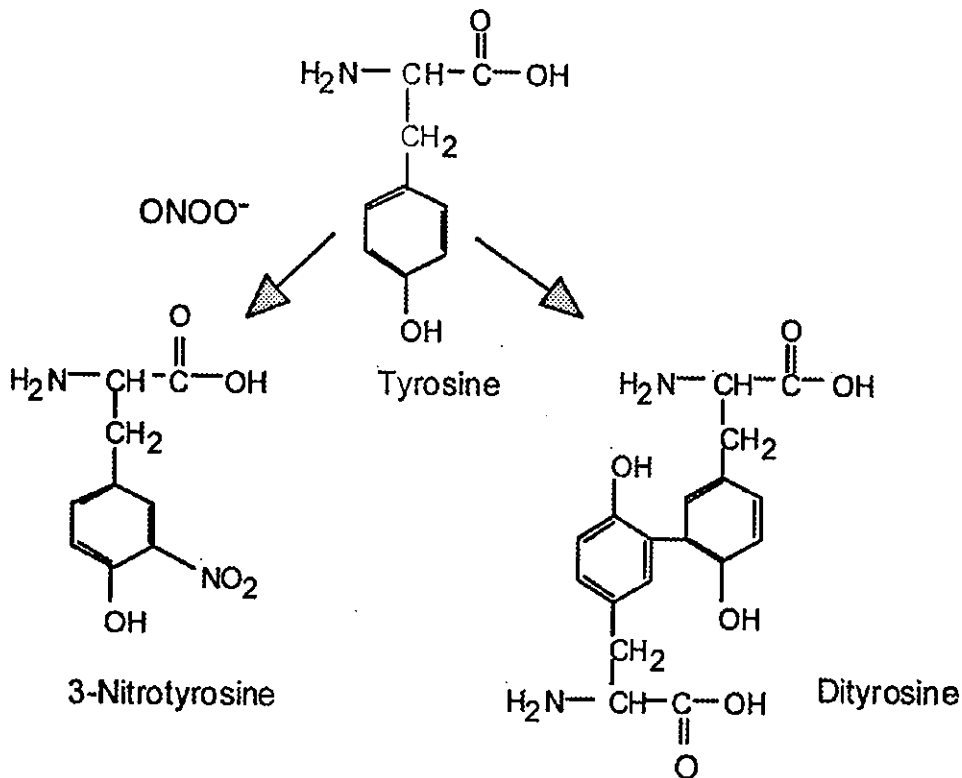
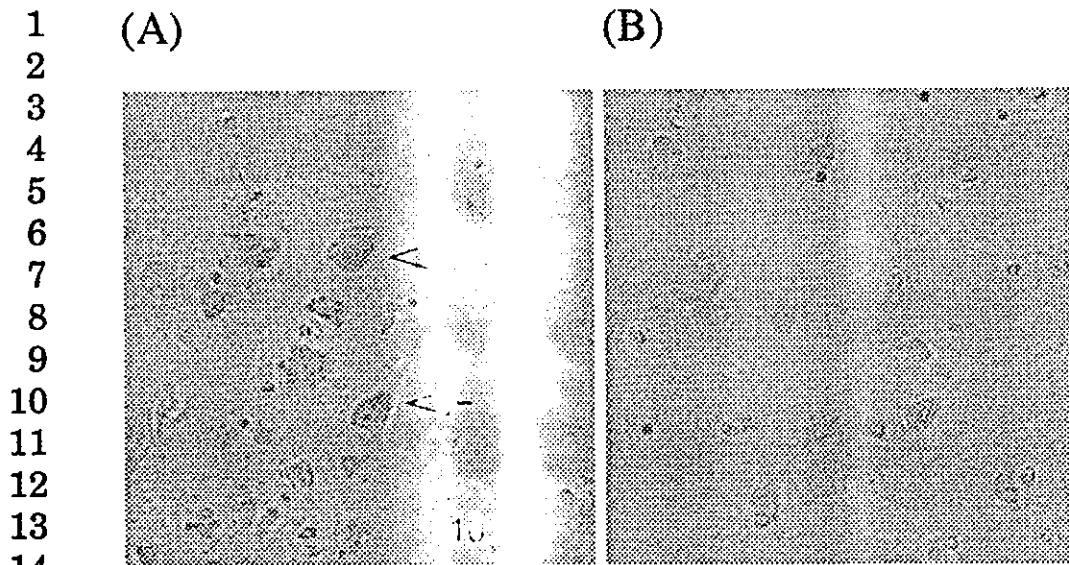


1 methionine, phenylalanine, and tyrosine. It inactivates the
 2 membrane function and key enzymes (see reviews 7, 10). As
 3 shown in Fig. 1, 3-nitrotyrosine (3-NT) is synthesized by the F1
 4 nitration of tyrosine residues in protein and a marker for
 5 the oxidative stress induced by peroxynitrite in vivo (11).
 6 3-Nitrotyrosine containing protein (3-NT protein) was detected
 7 in atherosclerosis (12) and neurodegenerative disorders, such
 8 as amyotrophic lateral sclerosis (ALS) (7), AD, (13,14), and
 9 PD (15). Figure 2 shows the immuno-histochemical detection F2
 10 of 3-NT protein in pyramidal hippocampal neurons, using
 11 anti-3-NT protein antibody (16). Another oxidation product
 12 of tyrosine is dityrosine, which is produced from free and
 13 protein-bound tyrosine in the presence of hydrogen peroxide
 14



