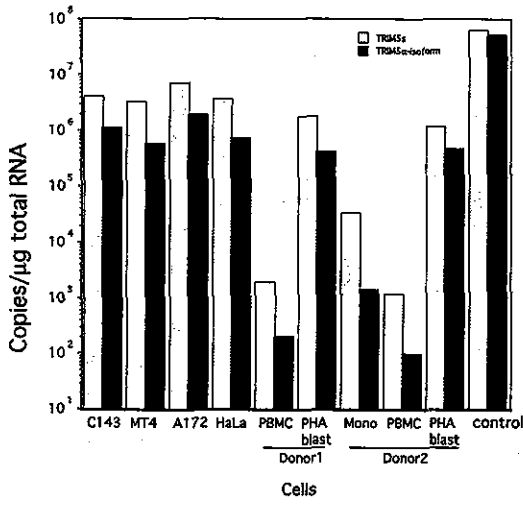
Copies/μg total RNA
C143
MT4
A172
HaLa
PBMC
PHA blast
Donor1
Mono
PBMC
PHA blast
Donor2
control
Cells

Antibody-Dependent Cellular Cytotoxicity via Humoral Immune Epitope of Nef Protein Expressed on Cell Surface[1]

Takeshi Yamada,[2]* Nobukazu Watanabe,† Tetsuya Nakamura,‡ and Aikichi Iwamoto[3]*‡

Antibodies against various proteins of HIV type 1 (HIV-1) can be detected in HIV-1-infected individuals. We previously reported that the level of Ab response against one Nef epitope is correlated with HIV-1 disease progression. To elucidate the mechanism for this correlation, we examined Ab-dependent cellular cytotoxicity (ADCC) against target cells expressing Nef. We observed efficient cytotoxicity against Nef-expressing target cells in the presence of patient plasma and PBMCs. This ADCC activity was correlated with the dilution of plasma from HIV-1-infected patients. Addition of a specific synthetic peptide (peptide 31: FLKEKGGLE) corresponding to the Nef epitope reduced cell lysis to ~50%. These results suggest that PBMCs of HIV-1-infected patients may exert ADCC via anti-Nef Abs in the patients' own plasma and serve as a mechanism used by the immune system to regulate HIV-1 replication. *The Journal of Immunology,* **2004, 172: 2401–2406.**

Highly active antiretroviral therapy dramatically suppresses HIV-1 replication and has thereby contributed to decrease the incidence of AIDS-related opportunistic infections and subsequent mortality (1, 2). However, elimination of HIV-1 from infected individuals has not yet been achieved by highly active antiretroviral therapy alone (3–5). Therefore, the development of different therapeutic approaches is mandatory.

Ab-dependent cellular cytotoxicity (ADCC)[4] as well as CTL play an important role in protective immunity against viral infections (6, 7). ADCC can inhibit viral replication and cell-to-cell infection by killing HIV-1-infected cells before maturation of virus particles (8, 9). Therefore, ADCC activity could benefit the prevention of disease progression. In early studies, Rook et al. (10) and Ljunggren et al. (11) demonstrated that sera from HIV-1-infected individuals were able to mediate ADCC against HIV-1-infected T cells, and there was a positive correlation between ADCC activity and disease progression. When HIV-1-infected cells produce virus particles, viral envelope glycoproteins are abundantly exposed to the cell surface through the plasma membrane. In fact, ADCC via Abs against gp120 or gp41, HIV-1 envelope protein, has been well documented (12–20). It has been described that gp120 or gp120/41-specific ADCC correlates with rate of disease progression (19, 21). But, in contrast, ADCC via envelope proteins could potentially kill the uninfected $CD4^+$ T cells with free viral envelopes on their surface, and therefore ADCC could contribute to depletion of $CD4^+$ T cells and AIDS pathogenesis (22, 23). In addition, gp120 is prone to high frequency of mutations; thereby, viral escape mutants may evolve easily (24–26). In view of these disadvantages, envelope proteins appear to be unsuitable as targets for ADCC against the progression of disease in HIV-1-infected patients. Conserved proteins may be better targets if one considers ADCC as a durable therapeutic weapon against HIV-1. With regard to this, Gag and Pol are very conserved proteins, and if their epitopes were expressed on the cell surface, these proteins could be good candidates for specific ADCC. Rook et al. (10) described that Ab reactivity with the p24 (Gag) protein of patient's serum correlates inversely with disease progression. It has been reported that Gag proteins are expressed on the cell surface (27, 28); nevertheless, the inductions of ADCC via Gag have never been succeeded (29). And, furthermore, there has been no evidence that Pol proteins are expressed on the HIV-1-infected cells; therefore, Pol Ags could not be exposed to the extracellular enviroment as ADCC target. Thus, the contribution of other HIV-1 proteins except envelope proteins to ADCC has remained unclear.

Nef protein is an HIV-1 accessory protein with important roles for pathogenesis of HIV-1 infection (30–35). Nef protein is partially expressed on the surface of HIV-1-infected cells (36–38). We previously reported that highly conserved amino acid residues (FLKEKGGLE) are expressed on the surface of HIV-1-infected cells. The peptide residues served as an epitope for Ab response, and the plasma level of the Abs against the epitope was correlated with HIV-1 disease progression (39, 40). To elucidate the mechanism of this correlation, we studied ADCC activities using patients' peripheral mononuclear cells (PBMCs) and a patient's plasma, which contained high amount of anti-Nef Abs. We also analyzed characteristics of patients' NK cells that should be the key player in ADCC against virus-induced target cells.

Materials and Methods

Cells

Five HIV-1-infected subjects whose PBMCs were used as effector cells for the ADCC assay are listed in Table I. PBMCs were freshly isolated by centrifuging heparinized blood over Ficoll-Hypaque (Meneki-seibutsuken, Gunma, Japan). PBMCs were counted and adjusted to the concentration of 2×10^6 cells/ml in RPMI 1640 medium supplemented with 10% heat-inactivated FCS (RPMI 10). A portion of the cells was used for phenotypic analysis using flow cytometry. For the flow cytometric analysis of NK

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[4] Abbreviations used in this paper: ADCC, Ab-dependent cellular cytotoxicity; LTNP, long-term nonprogressor.

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Table I. *Patient profiles*

Patient	Age	Sex	$CD4^+$ Count (cells/μl)	$CD8^+$ Count (cells/μl)	NK Cell Count (cells/μl)	% NK Cell in FBMC	HIV RNA (copies/ml)[a]	Antiretroviral Drugs[b]
P1	37	M	754	996	155	8.0	<400	d4T + 3TC + NFV
P2	32	M	63	214	20	3.7	770	d4T + 3TC + NFV
P3	45	M	204	620	220	12.6	<400	AZT + ddC + IDV
P4	37	M	638	1034	102	5.7	<400	d4T + 3TC + NFV
P5	35	M	372	877	73	5.0	2200	AZT + ddC + IDV

[a] Amplicor HIV monitor test (Roche Diagnostics Systems, Somerville, NJ).
[b] AZT, zidovudine; d4T, stavudine; 3TC, lamivudine; ddC, zalcitabine; NFV, nelfinavir; IDV, indinavir.

cells, PBMC samples from another 40 HIV-1-positive subjects and 16 uninfected donors were included in this study.

For the ADCC assay, we used CEM-NK^R cells that were obtained through the AIDS Research and Reference Reagent Program, Division of AIDS, National Institute of Allergy and Infectious Diseases, National Institutes of Health from J. Corbeil (41). Nef proteins were expressed in these cells by using a recombinant Sendai virus system, which has been shown to express large amounts of heterologous recombinant proteins in 24 h after infection in suspension cells (42). CEM-NK^R cells were infected with SeV-Nef to express HIV-1 (NL43 strain) Nef proteins or wild SeV at a multiplicity of infection of 10 for 1 h at 37°C, as previously described (43), and cultured for 24 h in RPMI 10. These cells were designated CEM-NK^R-Nef or CEM-NK^R-mock cells, respectively.

Subjects and reagents

For ADCC assay, we used the plasmas from long-term nonprogressor 2, 5, and 6 (LTNP 2, 5, and 6), whose characterization was published previously (39). $Na_2[^{51}Cr]O_4$ was obtained from NEN Life Science Products (Boston, MA). mAbs N901 (NKH-1) (anti-CD56; FITC conjugated) and 3G8 (anti-CD16; PE) were obtained from Coulter (Miami, FL). mAbs SJ25C1 (anti-CD19; PerCP) and SK7 (anti-CD3; allophycocyanin) were obtained from BD Immunocytometry Systems (San Jose, CA). mAb δG9 (anti-perforin) was a generous gift of E. Podack (University of Miami, Miami, FL). δG9 was conjugated with FITC in our laboratory. Nine-mer peptide 31 (=FLKEKGGLE) and control peptide (=GGGGGGGGG) were synthesized using a Multipin peptide synthesis kit (Chiron Mitotopes, Clayton, Victoria, Australia). The yields were analyzed by gas-liquid chromatography to confirm the correct synthesis.

Immunofluorescent staining

For analysis of Sendai virus-infected CEM-NK^R cells, cells (10^5) were centrifuged over silan-coating glass coverslips (DAKO, Carpenteria, CA), fixed with 2% paraformaldehyde in PBS for 5 min, blocked with BlockAce (Snow-Brand, Tokyo, Japan) for 30 min, and incubated for 1 h with plasma of LTNP 5 1/2.5 diluted in PBS. Then cells were incubated for 30 min with FITC-conjugated goat anti-human Igs (IgG, IgA, and IgM) $F(ab')_2$ (BioSource International, Camarillo, CA) after wash with PBS, and were mounted in 85% glycerol, 10 mM of Tris-HCl (pH 8), and 5% *n*-propylgallate. These stained cells were inspected with a confocal microscope (MRC 1024; Bio-Rad, Hercules, CA).

ADCC assays

ADCC assays were performed in 200 μl, total volume. Patient plasma used in the ADCC assay was incubated for 30 min at 56°C to inactivate the complement system. Plasmas from randomly selected healthy donors were used as control. A total of 1×10^6 target cells was labeled by incubation with medium containing $Na_2[^{51}Cr]O_4$ (0.5 mCi/ml) at 37°C for 1 h. Cells were washed three times with plain RPMI 1640 medium and resuspended in RPMI 10 at 2×10^5 cells/ml. A total of 50 μl of resuspended cells was added to each well of a 96-well microtiter plate (U bottom). Then, 50 μl of heat-inactivated healthy or patient's plasma diluted to 1/2.5 (thus, final concentration equals to 10^{-1} of original in 200 μl, total volume) in RPMI 10 was added to the plate before incubating for 30 min at 37°C. For the dilution assay of plasma, final concentration of plasma was adjusted to 10^{-1}, 10^{-2}, 10^{-3}, and 10^{-4} of original with RPMI 10, respectively. After incubation, either 100 μl of patients' PBMCs (2×10^6 cells/ml) (for sample count), 100 μl of RPMI 10 containing 2% Triton solution (for maximum count), or 100 μl of RPMI 10 (for spontaneous release count) was added to each well. The mixtures of reaction were incubated at 37°C in a humidified 5% CO_2 atmosphere for 4 h as in previous reports (8, 41). A total of 100 μl of supernatant was collected from each well, and γ emission was counted using a gamma counter. The percentage of dead cells was calculated using the following formula: cell death (%) = 100 × (sample count − spontaneous release)/(maximum count − spontaneous release).

Blocking of ADCC by peptide 31

After diluted plasma was added with 0, 10, or 100 μg/ml peptide 31 (=FLKEKGGLE) or 100 μg/ml of control peptide (=GGGGGGGGG), 50 μl of the solution was added to resuspended target cells. ADCC assay was performed as above.

