

Cell culture, reporter assay and expression of human BRM and Brg1 in insect cell

293T, SW13 and C33A cells were maintained in Dulbecco's modified Eagle's medium (DMEM). Cells were grown supplemented with 10% fetal calf serum. G401 cell was cultured in RPMI medium-1640 (Invitrogen) supplemented with 10% calf serum. Transfections of all expression vectors, pG5luc and pRL-TK, were carried out by using Lipofectamine Plus reagent (Invitrogen). After 36 h of transfection, the cells in 24-well plates were washed by phosphate-buffered saline (PBS) and analysed for luciferase activities in triplicate in each transfection experiment by using Dual-Luciferase Reporter Assay System (Promega). Sf9 insect cell was grown in Grace's Insect Medium (Invitrogen) supplemented with 10% fetal calf serum. Expression procedure of human BRM or Brg1 in Sf9 insect cell was performed using Bac-to-Bac Baculovirus Expression System (Invitrogen).

GST pull-down assay

GST-Gal4-SYT mutant DNAs were transformed in BL21 pLys-S (Novagen). Bacterial cells that expressed GST fusion proteins were harvested and re-suspended in buffer A [50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 500 mM NaCl, protease inhibitors]. After sonication, the lysates were centrifuged for 30 min at 8000 rpm. The supernatants were incubated with glutathione sepharose 4B beads (Amersham Pharmacia Biotech) at 4 °C for 1 h and the beads were washed with buffer A for four times. The Sf9 cells expressed flag tagged BRM or Brg1 were washed with PBS and resuspended in buffer B [10 mM Tris-HCl (pH 8.0), 10 mM glycerol, 0.15 M NaCl, 0.1% nonidet P-40, 5 mM 2-mercaptoethanol, protease inhibitors]. After sonication, the lysates were centrifuged for 30 min at 15 000 rpm. The supernatants were incubated with 30 µL of anti-flag M2 Agarose (Sigma) at 4 °C for overnight and washed five times with buffer B. Bound proteins were eluted with buffer B containing 0.2 mg/mL of flag peptide (Sigma). The elutes were incubated with GST-beads immobilized GST-Gal4-SYT mutants in buffer B at 4 °C for 4 h. The beads were washed four times with buffer B without 2-mercaptoethanol and re-suspended in sample buffer. The bound proteins were separated by 8% SDS-PAGE (sodium dodecyl sulfate—polyacrylamide gel electrophoresis) and analysed with anti-BRM (sc-6450, Santa Cruz) and anti-Brg1 (sc-8749, Santa Cruz) antibodies, respectively. The ECL System (Amersham Pharmacia Biotech) was used for detection on RX-U X-ray films (Fuji).

Immunoprecipitation

The expression vectors of flag-tagged SYT mutants (without GAL4 DNA binding domain), no-tagged BRM and Brg1 were transfected to 293T by Lipofectamine Plus reagent. After 36 h, the cells were washed and suspended in NP-40 lysis buffer [10 mM Tris-HCl (pH 7.8), 1% nonidet P-40, 0.15 M NaCl, 1 mM EDTA, protease inhibitors]. Then lysates were centrifuged for 30 min at 15 000 rpm. The supernatants were incubated with 30 µL of anti-flag M2 agarose at 4 °C for overnight and washed four times with NP-40 lysis buffer. The bound proteins were detected by Western blot.

Immunofluorescence microscopy

Expression vectors of GFP-SYT-WT, HA-tagged BRM or Brg1 were transfected in SW13 using the same method of immunoprecipitation. After 24 h, the cells were removed to a chamber slide, incubated a further 24 h, fixed with 3% formaldehyde in PBS for 10 min. After washing by PBS, the cells were permeabilized with 0.1% polyoxyethylene octylphenyl ether (10) in PBS for 5 min and washed with PBS. The slide was incubated in blocking buffer (2% normal swine serum in PBS) for 20 min. Cells were incubated overnight with primary antibody at 4 °C, washed three times with PBS and incubated with secondary antibody for 40 min at 25 °C. As primary antibodies, we used anti-BRM (sc-6450 and sc-6449, Santa Cruz), anti-Brg1 (sc-8749, Santa Cruz). As secondary antibodies, we used rhodamine-conjugated donkey anti-goat (sc-2094, Santa Cruz). Finally, the slide was mounted with DAPI and visualized under Leica DM IBD microscope with appropriate filters and images were acquired by Leica Qfluoro.

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