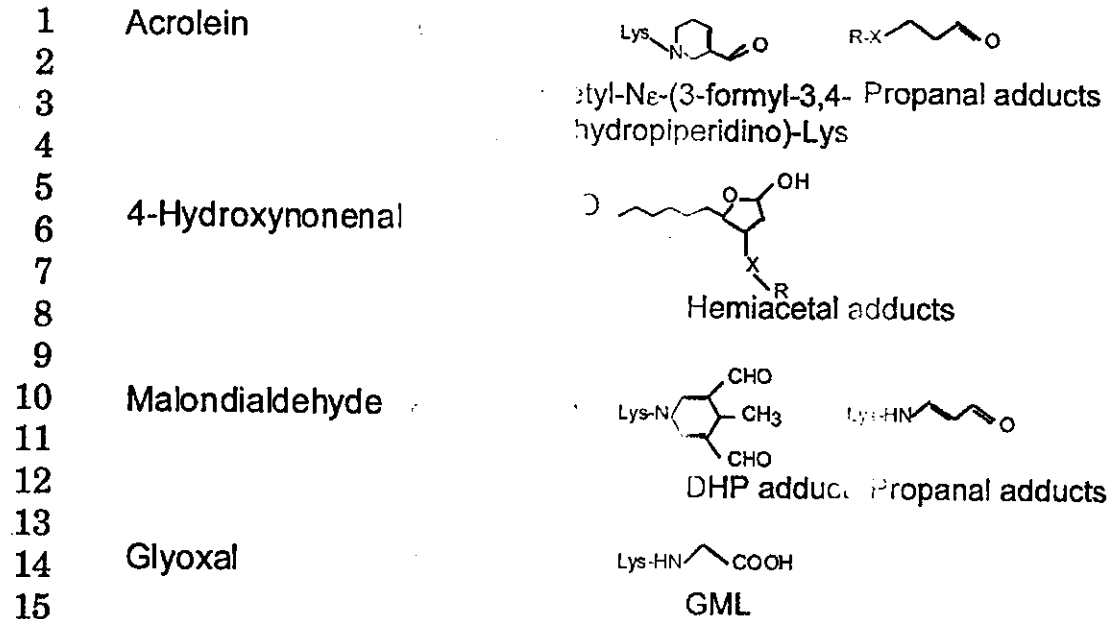
35 **Figure 1** Modification of tyrosine residue in protein by NOS and
 36 RNS. Peroxynitrite modifies tyrosine residues to 3-nitrotyrosine
 37 in protein, and hydrogen peroxide and peroxidase, and irradiation
 38 produces tyrosyl radicals, yielding dityrosine with tyrosine residues
 39 in protein or free tyrosine.



15 **Figure 2** Immunohistochemical detection of 3-NT-containing
16 protein in lipofuscin in pyramidal hippocampal neurons. The brain
17 was obtained from a 72-year-old male patient without neurological
18 or psychiatric disorders. The tissue samples were incubated with
19 (A) anti-3-NT protein antibody (diluted 1:200 with bovine serum
20 albumin) or (B) bovine serum albumin alone, followed by the treat-
21 ment of peroxidase-labeled anti-rabbit IgG. Lipofuscin stained was
22 observed as brown granules as indicated by arrows.

23
24 and myeloperoxidase (MPO) (Fig. 1), and is detected in athero-
25 sclerotic plaques (18) and lipofuscin pigments in the aged
26 human brain (19).

27 On the other hand, lipid peroxidation generates various
28 reactive aldehydes, including 4-hydroxynonenal (4-HNE) and
29 acrolein as shown in Fig. 3 (20). 4-Hydroxynonenal reacts
30 with sulfhydryl and amino groups and leads to inactivation
31 of DNA polymerases, dehydrogenases, and various trans-
32 porters, and also to cell cycle arrest and apoptosis. Proteins
33 modified with 4-HNE and malondialdehyde were detected
34 in nigro-striatal dopamine neurons in PD (21), neurofibrillar
35 tangles in AD (22,23), and the spinal cord of ALS patients
36 (24). Acrolein, $\text{CH}_2=\text{CH}-\text{CHO}$, is ubiquitously generated in
37 the biological system and is the most reactive α,β -unsaturated
38 aldehyde product of lipid peroxidation. It is incorporated
39 into proteins easily and accumulates as protein adducts



16 **Figure 3** Modification of protein by aldehyde products of lipid
 17 peroxidation. Aldehyde products of lipid and carbohydrate peroxida-
 18 tion, acrolein, 4-hydroxynonenal, malondialdehyde, and glyoxal, mod-
 19 ify lysine residues in protein.

20

21 after reacting with lysine and histidine residues by forming
 22 Michaelis-type amino acid complexes (25). Acro-
 23 lein-modified protein was detected in oxidized low-density
 24 lipoproteins (26) and in the brain of patients with AD (27). Fig-
 25 ure 4 shows the histochemical staining of the substantia nigra
 26 in a parkinsonian brain with an antibody against 4-HNE-
 27 modified protein. The neurons containing neuromelanin
 28 from parkinsonian patients were stained more markedly
 29 than those in normal control and nondopaminergic cells.
 30 These results indicate that the oxidative stress increases
 31 markedly in nigro-striatal dopamine neurons of a parkinson-
 32 nian brain.

35 II. MITOCHONDRIAL COMPLEX I SUBUNITS 36 ARE NITRATED BY NO⁺

37

38 In the brain, NO has been considered to be produced in micro-
 39 glia and astrocytes and transported to neurons, where it

