



## Discussion

In the present study, immunohistochemical and biochemical methods were used to investigate the localization and age-related changes of intracellular A $\beta$  in brains from young, adult and aged cynomolgus monkeys.

Recent studies have shown that intracellular A $\beta$  is found only in the brains of aged individuals or brains of AD patients, but not in brains of nonhuman primates [9,10,12,16]. However, in the present study, we found intracellular A $\beta$  in cortical neurones of monkeys of various ages (Figures 1 and 2). One possible technical explanation for this apparent discrepancy is that in previous studies, material came from brains fixed with neutral buffered formalin, whereas in the present study material was derived from brains fixed with paraformaldehyde [9,10,12,16]. For immunohistochemical studies we fixed brain samples, derived from the same monkeys, with either neutral buffered formalin or paraformaldehyde. Anti-A $\beta$  antibodies failed to immunostain intracellular A $\beta$  in cortical neurones from samples fixed with the formalin protocol (data not shown). On the other hand, all anti-A $\beta$  antibodies successfully immunostained neurones from samples fixed with paraformaldehyde (Figures 1 and 2). It is widely known that the cross linkages in paraformaldehyde-fixed tissues are much weaker than those in tissues fixed with neutral buffered formalin, leading to a kind of fixation with the former that more effectively preserves tissues and cell surface antigens [22]. Thus, the antigens of intracellular A $\beta$  would be preserved well and might be recognized by antibodies without formic acid treatment. It is also known that periodic acid pretreatment can improve immunoreactivity. These two factors may explain why we were able to detect intracellular A $\beta$  even in brains from young monkeys.

In the brains of young monkeys, A $\beta$  immunoreactivity was granular in nature. This diffuse pattern of immunoreactivity indicates that the intracellular A $\beta$  in young monkeys may represent A $\beta$  generated biologically in neurones, rather than accumulated A $\beta$  that is typically found in neurones of aged animals (Figure 1a,b).

In the brains of aged monkeys, A $\beta$  immunoreactivity in some cortical neurones tended to cluster together in clumps (Figure 1c,d). This clustering of immunoreactivity may represent A $\beta$  accumulated in neurones and is consistent with previous hypotheses that A $\beta$  accumulates in lipid rafts and that clustering of GM1 ganglioside within lipid raft-like membranes may facilitate the aggregation of

A $\beta$  (fibrils) in these rafts [23]. Although the pattern of A $\beta$  immunostaining (i.e. intracellular A $\beta$ ) changed with age, we failed to find clear evidence that intracellular A $\beta$  colocalizes with SPs.

In our previous studies, we described age-related changes of AD-related proteins in the cynomolgus monkey brain [19,20]. In the present study, we determined with biochemical methods whether intracellular A $\beta$  accumulates with advancing age. In the microsomal fraction, the amount of total A $\beta$  increased with age, whereas the amount of A $\beta$ 40 did not change with age (Figures 3 and 4a). Both A $\beta$ 42 and A $\beta$ 43, however, showed very interesting age-related changes. The amount of A $\beta$ 42 in the microsomal fraction increased significantly with age (Figures 3 and 4a). On the contrary, the amount of A $\beta$ 43 significantly decreased in this fraction (Figures 3 and 4a). This is the first study to show that the amounts of different A $\beta$  molecules generated in the brain change with age of the individual. These results suggest that during ageing the cleavage of APP by  $\gamma$ -secretase may shift to generate more A $\beta$ 42 than A $\beta$ 43, which in turn may lead to increased accumulation of A $\beta$ 42 in the brain. Several studies have shown that A $\beta$ 42 is more closely associated with AD pathogenesis than is A $\beta$ 40 [24–27]. The age-related increase of A $\beta$ 42 observed in the present study supports these previous findings and is consistent with the hypothesis that increased A $\beta$ 42 generation or accumulation may promote, or even cause, sporadic AD pathogenesis in the aged brain.

As the nerve ending fraction prepared in the standard way contains both synaptic vesicles and synaptic plasma membranes, the amount of A $\beta$  detected in this fraction may be extracellular A $\beta$  associated with the outer part of the synaptic plasma membrane. To avoid potential contamination with extracellular A $\beta$ , we pretreated each nerve ending fraction with Pro-K (this resulting fraction was termed the Pro-NE fraction). In the Pro-NE fraction, the amount of total intracellular A $\beta$  significantly increased with age (Figures 3 and 4b). Our previous study showed that both full-length APP and  $\beta$ CTF accumulated in the nerve ending fraction with age [20]. It is noteworthy that AD-related proteins also significantly accumulate in the nerve ending fraction [19,20]. Thus, one possible explanation for this age-related accumulation of A $\beta$  in the nerve ending fraction is that increased APP and  $\beta$ CTF in this fraction would subsequently cause intracellular A $\beta$  to also accumulate in this fraction. Alternatively, the

