling between NMDA and ryanodine receptors underlies caffeine-dependent oscillation [92,93]. In the absence of caffeine, the strength of functional coupling between NMDA and ryanodine receptors in hippocampal neurons depends on the magnitude of NMDA receptor activation [42]. In the presence of caffeine, caffeine activates ryanodine receptors and potentiates presynaptic glutamate release, resulting in an increased likelihood of functional coupling between NMDA and ryanodine receptors.

Strictly speaking, caffeine-dependent oscillation comprises initial propagating components and subsequent oscillatory components. These subsequent oscillatory components emerge from the local area in the visual cortex, showing that the oscillator is localized. Although synchrony is a local event, the neural oscillator delivers NMDA receptor-dependent signals to the surrounding areas [90,94]. These signal deliveries finally cause strengthening of non-NMDA receptor-dependent inter-cortical connections between long-range discrete areas [94]. The oscillators are separately located in the medial and lateral secondary visual cortices. Horizontal connections in layer II/III between the primary and secondary visual cortices are strengthened after repetitive NMDA receptor-dependent signal delivery originating from the oscillators.

Another study revealed that the oscillator is also located outside the visual cortex. The retrosplenial cortex is located

at a critical position between the visual cortex and hippocampal formation. In the area of the retrosplenial cortex, the oscillator is present in the retrosplenial granular cortex (RSGa). Activation of oscillators in both the secondary visual cortex and RSGa under application of caffeine finally opens functional connections from primary visual cortex to the postsubiculum [91]. Hence, in the presence of caffeine, an oscillator with local synchronization can induce spatially wide-ranging synapse plasticity from the visual cortex to the hippocampal formation.

These studies resulted in the “oscillator-dependent plasticity hypothesis” [90,91,94]. This hypothesis is illustrated in Fig. 2, showing the induction of plastic changes in the visual cortex in the presence of caffeine. Caffeine, in combination with low-frequency electrical stimulation, promotes the voltage oscillator delivering NMDA receptor-dependent signals at a frequency of 8-10 Hz from the secondary visual cortex to surrounding cortical areas. This induces opening and strengthening of non-NMDA receptor-dependent signal pathways. Repetitive activities of an NMDA receptor-dependent voltage oscillator thus induce use-dependent network plasticity in the cortical regions.

The same mechanism is present between the gustatory insular cortex and somatosensory parietal cortex. In these areas, the oscillator that delivers NMDA receptor-dependent signals is located in the parietal cortex, and is driven under application of caffeine by repetitive low-frequency stimulation. Oscillatory NMDA receptor-dependent signal delivery causes strengthening of functional connections between the insular and parietal cortices [88,89].

Theoretical investigation has demonstrated that oscillation-dependent mechanisms underlie the establishment of working memory. The study showed that NMDA receptor-mediated synaptic transmission at a frequency of 8 Hz is required to sustain persistent network activities of the prefrontal cortex [86]. Results collected by experimental and theoretical studies thus suggest that NMDA receptor-mediated α -range signal delivery plays a critical role in the generation and stabilization of functional networks *via* plastic changes from synapses to networks.

5. CONCLUSION

Caffeine displays various general pharmacological actions: 1) blockade of presynaptic A1 and A2A receptors, resulting in modulation of transmitter release; 2) activation of internal ryanodine receptors, resulting in reduction of CICR threshold; 3) blockade of PDEs, resulting in intracellular cAMP accumulation; and 4) blockade of GABA-A receptors, resulting in depression of inhibitory synaptic activities. In the brain, these general actions of caffeine are liable to take place at the synapses, as the targets of caffeine and its effects are concentrated at the synapses. Particularly in the regions of the hippocampal formation and neocortex, where use-dependent synapse plasticity is liable to be established, caffeine is able to enhance synaptic NMDA receptor activities and intracellular calcium signaling pathways, through which plastic changes in synaptic morphology and transmission efficiency are induced. In cortical neuron networks, the actions of caffeine in combination with adequate input fiber activation produce opening and strengthening of long-range inter-cortical signal

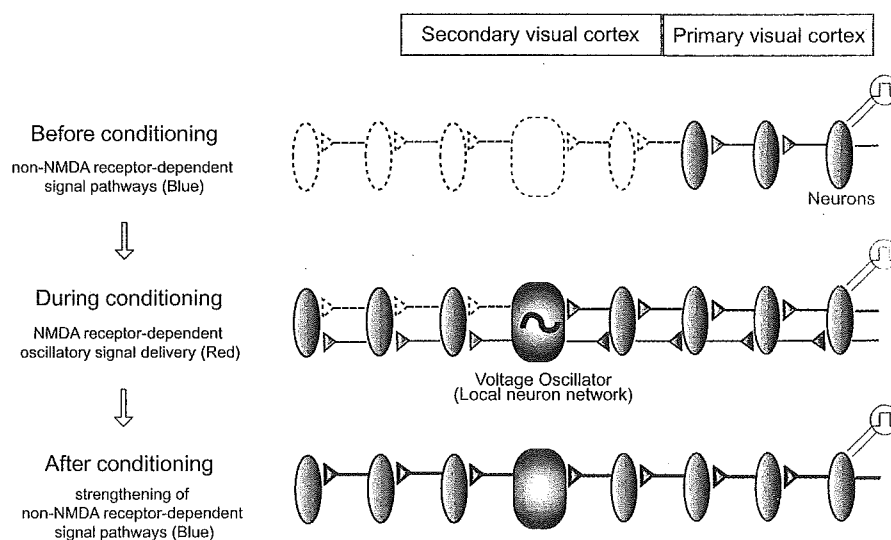


Fig. (2). Oscillator-dependent plasticity hypothesis.

Each ellipse indicates a postsynaptic neuron or neuron cluster, and small triangles indicate presynaptic terminals. In the presence of caffeine, low-frequency electrical stimulation is applied to the primary visual cortex as “conditioning”. Horizontal pathways in blue represent non-NMDA receptor-dependent pathways. A voltage oscillator comprising local neuron networks is equipped in the secondary visual cortex, and horizontal pathways in red represent *N*-methyl-D-aspartate (NMDA) receptor-dependent pathways. (Upper) Before conditioning, signals elicited by the primary visual cortex stimulation propagate within a short distance. (Middle) During conditioning, non-NMDA receptor-dependent signals switch on the oscillator that delivers NMDA receptor-dependent oscillatory signals back and forth. (Lower) After conditioning, non-NMDA receptor-dependent pathways are strengthened, and signals propagate a long distance. Note that when NMDA receptor activities are blocked from the beginning, strengthening of non-NMDA receptor-dependent pathways is not induced [90,91,94].

communications *via* activation of cortical neural oscillators that deliver NMDA receptor-dependent signals to surrounding areas. The actions of caffeine at a synapse level thus cause plastic changes at the cortical network level.