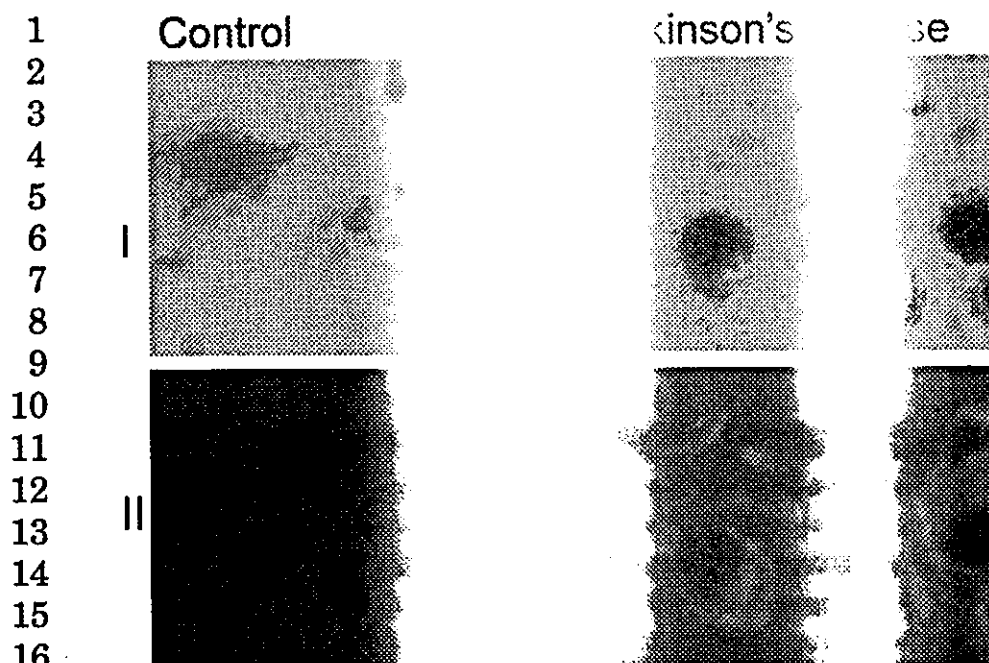


Figure 4 Occurrence of HNE-modified protein in dopamine neurons in substantia nigra of the brain from the patient with Parkinson's disease. On the left, substantia nigra of the brain from the parkinsonian patient. The dopamine neurons containing neuromelanin are positively stained with antibody against HNE modified protein. In control, all cells other than dopamine neurons are not stained with antibody.

reacts with the superoxide radical. SH-SY5Y cells produce superoxide anion by the use of 2',7'-dichlorofluorescein diacetate (28) and inhibitors of nitric oxide synthase (NOS); H₂DCFDA is cleaved into 2',7'-dichlorofluorescein by hydroxyl radical and ONOO⁻; and NO is inhibited by N⁵-(l-iminocetyl)-L-ornithine (L-NIO) and N⁵-methyl-L-lysine methyl ester (L-NAME), reduced DCF to a fluorescent product.

Using an antibody against the 3-NT protein (16), nitrated proteins were detected in human dopaminergic SH-SY5Y cells. The lysate from the cells was subjected to western blot analysis as shown in Figure 5, and the molecular weight of major 3-NT proteins was estimated to be 33, 21, 15, and 11 kDa. The nitrated protein bands were detected even in control under physiological conditions, suggesting that the cells were under

oxidative stress. However, SH-SY5Y cells produce ONOO⁻ in situ, as confirmed by the use of 2',7'-dichlorofluorescein diacetate (H₂DCFDA) and inhibitors of nitric oxide synthase (NOS); H₂DCFDA is cleaved into 2',7'-dichlorofluorescein by hydroxyl radical and ONOO⁻; and NO is inhibited by N⁵-(l-iminocetyl)-L-ornithine (L-NIO) and N⁵-methyl-L-lysine methyl ester (L-NAME), reduced DCF to a fluorescent product.

Using an antibody against the 3-NT protein (16), nitrated proteins were detected in human dopaminergic SH-SY5Y cells. The lysate from the cells was subjected to western blot analysis, and the molecular weight of major 3-NT proteins was estimated to be 33, 21, 15, and 11 kDa. The nitrated protein bands were detected even in control under physiological conditions, suggesting that the cells were under

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39

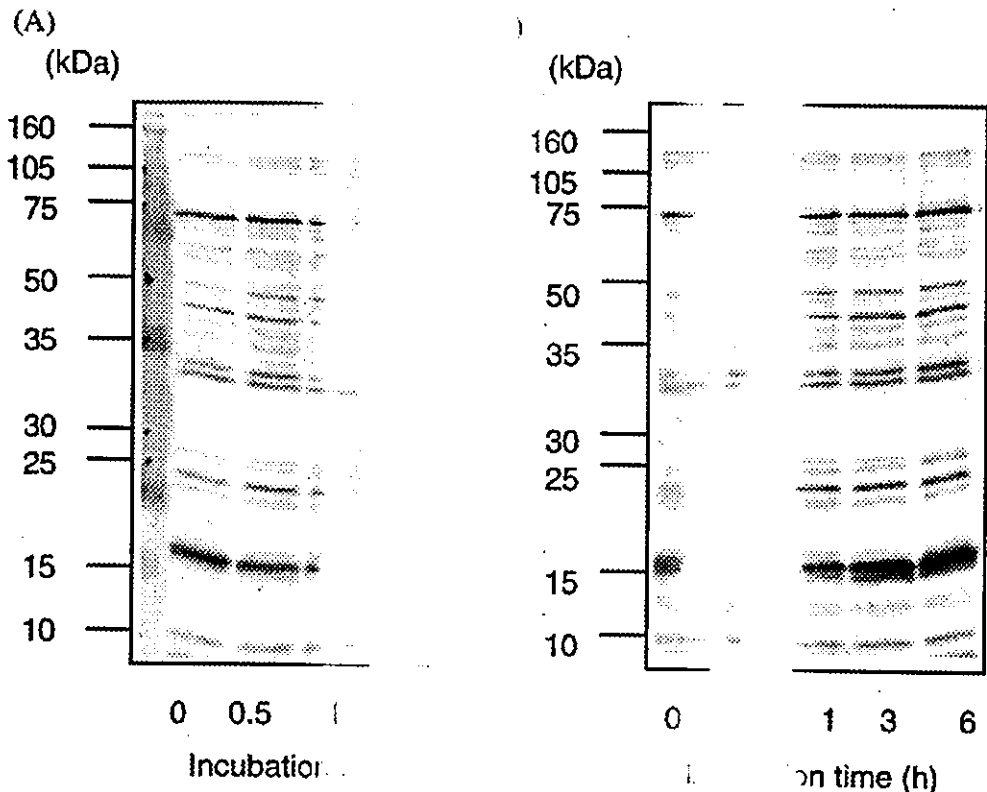


Figure 5 Formation of 3-Nitrotyrosine protein in SH-SY5V cells and the effects of peroxynitrite and proteasome inhibitor, PSI. (A) Cells were treated with 250 μM of SIN-1, then applied to the immunoblotting with anti-3-Nitrotyrosine antibody against 3-Nitrotyrosine proteins. After the treatment with SIN-1, the intensity of 3-Nitrotyrosine proteins increased, but the number did not change markedly. As shown in the left lane in A, the molecular marker is 15 kDa. (B) The cells were treated with 10 μM of proteasome inhibitor, PSI (10 μM). The intensity of 3-Nitrotyrosine protein increased according to the incubation time.