Flow cytometric analysis

For analysis of NK cell subsets, we used the following Ab combinations: 1) FITC-conjugated anti-CD56, PE anti-CD16, PerCP anti-CD19, allophycocyanin anti-CD3; 2) FITC anti-perforin, PE anti-CD56, PE anti-CD16, PerCP anti-CD19, allophycocyanin anti-CD3. For phenotypic analysis of NK cells, PBMCs were suspended in 50 μl of culture medium, and stained with Ab combination 1, for 20 min on ice. After incubation, cells were washed twice with cold PBS. Cells were resuspended in 200 μl of PBS containing 0.5% formaldehyde. For intracellular staining of perforin, cells were stained with Ab combination 2 (without anti-perforin Ab) for 20 min. After incubation, cells were washed twice with cold PBS, and resuspended in 100 μl of PBS. After addition of 100 μl of 4% formaldehyde and incubation for 20 min at room temperature, cells were pelleted and supernatants were removed. Cells were washed once with PBS/0.5% BSA/1 mM of sodium azide (PBS/BSA/azide buffer), and resuspended in 150 μl of permeabilization buffer (PBS/BSA/azide buffer containing 0.5% saponin). After pipetting gently to mix and incubating for 10 min at room temperature, cells were pelleted and supernatant was removed. A total of 25 μl of permeabilization buffer containing the appropriate amount of Abs against intracellular perforin was added to the cell pellets and incubated at room temperature for 30 min in the dark. Cells were washed once with 0.5 ml of permeabilization buffer and once with 1 ml of PBS/BSA/azide buffer. Finally, cells were suspended in 200 μl of PBS/BSA/azide buffer. All samples were kept at 4°C and protected from light until analysis on the flow cytometer.

Six-parameter flow cytometric analysis was done on a FACSCalibur flow cytometer (BD Immunocytometry Systems) using CellQuest software (BD Immunocytometry Systems) with FITC, PE, PerCP, and allophycocyanin as the four fluorescent parameters. FlowJow software (Tree Star, San Carlos, CA) was used to make configurations. Light scatter gates were designed to include only lymphocytes, and up to 100,000 events in this gate were collected. The absolute lymphocyte count was determined from the complete blood count. The number of NK cells per microliter of whole blood was calculated by multiplying the fraction of lymphocytes that were $CD16^+$ or $CD56^+$ by the absolute lymphocyte per microliter of blood. For analysis and display of statistical comparisons, we used JMP software for the Apple Macintosh (SAS Institute, Cary, NC). Comparisons of distributions were performed by the nonparametric two-sample Wilcoxon rank test.

Results

Nef protein expression on the cell surface infected with SeV-Nef

LTNP 5 in the previous study had a high titer of the Abs against peptide 31 (39). When CEM-NK^R-Nef cells fixed with paraformaldehyde were stained with diluted plasma from healthy donor or LTNP 5, and FITC-conjugated anti-human Ig secondary Abs, positive fluorescent signals were given on the surface of CEM-NK^R-Nef cells by plasma from LTNP 5, but not from a healthy donor

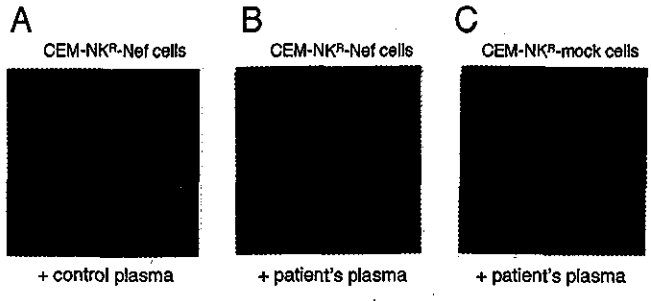

FIGURE 1. Immunological staining of CEM-NK^R cells infected with SeV-Nef (CEM-NK^R-Nef cells). Cells were stained with 1/2.5 diluted plasma and FITC-conjugated anti-human Ig secondary Abs. The stained cells were observed by confocal microscopy. *A*, CEM-NK^R-Nef cells stained with plasma of a healthy donor. *B*, CEM-NK^R-Nef cells stained with plasma from LTNP 5. *C*, CEM-NK^R-mock cells stained with plasma from LTNP 5.

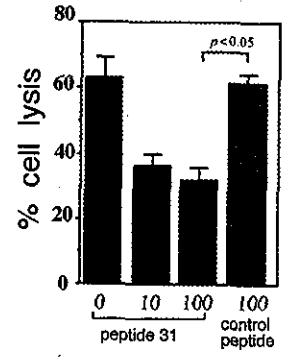

FIGURE 3. Inhibition of ADCC by peptide 31. Percentage of cell lysis by PBMCs of P3 was examined by ADCC assay in the presence of peptide 31 (■) or control peptide (hatched column) at an E:T ratio of 20:1. Data are shown as the mean of triplicate determinations (bars represent SDs). There is a significant difference between peptide 31 and control peptide at the concentration of 100 μg/ml (Student's *t* test, $p < 0.05$).

(Fig. 1, *A* and *B*). Plasma from LTNP 5 did not recognize proteins on the cell surface of CEM-NK^R-mock cell (Fig. 1*C*).

ADCC assay

An ADCC assay was conducted using plasma from LTNPs (LTNP 2, 5, and 6) (39) and PBMCs of either a healthy volunteer or from a patient 1–5 whose profiles are provided in Table I. As shown in Fig. 2*A*, CEM-NK^R-Nef incubated with plasma of LTNP 5 (final concentration, 10^{-1} of original) was efficiently lysed with PBMCs of a healthy volunteer at an E:T ratio of 20:1 (mean percentage of cell lysis, 58%) and 50:1 (66%). When the E:T ratio was lowered to 5:1, percentage of cell lysis decreased to 30% (Fig. 2*A*). The plasmas from LTNP 2, 5, and 6 (final concentration, 10^{-1} of original) induced ADCC activity via Nef, and the plasma of LTNP 6 indicated lower activity compared with that of LTNP 2 or LTNP 5 (Fig. 2*B*). Cytotoxic activity against CEM-NK^R-Nef was observed when PBMCs of five HIV-1-infected patients (p1–5) were used as effector cells at an E:T ratio of 20:1 (Fig. 2*C*). This cytotoxicity was specific to plasma of HIV-1-infected patients, because cell lysis was less than 10% when plasma from a healthy donor was used instead of patient plasma (Fig. 2*C*). In addition, the observation that dilution of patient plasma reduced the percentage of CEM-NK^R-Nef cell lysis (Fig. 2*D*) also suggested that lysis was mediated by the Ab in the plasma. To examine whether the cell lysis is specific to Nef, we added synthetic peptide 31 to the mixture of ^{51}Cr-labeled CEM-NK^R-Nef, PBMCs of patient 3, and LTNP 5 plasma at an E:T ratio of 20:1. Addition of 10 or 100 μg/ml peptide 31 decreased the percentage of cell lysis by 42 or 48% when compared with cell lysis without peptide 31, respectively, whereas addition of 100 μg/ml of control peptide did not show any effect on cytotoxicity (Fig. 3).

NK cells of HIV-1-infected patients

We analyzed NK cells in the peripheral blood using flow cytometry. NK cells were defined as CD3$^-$, CD19$^-$, CD16$^+$, or CD56$^+$ lymphocyte (44). PBMCs from 41 HIV-1-infected patients and 16 healthy donors were examined. There was a significant difference between HIV-1-infected patients and normal controls in total counts of NK cells (mean ± SD = 131 ± 85 and 198 ± 87 cells/μl, respectively, $p = 0.014$) (Fig. 4*A*). When HIV-1-infected individuals were divided into two groups by CD4$^+$ T cell counts (CD4 ≥ 200 or CD4 < 200 cells/μl), there was no significant difference between these two groups in absolute counts of NK cells (CD4 ≥ 200 and CD4 < 200 cells/μl; mean ± SD = 125 ± 94 and 142 ± 82 cells/μl, respectively, $p = 0.643$). For the functional analysis of NK cells, we next examined the expression of intracellular perforin in NK cells of HIV-1-infected patients. As shown in Fig. 4*B*, there was no significant difference between HIV-1-infected patients and healthy controls in frequency of perforin high-positive cell (%) of total NK cells (CD4 ≥ 200, CD4 < 200 cells/μl, and healthy controls; mean ± SD = 83 ± 12, 90 ± 6, and

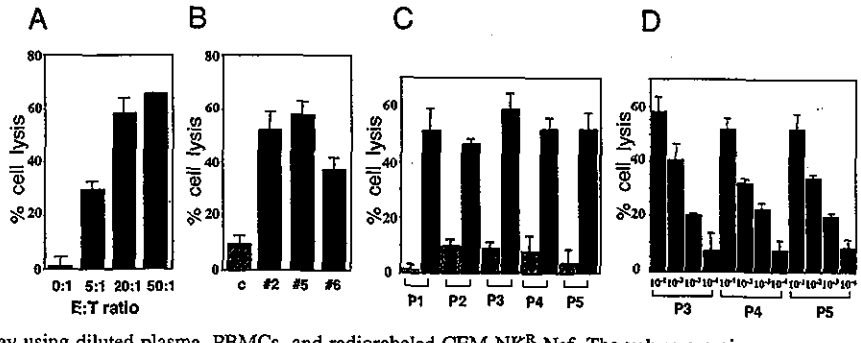

FIGURE 2. ADCC assay using diluted plasma, PBMCs, and radiorabeled CEM-NK^R-Nef. The values are given as percentage of specific cell lysis = 100 × (sample count − spontaneous release)/(maximum count − spontaneous release). *A*, Various E:T ratio with healthy donor PBMCs in the presence of plasma from LTNP 5. *B*, Plasma from a healthy donor (hatched column) or LTNPs (LTNP 2, 5, and 6) (■) at an E:T ratio of 20:1 with healthy donor PBMCs. *C*, PBMCs from five patients (P1-P5, Table I) at an E:T ratio of 20:1 in the presence of either plasma from a healthy donor (hatched column) or LTNP 5 (■) in *C*. *D*, Plasma Ab titration. Percentage of cell lysis by PBMCs from patient P3, P4, or P5 was examined with serially diluted plasma from LTNP 5 at an E:T ratio of 20:1. The values along the *x*-axis represent final concentration, 10^{-1}~10^{-4} of original plasma. Data are shown as the mean of triplicate determinations (bars represent SDs).

