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## ***In vitro* induction of the pronephric duct in *Xenopus* explants**

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The earliest form of embryonic kidney, the pronephros, consists of three components: glomus, tubule and duct. Treatment of the undifferentiated animal pole ectoderm of *Xenopus laevis* with activin A and retinoic acid (RA) induces formation of the pronephric tubule and glomus. In this study, the rate of induction of the pronephric duct, the third component of the pronephros, was investigated in animal caps treated with activin A and RA. Immunohistochemistry using pronephric duct-specific antibody 4A6 revealed that a high proportion of the treated explants contained 4A6-positive tubular structures. Electron microscopy showed that the tubules in the explants were similar to the pronephric ducts of normal larvae, and they also expressed *Gremlin* and *c-ret*, molecular markers for pronephric ducts. These results suggest that the treatment of *Xenopus* ectoderm with activin A and RA induces a high rate of differentiation of pronephric ducts, in addition to the differentiation of the pronephric tubule and glomus, and that this *in vitro* system can serve as a simple and effective model for analysis of the mechanism of pronephros differentiation.

**Key words:** activin, pronephric duct, retinoic acid, *Xenopus*.

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### **Introduction**

Vertebrates develop a succession of three forms of kidney during embryonic and adult life: the pronephros (first kidney), mesonephros (middle kidney) and metanephros (final kidney). All three kidneys have the same basic functional unit, the nephron, and similar mechanisms and regulatory genes are probably involved in nephron development in all three (Vize *et al.* 1997). The simplicity of the pronephros makes its developmental system an attractive model for studying kidney morphogenesis. The pronephros consists of three components: (i) the pronephric glomus, which filters waste from the bloodstream into the coelom; (ii) the pronephric tubule, which collects the waste from the coelom; and (iii) the pronephric duct, which carries the waste to the exterior via the cloaca (Vize *et al.* 1997). Two monoclonal antibodies that specifically recognize different parts of the pronephros have been developed (Vize *et al.* 1995): antibody 3G8 recognizes the pronephric tubule and otic vesicle, and antibody 4A6 specifically recognizes the pronephric duct. The

molecular nature of the antigens targeted by these antibodies remains unknown, except that by western blotting it has been found that antibody 4A6 recognizes a 50 kDa protein. Investigations using these antibodies have elucidated mechanisms of pronephros development, such as the timing of specification (Brennan *et al.* 1998) and the spatial relationship of the primordia of the pronephric tubules and ducts in normal embryos (Vize *et al.* 1995).

The *in vitro* inducibility of these pronephric components has been analyzed using an animal cap model. Animal cap ectoderm, fated to become epidermis and neural tissue in normal development, differentiates to form the pronephric tubule and glomus when treated with activin A and retinoic acid (RA; Moriya *et al.* 1993; Brennan *et al.* 1999). In this report, we used antibody 4A6 to show that the *in vitro* treatment of animal cap ectoderm with activin A and RA very frequently induces differentiation of the pronephric duct.

### **Materials and Methods**

#### *Animal cap assay*

*Xenopus* late blastulae (stage 9; Nieuwkoop & Faber 1956) were de-jellied by treatment with 2% cysteine hydrochloride in Steinberg's solution (58 mM NaCl,