age-related reduction of axonal transport to the soma for degradation may also cause the accumulation of these proteins, including A $\beta$ , in the nerve ending fraction. Although the amounts of intracellular A $\beta$ 40, A $\beta$ 42 and A $\beta$ 43 significantly increased with age, the magnitude of increase varied for all three isoforms (Figures 3 and 4b). Of these three A $\beta$  molecules, intracellular A $\beta$ 43 increased much less than did A $\beta$ 40 and A $\beta$ 42 (Figure 4b). This may explain why, in general, the generation of A $\beta$ 43 decreased with age (Figure 4a).

In summary, we found that intracellular A $\beta$  exists in cortical neurones of monkeys as young as 4 years old, albeit in the form of diffuse granules. With advancing age (24–36 years old), however, intracellular A $\beta$  tended to localize in neurones in the form of clusters or clumps (Figures 1 and 2). Our biochemical assays revealed that A $\beta$ 40 and A $\beta$ 43 were mainly generated in the brains of young monkeys. During ageing, the generation of intracellular A $\beta$  shifted such that A $\beta$ 42 generation increased, while A $\beta$ 40 generation remained unchanged (Figures 3 and 4).

It is also noteworthy that the amount of intracellular A $\beta$  increased in the nerve ending fraction with age (Figures 3 and 4), and this result closely parallels our findings with other AD-related proteins reported in our previous studies [19,20]. The accumulation of APP and  $\beta$ CTF in synaptosomes and/or synaptic plasma membranes may lead to A $\beta$  overproduction in these compartments, and the accumulated intracellular A $\beta$ , in turn, may be secreted extracellularly and accelerate SP formation. Thus, the nerve ending may be where SPs are initially formed or may be significantly associated with spontaneous SP formation that occurs with advancing age. Taken together, these results support the line of research that focuses in on age-related changes in the nerve ending. Findings from this line of research should reveal more details about the association between AD pathogenesis and other age-related neurodegenerative disorders.

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### References

- 1 Citron M, Oltersdorf T, Haass C, McConlogue L, Hung AY, Seubert P, Vigo-Pelfrey C, Lieberburg I, Selkoe DJ. Mutation of the beta-amyloid precursor protein in familial Alzheimer's disease increases beta-protein production. *Nature* 1992; **360**: 672–4
- 2 Glenner GG. Alzheimer's disease: its proteins and genes. *Cell* 1988; **52**: 307–8
- 3 Chartier-Harlin MC, Crawford F, Houlden H, Warren A, Hughes D, Fidani L, Goate A, Rossor M, Roques P, Hardy J, Mullan M. Early-onset Alzheimer's disease caused by mutations at codon 717 of the  $\beta$ -amyloid precursor protein gene. *Nature* 1991; **353**: 844–6
- 4 Murell J, Farlow M, Ghetti B, Benson MD. A mutation in the amyloid precursor protein associated with hereditary Alzheimer's disease. *Science* 1991; **254**: 97–9
- 5 Scheuer D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird TD, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad E, Viitanen M, Peskind E, Poorkaj P, Schellenberg G, Tanzi R, Wasco W, Lannfelt L, Selkoe DJ, Younkin S. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nature Med* 1996; **2**: 864–70
- 6 De Strooper B, Saftig P, Craessaerts K, Vanderstichele H, Guhde G, Annaert W, Von Figura K, Van Leuven F. Deficiency of presenilin-1 inhibits the normal cleavage of amyloid precursor protein. *Nature* 1998; **391**: 387–90
- 7 Wolfe MS, Xia W, Ostaszewski BL, Diehl TS, Taylor Kimberly W, Selkoe DJ. Two transmembrane aspartates in presenilin-1 required for presenilin endoproteolysis and  $\gamma$ -secretase activity. *Nature* 1999; **398**: 513–17
- 8 Xia X, Zhang J, Perez R, Koo EH, Selkoe DJ. Interactions between amyloid precursor protein and presenilins in mammalian cells. Implications for pathogenesis of Alzheimer's disease. *Proc Natl Acad Sci U S A* 1997; **94**: 8208–13
- 9 D'Andrea MR, Nagele RG, Wang H.-Y, Peterson PA, Lee DHS. Evidence that neurons accumulating amyloid can undergo lysis to form amyloid plaques in Alzheimer's disease. *Histopathology* 2001; **38**: 120–34
- 10 D'Andrea MR, Nagele RG, Wang H.-Y, Lee DHS. Consistent immunohistochemical detection of intracellular  $\beta$ -amyloid42 in pyramidal neurons of Alzheimer's disease entorhinal cortex. *Neurosci Lett* 2002; **333**: 163–6
- 11 Echeverria V, Cuello AC. Intracellular A-beta amyloid, a sign for worse things to come? *Mol Neurobiol* 2002; **26**: 299–316
- 12 Gouras GK, Tsai J, Naslund J, Vincent B, Edgar M, Checler F, Greenfield JP, Haroutunian V, Buxbaum JD, Xu H, Greengard P, Relkin NR. Intraneuronal A $\beta$ 42 accumulation in human brain. *Am J Pathol* 2000; **156**: 15–20
- 13 Klein WL, Krafft GA, Finch CE. Targeting small Abeta oligomers: the solution to an Alzheimer's disease conundrum? *Trends Neurosci* 2001; **24**: 219–24