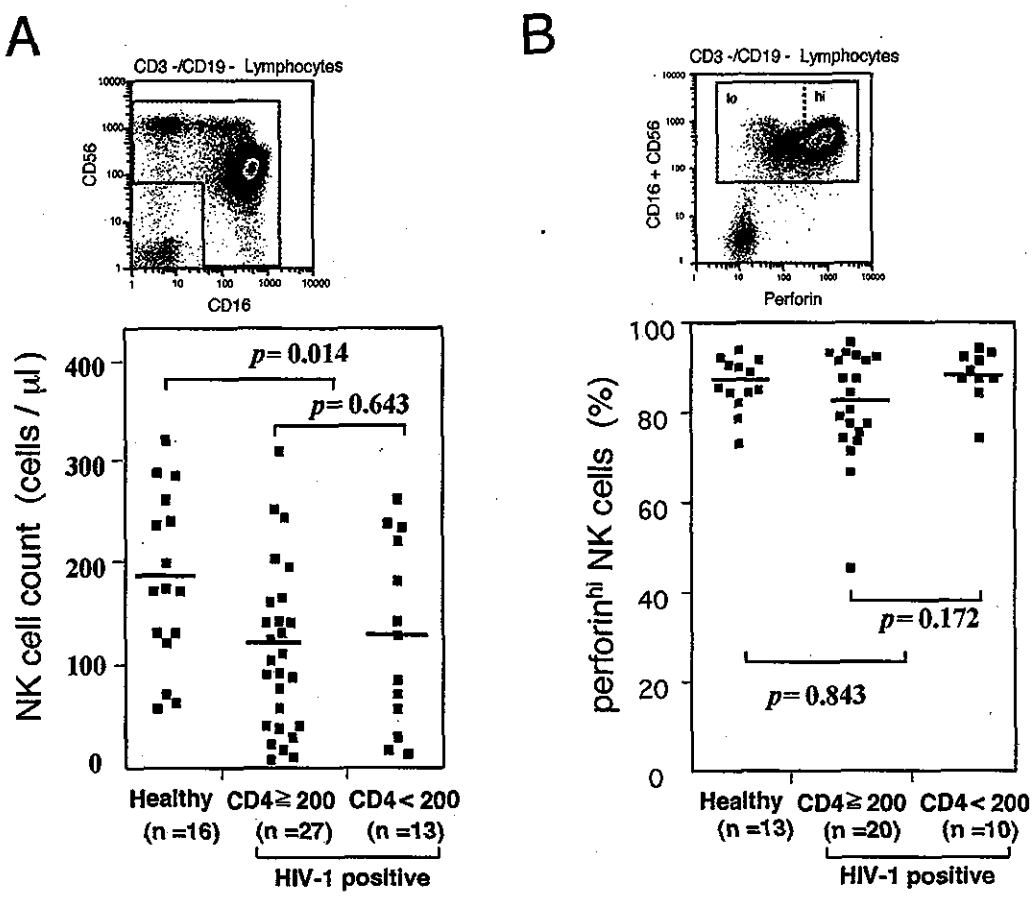

FIGURE 4. Flow cytometric analysis of NK cells. NK cells were defined by $CD3^-$, $CD19^-$, $CD16^+$, or $CD56^+$ expression. *Upper panels*, Show flow cytometry profiles gated on $CD3^-$ and $CD19^-$ lymphocytes. NK cells were gated by red filled line. *A*, *Lower panel*, Comparison of NK cell counts was conducted between 16 healthy donors and 40 HIV-1-positive individuals. *B*, *Upper panel*, NK cells are distingushed between perforin high/positive (hi) and low (lo) populations by red dotted line. *Lower panel*, Frequency of perforin high-positive cells (%) of total NK cells for each donor was calculated. Comparison was conducted between 13 healthy donors and 30 HIV-1-positive individuals. Median values are shown as bars.

88 ± 6%, respectively), suggesting that NK cells in HIV-1-infected patients were as functionally active as those in non-HIV-1-infected individuals.

Discussion

In a previous report, we showed that the progression of disease in HIV-1-infected patients was correlated with Ab titers against peptide 31 (39). In an effort to elucidate the mechanism for this correlation, we studied the role of ADCC against peptide 31 in this study. The interaction between plasma Abs of LTNP 5 and Nef proteins was specific (Fig. 1). We showed that PBMCs from HIV-1-infected donors as well as healthy donors could exert specific ADCC against the cells expressing Nef protein (CEM-NK^R-Nef cells) with patient's plasma even in the face of less than normal NK cell count (Table 1; Fig. 2, *A*, *B*, and *C*). Thus, the ADCC activity may contribute to the elimination of HIV-1-infected cells in vivo. Because ADCC activity is dependent on the titer of plasma Ab (Fig. 2*D*), the lower activity of LTNP 6 (Fig. 2*B*) could be attributed to the lower titer of Ab against Nef epitope compared with LTNP 2 or 5, based on our previous data (39). The ADCC activity was inhibited up to ~50% by peptide 31 compared with control peptide (Fig. 3), suggesting that specific Abs against peptide 31 may contribute substantially to eliminate the HIV-1-infected cells. However, other Nef-derived peptides may also contribute to the residual 50% activity as epitopes we have not yet isolated. It was previously shown that selective down-regulation of MHC class I molecules protects HIV-1-infected cells from CTL and NK cells (45–49). In contrast, ADCC via Abs against the conserved cell surface HIV-1 epitopes such as peptide 31 may be an alternative armor against HIV-1 infection.

Although percentages of NK cells varied in the five patients examined (3.7~12.6%) (Table I), they showed almost the same levels of ADCC activity (Fig. 2*C*). This result may be due to the high E:T ratio that we used in the cytotoxicity assay (Fig. 2*A*); however, it is possible that ADCC activity may be retained until late in the clinical stage, as previously reported (50, 51). Flow cytometric analysis revealed a reduction of total NK cell counts in HIV-1-infected individuals, similar to the previous reports (52, 53) (Fig. 4*A*). There was no significant difference between the two groups of HIV-1-positive patients (CD4 ≥ 200 cells/μl and CD4 < 200 cells/μl); therefore, NK cells appear to be retained even late in the disease progression. With regard to Nef epitope expressing on the cell surface, we previously documented that HIV-1-infected cells were lysed by the combination of rabbit polyclonal Abs against peptide 31 and rabbit complements (39). Thus, we speculate that the level of Nef expression could be sufficient for the induction of ADCC via Nef epitope on the cell surface. However, it could be too difficult to estimate ADCC via Nef epitope with HIV-1-infected cells and patient's plasma because of the existence of abundant anti-envelope Abs as well as anti-Nef Abs in the plasma from HIV-1-infected patient.

We and others showed that HIV-1-specific CD8 T cells contain less perforin (54–56). NK cells may function as better effector cells in the HIV-1-infected individuals. Although the number of

NK cells was lower in HIV-1-infected patients than healthy controls, NK cells retained the high expression of perforin until late in the clinical course (Fig. 4*B*). Rukavina et al. (57) demonstrated that perforin expression significantly correlates with NK cytotoxicity against K562 cells. The fact that LTNPs had higher anti-peptide 31 Abs than progressors may indicate that ADCC against conserved cell surface HIV-1 epitopes such as peptide 31 may have favorable influence on the clinical course. Finally, therapeutic intervention that contributes to raise specific Ab levels against the conserved cell surface HIV-1 epitopes may prove to have a clinical benefit.

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Frequent Transmission of Cytotoxic-T-Lymphocyte Escape Mutants of Human Immunodeficiency Virus Type 1 in the Highly HLA-A24-Positive Japanese Population

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Although Japan is classified as a country with a low prevalence of human immunodeficiency virus type 1 (HIV-1), domestic sexual transmission has been increasing steadily. Because 70% of the Japanese population expresses HLA-A24 (genotype HLA-A*2402), we wished to assess the effect of the dominant HLA type on the evolution and transmission of HIV-1 among the Japanese population. Twenty-three out of 25 A24-positive Japanese patients had a Y-to-F substitution at the second position [Nef138-10(2F)] in an immunodominant A24-restricted CTL epitope in their HIV-1 *nef* gene (Nef138-10). None of 12 A24-negative Japanese hemophiliacs but 9 out of 16 patients infected through unprotected sexual intercourse had Nef138-10(2F) ($P < 0.01$). Two of two A24-positive but none of six A24-negative Australians had Nef138-10(2F). Nef138-10(2F) peptides bound well to the HLA-A*2402 heavy chain; however, Nef138-10(2F) was expressed poorly on the cell surface from the native protein. Thus, HIV-1 with Nef138-10(2F) appears to be a cytotoxic-T-lymphocyte escape mutant and has been transmitted frequently by sexual contact among the highly A24-positive Japanese population.

While cytotoxic T lymphocytes (CTLs) exert immune pressure on human immunodeficiency virus type 1 (HIV-1) throughout the course of primary and chronic infection (4, 24, 30), HIV-1 escapes through a variety of immune evading mechanisms such as downregulation of HLA class I molecules by Nef (7, 32, 33, 36) and defects in differentiation and maturation of CTLs (2, 6, 27, 35). Viral mutation also plays a crucial role in immune escape, and CTL escape mutant viruses may appear early or late in the clinical course of infection (5, 14, 31). Mutations leading to CTL escape may occur at amino acid residues essential for major histocompatibility complex binding (8), for T-cell-receptor recognition (10), or in flanking regions that affect antigen processing (3, 26).

HIV-1 CTL escape mutants may be stable. One such example at the HLA-B27-restricted Gag epitope, which is related to slower disease progression in adults, could be transmitted vertically from mother to child (12). Although significant association between HLA alleles and polymorphism in reverse transcriptase sequences in a large cohort of patients indicated HIV-1 adaptation at a population level (28), direct horizontal transmission of CTL escape mutants is yet to be shown.

Japan is classified as a country of low HIV-1 prevalence; however, national HIV-1 and AIDS surveillance has shown a steady increase of HIV-1 and AIDS cases mainly through unprotected sexual intercourse (USI) (84% of HIV-1 patients and 71% of AIDS patients were infected through USI within the country) (1). The Japanese population is less polymorphic than other populations in that 70% express HLA-A24 (genotype HLA-A*2402) (13). We speculated that stable CTL escape mutants from HLA-A24 might be transmitted more frequently in Japan than in other countries where the prevalence of HLA-A24 is much lower. We postulated that Japanese hemophiliacs with HIV-1 infection might be a good comparator group since they were infected directly by contaminated blood products from abroad. We therefore examined an immunodominant CTL epitope in the *nef* gene (Nef138-10) in HLA-A24-positive and -negative hemophiliacs and compared the sequence with sequences from those patients infected through USI (13, 18). We included Caucasian Australians infected through USI as another control of transmission of CTL escape mutants in a country where HLA-A24 is less prevalent (19).

MATERIALS AND METHODS

Patient samples. For sequence analysis, blood specimens were collected in EDTA. Plasmas were separated and preserved at −80°C until use. For enzyme-linked immunospot (ELISPOT) assay, peripheral blood mononuclear cells (PBMCs) were separated from heparinized whole blood and used on the day of the assay. Patient HLA was typed serologically. In selected patients, HLA genotype was determined after written informed consent was obtained. The study was approved by institutional review boards. All patients serologically typed as A24 positive proved to be positive for HLA-A*2402.

RNA extraction and reverse transcription. Viral RNA was extracted from 140 μl of plasma from patients by using the QIAamp viral RNA Mini kit (QIAGEN) and subjected to reverse transcription according to the manufacturer's protocol with SuperScript II RNase H⁻ reverse transcriptase (Invitrogen) and 5 μM random primers (Takara).

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PCR amplification and sequencing. Fifteen microliters of cDNA (a one-sixth volume of the reverse transcription reaction) was subjected to the first PCR. One-tenth of the first PCR was subjected to the nested PCR. PCR was performed by using Ex-Taq (Takara) with 35 cycles of 30 s at 94°C, 30 s at 58°C, 30 s at 72°C, and a final extension for 7 min at 72°C. The primer sets are as follows (all nucleotide positions are in accordance with the HIV-1 SF2 strain). For the *env* V3 region, first PCR primer set 1, primers CBE297P (5′-GGTAGAACAG ATGCATGAGGAT-3′) (consensus B *env*, nucleotides [nt] 297 to 318) and E7668 M (5′-TTCTCCAATTGTCCCTCATATCTCCTCCTCCA-3′) (SF2, nt 7668 to 7636) were used; and for the second PCR primer set 1, primers E6554P (5′-ATCAGTTTATGGGATCAAAGCC-3′) (SF2, nt 6554 to 6575) and E7353 M (5′-ACAATTTCTGGGTCCCCTCCTGAGGA-3′) (SF2, nt 7353 to 7328) were used. For the first PCR primer set 2, primers E6984P (5′-ACATGGAAT TAGGCCA-3′) (SF2, nt 6984 to 7000) and E7395 M (5′-TTACAGTAGAAA AATTCCCC-3′) (SF2, nt 7395 to 7375) were used; and for the second PCR primer set 2, primers E7028P (5′-GGCAGTCTAGCAGAAGAAGA-3′) (SF2, nt 7028 to 7047) and E7353 M (5′-ACAATTTCTGGGTCCCCTCCTGAGGA-3′) (SF2, nt 7353 to 7328) were used. For the first PCR primer set 3, primers P6951 (5′-GACCATGTACAAATGTCAGC-3′) (SF2, nt 6951 to 6970) and M7592 (5′-CTCTTGTTAATAGCAGCCCT-3′) (SF2, nt 7592 to 7573) were used; and for the second PCR primer set 3, primers E6984P (5′-ACATGGAA TTAGGCCA-3′) (SF2, nt 6984 to 7000) and E7353 M (5′-ACAATTTCTGGG TCCCCTCCTGAGGA-3′) (SF2, nt 7353 to 7328) were used.