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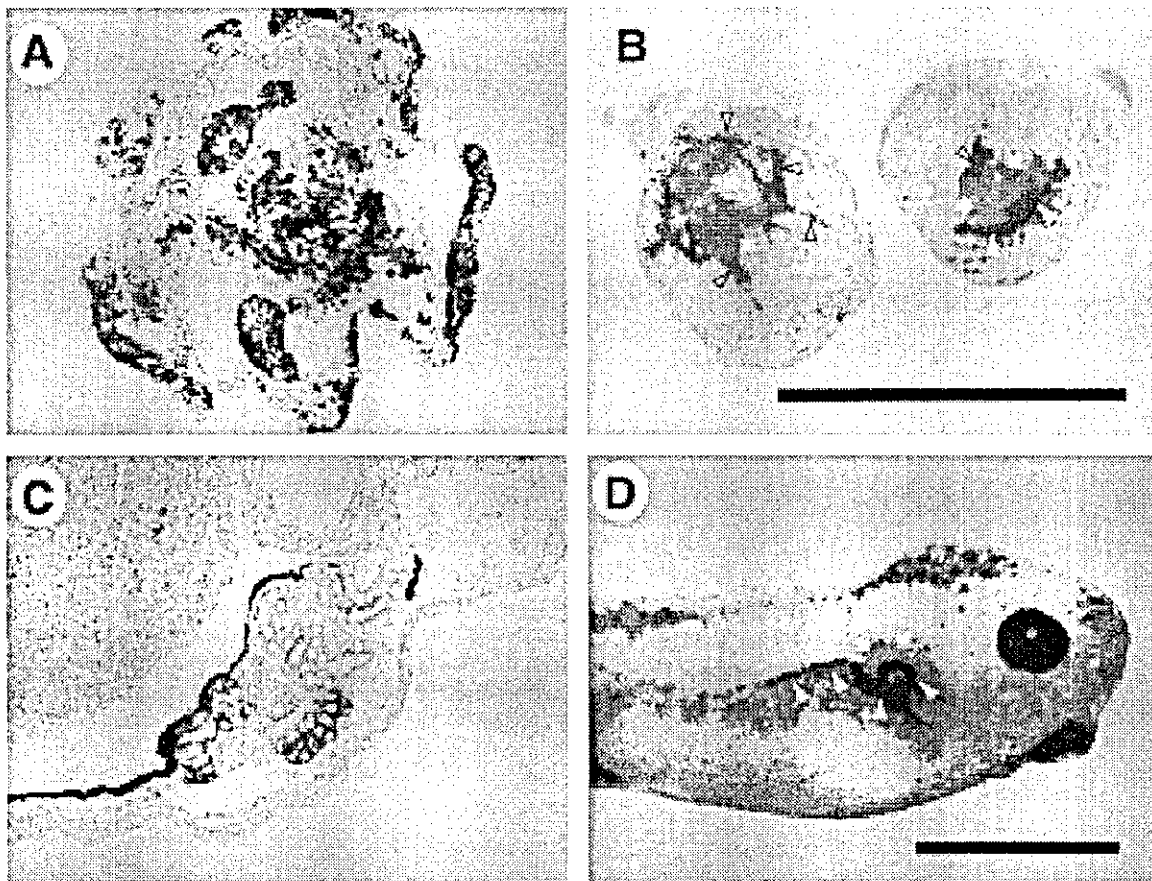
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0.67 mM KCl, 0.34 mM Ca(NO<sub>3</sub>)<sub>2</sub>, 0.83 mM MgSO<sub>4</sub>, 3 mM HEPES and 100 mg/L kanamycin sulfate, pH 7.8). The vitelline membrane of the blastula embryo was removed with fine forceps, and the presumptive ectoderm (animal cap) was isolated with tungsten needles. The ectoderm was removed in 0.6–0.7 mm squares, unless indicated otherwise. The explants were incubated for 1 h in Steinberg's solution containing recombinant human activin A (10 ng/mL; kindly provided by Dr Y. Eto, Ajinomoto Co. Inc., Kawasaki, Japan), and all-*trans* RA (10<sup>-4</sup> M; Sigma, St Louis, MO, USA). After washing, the explants were cultured in Steinberg's solution at 20°C until they were equivalent to stage 40–42 in normal embryos, unless indicated otherwise.

#### Immunohistochemistry

Embryos and explants were fixed in MEMFA (0.1 M MOPS, pH 7.4, 2 mM EGTA, 1 mM MgSO<sub>4</sub> and 3.7%

formaldehyde) and subsequently dehydrated in methanol and stored at -20°C. Both embryo and explant tissues were rehydrated with phosphate-buffered saline (PBS), then blocked with PBT (PBS, 0.1% Triton X)/20% goat serum for 1 h at room temperature. Hybridoma supernatant (3G8 or 4A6; kindly provided by Dr E. A. Jones, Warwick University, Coventry, UK), was diluted with PBT/20% goat serum, 1:2 for 3G8 and 1:1 for 4A6, and incubated overnight at 4°C. After washing the samples five times with PBT at room temperature, secondary incubation with a 1:4000 dilution of goat antimouse IgG + IgM conjugated to alkaline phosphatase (Biosource, Camarillo, CA, USA) was done overnight at 4°C. Samples were washed five times with PBT at room temperature, and an alkaline phosphatase blue color reaction was done by using 4-nitroblue tetrazolium chloride/5-bromo-4-chloro-3-indolylphosphate substrate (NBT/BCIP; Roche Diagnostics Corporation,



**Fig. 1.** Double immunostaining of a stage 42 equivalent *Xenopus* explant (A,B) and stage 40 larvae (C,D) using pronephric duct-specific antibody 4A6 (blue) and pronephric tubule-specific antibody 3G8 (red). Whole-mount immunostaining patterns (B,D) and photomicrographs of sections (A,C) are shown. (A,B) Explants treated with activin A and retinoic acid contain both 3G8-positive (pronephric tubule, arrows) and 4A6-positive tubular structures (pronephric duct, arrowheads). Bar, 1 mm.