- 14 Tabira T, Chui DH, Kuroda S. Significance of intracellular Abeta42 accumulation in Alzheimer's disease. *Front Biosci* 2002; 7: 44–9
- 15 Tabira T, de Chui H, Nakayama H, Kuroda S, Shibuya M. Alzheimer's disease with spastic paresis and cotton wool type plaques. *J Neurosci Res* 2002; 70: 367–72
- 16 Wang H.-Y, D'Andrea MR, Nagele RG. Cerebellar diffuse amyloid plaques are derived from dendritic A $\beta$ 42 accumulations in Purkinje cells. *Neurobiol Aging* 2002; 23: 213–23
- 17 Xu G, Gonzales V, Borchelt DR. Abeta deposition does not cause the aggregation of endogenous tau in transgenic mice. *Alzheimer Dis Assoc Disord* 2002; 16: 196–201
- 18 Nakamura S, Nakayama H, Goto N, Ono F, Sakakibara I, Yoshikawa Y. Histopathological studies of senile plaques and cerebral amyloidosis in cynomolgus monkeys. *J Med Primatol* 1998; 27: 244–52
- 19 Kimura N, Nakamura S, Honda T, Takashima A, Nakayama H, Ono, Sakakibara I, Doi K, Kawamura S, Yoshikawa Y. Age-related changes in the localization of presenilin-1 in cynomolgus monkey brain. *Brain Res* 2001; 922: 30–41
- 20 Kimura N, Tanemura K, Nakamura S, Takashima A, Ono F, Sakakibara I, Ishii Y, Kyuwa S, Yoshikawa Y. Age-related changes of Alzheimer's disease-associated proteins in cynomolgus monkey brains. *Biochem Biophys Res Comm* 2003; 310: 303–11
- 21 Tamai Y, Kojima H, Ohtani Y, Uchida K, Taguchi F, Kawaguchi T, Miura S, Tateishi J. Subcellular distribution of the transmissible agent in Creutzfeldt-Jakob disease mouse brain. *Microbiol Immunol* 1989; 33: 35–42
- 22 Smit JW, Meijer CJ, Decary F, Feltkamp-Vroom TM. Paraformaldehyde fixation in immunofluorescence and immunoelectron microscopy. Preservation of tissue and cell surface membrane antigens. *J Immunol Meth* 1974; 6: 93–8
- 23 Kakio A, Nishimoto SI, Yanagisawa K, Kozutsumi Y, Matsuzaka K. Cholesterol-dependent formation of GM1 ganglioside-bound amyloid-protein, an endogenous seed for Alzheimer amyloid. *J Biol Chem* 2001; 276: 24985–90
- 24 Burdick D, Soreghan B, Kwon M, Kosmoski J, Knauer M, Henshen A, Yates J, Cotman C, Glabe C. Assembly and aggregation properties of synthetic Alzheimer's A4/beta amyloid peptide analogs. *J Biol Chem* 1992; 267: 546–54
- 25 Jarrett JT, Berger EP, Lansbury PT Jr. The carboxy terminus of the beta amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease. *Biochemistry* 1993; 32: 4693–7
- 26 Suzuki N, Cheung TT, Cai XD, Odaka A, Otvos L Jr, Eckman C, Golde TE, Younkin SG. An increased percentage of long amyloid beta protein secreted by familial amyloid beta protein precursor (beta APP717) mutants. *Science* 1994; 264: 1336–40
- 27 Younkin SG. The amyloid beta protein precursor mutations linked to familial Alzheimer's disease alter processing in a way that fosters amyloid deposition. *Tohoku J Exp Med* 1994; 174: 217–23

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