For the Nef138-10 epitope, first PCR primer set 1, primers n226p (5′-CTCA GGTACCTTTAAGACCAATG-3′) (nt 9028 to 9050) and n650m (5′-GAAAG TCCCCAGCGGAAAGTCCC-3′) (nt 9474 to 9452) were used; and for the second PCR primer set 1, primers n296p (5′-GGGACTGGAAGGGCTAATT TGGT-3′) (nt 9098 to 9120) and n564m (5′-GAAATGCTAGTTTGCTGTCA AAC-3′) (nt 9387 to 9365) were used. For the first PCR primer set 2, primers P8923 (5′-TGGAAAAACATGGAGCAATCA-3′) (nt 8923 to 8944) and M9290 (5′-TCCTTCATTGGCCTCTTCTAC-3′) (nt 9290 to 9270) were used; and for the second PCR primer set 2, primers P8924 (5′-GGAAAAACATGGAGCAA TCAC-3′) (nt 8924 to 8945) and M9288 (5′-CTTCATTGGCCTCTTCTACCT-3′) (nt 9288 to 9268) were used. For the first PCR primer set 3, primers P8923 (5′-TGGAAAAACATGGAGCAATCA-3′) (nt 8923 to 8944) and n694m (5′-C AGCATCTGAGGGACGCCAC-3′) (nt 9525 to 9506) were used; and for the second PCR primer set 3, primers n226p (5′-CTCAGGTACCTTTAAGACCA ATG-3′) (nt 9028 to 9050) and n532m (5′-TCTCCGCGTCCTCCATCCCA-3′) (nt 9345 to 9326) were used.

The PCR products were electrophoresed through agarose gels and purified with a Minielute gel extraction kit (QIAGEN) before sequencing. Purified PCR products were directly sequenced. When sequence ambiguities resulted, DNA fragments were subcloned into the pGEM-T vector (Promega) and sequenced. DNA sequencing was performed by using an ABI Prism dye terminator cycle sequencing ready reaction kit (Applied Biosystems) on a Perkin-Elmer ABI-377 sequencer.

Cells and media. T2-A24, a kind gift from K. Kuzushima, was cultured in RPMI 1640 (Sigma) supplemented with 10% heat-inactivated fetal calf serum (FCS) (Sigma) and 0.8 mg of G418 (Invitrogen)/ml (25). We transformed PBMCs from an HLA-A*2402-positive person with human T-cell leukemia virus type 1 (HTLV-1) and established an HLA-A*2402- and CD4-positive-T-cell line (KWN-T4). KWN-T4 was cultured with RPMI 1640 supplemented with 25 U of interleukin-2 (Wako)/ml, 100 U of penicillin/ml, 100 U of streptomycin (Invitrogen)/ml, and 10% heat-inactivated FCS (JRH Bioscience). We also established Nef138-10-specific CTL clones as previously described (22). CTL clones were cultured with RPMI 1640 supplemented with 50 U of interleukin-2/ml, 100 U of penicillin/ml, 100 U of streptomycin/ml, and 10% heat-inactivated FCS.

Peptides. Synthetic peptides Nef138-10 (RYPLTFGWCF), 2F (RFPLTFGW CF), 5C (RYPLCFGWCF), and 2F5C (RFPLCFGWCF) were purchased from Sigma-Genosys. All peptides were more than 95% pure as determined by high-performance liquid chromatography and mass spectroscopy.

Peptide binding assays. Peptide binding to HLA-A*2402 was quantified by using a T2-A24 stabilization assay as previously described (25). T2-A24 cells were incubated at 26°C for 16 h, and then 2×10^5 cells were incubated with peptides at concentrations from 10^{-4} to 10^{-9} M for 1 h at 4°C. After incubation for 3 h at 37°C, the cells were stained with anti-HLA-A24 monoclonal antibody, A11.1 M (11), and an R-phycoerythrin (RPE)-conjugated $F(ab')_2$ fragment of anti-mouse immunoglobulin (DAKO). The mean fluorescence intensity was measured by FACSCalibur (Becton Dickinson).

ELISPOT assay and functional avidity analysis. Freshly prepared PBMCs (20,000 to 50,000) were added to 96-well multiscreen plates (Millipore) which had been precoated with 100 μl of 5 μg of anti-gamma interferon (IFN-γ) monoclonal antibody 1-D1K (Mabtech)/ml at room temperature for 3 h and blocked with RPMI 1640 medium containing 10% FCS for 1 h. The cells were cultured with synthetic peptide Nef138-10 or its derivatives at concentrations from 10^{-5} to 10^{-11} M for 18 h. After the plates were washed, 100 μl of 1 μg of biotinylated anti-IFN-γ monoclonal antibody 7-B6-1 (Mabtech)/ml was added and incubated at room temperature for 90 min. After the plates were washed again, 100 μl of 1:1,000-diluted streptavidin-alkaline phosphatase conjugate (Mabtech) was added and incubated at room temperature for 60 min. Spots were developed with an alkaline phosphatase conjugate substrate kit (Bio-Rad) and counted with a KS ELISPOT compact (Carl Zeiss). The IFN-γ responses to peptide dilutions were expressed as a percentage of the maximal IFN-γ response seen in each individual assay.

Expression of recombinant Nef protein. Mutations were introduced into *nef* derived from HIV-1 strain SF2 by site-directed mutagenesis based on overlap extension (16). Four proline residues in the Nef proline-rich domain that are important for HLA class I down-regulation were replaced by alanine as described previously (36). The wild type and various *nef* mutants were tagged by His_6 and introduced into a Sendai virus vector (SeV) as previously described (36). For Western blot analysis, KWN-T4 cells were infected with various SeVs at a multiplicity of infection of 10 and lysed 20 h after infection. Western blot analysis was performed according to the standard procedure. Anti-His_6 antibody (QIAGEN) and anti-SeV mouse antiserum were used to detect Nef and SeV proteins, respectively.

^{51}Cr release assay. Cytotoxicity was measured with a standard ^{51}Cr release assay as previously described (21). Briefly, KWN-T4 was labeled with 100 μCi of $Na_2{}^{51}CrO_4$ for 2 h and washed three times with R10. Labeled cells (2×10^3) were added to a 96-well round-bottom microtiter plate with a corresponding amount of peptide. After 1 h of incubation, Nef138-10-specific CTL clones were added and incubated for 4 h. When SeV-infected cells were used as target cells, the cells were infected with SeVs at a multiplicity of infection of 10, 20 h before adding the CTLs.

The supernatants were collected and analyzed with a microbeta counter. Spontaneous ^{51}Cr release was determined by measuring counts per minute in the supernatant of wells containing only target cells (cpm_{spn}). The maximum release (cpm_{max}) was determined by measuring the release of ^{51}Cr from target cells in the presence of 2% Triton X-100. Specific lysis was determined as follows: specific lysis = $(cpm_{exp} - cpm_{spn})/(cpm_{max} - cpm_{spn}) \times 100$, where cpm_{exp} represents the counts per minute in the supernatant of wells containing target and effector cells.

RESULTS

Sexual transmission of HIV-1 with stereotypic amino acid substitution among the Japanese population. Only patients infected with virus subtyped as B by phylogenic comparison of envelope sequences were included to avoid potential bias introduced by sequence differences across subtypes (data not shown). We extensively sequenced the Nef138-10 epitope and its flanking region from plasma HIV-1 RNA of 23 Japanese hemophiliacs (11 A24-positive and 12 A24-negative individuals) and 30 Japanese (14 A24-positive and 16 A24-negative individuals) and 8 Caucasian Australians (2 A24-positive and 6 A24-negative individuals) infected through USI (Table 1). Ten out of 11 A24-positive but none of A24-negative Japanese hemophiliacs had a Y-to-F amino acid substitution at the second position [Nef138-10(2F)] (Fig. 1A) ($P < 0.01$), suggesting that HLA-A24 selected for Nef138-10(2F). In the case of patients infected through USI, 13 out of 14 A24-positive and 9 out of 16 A24-negative Japanese patients had Nef138-10(2F) by direct sequencing (Fig. 1B) (data not significant). The frequency of Nef138-10(2F) was significantly higher in Japanese A24-negative patients infected through USI than A24-negative hemophiliacs ($P < 0.01$). Two out of two A24-positive but none of six A24-negative Caucasian Australians had Nef138-10(2F) (Fig. 1C). The frequency of Nef138-10(2F) in A24-negative patients infected through USI was significantly higher for Japanese patients than for Australian patients ($P < 0.05$), suggesting that sexual transmission of the variant was more

TABLE 1. Patient profile[a]

Patient ID	Sex	HLA type	No. of CD4 cells/μl	Viral load (copies/ml)	Sample date (mo/day/yr)	HIV subtype
A24-positive Japanese hemophiliacs						
A24-J037	M	A24/26, B35/51	207	180,000[b]	03/09/95	B
A24-J041	M	A24/26, B44/61	261	7,500[bd]	03/09/95	B
A24-J033	M	A24/26, B46/52	27	200,000[b]	03/27/95	B
A24-J035	M	A24, B40/48	148	360,000	04/10/95	B
A24-J031	M	A24/31, B51/60	29	180,000[b]	10/23/95	B
A24-J030	M	A11/24, B13/62	3	380,000[bd]	02/26/96	B
A24-J029	M	A24/31, B35/61	38	ND	04/01/96	B
A24-J036	M	A2 /24, B35/51	60	74,000[b]	05/13/96	B
A24-J034	M	A24, B46/52	180	74,000[bd]	05/20/96	B
A24-J038	M	A2 /24, B51/62	356	29,000[b]	03/03/97	B
A24-J005	M	A24, B52/70	39	220,000[b]	06/19/97	B
A24-negative Japanese hemophiliacs						
NA24-J037	M	A26, B40	8	>1,600,000[bd]	06/08/95	B
NA24-J035	M	A11/26, B54/56	342	100,000[b]	09/07/95	B
NA24-J031	M	A2/26, B51/61	521	130,000[b]	09/18/95	B
NA24-J041	M	A26, B39/54	12	700,000[bd]	10/05/95	B
NA24-J032	M	A2/11, B46/54	1[d]	150,000[b]	11/10/95	ND
NA24-J030	M	A31/33, B44/51	363	65,000[b]	03/21/96	B
NA24-J040	M	A2/33, B17/54	101	74,000[b]	03/21/96	ND
NA24-J033	M	A26, B61	143	140,000[b]	04/18/96	B
NA24-J029	M	A11/33, B44/51	401	<10,000	07/15/96	B
NA24-J034	M	A11/33, B17/56	38	81,000[b]	08/15/96	B
NA24-J039	M	A11/26, B51/62	3	88,000[b]	09/01/97	B
NA24-J006	M	A2/26, B39/61	335	9,200	10/30/00	B
A24-positive Japanese infected through USI						
A24-J006	M	A2/24, B7/54	212	33,000	09/19/97	B
A24-J007	M	A24/26, B17/56	103	120,000	11/06/97	B
A24-J009	M	A24, B48/52	278	4,500	01/19/98	B
A24-J010	M	A24, B52	393	18,000	03/09/98	B
A24-J024	M	A24, B35/61	274	110,000	10/27/98	B
A24-J012	M	A24/26, B46/60	253	24,000	07/19/99	B
A24-J013	M	A24/26, B35/48	168	15,000	9/20/99	B
A24-J016	M	A11/24, B7/55	245	150,000	05/15/00	B
A24-J017	M	A1/24, B54/70	255	70,000	10/17/00	B
A24-J018	M	A24/31, B37/61	185	8,300	01/04/01	B
A24-J025	M	A24, B51/52	282	130,000	06/07/01	B
A24-J023	M	A2/24, B51/54	856[d]	17,000[d]	08/06/01	B
A24-J021	M	A2/24, B46/52	344	35,000	11/26/01	B
A24-J026	M	A2/24, B13/51	381	110,000	11/28/01	B
A24-negative Japanese infected through USI						
NA24-J025	M	A2/31, B51/61	352	18,000[b]	03/23/95	B
NA24-J023	M	A11/26, B35/51	23	5,000[b]	04/01/96	ND
NA24-J021	M	A26, B52/54	9	44,000	08/04/97	B
NA24-J018	M	A2, B39/60	378	72,000	04/06/98	B
NA24-J017	M	A11/31, B51/56	197	72,000	04/16/98	B
NA24-J016	M	A3/31, B51/58	257	200,000	05/25/98	B
NA24-J015	M	A2/26, B51/62	543	13,000	06/26/98	B
NA24-J012	M	A31, B13/51	268	26,000	10/19/98	B
NA24-J011	M	A2, B55/60	408	12,000	10/22/98	B
NA24-J010	M	A2/26, B51/61	206	16,000	12/17/98	B
NA24-J009	M	A2, B52/60	115	850,000	05/24/99	B
NA24-J008	M	A11/33, B44/60	312	2,600	07/08/99	ND
NA24-J007	M	A26, B7/52	396	450	08/09/00	B
NA24-J005	M	A2/31, B48/52	604	17,000	01/18/01	B
NA24-J003	M	A31/33, B44/51	308	20,000	06/04/01	B
NA24-J002	M	A2/33, B44/46	496	14,000	09/27/01	ND
A24 positive Australian infected through USI						
A24-A001	M	A3/24, B7	255	38,000	08/16/96	ND
A24-A002	M	A24/30, B13	598	21,700	03/22/01	B
A24-negative Australian infected through USI						
NA24-A007	M	A2/3, B7	704	ND[c]	11/02/95	B
NA24-A005	M	A1/3, B8/70	620	7,700	05/26/96	B
NA24-A013	M	A32, B13/64	851	23,200	09/28/98	B
NA24-A008	M	A2/3, B39/44	543	52,836	01/04/99	B
NA24-A003	M	A2, B18/62	575	19,400	11/06/99	B
NA24-A006	M	A3/26, B18/27	594	18,200	04/13/00	B