Most of the experimental evidence has been collected from basic studies using peculiar conditions *in vitro*. However, these basic studies have elicited the potential of caffeine, and the evidence indicates that caffeine exerts profound actions from synapse to neuron networks in the cortical regions. Caffeine might thus provide the potential for use-dependent reorganization of brain function.

ACKNOWLEDGEMENTS

We wish to thank Drs. N. Onoda and N. Segami for providing valuable advice on the manuscript. This work was supported by grants from the Ministry of Health, Labor and Welfare of Japan (Comprehensive Research on Aging and Health, H-17-Chouju-018).

REFERENCES

- [1] Akopian, A., Gabriel, R., Witkovsky, P. (1998) Calcium released from intracellular stores inhibits GABAA-mediated currents in ganglion cells of the turtle retina. *J. Neurophysiol.*, **80**, 1115-1115.
- [2] Ault, B., Olney, M.A., Joyner, J.L., Boyer, C.E., Notrica, M.A., Soroko, F.E., Wang, C.M. (1987) Pro-convulsant actions of theophylline and caffeine in the hippocampus: implications for the management of temporal lobe epilepsy. *Brain Res.*, **426**, 93-102.
- [3] Bailey, C.H., Kandel, E.R. (1993) Structural changes accompanying memory storage. *Ann. Rev. Physiol.*, **55**, 397-426.
- [4] Beavo, J.A. (1995) Cyclic nucleotide phosphodiesterases: functional implications of multiple isoforms. *Physiol. Rev.*, **75**, 725-748.
- [5] Beavo, J.A., Reifsnnyder, D.H. (1990) Primary sequence of cyclic nucleotide phosphodiesterase isozymes and the design of selective inhibitors. *Trends Pharmacol. Sci.*, **11**, 150-155.
- [6] Berardi, N., Pizzorusso, T., Ratto, G.M., Maffei, L. (2003) Molecular basis of plasticity in the visual cortex. *Trends Neurosci.*, **26**, 369-378.
- [7] Berridge, M.J. (1998) Neuronal calcium signaling. *Neuron*, **21**, 13-26.
- [8] Bhalla, U.S., Iyengar, R. (1999) Emergent properties of networks of biological signaling pathways. *Science*, **283**, 381-387.
- [9] Bi, G., Poo, M. (2001) Synaptic modification by correlated activity: Hebb's postulate revisited. *Ann. Rev. Neurosci.*, **24**, 139-166.
- [10] Bianchi, R., Young, S.R., Wong, R.K.S. (1999) Group I mGluR activation causes voltage-dependent and -independent Ca^{2+} rise in hippocampal pyramidal cells. *J. Neurophysiol.*, **81**, 2903-2913.
- [11] Bliss, T.V., Collingridge, G.L. (1993) A synaptic model of memory: long-term potentiation in the hippocampus. *Nature*, **361**, 31-39.
- [12] Bliss, T.V.P., Lomo, T. (1973) Long-lasting potentiation of synaptic transmission in the dentate of anesthetized rabbit following stimulation of the perforant path. *J. Physiol.*, **232**, 331-356.
- [13] Buonomano, D.V., Merzenich M.M. (1998) Cortical plasticity: From Synapse to maps. *Ann. Rev. Neurosci.*, **21**, 149-186.
- [14] Burnstock, G. (1990) Overview. Purinergic mechanisms. *Annals N.Y. Acad. Sci.*, **603**, 1-17.
- [15] Butcher, R.W., Sutherland, E.W. (1962) Adenosine-3', 5'-phosphate in biological materials. *J. Biol. Chem.*, **273**, 1244-1250.
- [16] Carroll, R.C., Zukin R.S. (2002) NMDA-receptor trafficking and targeting: implications for synaptic transmission and plasticity. *Trends Neurosci.*, **25**, 571-577.
- [17] Carter, A.J., O'Connor, W.T., Carter, M.J., Ungerstedt, U. (1995) Caffeine enhances acetylcholine release in the hippocampus *in vitro* by a selective interaction with adenosine A1 receptors. *J. Pharmacol. Exp. Therapeut.*, **273**, 637-642.
- [18] Cho, K., Brown, M.W., Bashir, Z.I. (2002) Mechanisms and physiological roles of enhancement of mGlu5 receptor function by group II mGlu receptor activation in rat perirhinal cortex. *J. Physiol.*, **540**, 895-906.
- [19] Compte, A., Brunel, N., Goldman-Rakic, P., Wang, X.J. (2000) Synaptic mechanisms and network dynamics underlying spatial working memory in a cortical network model. *Cereb. cortex*, **10**, 910-923.
- [20] Daly, J.W., Butts-Lamb, P., Padgett, W. (1983) Subclasses of adenosine receptors in the central nervous system: interaction with caffeine and related methylxanthines. *Cell. Mol. Neurobiol.*, **3**, 69-80.
- [21] De Koninck, Y., Mody, I. (1996) The effects of rising intracellular calcium on synaptic GABAA receptor-channels. *Neuropharmacology*, **35**, 1365-1374.
- [22] Dixon, A.K., Gubitz, A.K., Sirinathsinghji, D.J., Richardson, P.J., Freeman, T.C. (1996) Tissue distribution of adenosine receptor mRNAs in the rat. *Br. J. Pharmacol.*, **118**, 1461-1468.
- [23] Douglas, R.J., Martin K.A. (2004) Neuronal circuits of the neocortex. *Ann. Rev. Neurosci.*, **27**, 419-451.
- [24] Dunwiddie, T.V., Masino, S.A. (2001) The role and regulation of adenosine in the central nervous system. *Ann. Rev. Neurosci.*, **24**, 31-55.
- [25] Edwards, J.A., Cline, H.T. (1999) Light-induced calcium influx into retinal axons is regulated by presynaptic nicotinic acetylcholine receptor activity *in vivo*. *J. Neurophysiol.*, **81**, 895-907.
- [26] Ehrlich, B.E. (1995) Functional properties of intracellular calcium-release channels. *Curr. Opin. Neurobiol.*, **5**, 304-349.
- [27] Ermentrout, G.B., Kleinfeld, D. (2001) Traveling electrical waves in cortex: insight from phase dynamics and speculation on a computational role. *Neuron*, **29**, 33-44.
- [28] Ferre, S., Fredholm, B.B., Morelli, M., Popoli, P., Fuxe, K. (1997) Adenosine-dopamine receptor-receptor integrations as an integrative mechanism in the basal ganglia. *Trends Neurosci.*, **20**, 482-487.
- [29] Fink, J.S., Weaver, D.R., Rivkees, S.A., Peterfreund, R.A., Pollack, A.E., Adler, E.M., Reppert, S.M. (1992) Molecular cloning of the rat A2 adenosine receptor: selective co-expression with D2 dopamine receptors in rat striatum. *Brain Res. Mol. Brain Res.*, **14**, 186-195.
- [30] Francis, S.H., Turko, I.V., Corbin, J.D. (2001) Cyclic nucleotide phosphodiesterases: relating structure and function. *Prog. Nucl. Acid Res. Mol. Biol.*, **65**, 1-52.
- [31] Fredholm, B.B., Abbracchio, M.P., Burnstock, G., Daly, G.W., Harden, T.K., Jacobson, K.A., Leff, P., Williams, M. (1994) Nomenclature and classification of purinoceptors. *Pharmacological Rev.*, **46**, 143-156.
- [32] Fredholm, B.B., Bättig, K., Holmén, J., Nehlig, A., Zuvartau, E.E. (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol. Rev.*, **51**, 83-133.
- [33] Fredholm, B.B., Dunwiddie, T.V. (1988) How does adenosine inhibit transmitter release? *Trends Pharmacol. Sci.*, **9**, 130-134.
- [34] Fredholm, B.B., Hedqvist, P. (1980) Modulation of neurotransmission by purine nucleotides and nucleosides. *Biochem. Pharmacol.*, **29**, 1635-1643.
- [35] Fuxe, K., Ferre, S., Zoli, M., Agnati, L.F. (1998) Integrated events in central dopamine transmission as analyzed at multiple levels. Evidence for intramembrane adenosine A2A/dopamine D2 and adenosine A1/dopamine D1 receptor interactions in the basal ganglia. *Brain Res. Brain Res. Rev.*, **26**, 258-273.
- [36] Garaschuk, O., Yarri, Y., Konnerth, A. (1997) Release and sequestration of calcium by ryanodine-sensitive stores in rat hippocampal neurones. *J. Physiol.*, **502**, 13-30.
- [37] Greene, R.W., Hass, H.L., Hermann, A. (1985) Effects of caffeine on hippocampal pyramidal cells *in vitro*. *Br. J. Pharmacol.*, **85**, 163-169.
- [38] Harris, K. (1999) Calcium from internal stores modifies dendritic spine shape. *Proc. Natl. Acad. Sci. USA*, **96**, 12213-12215.
- [39] He, S. J., Xioa, C., Ruan, D. Y. (2004) Caffeine-dependent stimulus-triggered oscillations in the CA3 region of hippocampal slices from rats chronically exposed to lead. *Exp. Neurol.*, **190**, 525-534.
- [40] Hebb, D.O. (1949) *The Organization of behavior*. New York: Wiley.

