

celecoxib plus indomethacin. Since many factors can affect the production of gastric lesions *in vivo* (mucosal blood flow and gastric motility for example), a number of interpretations for this phenomenon are possible. However, we consider that the direct cytotoxicity of NSAIDs, or direct cell damage at the gastric mucosa by NSAIDs in other words, can explain this phenomenon. In gastric lesions produced by a combination of the oral administration of celecoxib with the intraperitoneal administration of indomethacin or NCX 530, the direct cell damage at gastric mucosa should occur on account of the orally administered celecoxib. As shown *in vitro*, NCX 530 may suppress celecoxib-induced cell death at the gastric mucosa, meaning that NCX 530 does not actually produce gastric lesions when administered in conjunction with the celecoxib. This idea can also be used to explain the NCX 530-dependent suppression of the production of gastric lesions by ethanol or celecoxib plus indomethacin, given that, *in vitro*, NCX 530 protected the gastric mucosal cells not only from celecoxib but also from ethanol. Furthermore, observations that NCX 530 did not protect gastric mucosal cells from indomethacin *in vitro* may explain why the production of gastric lesions by the oral administration of high doses of indomethacin was not suppressed by NCX 530 *in vivo*. However, in Fig. 8, NCX 530 almost completely inhibited the production of gastric lesions by celecoxib *in vivo*, whereas the effect of this drug on celecoxib-induced cell death is partial *in vitro* (Fig. 2A and 3A). Previous papers reported that NCX 530 stimulated mucosal blood flow and mucus synthesis and did not so clearly increase gastric motility and adhesion of neutrophil as indomethacin (17, 39). We consider that these phenomenon are involved in the safety of NCX 530 on gastric mucosa *in vivo*.

Acknowledgements

We thank NicOx S. A. for providing indomethacin and NCX 530.

REFERENCES

1. Smalley WE, Ray, WA, Daugherty, JR, Griffin, MR: Nonsteroidal anti-inflammatory drugs and the incidence of hospitalizations for peptic ulcer disease in elderly persons. *Am J Epidemiol* 141: 539-545, 1995
2. Hawkey CJ: Nonsteroidal anti-inflammatory drug gastropathy. *Gastroenterology* 119: 521-535, 2000
3. Barrier CH, Hirschowitz, BI: Controversies in the detection and management of nonsteroidal antiinflammatory drug-induced side effects of the upper gastrointestinal tract. *Arthritis Rheum* 32: 926-932, 1989
4. Gabriel SE, Jaakkimainen, L, Bombardier, C: Risk for serious gastrointestinal complications related to use of nonsteroidal anti-inflammatory drugs. A meta-analysis. *Ann Intern Med* 115: 787-796, 1991
5. Fries JF, Miller, SR, Spitz, PW, Williams, CA, Hubert, HB, Bloch, DA: Toward an epidemiology of gastropathy associated with nonsteroidal antiinflammatory drug use. *Gastroenterology* 96: 647-655, 1989
6. Kurata JH, Abbey, DE: The effect of chronic aspirin use on duodenal and gastric ulcer hospitalizations. *J Clin Gastroenterol* 12: 260-266, 1990
7. Singh G: Recent considerations in nonsteroidal anti-inflammatory drug gastropathy. *Am J Med* 105: 31S-38S, 1998

8. Vane JR, Botting, RM: Mechanism of action of anti-inflammatory drugs. *Scand J Rheumatol Suppl* 102: 9-21, 1996
9. Miller TA: Protective effects of prostaglandins against gastric mucosal damage: current knowledge and proposed mechanisms. *Am J Physiol* 245: G601-623, 1983
10. Ligumsky M, Golanska, EM, Hansen, DG, Kauffman, GJ: Aspirin can inhibit gastric mucosal cyclo-oxygenase without causing lesions in rat. *Gastroenterology* 84: 756-761, 1983
11. Ligumsky M, Sestieri, M, Karmeli, F, Zimmerman, J, Okon, E, Rachmilewitz, D: Rectal administration of nonsteroidal antiinflammatory drugs. Effect on rat gastric ulcerogenicity and prostaglandin E2 synthesis. *Gastroenterology* 1245-1249, 1990
12. Lichtenberger LM: Where is the evidence that cyclooxygenase inhibition is the primary cause of nonsteroidal anti-inflammatory drug (NSAID)-induced gastrointestinal injury? Topical injury revisited. *Biochem Pharmacol* 61: 631-637, 2001
13. Lichtenberger LM, Wang, ZM, Romero, JJ, Ulloa, C, Perez, JC, Giraud, MN, Barreto, JC: Non-steroidal anti-inflammatory drugs (NSAIDs) associate with zwitterionic phospholipids: insight into the mechanism and reversal of NSAID-induced gastrointestinal injury. *Nat Med* 1: 154-158, 1995
14. Somasundaram S, Rafi, S, Hayllar, J, Sigthorsson, G, Jacob, M, Price, AB, Macpherson, A, Mahmud, T, Scott, D, Wrigglesworth, JM, Bjarnason, I:

- Mitochondrial damage: a possible mechanism of the "topical" phase of NSAID induced injury to the rat intestine. *Gut* 41: 344-353, 1997
15. Tomisato W, Tsutsumi, S, Rokutan, K, Tsuchiya, T, Mizushima, T: NSAIDs induce both necrosis and apoptosis in guinea pig gastric mucosal cells in primary culture. *Am J Physiol Gastrointest Liver Physiol* 281: G1092-1100, 2001
 16. Tomisato W, Tsutsumi, S, Hoshino, T, Hwang, HJ, Mio, M, Tsuchiya, T, Mizushima, T: Role of direct cytotoxic effects of NSAIDs in the induction of gastric lesions. *Biochem Pharmacol* 67: 575-585, 2004
 17. Takeuchi K, Mizoguchi, H, Araki, H, Komoike, Y, Suzuki, K: Lack of gastric toxicity of nitric oxide-releasing indomethacin, NCX-530, in experimental animals. *Dig Dis Sci* 46: 1805-1818, 2001
 18. Hawkey CJ, Jones, JI, Atherton, CT, Skelly, MM, Bebb, JR, Fagerholm, U, Jonzon, B, Karlsson, P, Bjarnason, IT: Gastrointestinal safety of AZD3582, a cyclooxygenase inhibiting nitric oxide donator: proof of concept study in humans. *Gut* 52: 1537-1542, 2003
 19. Johnson AJ, Hsu, AL, Lin, HP, Song, X, Chen, CS: The cyclo-oxygenase-2 inhibitor celecoxib perturbs intracellular calcium by inhibiting endoplasmic reticulum Ca²⁺-ATPases: a plausible link with its anti-tumour effect and cardiovascular risks. *Biochem J* 366: 831-837, 2002

20. Fiorucci S, Santucci, L, Gresele, P, Faccino, RM, Del Soldato, P, Morelli, A:
Gastrointestinal safety of NO-aspirin (NCX-4016) in healthy human volunteers: a
proof of concept endoscopic study. *Gastroenterology* 124: 600-607, 2003
21. Wallace JL, Reuter, B, Cicala, C, McKnight, W, Grisham, MB, Cirino, G: Novel
nonsteroidal anti-inflammatory drug derivatives with markedly reduced ulcerogenic
properties in the rat. *Gastroenterology* 107: 173-179, 1994
22. Wallace JL, McKnight, W, Del Soldato, P, Baydoun, AR, Cirino, G: Anti-
thrombotic effects of a nitric oxide-releasing, gastric-sparing aspirin derivative. *J Clin
Invest* 96: 2711-2718, 1995
23. Wallace JL, Miller, MJ: Nitric oxide in mucosal defense: a little goes a long way.
Gastroenterology 119: 512-520, 2000
24. Johal K, Hanson, PJ: Opposite effects of flurbiprofen and the nitroxybutyl ester of
flurbiprofen on apoptosis in cultured guinea-pig gastric mucous cells. *Br J Pharmacol*
130: 811-818, 2000
25. Fiorucci S, Santucci, L, Federici, B, Antonelli, E, Distrutti, E, Morelli, O, Renzo,
GD, Coata, G, Cirino, G, Soldato, PD, Morelli, A: Nitric oxide-releasing NSAIDs
inhibit interleukin-1beta converting enzyme-like cysteine proteases and protect
endothelial cells from apoptosis induced by TNFalpha. *Aliment Pharmacol Ther* 13:
421-435, 1999