[a] ND, not determined.
[b] Data were obtained by Branch DNATM version 1.0.
[c] Nearest data were 17,000 with CD4 counts of 638.
[d] Nearest data were within 6 months of sample collection.

A

A24-positive Japanese hemophiliacs

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV
A24-J041	----------V	-F----------	----------M
A24-J033	-----E---T	-F--------Y	--------D--
A24-J031	-H--------T	-F----------	-
A24-J030	----------T	-F--C------	----------
A24-J034	----------T	-F----------	-----DQ-Q-
A24-J038	----------	----C------	-----D-D--
A24-J005	-D/E--------T	-F----------	-
A24-J029	----------V/T	-F----------	--------Q-
A24-J037	--C-------T	-F----------	-----D----
A24-J035	----------T	-F----------	----------
A24-J036	--C-------T	-F----------	----------

A24-negative Japanese hemophiliacs

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV
NA24-J037	----------	----------	----------
NA24-J035	----------	----------	----M-----
NA24-J031	----------	----------	--------G/E-V/I
NA24-J041	----------	----------	--------DE
NA24-J032	----------	----------	----M-----
NA24-J030	--S-------V	----C-----	----------
NA24-J040	----------	----------	---------I
NA24-J033	----------	----------	-L/V--------
NA24-J029	-H--------	----------	--------D-
NA24-J034	----------	----------	----V/L-----
NA24-J039	----------	----C-----	-----D-D--
NA24-J006	----------V	----C-----	-----D----

B

A24-positive Japanese infected through USI

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV
A24-J006	----------V	-F----------	-----E/D--Q-
A24-J007	----------T	-F--C------	----A---E-
A24-J009	----------T	-F----------	-
A24-J010	----------T	-F----------	--------QR-
A24-J012	----------T	-F----------	--------D-
A24-J013	----------T	-F----------	-----D-DQ-
A24-J016	-D--------V	----C------	-----DQD--
A24-J017	-D--------T	-F--C------	----------I
A24-J018	----------T	-F----------	----------I
A24-J023	----------T	-F----------	----L--GEA
A24-J021	----------T	-F----------	-----D-DQ-
A24-J024	----------T	-F----------	-----D-D--
A24-J025	-D--------T	-F----------	-----DQDQ-
A24-J026	----------T	-F----------	--------KQ-

A24-negative Japanese infected through USI

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV	
NA24-J025	-H--------V	----C------	-----D-D/AQ-	
NA24-J023*	----------T	-Y/W/F---------	----------I	Y 2/11,F 3/11,W 5/11
NA24-J021	----------	----------	-----N--Q-	
NA24-J018*	----------T	-Y/F--C------	----------	Y 8/10,F 2/10
NA24-J017*	----------T	-Y/F---------	----L-----	Y 5/13,F 8/13
NA24-J016	----------V	----------	----L---Q-	
NA24-J015	----------T	-F----------	-----D-DQ-	
NA24-J012	-H/QS------T	----------	-----D-DQ-	
NA24-J011	----------T	-F----------	--------NQ-	
NA24-J010	-	----------	-	
NA24-J009	----------T	-F----------	--------NQ-	
NA24-J008	-D--------T	-F----------	----L---Q-	
NA24-J007	----------T	-F----------	--------NQ-	
NA24-J005	-G/D--------T	-F----------	-----DQDQ-	
NA24-J003	-H---------	----------	-----DQ---	
NA24-J002	-Q/HG-------	----------	-----D-DQ-	

C

A24-positive Australian infected through USI

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV
A24-A001	----------T	-F----------	----------
A24-A002	----------T	-F----------	----M-----

A24-negative Australian infected through USI

Patient ID	flanking WQNYTPGPGI	CTL epitope RYPLTFGWCF	flanking KLVPVEPEKV
NA24-A007	----------V	----------	-
NA24-A005	----------V	----------	----------
NA24-A013	----------	----------	----------
NA24-A008	-H---------	----------	----M-P/Q---
NA24-A003	-H---------	----------	-----D-D--
NA24-A006	----------	----C-----	--------E-

FIG. 1. Nef138-10 epitope and its flanking sequences. Amino acid sequences deduced from the direct DNA sequencing of Nef138-10 CTL epitope and both flanking regions are presented. Wild-type sequences (HIV-1 strain SF2) are presented on the top. Dashes indicate the same amino acid as that of the wild type. Sequence substitutions are presented by single amino acid characters. Where a mixture of two or three amino acids was plausible, two or three amino acids were shown together separated by a shill. A single dash indicates that the sequences could not be determined by ambiguities. (A) Sequences from A24-positive and -negative Japanese hemophiliacs. (B) Sequences from A24-positive and -negative Japanese patients infected through USI. Asterisks indicate samples for which sequence ambiguities were found by direct sequence analysis. We cloned these PCR fragments into the pGEM-T vector and sequenced each 10 to 13 colonies. All amino acid sequences are indicated. (C) Sequences from A24-positive and -negative Australians infected through USI.

frequent among the Japanese population, which has a higher prevalence of HLA-A*2402.

Nef138-10(2F) accompanied a particular amino acid substitution in the N-terminal flanking region. We detected an I-to-T substitution at the −1 position (−1T) in 32 flanking sequences out of 34 accompanying Nef138-10(2F) sequences (94%), while others were two I-to-V substitutions (Fig. 1). The −1T substitution was quite unusual in the flanking region of the wild-type Nef138-10 CTL epitope in our cohort (Fig. 1).

Reversion of CTL escape mutants. Since three acutely infected A24-positive patients (A24-J023, A24-J024, and A24-J025) had Nef138-10(2F) in their earliest plasma samples available, we could not demonstrate the evolution of Nef138-10(2F) from the wild type under the selective pressure of HLA-A*2402 (data not shown). However, we could analyze serial samples from chronically infected A24-negative patients who had been followed without treatment over years. All the 12 cloned sequences in the earliest plasma samples available from NA24-J015 had F at the second position; however, 11 out of 11 clones displayed wild-type sequence within a year (Fig. 2A). It is interesting that the −1T substitution in the flanking region was present for at least a further two years before reverting to the wild type (Fig. 2A). In another chronically infected A24-negative patient (NA24-J018), we observed that the proportion of Nef138-10(2F) decreased gradually but persisted in the plasma for almost two years after the start of the follow-up (Fig. 2B). This patient had a T-to-C substitution at the fifth position with [Nef138-10(2F5C)] or without [Nef138-10(5C)] a substitution at the second position (Fig. 2B). Interestingly, the ratio of Nef138-10(2F5C) to Nef138-10(5C) decreased as time went by (Fig. 2B), suggesting that Nef138-10(5C) is more stable than Nef138-10(2F5C). Actually, we observed Nef138-10(5C) in both A24-positive and -negative patients (Fig. 1).

In order to elucidate the higher stability of the 5C rather than the 2F substitution, we examined the codon usage at these positions (data not shown). The wild-type codon for the second tyrosine (Y) residue in Nef138-10 was coded by TAT or TAC in 23 (77%) and 12 (40%) out of 30 patients, respectively. Five patients (17%) had a mixture of TAT and TAC for the codon (data not shown). Mutated nucleotide triplet TTT or TTC was responsible for the Y-to-F amino acid substitution in 27 (80%) and 9 (26%) out of 34 patients, respectively (data not shown).

A

	Sample Date		nef138-10		
Patient ID	M/D/Y	WQNYTPGPGI	RYPLTFGWCF	KLVPVEPEKV	cloning
NA24-J015	06/26/98	---------T	-F--------	-----D-DQ-	direct
		---------T	-F--------	-----D-DQ-	11/12
		---------T	-F-----R--	-----D-DQ-	1/12
NA24-J015	06/07/99	---------T	-Y--------	-----D-DQ-	direct
		---------T	-Y--------	-----D-DQ-	9/11
		-R-------T	-Y--------	-----D-DQ-	1/11
		---------T	-Y--------	--I--D-DQ-	1/11
NA24-J015	03/09/00	---------T	-Y--------	-----D-DQ-	direct
NA24-J015	04/16/01	---------T	-Y--------	-----D-DQ-	direct
NA24-J015	01/16/03	----------	-Y--------	-----D-DQ-	direct
		----------	-Y--------	-----D-DQ-	10/10

B

	Sample Date		nef138-10		
Patient ID	M/D/Y	WQNYTPGPGI	RYPLTFGWCF	KLVPVEPEKV	cloning
NA24-J018	04/08/96	---------T	-F--C-----	--------Q-	direct
		---------T	-F--C-----	--------Q-	7/11
		---------P	-F--C-----	--------Q-	3/11
		---------P	-Y--C-----	--------Q-	1/11
NA24-J018	06/02/97	---------T	-F/Y--C-----	--------Q-	direct
		---------T	-F--C-----	--------Q-	7/13
		---------T	-Y--C-----	--------Q-	3/13
		---------A	-Y--C-----	--------Q-	1/13
NA24-J018	04/06/98	---------T	-Y/F--C-----	----------	direct
		---------T	-Y--C-----	--------Q-	6/10
		---------T	-F--C-----	--------Q-	2/10
		---------T	-Y--C-----	----------	2/10

FIG. 2. Serial Nef138-10 epitope and its flanking sequences in two A24-negative patients. Data are shown as described in the legend to Fig. 1. Fractional numbers in the right-most column indicate clone numbers with the sequences shown in the numerator and total clone numbers sequenced shown in denominator. "Direct" indicates the result of direct sequencing. (A) Patient NA24-Jo15. (B) Patient NA24-J018.