- [41] Hering, H., Sheng, M. (2001) Dendritic spines: structure, dynamics and regulation. *Nat. Rev. Neurosci.*, **2**, 880-888.
- [42] Isokawa, M. (2005) *N*-methyl-D-aspartic acid-induced and Ca-dependent neuronal swelling and its retardation by grain-derived neurotrophic factor in the epileptic hippocampus. *Neuroscience*, **131**, 801-812.
- [43] Katz, L.C., Shatz, C.J. (1996) Synaptic activity and the construction of cortical circuits. *Science*, **274**, 1133-1138.
- [44] Korkotian, E., Segal, M. (1998) Fast confocal imaging of calcium released from stores in dendritic spines. *Eur. J. Neurosci.*, **10**, 2076-2084.
- [45] Korkotian, E., Segal, M. (1999) Release of calcium from stores alters the morphology of dendritic spines in cultured hippocampal neurons. *Proc. Natl. Acad. Sci. USA*, **96**, 12068-12072.
- [46] König, P., Engel, A. K., Singer, W. (1995) Relation between oscillatory activity and long-range synchronization in cat visual cortex. *Proc. Natl. Acad. Sci. USA*, **92**, 290-294.
- [47] Koryntova, H., Kubova, H., Tutka, P., Mares, P. (2002) Changes of cortical epileptic after discharges under the influence of convulsant drugs. *Brain Res. Bulletin*, **58**, 49-54.
- [48] Lamprecht, R., LeDoux, J. (2004) Structural plasticity and memory. *Nat. Rev. Neurosci.*, **5**, 45-54.
- [49] Le Moine, C., Svenningsson, P., Fredholm, B.B., Bloch, B. (1997) Dopamine-adenosine interactions in the striatum and the globus pallidus: inhibition of striatopallidal neurons through either D2 or A2A receptors enhances D1 receptor-mediated effects on c-fos expression. *J. Neurosci.*, **17**, 8038-8048.
- [50] Linden, D.J., Conner, J.A. (1995) Long-term synaptic depression. *Ann. Rev. Neurosci.*, **18**, 319-357.
- [51] Lu, K. T., Wu, S. P., Gean, P. W. (1999) Promotion of forskolin-induced long-term potentiation of synaptic transmission by caffeine in area CA1 of the rat hippocampus. *Chin. J. Physiol.*, **42**, 249-253.
- [52] Luscher, C., Nicoll, R.A., Malenka, R.C., Muller, D. (2000) Synaptic plasticity and dynamic modulation of the postsynaptic membrane. *Nat. Neurosci.*, **3**, 545-550.
- [53] Malenka, R.C., Nicoll, R.A. (1999) Long-term potentiation—a decade of progress? *Science*, **285**, 1870-1874.
- [54] Malinow, R., Malenka, R.C. (2002) AMPA receptor trafficking and synaptic plasticity. *Ann. Rev. Neurosci.*, **25**, 103-126.
- [55] Mao, L., Wang, J.Q. (2002) Glutamate cascade to cAMP response element-binding protein phosphorylation in cultured striatal neurons through calcium-coupled group I metabotropic glutamate receptors. *Mol. Pharmacol.*, **62**, 473-484.
- [56] Margineanu, D.G., Klitgaard, H. (2004) Caffeine-induced epileptiform field potentials in rat hippocampal slices: a pharmacological characterization. *Neuropharmacology*, **47**, 926-934.
- [57] Martín, E. D., Buño, W. (2003) Caffeine-mediated presynaptic long-term potentiation in hippocampal CA1 pyramidal neurons. *J. Neurophysiol.*, **89**, 3029-3038.
- [58] Martín, E. D., Buño, W. (2005) Stabilization effects of extracellular ATP on synaptic efficiency and plasticity in hippocampal pyramidal neurons. *Eur. J. Neurosci.*, **21**, 936-944. not yet in [ref].
- [59] Martin, S.J., Grimwood, P.D., Morris, R.G.M. (2000) Synaptic plasticity and memory: An elevation of the hypothesis. *Ann. Rev. Neurosci.*, **23**, 649-711.
- [60] Mayer, M.L., Westbrook, G.L., Guthrie, P.B. (1984) Voltage-dependent block by Mg^{2+} of NMDA responses in spinal cord neurones. *Nature*, **309**, 261-263.
- [61] McPherson, P.S., Kim, Y.K., Valdivia, H., Knudson, C.M., Takekura, H., Franzini-Armstrong, C., Coronado, R., Campbell, K.P. (1991) The brain ryanodine receptor: a caffeine-sensitive calcium release channel. *Neuron*, **7**, 17-25.
- [62] Moraidis, I., Bingmann, D. (1994) Epileptogenic actions of xanthines in relation to their affinities for adenosine A1 receptors in CA3 neurons of hippocampal slices (guinea pig). *Brain Res.*, **640**, 140-145.
- [63] Moraidis, I., Bingmann, D., Lehmenkuhler, A., Speckmann, E.J. (1991) Caffeine-induced epileptic discharges in CA3 neurons of hippocampal slices of guinea pig. *Neurosci. Lett.*, **129**, 51-54.
- [64] Moser, M.-B., Trommald, M., Andersen P. (1994) An increase in dendritic spine density on hippocampal CA1 pyramidal cells following spatial learning in adult rats suggests the formation of new synapses. *Proc. Natl. Acad. Sci. USA*, **91**, 12673-12675.
- [65] Nakano M., Yamada, S., Udagawa, R., Kato, N. (2004) Frequency-dependent requirement for calcium store-operated mechanisms in induction of homosynaptic long-term depression at hippocampus CA1 synapses. *Eur. J. Neurosci.*, **19**, 2881-2887.
- [66] Nehlig, A., Daval, J.-L., Debry, G. (1992) Caffeine and the central nervous system: mechanisms of action, biochemical, metabolic and psychostimulant effects. *Brain Res. Brain Res. Rev.*, **17**, 139-170.
- [67] Nowak, L., Bregestovski, P., Ascher, P., Herbet, A., Prochiantz, A. (1984) Magnesium gates glutamate-activated channels in mouse central neurones. *Nature*, **307**, 462-465.
- [68] Rae, M.G., Martin, D.J., Collingridge, G.L., Irving, A.J. (2000) Role of Ca^{2+} stores in metabotropic L-Glutamate receptor-mediated supralinear Ca^{2+} signaling in rat hippocampal neurons. *J. Neurosci.*, **20**, 8628-8636.
- [69] Ribeiro, J.A., Sebastiao, A.M., de Mendonca, A. (2002) Adenosine receptors in the nervous system: pathophysiological implications. *Prog. Neurobiol.*, **68**, 377-392.
- [70] Salinas, E., Sejnowski, T. J. (2001) Correlated neuronal activity and the flow of neural information. *Nat. Rev. Neurosci.*, **2**, 539-550.
- [71] Sandler, V.M., Barbara, J.G. (1999) Calcium-induced calcium release contributes to action potential-evoked calcium transients in hippocampal CA1 pyramidal neurons. *J. Neurosci.*, **19**, 4325-4336.
- [72] Savić, N., Sciancalepore, M. (1998) Intracellular calcium stores modulate miniature GABA-mediated synaptic currents in neonatal rat hippocampal neurons. *Eur. J. Neurosci.*, **10**, 3379-3386.
- [73] Shi, S.H., Hayashi, Y., Petralia, R.S., Zaman, S.H., Wenthold, R.J., Svoboda, K., Malinow, R. (1999) Rapid spine delivery and redistribution of AMPA receptors after NMDA receptor activation. *Science*, **284**, 1811-1816.
- [74] Simkus, C. R., Stricker, C. (2002) The contribution of intracellular calcium stores to mEPSCs recorded in layer II neurones of rat barrel cortex. *J. Physiol.*, **545**, 521-535.
- [75] Singer, W. (1995) Development and plasticity of cortical processing architectures. *Science*, **270**, 758-764.
- [76] Singer, W., Gray, C.M. (1995) Visual feature integration and the temporal correlation hypothesis. *Ann. Rev. Neurosci.*, **18**, 555-586.
- [77] Sorra, K.E., Harris, K.M. (2000) Overview on the structure, composition, function, development, and plasticity of hippocampal dendritic spines. *Hippocampus*, **10**, 51-511.
- [78] Sur, M., Leamey, C.A. (2001) Development and plasticity of cortical areas and networks. *Nat. Rev. Neurosci.*, **2**, 251-262.
- [79] Sutor, B., Mantell, K., Bacher, B. (1998) Evidence for the activity of five adenosine-3', 5'-monophosphate-degrading phosphodiesterase isozymes in the adult rat neocortex. *Neurosci. Lett.*, **252**, 57-60.
- [80] Svenningsson, P., Le Moine, C., Kull, B., Sunahara, R., Bloch, B., Fredholm, B.B. (1997) Cellular expression of adenosine A2A receptor messenger RNA in the rat central nervous system with special reference to dopamine innervated areas. *Neuroscience*, **80**, 1171-1185.
- [81] Taketo, M., Matsuda, H., Yoshida, T. (2004) Calcium-independent inhibition of GABA_A current by caffeine in hippocampal slices. *Brain Res.*, **1016**, 229-239.
- [82] Thümler, S., Dunwiddie, T.V. (2000) Adenosine receptor antagonists induce persistent bursting in the rat hippocampal CA3 region via an NMDA receptor-dependent mechanism. *J. Neurophysiol.*, **83**, 1787-1795.
- [83] Traub, R. D., Bibbig, A., LeBeau, F. E. N., Buhl, E. H., Whittington, M. A. (2004) Cellular mechanisms of neuronal population oscillations in the hippocampus *in vitro*. *Ann. Rev. Neurosci.*, **27**, 247-278.
- [84] Volfovsky, N., Parnas H., Segal M., Korkotian, E. (1999) Geometry of dendritic spines affects calcium dynamics in hippocampal neurons: theory and experiments. *J. Neurophysiol.*, **82**, 450-462.
- [85] Wang, S.J., Cheng, L.L., Gean, P.W. (1999) Cross-modulation of synaptic plasticity by beta-adrenergic and 5-HT_{1A} receptors in the rat basolateral amygdala. *J. Neurosci.*, **19**, 570-577.
- [86] Wang, X.J. (1999) Synaptic basis of cortical persistent activity: the importance of NMDA receptors to working memory. *J. Neurosci.*, **19**, 9587-9603.
- [87] Whiteford, K.L., Dijkhuizen, P., Polleux, F., Gosh, A. (2002) Molecular control of cortical dendrite development. *Ann. Rev. Neurosci.*, **25**, 127-149.