constant oxidative stress, which is essential for cell growth and proliferation. The active mechanism to eliminate oxidative stress is the mitochondrial fractionation. The mitochondrial fractionation is the most intensively studied method, and was subjected to immunoblotting against 3-NT protein. The immunoblotting results showed that complex I, II, III, and IV (29). Some of the identified proteins were intact in the immunoblotting of an oxidized protein from the cells. The SH-SY5V cell line was the one in which the anti-3-NT antibody, lot analysis of the antibodies against mitochondrial complex I, II, III, and IV were identified.

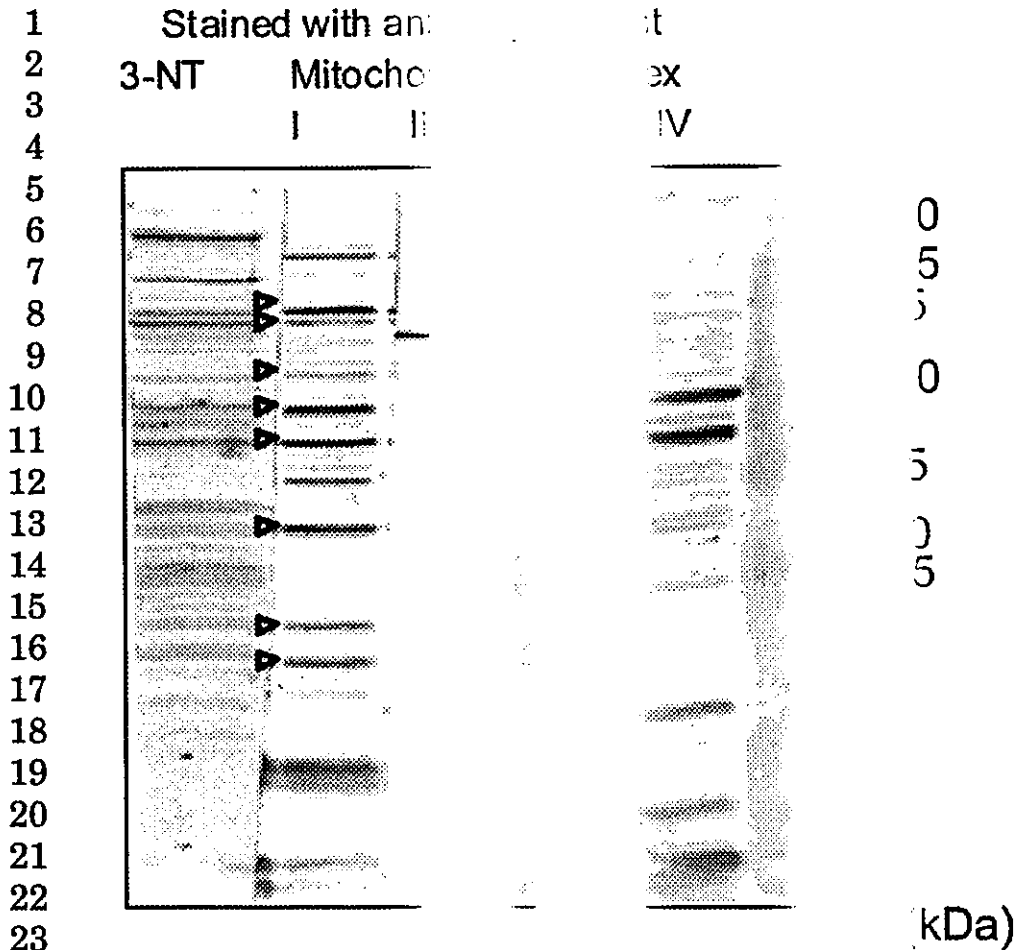


Figure 6 Detection of mitochondrial proteins in P1 and P2 fractions. The P1 and P2 fraction was separated by SDS-PAGE and detected by Western blot analysis with anti-3-NT (samples I-IV) and anti-complex I-IV (samples I-IV) antibodies, respectively. Molecular weight markers are shown in the right column. Arrows show the positions of the bands positively stained with anti-complex I-IV antibody.

to be the subunits of complex I (Fig. 6). The results clearly show that the preferential loss of complex I subunits may contribute to neuronal dysfunction observed in the nigro-striatum of parkinsonian brain (30,31). Treatment of C57BL/6J mice with 3-methyl-6-tyrosine (MPTP) or 1-methyl-4-phenylpyridinium (MPP+) inducing parkinsonism in the SH-SY5Y cells (32-35).

F6

1 A step-wise activation of the apoptotic cascade was observed:
 2 decline in $\Delta\Psi_m$, activation of caspase 3, and phosphorylation
 3 of p38 mitogen-activated protein kinase (MAPK) (33). Nitric
 4 oxide and ONOO⁻ were shown to induce apoptosis by nitra-
 5 tion of tyrosine residues of the cytochrome c (36), or of cyto-
 6 chrome c itself (37). The mitochondrial ATP synthesis
 7 was inhibited markedly by ONOO⁻, as shown in Fig. 7, which
 8 may be due to its reaction leading to cytochrome oxidase or
 9 inactivation of components of the electron transport chain and ATP synthase (38-41).