In two patients (6%) Nef138-10(2F) was coded by a mixture of HIV-1 using TTT and TTC for the codon. It appeared that at least one point mutation was necessary for the Y-to-F amino acid substitution. The wild-type codon for the fifth threonine (T) residue in Nef138-10 was coded by ACC or ACT in 49 (98%) and 2 (4%) out of 50 patients. One patient (2%) had a mixture of ACC and ACT. Mutated nucleotide triplet TGC or TGT was responsible for the T-to-C amino acid substitution in 5 (45%) and 6 (55%) out of 11 patients, respectively (data not shown). It appeared that at least two nucleotides had to be mutated for the T-to-C amino acid substitution, although we could not exclude the possibility of a three-nucleotide mutation for the amino acid substitution. Therefore, a Y-to-F amino acid substitution, or vice versa, at the second position required less nucleotide mutations than did the T-to-C substitution at the fifth position.

Peptide-based analysis of Nef138-10 and its variants. We measured the binding affinities of Nef138-10 and its variants to HLA-A*2402 (Fig. 3). Although a Y-to-F substitution occurred at the amino acid crucial for peptide affinity with the binding groove, Nef138-10(2F) binding to the HLA-A*2402 heavy chain was essentially preserved. However, the acquisition of a T-to-C substitution at the fifth position, such as Nef138-10(2F5C) and Nef138-10(5C), greatly reduced the binding affinity (Fig. 3). A functional avidity assay in which PBMCs from five patients were used confirmed the results of the binding assay (Fig. 4). Namely, the patients' PBMCs recognized Nef138-10(2F) at a very low concentration (one-half maximum response <1 nM) and had equivalent or even higher functional avidity than did the wild-type peptide. On the contrary, patients' PBMCs showed very low functional avidity against Nef138-10(2F5C) and Nef138-10(5C) (one-half maximum response >100 nM).

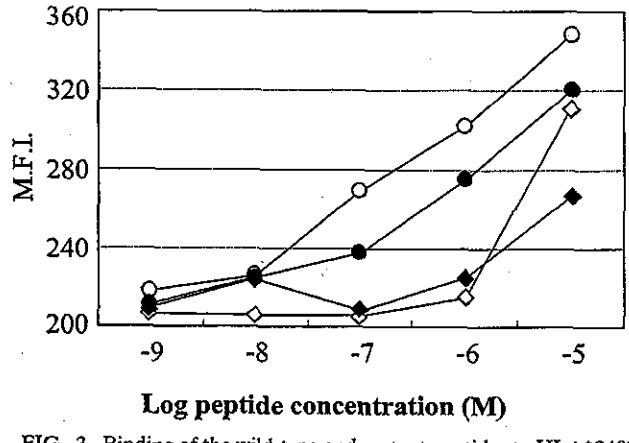

FIG. 3. Binding of the wild-type and mutant peptides to HLA*2402 molecules. Peptide binding to HLA-A*2402 was quantified by using a T2-A24 stabilization assay. Symbols: ○, wild type; ●, 2F; ◇, 5C; ◆, 2F5C. M.F.I., mean fluorescence intensity.

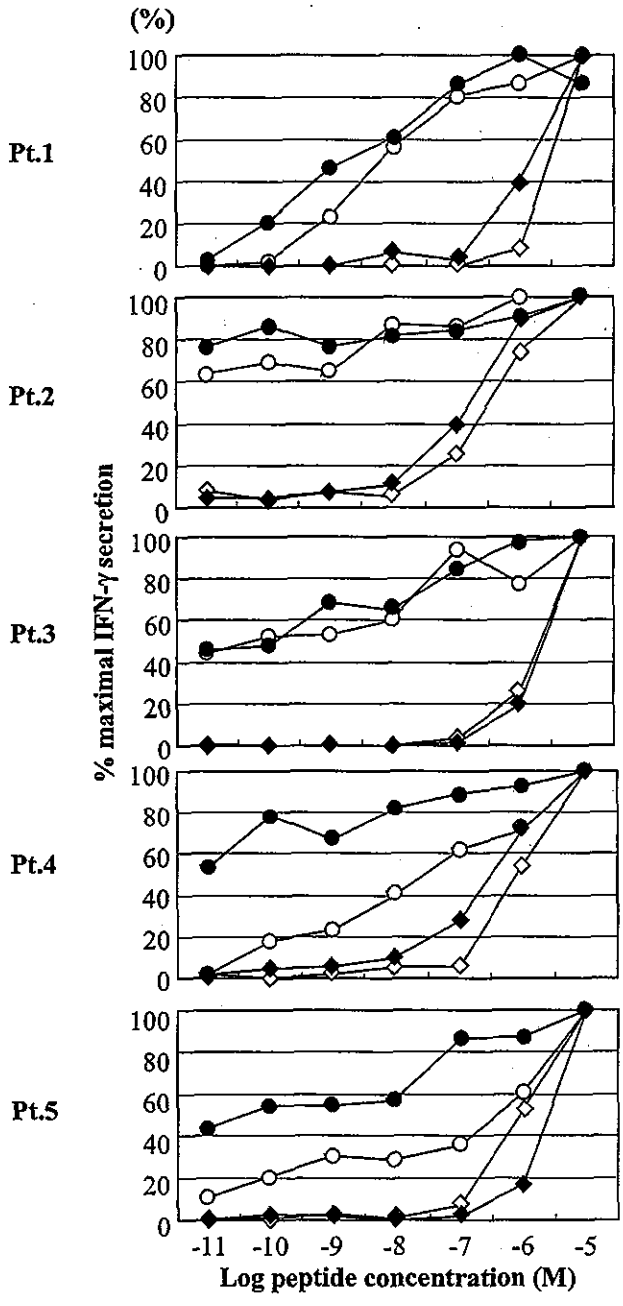

FIG. 4. Functional avidity assay. The reactivity of peptide-specific cells in PBMCs from five patients against log-fold dilutions of peptide was determined. Symbols: ○, wild type. ●, 2F; ◇, 5C; ◆, 2F5C.

Epitope presentation from native Nef protein. Strong selection for Nef138-10(2F) in the presence of CTLs with high in vivo functional avidity against the peptide prompted us to examine the processing and presentation of the Nef138-10 CTL epitope from the native protein. Native Nef proteins containing wild-type or variant CTL epitopes were expressed in an HLA-A*2402-positive-T-cell line (KWN-T4) via SeV. CTL epitope presentation was examined by two CTL clones established from A24-positive patients outside these cohorts. Although the two CTL clones were established by stimulation with the wild-type peptide (Nef138-10), they killed the target cells pulsed with Nef138-10(2F) peptides almost as well as the wild type (Fig. 5A and B). Both CTL clones efficiently killed the target cells expressing either wild-type Nef or Nef with −1T substitution in the flanking region (−1T2Y5T). However, these CTL clones failed to kill the target cells infected with vectors expressing Nef138-10(2F) with (−1T2F5T) or without (−1I2F5T) the −1T substitution in the flanking region. As expected, the CTL clones did not kill the target cells infected with a vector coding Nef138-10(2F5C), a nonbinding mutant (−1I2F5C) (Fig. 5A and B). Western blot analysis revealed that Nef proteins with wild-type or variant CTL epitopes were expressed abundantly in the target cells. Taken together, these data indicate that a Y-to-F substitution within the CTL epitope itself but not the −1T substitution in the flanking region resulted in the poor antigen presentation against CTL, which resulted in the escape.

DISCUSSION

We showed a significantly higher prevalence of a stereotypic amino acid substitution [Nef138-10(2F)] at an A24-restricted CTL epitope in Nef among A24-positive Japanese hemophiliacs compared with A24-negative counterparts. The origin of their HIV-1 infection was from the plasma collected and processed in Western countries where HLA-A*2402 was less prevalent (19). It is inferred that Nef138-10(2F) might be rare in a population where HLA-A*2402 is not prevalent but that it has a selective advantage in the presence of HLA-A*2402. Our findings with Australians are consistent with this notion. Although we examined only two HIV-1-infected A24-positive Caucasian Australians, both had Nef138-10(2F). On the other hand, Nef138-10(2F) was rare in A24-negative Australians. Japanese and Australians are distinctly different in the frequency of HLA-A*2402 within their respective populations (allele frequency of HLA-A24 is 35.1 and 7.8%, respectively) (19). Nef138-10(2F) was also positively selected among Japanese patients who were infected through USI. Interestingly, we detected Nef138-10(2F) frequently among A24-negative Japanese who were infected through USI. The result suggests that HIV-1 that went through selective pressure by HLA-A*2402 is actually circulating among the Japanese population because of the high prevalence of HLA-A24. Although we showed the reversion of Nef138-10(2F) to the wild type, it occurred very slowly over years, allowing the horizontal spread via sexual contact. In this study, we showed that HIV-1 with Nef138-10(2F) is actually a CTL escape mutant. Although the stereotypic Y-to-F substitution occurred at an anchor residue, Nef138-10(2F) peptide did bind to HLA-A*2402 heavy chain with almost the same efficiency as did the wild type (Fig. 3). This result is consistent with the algorithm prediction of the published binding motif (http://hiv-web.lanl.gov/content/immunology/motif_scan/motif.html). When native Nef proteins with or without a substitution were overexpressed in the A24-positive target cells via SeV, the Y-to-F substitution at the second position of the CTL epitope virtually abolished the killing by the CTL clones. The substitution in the flanking region did not affect the killing substantially. Therefore, the mechanism for the CTL escape appeared to reside in the processing of native Nef proteins and subsequent antigen presentation rather than HLA binding. A proteosomal cleavage

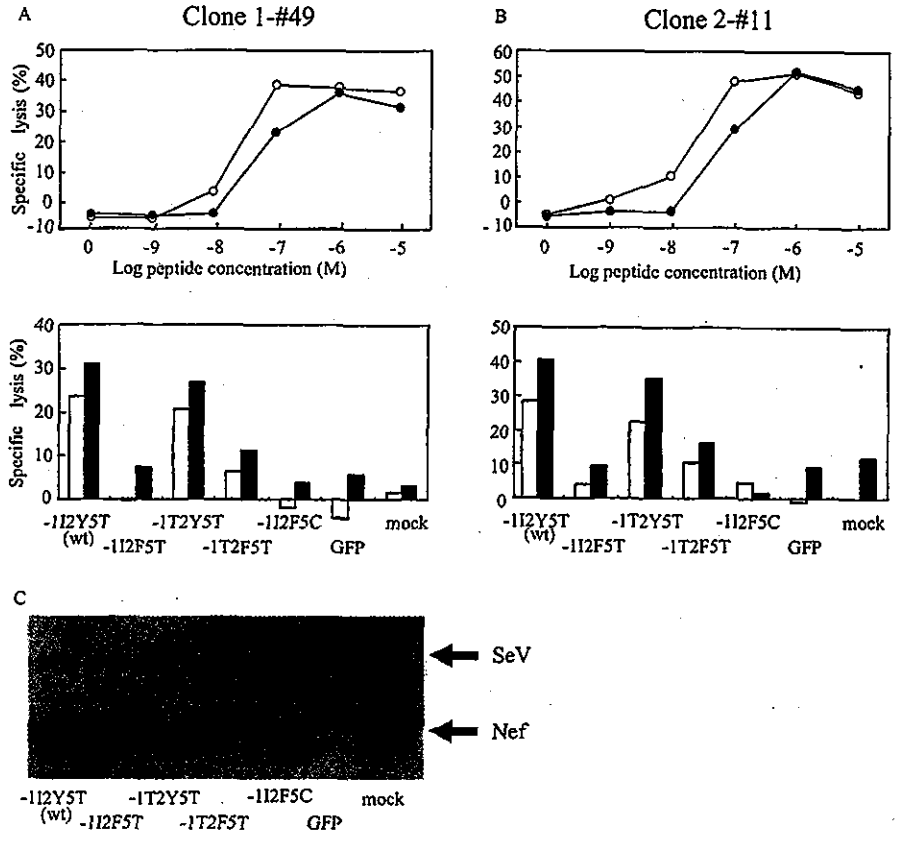

FIG. 5. Killing activity of clone 1-#49 (A) against KWN-T4 target cells pulsed with log-fold dilutions of peptide (top) and expressing native Nef proteins containing wild-type sequences (−1I2Y5T), a Y-to-F substitution at the second position of the CTL epitope (−1I2F5T), an I-to-T substitution at the −1 flanking position (−1T2Y5T), double substitutions at the −1 and second positions (−1T2F5T), and double substitutions at the second and fifth positions (−1I2F5C) (bottom). The effector-versus-target ratio was 1:1 (□) or 2.5:1 (■) in panel A and 1:1 (□) or 4:1 (■) in panel B. Killing activity against KWN-T4 cells infected with control vector expressing green fluorescent protein (GFP) and mock infected (mock) are also shown. (C) Western blot analysis of intracellular expression of various Nef mutants in KWN-T4 target cells. KWN-T4 target cells expressing native Nef proteins containing wild-type sequences (−1I2Y5T), a Y-to-F substitution at the second position of the CTL epitope (−1I2F5T), an I-to-T substitution at the −1 flanking position (−1T2Y5T), double substitutions at the −1 and second positions (−1T2F5T), and double substitutions at the second and fifth positions (−1I2F5C) were examined. KWN-T4 cell lysates infected with control vector expressing GFP and mock infected are also shown. An aliquot (3 μg) of the same KNW-T4 target cells used for the killer assay in the upper panel was used for the Western blot. Symbols: ○, Nef138-10; ●, Nef138-10(2F).