- [88] Yoshimura, H., Kato, N., Honjo, M., Sato, J., Sugai, T., Segami, N., Onoda, N. (2004) To-and-fro voltage signal propagation between the insular gustatory and parietal oral somatosensory areas in rat cortex slices. *Brain Res.*, **1015**, 114-121.
- [89] Yoshimura, H., Kato, N., Sugai, T., Segami, N., Onoda, N. (2003) Age-dependent appearance of an insulo-parietal cortical signal propagation that elicits a synchronized population oscillation in the parietal cortex in rats. *Brain Res. Development. Brain Res.*, **143**, 245-251.
- [90] Yoshimura, H., Kato, N., Sugai, T., Segami, N., Onoda, N. (2003) Age-dependent emergence of oscillatory signal flow between the primary and secondary visual cortices in rat brain slices. *Brain Res.*, **990**, 172-181.
- [91] Yoshimura, H., Sugai, T., Honjo, M., Segami, N., Onoda, N. (2005) NMDA receptor-dependent oscillatory signal outputs from the retrosplenial cortex triggered by a non-NMDA receptor-dependent signal input from the visual cortex *Brain Res.*, **1045**, 12-21.
- [92] Yoshimura, H., Sugai, T., Onoda, N., Segami, N., Kato, N. (2001) Synchronized population oscillation of excitatory synaptic potentials dependent on calcium-induced calcium release in rat neocortex layer II/III neurons. *Brain Res.*, **915**, 94-100.
- [93] Yoshimura, H., Sugai, T., Onoda, N., Segami, N., Kato, N. (2002) Age-dependent occurrence of synchronized population oscillation suggestive of a developing functional coupling between NMDA and ryanodine receptors in the neocortex. *Brain Res. Development. Brain Res.*, **136**, 63-68.
- [94] Yoshimura, H., Sugai, T., Segami, N., Onoda, N. (2005) Strengthening of non-NMDA receptor-dependent horizontal pathways between primary and lateral secondary visual cortices after NMDA receptor-dependent oscillatory neural activities. *Brain Res.*, **1036**, 60-69.
- [95] Yuste, R., Bonhoeffer, T. (2001) Morphological changes in dendritic spines associated with long-term synaptic plasticity. *Ann. Rev. Neurosci.*, **24**, 1071-1089.
- [96] Yuste, R., Majewska, A., Cash, S.S., Denk, W. (1999) Mechanisms of calcium influx into hippocampal spines: heterogeneity among spines, coincidence detection by NMDA receptors, and optical quantal analysis. *J. Neurosci.*, **19**, 1976-1987.