26. Hirakawa T, Rokutan, K, Nikawa, T, Kishi, K: Geranylgeranylacetone induces heat shock proteins in cultured guinea pig gastric mucosal cells and rat gastric mucosa. *Gastroenterology* 111: 345-357, 1996
27. Tomisato W, Takahashi, N, Komoto, C, Rokutan, K, Tsuchiya, T, Mizushima, T: Geranylgeranylacetone protects cultured guinea pig gastric mucosal cells from indomethacin. *Dig Dis Sci* 45: 1674-1679, 2000
28. Tomisato W, Hoshino, T, Tsutsumi, S, Tsuchiya, T, Mizushima, T: Maturation-associated increase in sensitivity of cultured guinea pig gastric pit cells to hydrogen peroxide. *Dig Dis Sci* 47: 2125-2133, 2002
29. Tsutsumi S, Tomisato, W, Takano, T, Rokutan, K, Tsuchiya, T, Mizushima, T: Gastric irritant-induced apoptosis in guinea pig gastric mucosal cells in primary culture. *Biochim Biophys Acta* 1589: 168-180, 2002
30. Katsu T: Application of calcein-loaded liposomes for the determination of membrane channel size. *Biol Pharm Bull* 22: 978-980, 1999
31. New RRC: *Liposomes: a practical approach*, p. 105-161: IRL Press Oxford, 1990.
32. Tomisato W, Tanaka, K, Katsu, T, Kakuta, H, Sasaki, K, Tsutsumi, S, Hoshino, T, Aburaya, M, Li, D, Tsuchiya, T, Suzuki, K, Yokomizo, K, Mizushima, T: Membrane permeabilization by non-steroidal anti-inflammatory drugs. *Biochem Biophys Res Commun* 323: 1032-1039, 2004
33. Lee BS, Chen, J, Angelidis, C, Jurivich, DA, Morimoto, RI: Pharmacological modulation of heat shock factor 1 by antiinflammatory drugs results in protection

- against stress-induced cellular damage. *Proc Natl Acad Sci U S A* 92: 7207-7211, 1995
34. Tsutsumi S, Gotoh, T, Tomisato, W, Mima, S, Hoshino, T, Hwang, HJ, Takenaka, H, Tsuchiya, T, Mori, M, Mizushima, T: Endoplasmic reticulum stress response is involved in nonsteroidal anti-inflammatory drug-induced apoptosis. *Cell Death Differ*, 2004
35. Kim YM, Talanian, RV, Billiar, TR: Nitric oxide inhibits apoptosis by preventing increases in caspase-3-like activity via two distinct mechanisms. *J Biol Chem* 272: 31138-31148, 1997
36. Brunner F, Stessel, H, Kukovetz, WR: Novel guanylyl cyclase inhibitor, ODQ reveals role of nitric oxide, but not of cyclic GMP in endothelin-1 secretion. *FEBS Lett* 376: 262-266, 1995
37. Fiorucci S, Antonelli, E, Santucci, L, Morelli, O, Miglietti, M, Federici, B, Mannucci, R, Del Soldato, P, Morelli, A: Gastrointestinal safety of nitric oxide-derived aspirin is related to inhibition of ICE-like cysteine proteases in rats. *Gastroenterology* 116: 1089-1106, 1999
38. Yeh RK, Chen, J, Williams, JL, Baluch, M, Hundley, TR, Rosenbaum, RE, Kalala, S, Traganos, F, Benardini, F, del Soldato, P, Kashfi, K, Rigas, B: NO-donating nonsteroidal antiinflammatory drugs (NSAIDs) inhibit colon cancer cell growth more potently than traditional NSAIDs: a general pharmacological property? *Biochem Pharmacol* 67: 2197-2205, 2004

39. Mizoguchi H, Hase, S, Tanaka, A, Takeuchi, K: Lack of small intestinal ulcerogenicity of nitric oxide-releasing indomethacin, NCX-530, in rats. *Aliment Pharmacol Ther* 15: 257-267, 2001
40. Mukherjee D, Nissen, SE, Topol, EJ: Risk of cardiovascular events associated with selective COX-2 inhibitors. *Jama* 286: 954-959, 2001
41. Mukherjee D: Selective cyclooxygenase-2 (COX-2) inhibitors and potential risk of cardiovascular events. *Biochem Pharmacol* 63: 817-821, 2002
42. McAdam BF, Catella, LF, Mardini, IA, Kapoor, S, Lawson, JA, FitzGerald, GA: Systemic biosynthesis of prostacyclin by cyclooxygenase (COX)-2: the human pharmacology of a selective inhibitor of COX-2. *Proc Natl Acad Sci U S A* 96: 272-277, 1999
43. Catella LF, McAdam, B, Morrison, BW, Kapoor, S, Kujubu, D, Antes, L, Lasseter, KC, Quan, H, Gertz, BJ, FitzGerald, GA: Effects of specific inhibition of cyclooxygenase-2 on sodium balance, hemodynamics, and vasoactive eicosanoids. *J Pharmacol Exp Ther* 289: 735-741, 1999
44. Belton O, Byrne, D, Kearney, D, Leahy, A, Fitzgerald, DJ: Cyclooxygenase-1 and -2-dependent prostacyclin formation in patients with atherosclerosis. *Circulation* 102: 840-845, 2000
45. Hennen JK, Huang, J, Barrett, TD, Driscoll, EM, Willens, DE, Park, AM, Crofford, LJ, Lucchesi, BR: Effects of selective cyclooxygenase-2 inhibition on vascular