Indianapolis, IN, USA). Samples were fixed again with MEMFA, dehydrated through a graded series of ethanol and xylene, embedded in paraffin, and sectioned at 10 µm.

Whole-mount double staining was carried out by first-round staining with 4A6 antibody using NBT/BCIP and fixation in MEMFA for 30 min, followed by second-round staining with 3G8 antibody using the Fast Red Tablet substrate (Roche Diagnostics Corporation). For light microscopy, the first-round stained explants were sectioned at 10 µm and subjected to the second round of staining.

**Electron microscopy**

Stage 42 tadpoles and the explants cultured to stage 42 equivalence after treatment with activin A and RA were prefixed in 3% paraformaldehyde, 2.5% glutaraldehyde and 0.1 M cacodylate buffer (pH 7.4) for 1 day. The explants and tadpoles were then washed with the buffer and postfixed in 1% OsO<sub>4</sub> and 0.1M cacodylate buffer (pH 7.4) for 2 h. After washing with the buffer, the explants and tadpoles were dehydrated through a graded series of ethanol and acetone and embedded in epoxy resin. Ultrathin sections were cut and were double stained with uranyl acetate and lead citrate, and the sections were examined with a transmission electron microscope (JEM-200CX; Jeol, Tokyo, Japan).

**Whole-mount in situ hybridization**

Whole-mount *in situ* hybridization was performed using the method described by Harland (1991). 1.5 kb

partial fragments of *Gremlin* and *c-ret* amplified by the polymerase chain reaction (PCR) were subcloned into PCRII vector (Invitrogen Corporation, Carlsbad, CA, USA) and sequenced. The primers used for PCR were: 5'-TGTAGCAGCCTCACATGGAC-3' and 5'-CCACCCCTGACCTCAATAAA-3' for *Gremlin*; and 5'-AAGATGACACAGCCCCATTC-3' and 5'-GGGATGCACTCTCTTCAGC-3' for *c-ret*. After linearization of the templates, RNA probes were transcribed with Sp6 and T7 RNA polymerases (Promega, Madison, WI, USA). For histological analysis, stained explants and embryos were sectioned at 10–20 µm.

**Results**

*Immunohistochemistry demonstrating the pronephric tubule and duct induced in explants*

Ectoderm explants treated with activin A and RA were examined by immunohistochemistry. Double staining using pronephric duct-specific antibody 4A6 and pronephric tubule-specific antibody 3G8 was positive for both in the tubular structure of the explants (Fig. 1A,B). Staining of 4A6 was seen around the entire cell surface of the tubular epithelia of the explants (Fig. 1A), similar to that in the pronephric duct cells in stage 40 tadpoles (Fig. 1C). In contrast, 3G8 stained the apical surface of the tubule cells in the explants (Fig. 1A), the same as in stage 40 tadpoles (Fig. 1C). These staining patterns are consistent with the original report of Vize *et al.* (1995). Whole-mount staining (Fig. 1B,D) and light microscopic examination (Fig. 1A,C) revealed that the two antibodies stained different portions of the tubular structure,

**Table 1.** Induction of 4A6-positive tubular structures in explants treated with activin A and retinoic acid

	0 ng/mL activin A		10 ng/mL activin A	
	0 M RA <sup>†</sup>	10 <sup>-4</sup> M RA <sup>†</sup>	0 M RA <sup>†</sup>	10 <sup>-4</sup> M RA <sup>†</sup>
0.4–0.5 mm	0/16 (0)	0/27 (0)	0/23 (0)	21/29 (72)
0.6–0.7 mm	0/14 (0)	0/29 (0)	3/30 (10)	30/33 (91)
Ventral side	0/15 (0)	0/26 (0)	4/33 (12)	26/27 (96)
Dorsal side	0/15 (0)	0/30 (0)	2/29 (7)	23/40 (58)

<sup>†</sup>Figures represent the ratios of the number of explants with 4A6-positive tubular structures to the total number of explants. Figures in parentheses show the ratios as percentages. RA, retinoic acid.

**Table 2.** Induction rate of 4A6- or 3G8-positive tubular structures in explants cultured until equivalent to various stages of normal larvae

	Stage 27 <sup>†</sup>	Stage 32 <sup>†</sup>	Stage 37/38 <sup>†</sup>	Stage 42 <sup>†</sup>	Stage 47 <sup>†</sup>
4A6-positive rate	0/25 (0)	0/25 (0)	12/33 (36)	23/25 (92)	11/11 (100)
3G8-positive rate	0/11 (0)	12/12 (100)	8/12 (67)	11/14 (79)	11/13 (85)

<sup>†</sup>Figures represent the ratios of the number of explants with 4A6- or 3G8-positive tubular structures to the total number of explants. Figures in parentheses show the ratios as percentages.