Received: 18 April, 2005

Accepted: 08 July, 2005



ELSEVIER

available at www.sciencedirect.com

SCIENCE @ DIRECT®

www.elsevier.com/locate/brainresBRAIN
RESEARCH

Research Report

Cyclic AMP-dependent attenuation of oscillatory-activity-induced intercortical strengthening of horizontal pathways between insular and parietal cortices

Hiroshi Yoshimura^{a,b,*}, Makoto Honjo^a, Natsuki Segami^a, Keiseki Kaneyama^a,
Tokio Sugai^b, Yuichi Mashiyama^a, Norihiko Onoda^b

^aDepartment of Oral and Maxillofacial Surgery, Kanazawa Medical University, Uchinada-cho, Ishikawa 920-0293, Japan

^bDepartment of Physiology, Kanazawa Medical University, Uchinada-cho, Ishikawa 920-0293, Japan

ARTICLE INFO

Article history:

Accepted 2 November 2005

Available online 4 January 2006

Keywords:

Insular cortex

Parietal cortex

Caffeine

cAMP

Oscillation

Optical recording

ABSTRACT

Cyclic AMP (cAMP) is a key intracellular second messenger, and the intracellular cAMP signaling pathway acts to modulate various brain functions. We have previously reported that low-frequency insular cortex stimulation in rat brain slices switches on a voltage oscillator in the parietal cortex that delivers signals horizontally back and forth under caffeine application. The oscillatory activities are N-methyl-D-aspartate (NMDA) receptor-dependent, and the role of oscillation is to strengthen functional intercortical connections. The present study investigated actions of the cAMP signaling pathway on caffeine-induced strengthening of intercortical connections and tried to confirm the role of oscillation on intercortical strengthening by focusing on the cAMP pathway. After induction of parietal oscillation by insular cortex stimulation in caffeine-containing medium, application of membrane-permeable cAMP analog, bromo-cAMP, diminished oscillatory signal delivery from the parietal cortex, but initial insulo-parietal signal propagation remained strong. When oscillatory activities were reduced with co-application of caffeine and bromo-cAMP from the beginning, initial insulo-parietal propagation was established, but amplitudes of propagating wavelets and propagating velocity were reduced. Thus, cAMP-dependent diminution of caffeine-induced NMDA-receptor-dependent oscillatory signal delivery causes attenuation of intercortical strengthening of horizontal pathways between insular and parietal cortices. This finding suggests that the intracellular cAMP signaling pathway has the ability to regulate extracellular communications at the network level, and also that full expression of strengthened intercortical signal communication requires sufficient NMDA-receptor-dependent oscillatory neural activities.

© 2005 Elsevier B.V. All rights reserved.

1. Introduction

In general, one of the roles of intracellular signaling pathways is to regulate extracellular signal communication (Bhella and

Iyengar, 1999; Butcher and Sutherland, 1962). Such signal regulation enables cells to make feedback or feedforward loops for cellular interaction and communication. Cyclic AMP (cAMP) is a critical intracellular second messenger, and the

* Corresponding author. Department of Oral and Maxillofacial Surgery, Kanazawa Medical University, Uchinada-cho, Ishikawa 920-0293, Japan. Fax: +81 76 286 2010.

E-mail address: hyoshimu@kanazawa-med.ac.jp (H. Yoshimura).

cAMP signaling pathway modulates various biological functions (Butcher and Sutherland, 1962; Cho et al., 2002; Crump et al., 2001; Frey et al., 1993; Mao and Wang, 2002; Renden and Broadie, 2003; Wang et al., 1999). In the brain, neurotransmitters and receptors dominantly intermediate extracellular signal communication at the synapses, and the cAMP signaling pathway largely operates at the synapses. The cAMP signaling pathway may thus have the ability to regulate extracellular signal communication in the brain via modulation of synaptic activities (Evans and Morgan, 2003; Ma et al., 1999; Thakur et al., 2004; Yoshimura and Kato, 2000).

Caffeine displays several pharmacological actions and can invade the synapses in the brain (Fredholm et al., 1999; Nehlig and Boyet, 2000). We recently demonstrated that caffeine enhances intercortical neural activities between primary and secondary visual cortices (Yoshimura et al., 2003a, 2005b). In those studies, we reported that intercortical horizontal connections are functionally opened and strengthened by caffeine-induced α -range voltage oscillations. The oscillatory

signal deliveries that activate N-methyl-D-aspartate (NMDA) receptor components to surrounding areas cause strengthening of functional neural connections (Yoshimura et al., 2003a, 2005a,b). One important action of caffeine is thus the opening and strengthening of intercortical signal communications by way of driving NMDA-receptor-dependent neural oscillators.