responses and thrombosis in canine coronary arteries. *Circulation* 104: 820-825, 2001

46. Dowd NP, Scully, M, Adderley, SR, Cunningham, AJ, Fitzgerald, DJ: Inhibition of cyclooxygenase-2 aggravates doxorubicin-mediated cardiac injury in vivo. *J Clin Invest* 108: 585-590, 2001
47. Katsu T, Kobayashi, H, Hirota, T, Fujita, Y, Sato, K, Nagai, U: Structure-activity relationship of gramicidin S analogues on membrane permeability. *Biochim Biophys Acta* 899: 159-170, 1987

FIGURE LEGENDS

Fig. 1. Necrosis and apoptosis induced by NCX 530 or indomethacin.

Cultured guinea pig gastric mucosal cells were incubated with indicated concentrations of NCX 530 or indomethacin for 1 h (A) (necrotic conditions) or 16 h (B, C) (apoptotic conditions). Cell viability was determined by the MTT method (A, B). Values are mean \pm S.E.M. (n=3). *** P <0.001; ** P <0.01. Chromosomal DNA was extracted and analyzed by 2% agarose gel electrophoresis (C).

Fig. 2. Effect of NCX 530 on apoptosis induced by various gastric irritants.

Cultured guinea pig gastric mucosal cells were pre-incubated with indicated concentrations of NCX 530 for 1 h and further incubated with 0.1 mM celecoxib (A), 3% ethanol (B), 0.6 mM indomethacin (C) or 0.4 mM hydrogen peroxide (D) in the presence of indicated concentrations of NCX 530 for 16 h (apoptotic conditions). Cell viability was determined by the MTT method. Values are mean \pm S.E.M. (n=3). ** P <0.001; ** P <0.01; * P <0.05.

Fig. 3. Effect of NCX 530 on necrosis induced by various gastric irritants.

Cultured guinea pig gastric mucosal cells were pre-incubated with indicated concentrations of NCX 530 for 1 h and further incubated with 0.18 mM celecoxib (A), 8% ethanol (B), 3 mM indomethacin (C) or 1 mM hydrogen peroxide (D) in the presence of

indicated concentrations of NCX 530 for 1 h (necrotic conditions). Cell viability was determined by the MTT method. Values are mean \pm S.E.M. (n=3). *** P <0.001; ** P <0.01; * P <0.05.

Fig. 4. Membrane permeabilization by NSAIDs.

Calcein-loaded liposomes were incubated for 10 min at 30°C with indicated concentrations of each NSAID (A) or 0.1 mM celecoxib plus indicated concentrations of NCX530 (B). The release of calcein from liposomes was determined by measuring fluorescence intensity. Melittin (10 μ M) was used to determine the 100% level of membrane permeabilization (47).

Fig. 5. Effect of cycloheximide on cell viability in the presence of NCX 530.

Cultured guinea pig gastric mucosal cells were pre-incubated with indicated concentrations of cycloheximide for 1 h. Cells were further incubated with 2 mM NCX 530 and indicated concentrations of cycloheximide for 1 h (A) (necrotic conditions). Cells were pre-incubated with indicated concentrations of cycloheximide and 1 mM NCX 530 for 1 h. Cells were further incubated with 1 mM NCX 530, 0.18 mM celecoxib and indicated concentrations of cycloheximide for 1 h (B) (necrotic conditions). Cell viability was determined by the MTT method. Values are mean \pm S.E.M. (n=3).

Fig. 6. Effect of ODQ on cell viability in the presence of NCX 530.