10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39

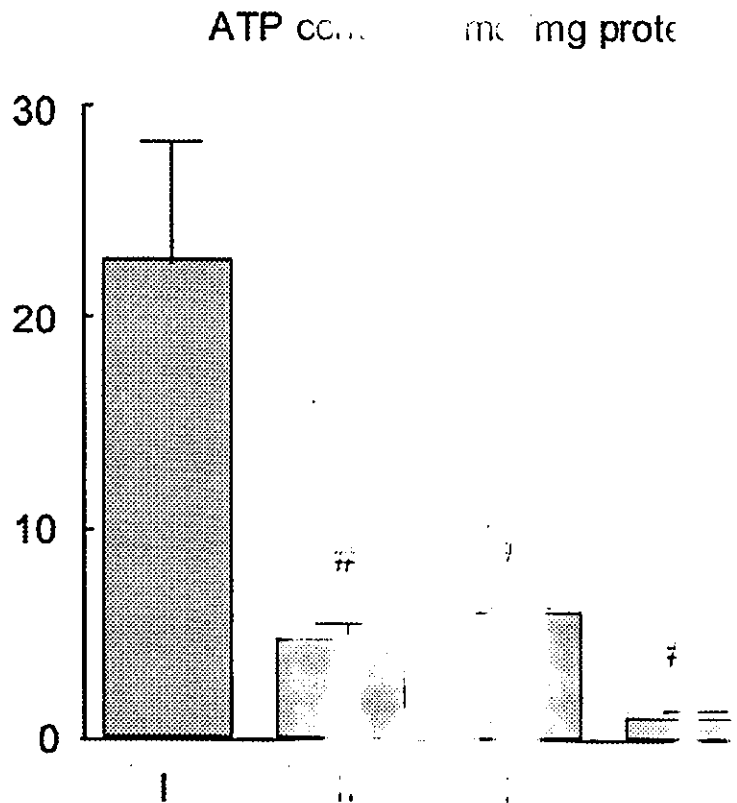


Figure 7 Effect of SIN-1 and PSI on the ATP content in SH-SY5Y cells. The cells were treated for 24 h without or with 100 μ M SIN-1 in the absence and presence of 100 μ M PSI for 24 h. The ATP content was measured by the luminescenceometric method. The column and bar represent the mean \pm SD of triplicate measurements of three independent experiments. I, control cells; II, cells treated with PSI alone; III, SIN-1 alone; IV, SIN-1 and PSI. The difference from control (I) is statistical significant ($p < 0.05$). The difference from SIN-1 alone (III) is statistical significant ($p < 0.05$).

1 As shown in Fig. 5A, the amount of 3-NT protein increased after treatment with 3-NT. The amount of 3-NT protein only in the cytosol fraction increased, suggesting the preference of some proteins for 3-NT modification. In addition, the amount of 3-NT protein in the nucleus was also increased significantly.

7 III. PROTEASOMES AND ACCUMULATION OF MODIFIED PROTEINS

10 Insoluble intracellular protein aggregates such as Lewy
11 body in PD and neurofibrillary tangles composed of β -amyloid in
12 AD, are hallmarks of neurodegeneration. Although it remains
13 unclear whether protein aggregates cause directly neuronal
14 cell death or are the result of deterioration of cellular homeo-
15 stasis in dying neurons, the analysis of the constituents of
16 inclusion bodies may provide a molecular mechanism lead-
17 ing their formation. The ubiquitin-proteasome system, a manifestation of
18 disturbed cellular proteostasis, is maintained by
19 the ubiquitin-proteasome system. In LB, ubiquitin, and pro-
20 teasome, subunits are major components (42,43), in addition
21 to α -synuclein, Parkin, and ubiquitin C-terminal hydrolase-
22 L1 (UCH-1) (41-44). Ubiquitin is modified by ROS-RNS as
23 nitrated synuclein and ubiquitin (45).

24 To clarify the interactions among oxidative stress,
25 dysfunction of the proteasome system, and formation of
26 the inclusion body, the effects of a proteasome inhibitor, car-
27 bobenzoxy-L-isoleucyl-leu-butyl-L-alanyl-L-leucinal (PSI) were
28 examined on the amount of modified proteins and the cell
29 vulnerability (34). PSI increased the amount of 3-NT
30 proteins in the SH-SY5Y cells, but the number of 3-NT protein
31 bands was almost the same as in the control (Fig. 5B). The func-
32 tional deterioration of mitochondria was also enhanced by PSI,
33 as shown by the severe reduction of ATP synthesis (Fig. 7). In
34 addition, the number of apoptotic cells increased significantly
35 by PSI, whereas that of necrotic cells remained almost the
36 same (Fig. 8A). At the same time, the acrolein-modified protein
37 increased significantly in subcellular fractions of the SH-SY5Y
38 cells after being treated with PSI (Fig. 8B). These results
39 clearly demonstrate that the inhibition of proteasome activity