Since synapses are major sites where both the cAMP signaling pathway and caffeine operate and since caffeine has the ability to affect the cAMP signaling pathway (Beavo and Reifsnnyder, 1990; Butcher and Sutherland, 1962; Fredholm et al., 1999), increased intracellular cAMP level by pharmacological manipulation in addition to application of caffeine might induce up- or down-regulation of synaptic activities. We have focused attention on areas between the gustatory insular cortex where chemosensory information is processed and the parietal oral somatosensory cortex where somatosensory information is processed (Hanamori et al., 1999; Katz et al., 2001; Kosar et al., 1986; Ogawa and Wang, 2002; Paxinos

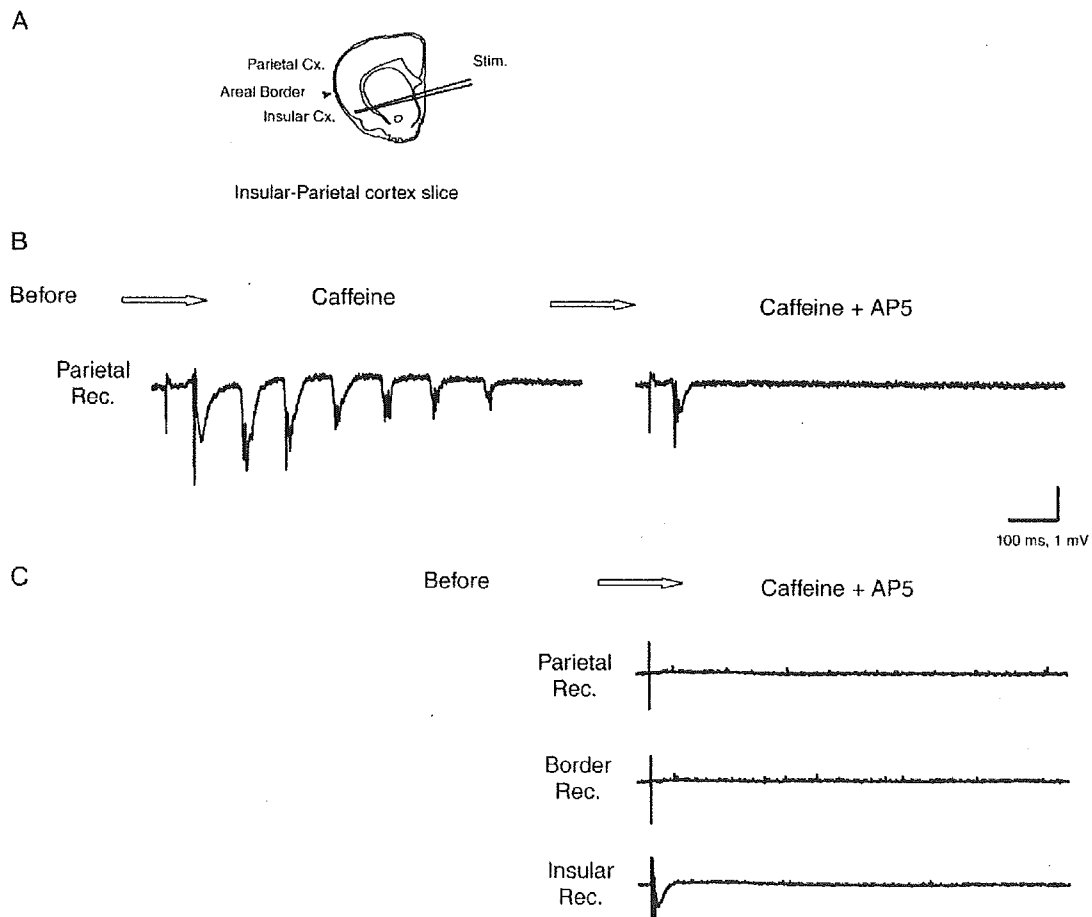


Fig. 1 – Comparing insulo-parietal signal propagation under caffeine application and NMDA receptor blockade after establishing voltage oscillation in the parietal cortex with propagation under application of caffeine and NMDA receptor blockade from the beginning. (A) Brain slices including insular and parietal cortices and electrical stimulation site. Low-frequency stimulation was delivered to the insular cortex, and field potentials were recorded from layer II/III in the insular cortex, parietal cortex and areal border. (B) After induction of parietal oscillation in caffeine-containing medium, D-AP5, an NMDA receptor blocker, was added. Note that initial insulo-parietal component remained. (C) Caffeine and D-AP5 co-applied to medium from the beginning. Note that initial component did not penetrate the parietal cortex.

et al., 1997; Remple et al., 2003; Shi and Cassell, 1998; Swanson et al., 1992; Yamamoto et al., 1981; Yoshimura et al., 2004b; Zilles and Wree, 1995) We recently reported that insulo-parietal neural connections are functionally strengthened after oscillatory activities in the parietal cortex (Yoshimura et al., 2003b, 2004a). The present study investigated how intracellular cAMP elevation affects opening and strengthening of insulo-parietal connections to examine whether cAMP pathways have the ability to regulate extracellular signal communications at the intercortical network level. In addition, we tried to verify that NMDA-receptor-dependent oscillatory activities cause strengthening of intercortical signal communications by focusing on the intracellular cAMP signaling pathway.

2. Results

Low-frequency intracortical electrical stimulation was delivered to layer IV in the gustatory insular cortex, and field potentials were recorded from layer II/III in the parietal cortex (Fig. 1A). In normal medium, a small solitary potential was observed only in the gustatory insular cortex and did not propagate horizontally toward the parietal cortex (24/24 slices, not shown). After application of caffeine into the medium, the evoked potential was enlarged, and the potential propagated from the insular cortex to the parietal cortex. However,

marked oscillation was only generated in the parietal cortex (22/24 slices). These propagation and oscillation have been precisely described in our previous report, and histological identification of the stimulation and recording sites has also been demonstrated (Yoshimura et al., 2003b, 2004a). The same kinds of propagation and oscillation are induced in both the insulo-parietal cortex and primary-secondary visual cortex (Yoshimura et al., 2003a, 2005b). We have repetitively reported that later oscillatory components are entirely NMDA-receptor-dependent in both the parietal and visual cortices (Yoshimura et al., 2003a,b, 2004a, 2005b). The present findings again confirmed this. Oscillation in the parietal cortex was induced in caffeine-containing medium after low-frequency stimulation to the insular cortex. After oscillation in the parietal cortex stabilized, D-2-amino-5-phosphonovaleric acid (D-AP5), a blocker of NMDA receptor, was applied to the medium. Approximately 30 min later, the insulo-parietal signal propagation remained, but oscillatory activities in the parietal cortex were depressed (Fig. 1B; "Caffeine + AP5"; 12/12 slices). The remaining initial response was non-NMDA-receptor-dependent since additional application of 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX), a blocker of non-NMDA receptor, completely depressed the initial response (15/15 slices, not shown).