prediction program, NetChop (23), suggested the possibility that the Y-to-F substitution in the second position creates a new cleavage site at the fifth T residue in the CTL epitope. Proteolytic cleavage within the epitope could be the cause of poor antigen presentation.

Although we could not show the process of positive selection for Nef138-10(2F), Nef138-10(2F5C), and Nef138-10(5C), the high prevalence of Nef138-10(2F) in A24-positive patients and the reversion in A24-negative patients suggested that one point mutant, Nef138-10(2F), was selected first, and then two or three point mutants, Nef138-10(2F5C), evolved. Once the T-to-C amino acid substitution at the fifth position is acquired, the binding capacity of the CTL epitope to the HLA-A*2402 heavy chain is abolished (Fig. 3), and the Y-to-F substitution at the second position may become dispensable even in the presence of HLA-A*2402.

In our cohort of patients, Nef138-10(2F) accompanied a −1T substitution in the flanking region very frequently. We observed sequential reversion in the CTL epitope and flanking region at least in one patient with an A24-negative background. As of 11 October 2003, the HIV-1 sequence database showed that the 2F substitution (74 sequences) accompanied the −1T substitution frequently (64.9%) but accompanied the wild-type residue (I) only rarely (9.5%). On the other hand, the wild-type residue (Y) in the second position of the CTL epitope (195 sequences) accompanied wild-type (I) residue more frequently (57.4%) than the −1T substitution (20.5%). Although the function of the region surrounding Nef138-10 has not been elucidated, there seems to be a compensation between these two residues.

In simian immunodeficiency virus infection, CTLs with high functional avidity select for escape variants (29). However, we found CTLs with high functional avidity not only against the wild type but also against Nef138-10(2F) in five patients studied. It is not known how these CTLs against Nef138-10(2F) are maintained in vivo. Very recently, new insights into the exogenous pathway for antigen presentation to CTLs have been elucidated (15, 17). Cross presentation by professional antigen-

presenting cells such as dendritic cells may be responsible. Our study underlines the difficulties in evaluating the effective CTL responses in vivo by CTL assays in which peptides are used, such as ELISPOT.

For example, a CTL escape variant of Epstein-Barr virus was demonstrated in a highly A11-positive population in New Guinea (9). HLA-restricted CTL responses appear to be driving HIV-1 evolution at a population level (20). As far as we know, this is the first direct demonstration of horizontal transmission of CTL escape mutants of HIV-1 at a population level. We previously reported stereotypic amino acid substitutions in HIV-1 at some CTL epitopes restricted by HLA-B35 (21). Stereotypically selected HIV-1 may become dominant through transmission where certain HLA types are highly prevalent. Recently, a rare HLA supertype was shown to have a selective advantage for the prognosis of HIV-1 infection (34). In a population with less diverse HLA types, such as that of Japan, HLA types may have a large impact on HIV-1 evolution and escape. Our study may prove to have important implications for understanding viral pathogenesis and vaccine development.

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Influence of single-nucleotide polymorphisms in the multidrug resistance-1 gene on the cellular export of nelfinavir and its clinical implication for highly active antiretroviral therapy

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Protease inhibitors (PIs) such as nelfinavir (NFV) suppress HIV replication. PIs are substrates of P-glycoprotein (P-gp), the product of the multidrug-resistance-1 (*MDR1*) gene. Three single-nucleotide polymorphisms (SNPs) are present in exons of the *MDR1* gene: *MDR1* 1236, *MDR1* 2677 and *MDR1* 3435. We speculated that these genetic polymorphisms affected PI concentration in the cell. To verify this hypothesis, we first genotyped these SNPs in 79 Japanese patients by the SNaPshot method and found incomplete linkage disequilibrium between the SNPs. Because the SNP at *MDR1* 3435 has been reported to be associated with P-gp expression, we evaluated the effect of that SNP on the export of NFV from HIV-positive patients' lymphoblastoid cell lines by measuring time-dependent decrease in the amount of intracellular NFV by high-performance liquid chromatography. We found the intracellular concentration of NFV in lymphoblastoid cell lines (LCLs) with the homozygous T/T genotype at *MDR1* 3435 were higher than that with C/C genotype with statistical significance. This suggests that the activity of P-gp in patients' LCL cells with the *MDR1* 3435 T/T genotype was lower. In a retrospective study we evaluated the effect of the SNPs on CD4 cell count recovery in response to antiretroviral treatment with PIs, and obtained statistically significant evidence that suggested marginal association of the SNP at *MDR1* 1236 but not at *MDR1* 2677 or *MDR1* 3435. As *in vitro* results were not consistent with the clinical evaluation, clinical importance of *MDR1* genotyping for antiretroviral therapy remains to be investigated in a larger, case-controlled study.

Introduction

Antiretroviral therapy with HIV protease inhibitors (PIs) in combination with reverse transcriptase inhibitors dramatically improved the prognosis of patients infected with HIV-1. However, some patients fail to achieve the maximal virological suppression. We speculate that such failure is partly because PIs do not accumulate in lymphocytes in their active free forms in a concentration high enough to inhibit viral replication [1,2], although the intracellular active PI levels have, to the best of our knowledge, not yet been determined. The activity of P-glycoprotein (P-gp), the product of the multidrug resistance-1 (*MDR1*) gene, appears to affect intracellular PI concentration, because PIs such as nelfinavir (NFV) are substrates of P-gp [2]. P-gp is a glycosylated membrane protein belonging to the ATP-binding cassette superfamily of membrane transporters. P-gp is expressed in many tissues and cell types including intestinal epithelial cells and lymphocytes, where it acts as an energy-dependent exporter [3–9]. The *MDR1* is polymorphic and at least three single-nucleotide polymorphisms (SNPs) have been identified in the exons in a healthy Japanese population [10] as well as in other ethnic groups [6]. *MDR1* 1236 and *MDR1* 3435 are silent mutations in exons 12 and 26 [3,11], respectively, whereas *MDR1* 2677 is a substitution mutation in exon 21 [11]. Reportedly, the SNP at *MDR1* 3435 is associated with the amount and activity of P-gp protein both *in vitro* and *in vivo* [3,12]. In addition, individuals with the T/T genotype at *MDR1* 3435 were found to express less P-gp in lymphocytes and in intestinal epithelial cells [3,13] and showed lower efflux of rhodamine from natural killer (NK)

cells than those with the C/C genotype [13]. According to these observations, *MDR1* polymorphisms seem to affect the intracellular PI concentration and the outcome of antiretroviral treatment. However, the role of *MDR1* 3435 SNP in the response to antiretroviral therapy is still controversial [12,14].

The objective of this study was to evaluate the effect of three *MDR1* SNPs on the intracellular concentrations of NFV and to evaluate the impact of those SNPs on virological and immunological response to antiretroviral treatment, including NFV and PIs. We genotyped the SNPs in 79 Japanese patients and compared the velocity of NFV efflux among selected patients' lymphoblastoid cell lines (LCLs) with different *MDR1* 3435 genotypes. We also analysed the viral loads and CD4 cell counts after initiation of antiretroviral treatment with prescriptions with PIs including NFV in 21 patients.

Materials and methods

Patients

A total of 79 HIV-positive Japanese patients were enrolled in this study. These patients attended a hospital AIDS clinic at the Institute of Medical Science, University of Tokyo (IMSUT). The patients provided their written informed consent to participate in the study and to supply blood samples for DNA analysis and cell culture. Of the 79 patients, 21 receiving highly active antiretroviral therapy (HAART) including PIs were divided into three groups: 11 patients receiving HAART with NFV, four patients receiving HAART with indinavir (IDV) and six patients receiving HAART with saquinavir (SQV) or lopinavir/ritonavir (LPV/RTV). CD4 cell counts and HIV-RNA of plasma were analysed for 9 months after the initiation of the antiretroviral treatment. The study has been approved by the ethics committee of IMSUT.

Single-nucleotide polymorphisms

We typed three single-nucleotide polymorphisms (SNPs) at *MDR1* 1236 (exon 12), *MDR1* 2677 (exon 21) and *MDR1* 3435 (exon 26) by polymerase chain reaction (PCR) followed by ABI PRISM SNaPshot Multiplex Kit (PE Biosystems, Foster City, Calif., USA) [15]. Information on primers and conditions for PCR was obtained at http://snp.ims.u-tokyo.ac.jp [10].

Cells and determination of uptake and efflux of NFV

Peripheral blood mononuclear cells (PBMCs) were separated from patients' whole blood with Ficoll-Conray gradient centrifugation. LCLs were obtained by transforming PBMCs with Epstein-Barr virus (EBV), which was obtained from cell-free supernatants of EBV-producing B95–8 cell lines [16]. LCLs were maintained in RPMI 1640 medium (Sigma-Aldrich, St. Louis, Mo., USA) supplemented with 10% heat-inactivated fetal calf serum.

To determine the time course of NFV uptake into LCL cells, LCL cells (1×10^6/10 ml, counted with a haematocytometer) were incubated at 37°C in a medium containing 10 μM NFV. Cells were harvested by centrifugation at 1500 ×*g* for 5 min at 4°C and immediately frozen at –80°C until high-performance liquid chromatography (HPLC) analysis. To determine the velocity of NFV efflux from LCL cells, these patients' LCL cells were incubated at 37°C in a medium containing 10 μM NFV for 3 h. The cells were then quickly washed twice with 10 ml ice-cold phosphate-buffered saline and cultured in 10 ml NFV-free medium for up to 3 h. After an interval, aliquot cells were harvested by centrifugation at 1500 ×*g* for 5 min at 4°C and immediately frozen at –80°C until HPLC analysis.

Reverse transcription-PCR (RT-PCR)

For quantification of *MDR1* transcript, RNA from 1×10^7 LCL cells was isolated using Trizol reagents (Invitrogen Corp, Carlsbad, Calif., USA). First strand cDNA was obtained by using ReverTra Ace (Toyobo, Osaka, Japan) with 1 μg of total RNA. cDNA was subjected to PCR. Information on primers and conditions for PCR was obtained as previously described [17]. We used human glyceraldehyde 3-phosphate dehydrogenase mRNA as a positive control.