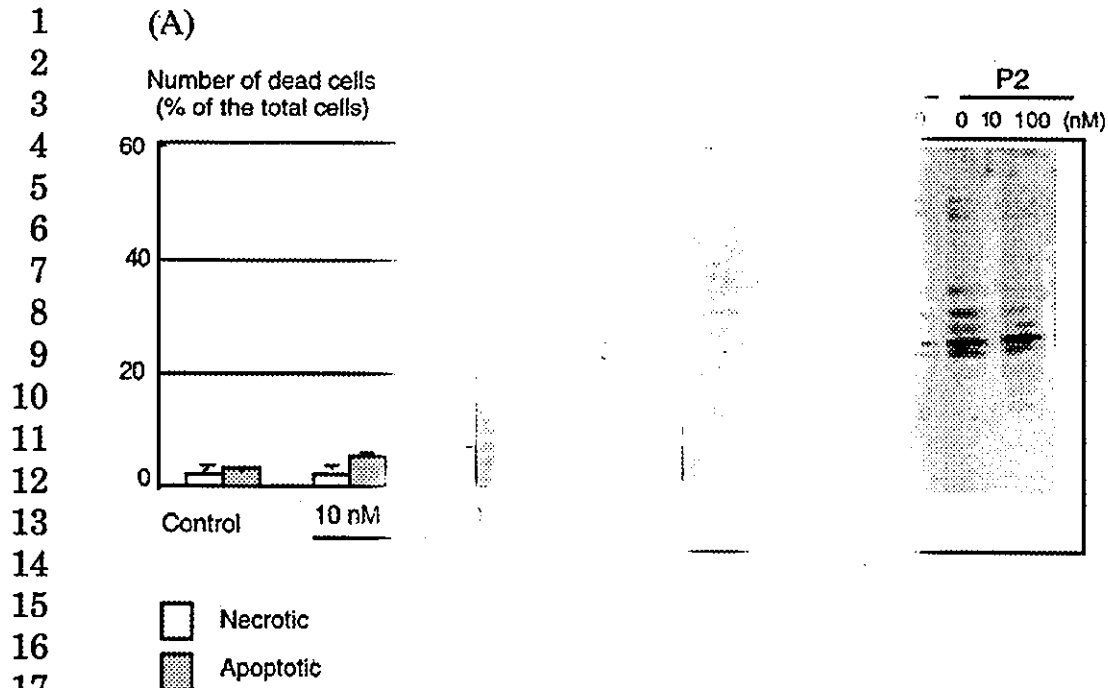


Figure 8 Effect of acrolein on cell death and protein levels. (A) Bar graph showing the percentage of necrotic and apoptotic cells in control and 10 nM treated groups. (B) Western blot showing P2 protein levels in control and 100 nM treated groups. The data indicate that treatment with 10 nM acrolein significantly increases the percentage of apoptotic cells and the level of acrolein-modified protein (P2) in the cytosol fraction of the cells. The results suggest that acrolein may play a key role in cell death and neuronal damage.

may play a key role in cell death and neuronal damage.

IV. MITOCHONDRIAL FUNCTION INHIBITS THE CYTOSOLIC SYSTEM BY OXIDATIVE DAMAGE

Systemic reduction of mitochondrial electron transfer chains was reported in the brain of patients with Alzheimer's disease.

1 with PD (30,31
2 none, was reported
3 fibrillar cytoplas-
4 α -synuclein we-
5 tion (52). The c-
6 teasome system
7 concentration t-
8 ATP levels red-
9 the cells after
10 tive modificatio-
11 body against the
12 the acrolein-me-
13 treatment (Fig.

14 The ubiquitin-proteasome system
15 remove damaged
16 proteins controlling
17 nigro-striatum
18 ity of the prote-
19 ment in the par-
20 is preferentially
21 ATP-independ-
22 rings to make a
23 of seven α and β
24 rings form each
25 mers form the t-
26 nit named 19S
27 20S cylinder pro-
28 activity than the
29 degrade polyubi-
30 case in an ATP-de-

31 In the SH-SY5Y
32 RNS levels only
33 oxidized proteins
34 degradation of
35 in the deposition
36 sured with an ar-
37 leucyl-L-leucyl-L-
38 (Z-Leu-Leu-Glu-
39 way, and virtual-

inhib-
kin-
com-
the
trial
y the
s in t-
d ap-
e wit-
follo-
d pro-
eased

system
proteins
signa-
an bra-
ted, s-
es (53)
tro by
teason
ature,
54). Nor
rings
Lindin
se, PA7
teasome
ome. Th
eins an
s.

one tre-
y and
d progre-
eins ma-
ic activi-
ent sub-
-4-met
in a tin-
ted afte

plex I, rote-
dents, and
quitin and
administra-
on the pro-
enone at a
Y cells (50).
induced in
The oxida-
use of anti-
he levels of
by rotenone

F9

major site to
regulatory pro-
tion. In the
deased activ-
its involve-
ized protein
some in an
osed of four
ing is made
heptameric
ytic β -hepta-
regulatory sub-
ends of the
mer catalytic
teasome can
decarboxy-

deased ROS-
whereas the
suggesting that
only involved
teasome mea-
tobenzoy-L-
ryl-7-amide)
e-dependent
reatment with

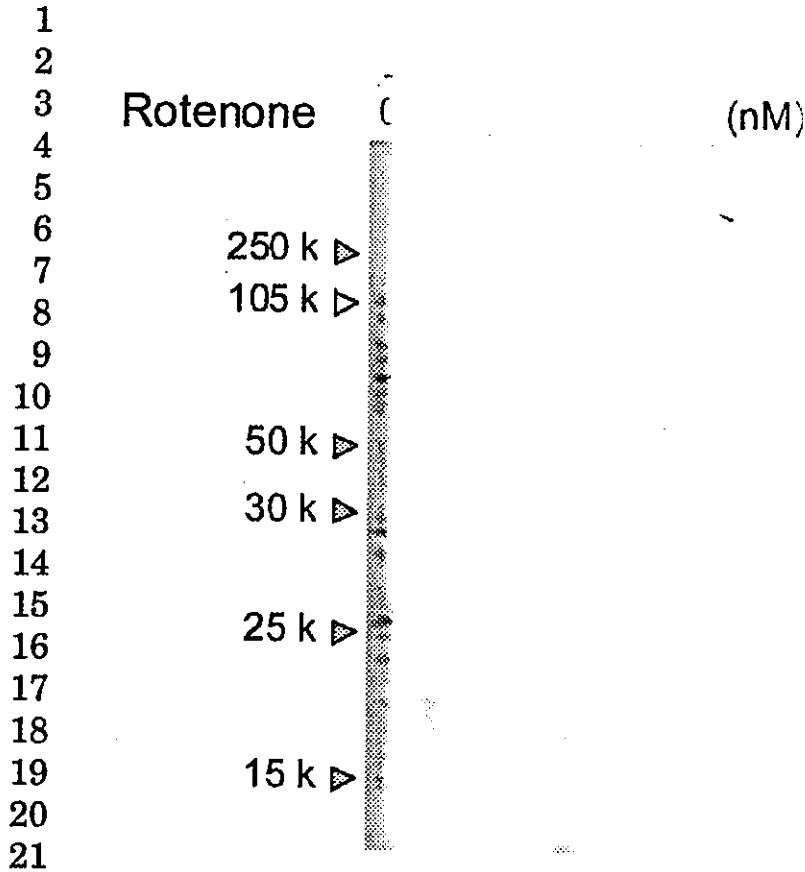


Figure 9 Effect of rotenone on SH-SY5Y cells. The cells were treated with rotenone for 5 days. The cells were fractionated into P1 and P2 fractions, and immunoblotted, using antibodies against aggregated proteins. The aggregated proteins were identified by Western blotting (arrow).

rotenone, as shown in Figure 9. The activity of the proteasome was not affected by rotenone, as shown by the fact that the antiacrolein precipitated with rotenone, suggesting that the aggregation of proteins is not due to a decrease in proteasome activity. In fact, immunoprecipitation analysis of the precipitant from cells treated with rotenone, showed that the aggregated protein

T2

Table 2 Effects of
of Proteasome in SH-SY5Y

Rotenone (25 nM)	ATP
Control	-
	+
Sample	-
	+

Proteasome activity in
rotenone was measured
added ATP (2 mM).