In the visual cortex, co-application of caffeine and D-AP5 from the beginning abolishes not only induction of oscillatory signal deliveries from the secondary visual cortex, but also

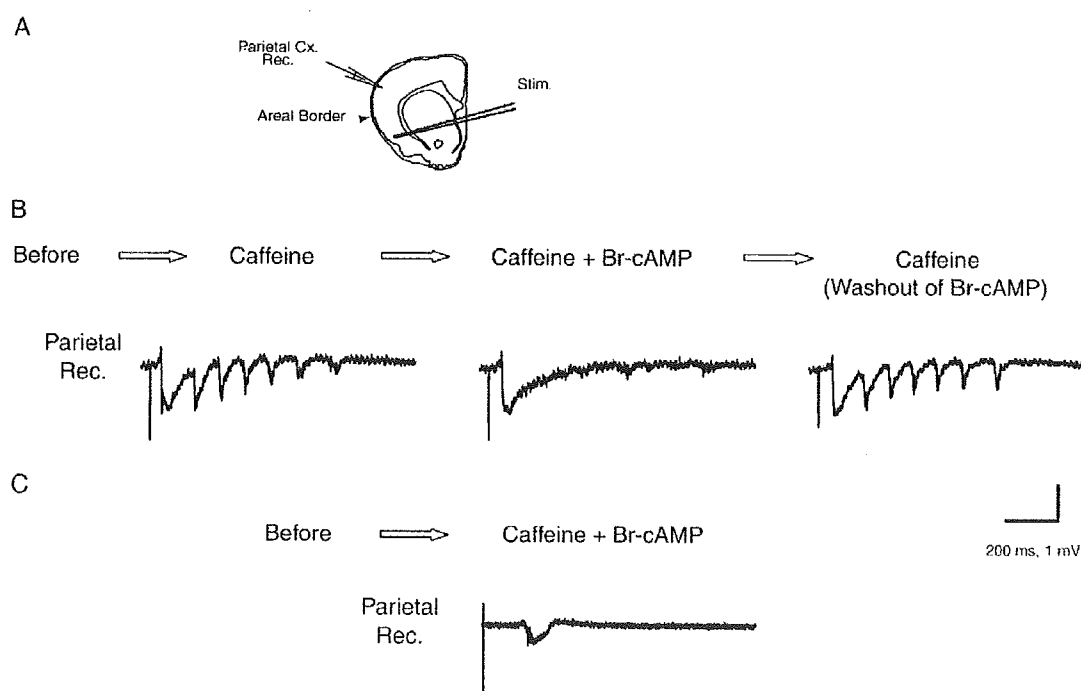


Fig. 2 – Comparing effects of bath application of bromo-cAMP on responses in parietal cortex evoked by insular cortex stimulation under co-application of caffeine and bromo-cAMP after caffeine conditioning with effects under co-application of caffeine and bromo-cAMP from the beginning. (A) Location of stimulation and recording electrodes. (B) After inducing marked parietal oscillations in caffeine-containing medium, bromo-cAMP was added to the medium. When oscillatory phases were depressed, bromo-cAMP was washed out from medium. (C) Bromo-cAMP co-applied with caffeine from the beginning. Note that induction of oscillatory activities was depressed, but depression of oscillatory activities was incomplete, and initial response with smaller amplitude and longer latency was observed.

opening horizontal signal pathways from the primary visual cortex to the secondary visual cortex (Yoshimura et al., 2005b). In the same way, co-application of caffeine and D-AP5 from the beginning in the present study abolished not only induction of parietal oscillation, but also opening horizontal signal pathways from the insular cortex to the parietal cortex (Fig. 1C). The present study offered new confirmation that NMDA-receptor-dependent oscillatory activities cause opening and strengthening of intercortical horizontal signal pathways between the insular and parietal cortices.

The intracellular cAMP pathway is a key second messenger pathway that modulates neuron functions. We have previously reported that synaptic efficiency in the visual cortex is modulated by increases in intracellular cAMP (Yoshimura and Kato, 2000). In the next experiment, effects of intercellular

cAMP elevation on propagation and oscillation between insular and parietal cortices were investigated. Oscillation in the parietal cortex was induced in the same way as described above, and, after oscillation in the parietal cortex stabilized, bromo-cAMP, a membrane permeable cAMP analog, was applied to the medium. Approximately 30 min later, insulo-parietal signal propagation remained, but oscillatory activities in the parietal cortex were depressed. Depressed oscillatory activities recovered with washout of bromo-cAMP from the medium (Fig. 2B; 7/7 slices). Application of bromo-cAMP thus reversibly diminished oscillatory activities in the parietal cortex.

NMDA-receptor-dependent oscillatory signal deliveries contribute to strengthening of corticocortical signal pathways between the primary and secondary visual cortices (Yoshimura