Determination of intracellular concentration of NFV by HPLC

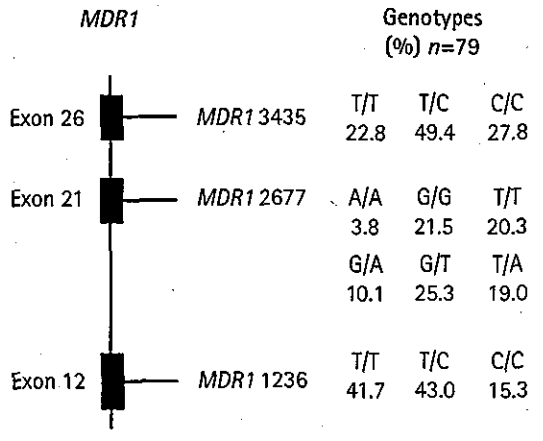
The patients' frozen LCL cells were extracted with 1.5 ml of ethanol. The extracts were then clarified by centrifugation at 2050 ×*g* for 10 min at 4°C. The ethanol extracts were evaporated at 30°C and dissolved in 180 μl of mobile phase, which was a mixture of phosphate buffer (containing 50 mM KH_2PO_4 and 50 mM Na_2HPO_4; pH 5.63) and acetonitrile (1:1, v:v) [18]. The amounts of NFV were measured using a Sensyu Pack ODS C_{18} column (5 μm particle size; 150×4.6 mm, Sensyu Scientific Co, Tokyo, Japan) at a flow rate of 1.5 ml/min by HPLC (Shimadzu Co, Tokyo, Japan). The UV detection wave length was 220 nm and efavirenz (EFV) was used as an internal standard. The lower limits of detection and quantification were 20 ng (30.1 pmole)/10^6 cells, and the calibration range was 20–2000 ng (30.1–3010 pmole/10^6 cells).

Results

We typed the three SNPs at *MDR1* 1236 (exon 12), *MDR1* 2677 (exon 21) and *MDR1* 3435 (exon 26) in DNA samples from 79 HIV-positive Japanese patients

(Figure 1). We found that it was consistent with the Hardy–Weinberg principle (Tables 1 and 2). Furthermore, in all possible two-way comparisons of the three SNPs at *MDR1* 1236 (exon 12), *MDR1* 2677 (exon 21, excluding the genotypes containing G) and *MDR1* 3435 (exon 26), we found significant linkage disequilibrium between *MDR1* 2677 A (T) and *MDR1* 1236 C (T), *MDR1* 2677 A (T) and *MDR1* 3435 C (T), and *MDR1* 1236 C (T) and *MDR1* 3435 C (T), respectively.

Figure 1. Frequency of SNPs in *MDR1*



The SNPs at *MDR1* 1236, *MDR1* 2677 and *MDR1* 3435 were typed by the SNaPshot method. Genotype frequencies at each site are shown as percentage among 79 HIV-infected Japanese patients. The thin vertical line at left represents the *MDR1* gene on human chromosome 7. The closed boxes represent exons 12, 21 and 26.

Table 1. Hardy–Weinberg principle at *MDR1* 1236 (*n*=79)

	T/T	T/C	C/C
Observed number of patients	33	34	12
Expected number of patients	31.7*	36.7†	10.6‡

p: Frequency for the T allele $\frac{33\times2+34}{2\times79}=0.633$

q: Frequency for the C allele 1–p=0.367

*$79\times p^2=31.7$

†$79\times2pq=36.7$

‡$79\times q^2=10.6$

Table 2. Hardy–Weinberg principle at *MDR1* 3435 (*n*=79)

	T/T	T/C	C/C
Observed number of patients	18	39	22
Expected number of patients	17.8*	39.4†	21.8‡

p: Frequency for the T allele $\frac{18\times2+39}{2\times79}=0.475$

q: Frequency for the C allele 1–p=0.525

*$79\times p^2=17.8$

†$79\times2pq=39.4$

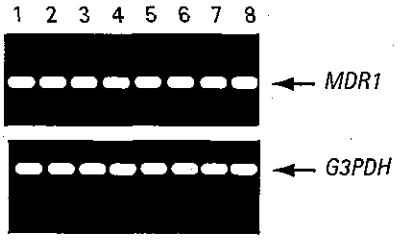
‡$79\times q^2=21.8$

Reportedly, *MDR1* 3435 T/T genotype was associated with lower expression of P-gp in leukocytes [13] so we hypothesized that the genotype was also associated with slower cellular export of NFV in patients' lymphocytes. To investigate this, we first established LCLs by immobilizing selected patients' PBMCs with EBV. We selected eight patients' LCLs with *MDR1* 3435 C/C (*n*=4) and T/T (*n*=4) and verified similar levels of *MDR1* in these LCLs by RT-PCR (Figure 2). We observed little variation in *MDR1* transcripts.

We found that uptake of NFV was rapid into LCLs reaching a steady-state within 5 min (Figure 3). We studied eight patients' LCLs with *MDR1* 3435 T/T and *MDR1* 3435 C/C to compare the steady-state intracellular concentration of NFV after 3 h incubation in a medium containing 10 μM NFV. The intracellular concentrations of NFV in LCLs with *MDR1* 3435 T/T and C/C genotypes were 2593 μM and 2411 μM, respectively (*n*=4), with no statistical difference. We calculated these values by hypothesizing that the LCLs are ideal spheres (10 μm diameter) and that NFV distributes uniformly in the cell.

We then compared NFV efflux from those LCLs with different genotypes at *MDR1* 3435. Before measuring export of NFV, LCLs were cultured with NFV to a saturated level. These NFV-loaded cells were transferred to NFV-free medium and cultured for 3 h with intermittent sampling of cell aliquots. We compared the efflux of NFV from the eight patients' LCLs with *MDR1* 3435 T/T and C/C (*n*=4 each), which had been verified to express *MDR1* mRNA by

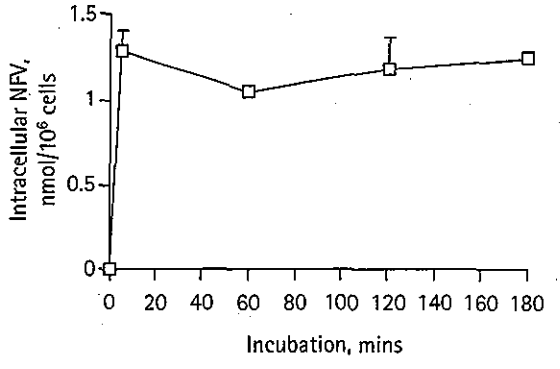
Figure 2. *MDR1* mRNA expression in LCLs



We selected eight patients' LCLs with *MDR1* 3435 C/C (lanes 1–4) and T/T (lanes 5–8) and measured the expression of *MDR1* mRNA. Total cellular RNA from LCLs was subjected to RT-PCR with primer sets for *MDR1* and *G3PDH* transcripts. Aliquots were subjected to agarose gel electrophoresis. The genotypes at *MDR1* 1236, 2677 and 3435: lanes 1 and 2, (T/T, G/G, C/C); lane 3, (T/C, G/A, C/C); lane 4 (C/C G/A C/C); lane 5 (T/T, G/T, T/T); and lanes 6–8 (T/T, T/T, T/T).

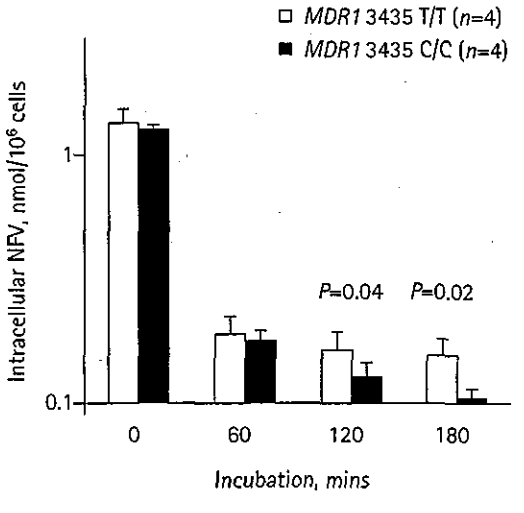
RT-PCR (Figure 2). The concentration of intracellular NFV in LCLs with the homozygous T/T genotype at *MDR1* 3435 was higher than in those with C/C genotype at 120 min and 180 min. This difference was statistically significant (P=0.04 and 0.02, respectively, Mann–Whitney U-test, Figure 4). This meant the NFV efflux in patients' LCL cells with the *MDR1* 3435 T/T genotype was slower than that with C/C genotype. Thus, we suspect the activity of P-gp in patients' LCLs with the *MDR1* 3435 T/T genotype is lower than that with the C/C genotype.

Figure 3. A typical time course of NFV uptake



LCL cells (1 ×10⁶/10 ml) were incubated in medium containing 10 μM of NFV. Cells were harvested at 0, 5, 60, 120 and 180 min and assayed for intracellular NFV by HPLC. The horizontal axis shows the incubation time in min. The vertical axis shows the intracellular amount of NFV per 10⁶ cells. The error bars represent the standard deviations.

Figure 4. NFV efflux from patients' LCLs



LCL cells were incubated in medium containing 10 μM of NFV for 3 h. Cells were then washed and cultured in NFV-free medium. Intracellular concentration of NFV was determined at 0, 60, 120 and 180 min by HPLC. The horizontal axis shows the incubation time in min. The vertical axis shows the intracellular amount of NFV per 10⁶ cells. We selected eight patients (described in the legend to Figure 2) and examined the velocity of NFV efflux from those cells. The intracellular concentration of NFV was measured several times in all patients' LCLs, and data were similar in every test. The error bars represent the standard deviations.

To examine the influence of *MDR1* 3435 genotypes on the response to treatment, we assessed increase in CD4 cell counts and viral suppression in 21 patients after initiation of HAART. At first, we hoped to analyse data obtained from a group of patients receiving NFV alone as a PI, but could not, due to the small number of NFV-receiving patients. Thus, we carried out the analysis in those patients receiving PIs including NFV (n=11), indinavir (n=4) and saquinavir/lopinavir/ritonavir (n=6). CD4 cell counts before treatment were similar among patients with various genotypes. Patients with various genotypes at *MDR1* 3435 showed similar changes in CD4 cell counts (Figure 5A) and viral suppression (Figure 6A) during 9 months of HAART. We found patients with the *MDR1* 1236 T/T showed higher increase in CD4 cell counts at 1 month (148 cells/μl) and 9 months (264 cells/μl) after initiation of therapy than those with *MDR1* 1236 C/C (20 cells/μl and 34 cells/μl, respectively) (Figure 5C). We suspected that *MDR1* 1236 T/T was associated with a higher rate of recovery of CD4 cell counts for patients receiving HAART with PI. We did not find differences in rates of viral suppression among the patients with various *MDR1* 1236 genotypes (Figure 6C). We did not observe a statistical difference in CD4 cell counts or viral loads among patients with different *MDR1* 2677 genotypes (Figures 5B and 6B).

Discussion

In this study, we genotyped three SNPs at *MDR1* 1236 (exon 12), *MDR1* 2677 (exon 21) and *MDR1* 3435 (exon 26) (Figure 1) in 79 HIV-positive Japanese patients and found incomplete linkage disequilibrium – as has also been reported in other ethnic groups [6]. We found that genotype frequencies of the SNPs at *MDR1* 1236 (exon 12) and *MDR1* 3435 (exon 26) in this population were in Hardy–Weinberg equilibrium. This suggested that the studied population was precisely genotyped and unbiased in terms of the *MDR1* gene. We compared the activity of P-gp among patients' LCLs with different *MDR1* 3435 genotypes by measuring NFV efflux from the cultured LCL cells by HPLC. We found that the intracellular concentration of NFV in LCLs with the homozygous T/T genotype at *MDR1* 3435 was higher than in those with the C/C genotype at 120 min and 180 min. This difference was statistically significant (P=0.04 and 0.02, respectively; Mann–Whitney U-test; Figure 4). In contrast, in the retrospective evaluation of 21 HIV-positive patients