*Mean ± SD of four experiments.

SH-SY5Y
rotenone
(protein)

per 96 hr
11* 19 ± 0.47
12 21 ± 0.11
13 96 ± 0.04
14 30 0.13

of the
-ence
ed with 25 nM
of exogenously

may be a substrate of 20S
are required to cleave
can directly inhibit
4-HNE proteins (Fig. 1)
rotenone treatment
the presence of PMS as de-

. How
acrole
ity, a
in ac
ble w
her.
ther studies
died protein
case of the
ducts after
observed in

V. CONCLUSION

The results of this study
tion may induce degenerative
modification and subsequent
subsequent aggregation
mechanism behind this
requires further investigation
to consider that the inactive
critical role in the activation
generative disorder, these
may induce a neurodegenerative
function, increased
proteasome in neurons,

at mitochondria
pamir
proteasome
ized proteasome
duced
it may
proteasome
ptotic
ental
ween
g, and
the t
dial dysfunction
ns through
system and
The precise
ative stress
ite relevant
may play a
In neurode-
netic factors
ndrial dys-
l activity of
pathological

1 characteristics, co
 2 Our recent studie
 3 the substantia nig
 4 to our hypothesis.
 5 nit precipitated wi
 6 acrolein, whereas
 7 studies may give a
 8 nal cell death in a
 9 us clues for a ne
 10 from apoptosis.

10 cision body.
 11 m system in
 12 er support
 13 OS β subu-
 14 dified with
 15 ed. Future
 16 n of neuro-
 17 d may give
 18 ct neurons

11
 12
 13 **ACKNOWLEDGEMENTS**

14
 15 This work was su
 16 Research on Priori
 17 Promotion of Scie
 18 for Evidence Bas
 19 Ministry of Health

15
 16
 17
 18
 19
 20
 21
 22
 23
 24
 25
 26
 27
 28
 29
 30
 31
 32
 33
 34
 35
 36
 37
 38
 39

20
 21 **REFERENCES**

22
 23 1. Halliwell B. R
 24 system. J Neu
 25
 26 2. Stadtman ER.
 27 1220-1224.
 28
 29 3. Kroemer G, D
 30 death/life reg
 31 Physiol 1998; (1
 32
 33 4. Thompson CG.
 34 disease. Scienc
 35
 36 5. Cotman CW,
 37 Alzheimer's dis
 38
 39 6. Kaytor MD, W
 40 disease. J Biol
 41
 42 7. Beckman JS. (1
 43 peroxynitrite. (1

23
 24
 25
 26
 27
 28
 29
 30
 31
 32
 33
 34
 35
 36
 37
 38
 39

AQ2

- 1 8. Pryor WA, Squibb RL. Nitric oxide: a
2 product from endothelial cells and
3 J Physiol 1998; 401:1-10.
- 4 9. Koppenol WE, Beckman JS. Nitric
5 oxide and superoxide: toxicology and
6 toxicology. *Chem Res Toxicol* 1990;
7 3:317-323.
- 8 10. Kröncke KD, Halliwell B. Nitric
9 oxide: cytotoxicity versus cytoprotection.
10 *Nitric Oxide* 1998; 2:1-10.
- 11 11. Halliwell B. Nitric oxide as a biomarker
12 of oxidative stress. *FEBS Lett* 1997;
13 411:157-160.
- 14 12. White R, Brodeur GM, Griendling
15 W, Gianturco S, Loscalzo J, Loscalzo
16 and peroxynitrite. *Proc Natl Acad Sci USA*
17 1994; 91:1044-1048.
- 18 13. Koppal T, Draetta GF, Hausen L, Butcher
19 synaptosomal membrane. *J Biol Chem*
20 1998; 273:19311-19317.
- 21 14. Smith M, Richey A. Widespread peroxynitrite
22 modification of proteins in Alzheimer's dis-
23 ease. *J Neurosci* 1998; 18:2657-2667.
- 24 15. Good PF, Hsu CY. Nitration in Parkin-
25 son's disease. *Neurosci Biobehav Rev* 1998;
26 22:338-342.
- 27 16. Kato K, Ogino T, Nishida K, Osawa T.
28 Phenolic anti-oxidant modification in
29 collagen. *J Biol Chem* 1998; 273:
30 3004-3009.
- 31 17. Heinecke JW, Eisinger M. Dityrosine,
32 a specific marker of myeloperoxidase-
33 oxidase-hydrogen peroxide metabolism in
34 macrophages. *J Biol Chem* 1993; 268:
35 359-363.
- 36 18. Leeuwenburgh S, Pannathur S, Heinecke
37 JW. Nitration of tyrosine residues: a
38 marker of oxidative stress. *Free Radic Biol Med*
39 1998; 35:1-10.

- 1 intracellular
2 species. *Ni*
- 3 29. Tanaka
4 Shimomura
5 defects of
6 cytochrome
7
8 30. Schapira
9 Marsden
10 son's disease
11
12 31. Mizuno Y,
13 Oya H, Oz
14 of the res
15 Biophys Res
16
17 32. Maruyama
18 human dop
19 tosis induc
20 70:2510-25
21
22 33. Oh-hash
23 Isobe M. N
24 peroxynitri
25 roblastoma
26 1999; 263:
27
28 34. Maruyama
29 Naoi M. Pe
30 age-associat
31
32 35. Yamamoto
33 Tanaka M,
34 drial compl
35 dysfunction
36 J Neural Tra
37
38 36. Hortelano S.
39 nitration an
of mitochon
FASEB J 19
and oxygen
Suzuki H,
Extensive
muscular
:447-454.
Clark JB,
n Parkin-
K, Sato T,
I subunits
Biochem
l protects
from apop-
hem 1998;
Naoi M,
mediates
ergic neu-
Commun
shizume Y,
aging and
:11-18.
-Nagai M,
mitochon-
chondrial
5Y5Y cells.
es tyrosine
n increase
acrophases.
Castro L,
e nitration
15.