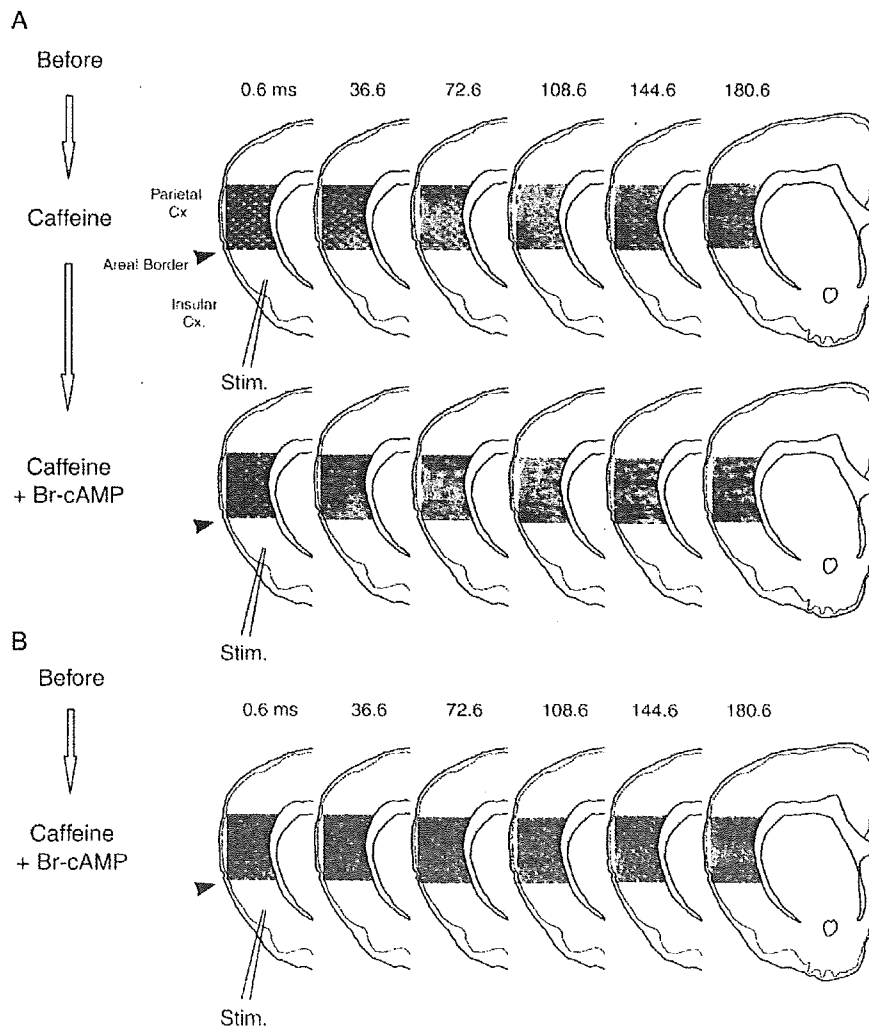


Fig. 3 – Comparing spatiotemporal dynamics of initial voltage signal propagation in parietal cortex under co-application of caffeine and bromo-cAMP after caffeine conditioning with dynamics under co-application of caffeine and bromo-cAMP from the beginning. (A) After caffeine conditioning, initial signal evoked by insular cortex stimulation propagated into parietal cortex along layer II/III (upper). After additional application of bromo-cAMP, initial evoked signal propagated into parietal cortex in the same way (lower). (B) When caffeine and bromo-cAMP were co-applied from the beginning, initial evoked signal managed to propagate into parietal cortex, but intensity of optical response and propagating velocity was reduced, compared with the case for caffeine application preceded by bromo-cAMP.

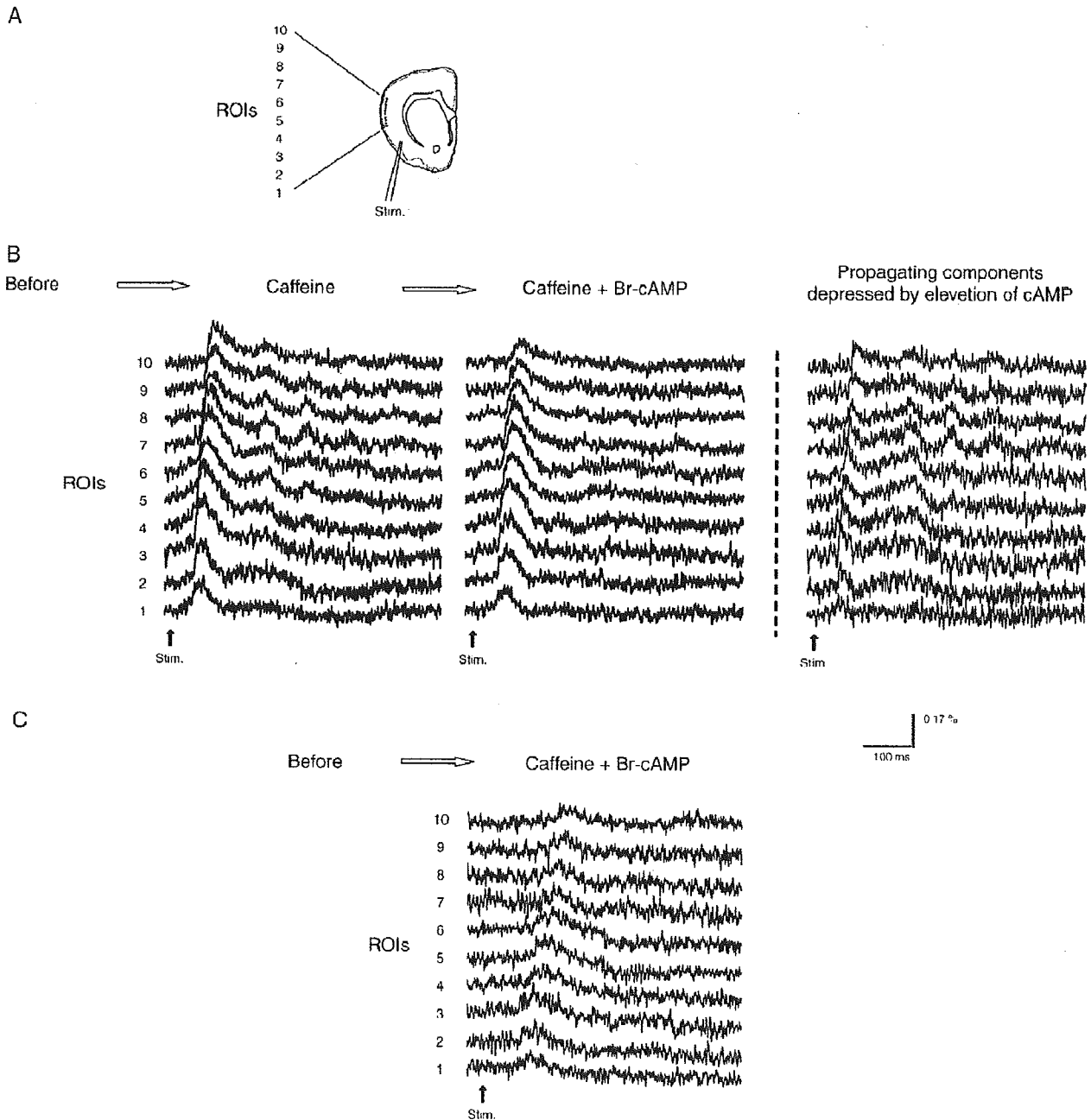


Fig. 4 – Comparing time courses of optical responses in parietal cortex obtained from optical recordings as shown in Fig. 3 under co-application of caffeine and bromo-cAMP after caffeine conditioning with time courses under co-application of caffeine and bromo-cAMP from the beginning. (A) Regions of interest (ROIs) 1–10 are shown in an illustration of insulo-parietal cortex slice. (B) Time courses of optical responses obtained from ROIs were arranged vertically. Left: after caffeine conditioning, initial signal evoked by insular cortex stimulation propagated into parietal cortex, and later oscillatory components were generated in parietal cortex and propagated toward insular cortex. Precise analysis of propagation is described in Thakur et al. (2004). Center: after additional application of bromo-cAMP, initial evoked signal propagated into parietal cortex in the same way, but later oscillatory components were largely depressed. Right: propagating components depressed by elevation of cAMP were obtained by subtracting “Caffeine” recordings from “Caffeine + bromo-cAMP” recordings. (C) When caffeine and bromo-cAMP were co-applied from the beginning, initial evoked signal managed to propagate into parietal cortex, but amplitudes of optical response and propagating velocity were reduced compared with the case of caffeine application preceded by bromo-cAMP.