1 38. Radi R, et al. Role of mito-
 2 chondria in oxidative stress. *Biochem*
 3 *Biophys Res Commun* 2000; 271:1-6.

4 39. Brorson J, et al. PTEN
 5 inhibits neuronal apoptosis by induc-
 6 ing mitochondrial apoptosis. *Neuron* 2000;
 7 27:103-113.

8 40. Bolanos JP, et al. Mitochondrial
 9 damage and oxidative stress in neuro-
 10 degeneration. *Neurobiol Aging* 2003;
 11 24:1019-1028.

12 41. Beltran M, et al. Mitochondrial
 13 Espluga M, et al. Mitochondrial
 14 dysfunction by oxidative stress: role of
 15 Proc Natl Acad Sci U S A 2003;
 16 100:1200-1205.

17 42. Iwatsubo T, et al. H, I, and J
 18 Trojanowski JQ, et al. Characterization
 19 of Lewy pathology in the brain. *Neuron* 1998;
 20 21:907-920.

21 43. Ii K, Ito T, et al. Histochemical
 22 localization of ubiquitin carboxyl-terminal
 23 hydrolase L1 in neurodegenerative disease.
 24 *Exp Neurol* 2000; 165:25-33.

25 44. Spillantini M, et al. Schmidt M,
 26 Jakes R, et al. *Proc Natl Acad Sci U S A* 1997;
 27 94:837-841.

28 45. Choi P, et al. N, S, and T. Lee JM,
 29 Wolozin L. Mitochondrial dysfunction and
 30 proteasome inhibition in neurodegeneration.
 31 *Neuroreport* 2003; 14:1-3.

32 46. Shimura S, et al. Machuga MT,
 33 Trockenbacher A, et al. Characterization
 34 of the ubiquitin-protein ligase complex
 35 containing parkin and ubiquitin-protein
 36 ligase 1. *Science* 2000; 287:243-249.

37 47. Tanaka Y, et al. Igarashi S, Igarashi
 38 RE, Sawada M, et al. Inducible
 39 expression of alpha-synuclein: role of
 40 proteasome activity in neurodegeneration.
 41 *Mol Cell Neurosci* 2003; 21:1-12.

1 48. Trojanov
2 attraction
3 in Parkin
4 Differ 199
5 49. Souza J
6 linkage
7 J Biol Ch
8 50. Shamoto-
9 Naoi M,
10 rotenone.
11 and indu
12 J Neuro
13 51. Hattori M
14 ical studi
15 Parkinson
16 52. Betarber
17 Panov A
18 sure rep.
19 2000; 3:1
20 53. McNaugh
21 substanti
22 297:191-
23 54. Hough I
24 molecula
25 J Biol Ch
26 55. Okada K,
27 Ucida K.
28 cellular p
29 274:2378
30
31
32
33
34
35
36
37
38
39

6-Y. Fatal
ron death
ell Death

line cross-
polymers.

Yanaka M,
complex I,
modification
Y5Y cells.

AQ4

histochem-
ondria in

Osuna M,
icide expo-
t Neurosci

mpaired in
Lett 2001;

two high
yte lysate.

, Tanak K,
at of intra-
hem 1999;

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39

23

Neuroprotection by Rasagiline and Related Propargylamines is Mediated by Suppression of Mitochondrial Death Signal and Induction of Antiapoptotic Genes

WAKAKO MARUYAMA

Laboratory of Biochemistry and
Metabolism, Department of Basic
Gerontology, National Institute for
Longevity Sciences, Aichi, Japan

ATSUMI NITTA

Department of Pharmacy, Nagoya
University School of Medicine,
Nagoya, Japan

**YUKIHIRO AKAO and
MAKOTO NAOI**

Gifu International Institute of
Biotechnology,
Gifu, Japan

1 **ABSTRACT**

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

In neurodegenerative disorders, such as Parkinson's and Alzheimer's diseases, apoptosis is a major type of neuronal cell death, and the apoptotic cascade has been proposed to be a target of "neuroprotection" through preventing and delaying cell death. A series of propargylamine derivatives have been confirmed to protect neurons against cell death induced by various insults. The mechanism underlying the neuroprotection has been clarified by use of rasagiline [*N*-propargyl-1(*R*) aminoindan], the most potent propargylamine, and human dopaminergic neuroblastoma SH-SY5Y cells. Rasagiline stabilizes the mitochondrial membrane potential, $\Delta\Psi_m$, prevents permeability transition, and suppresses the activation of following apoptotic signal transduction; release of cytochrome *c*, activation of caspase 3, nuclear translocation of glyceraldehydes-3-phosphate dehydrogenase (GAPDH) and fragmentation of nuclear DNA. In addition, rasagiline induces antiapoptotic Bcl-2 and glial cell-line-derived neurotrophic factor (GDNF) in SH-SY5Y cells. In this review, we summarize our recent advances in understanding the mechanism behind the neuroprotection by rasagiline. Rasagiline was found to activate NF- κ B, a nuclear transcription factor playing a critical role in determining cell death/survival pathway. Rasagiline activated I κ B kinase, and active NF- κ B p65 subunit was translocated into nuclei. In addition, gene array analysis revealed that rasagiline increased the expression of the genes coding mitochondrial energy synthesis, apoptosis, transcription, kinases, and ubiquitin-proteasome system, the involvement of which has been proposed in neuronal cell death and accumulation of inclusion bodies in various neurodegenerative disorders. These results are discussed in concern to possibility of neuroprotection by propargylamines in Parkinson's and Alzheimer's diseases.