Cultured guinea pig gastric mucosal cells were pre-incubated with indicated concentrations of ODQ for 1 h. Cells were further incubated with indicated concentrations of NCX 530 and ODQ (A, C). Cells were pre-incubated with indicated concentrations of ODQ and 1 mM NCX 530 for 1 h. Cells were further incubated indicated concentrations of ODQ, NCX530 and celecoxib (B, D). Incubation was performed for 1 h (A, B) (necrotic conditions) or for 16 h (C, D) (apoptotic conditions). Cell viability was determined by the MTT method. Values are mean \pm S.E.M. (n=3). *** P <0.001; * P <0.05.

Fig. 7. Production of gastric lesions by NCX 530 or indomethacin.

Rats were orally administered with NCX 530 or indomethacin as indicated. After 6 h, the stomach was removed and scored for hemorrhagic damage. Values are mean \pm S.E.M. (n=5 - 6). ** P <0.01. n. d.; not detected

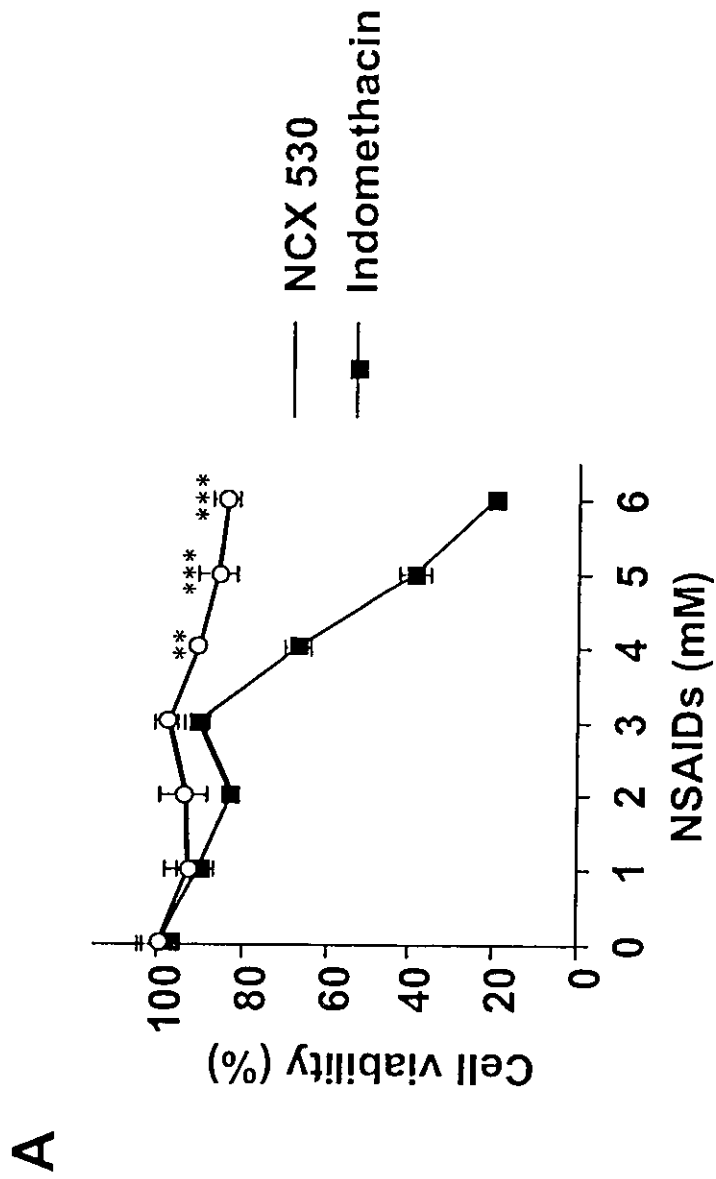
Fig. 8. Production of gastric lesions by NCX 530 or indomethacin in combination with celecoxib.

Rats were intraperitoneally administered with 5 mg/kg indomethacin or 7.1 mg/kg NCX 530 or vehicle. After 1 h, animals were administered orally with 15 mg/ml celecoxib or vehicle. After 6 h, the stomach was removed and scored for hemorrhagic damage. Values are mean \pm S.E.M. (n=5 - 6). ** P <0.01; * P <0.05. n. d.; not detected

Fig. 9. Effect of NCX 530 on production of gastric lesions by other gastric irritants.

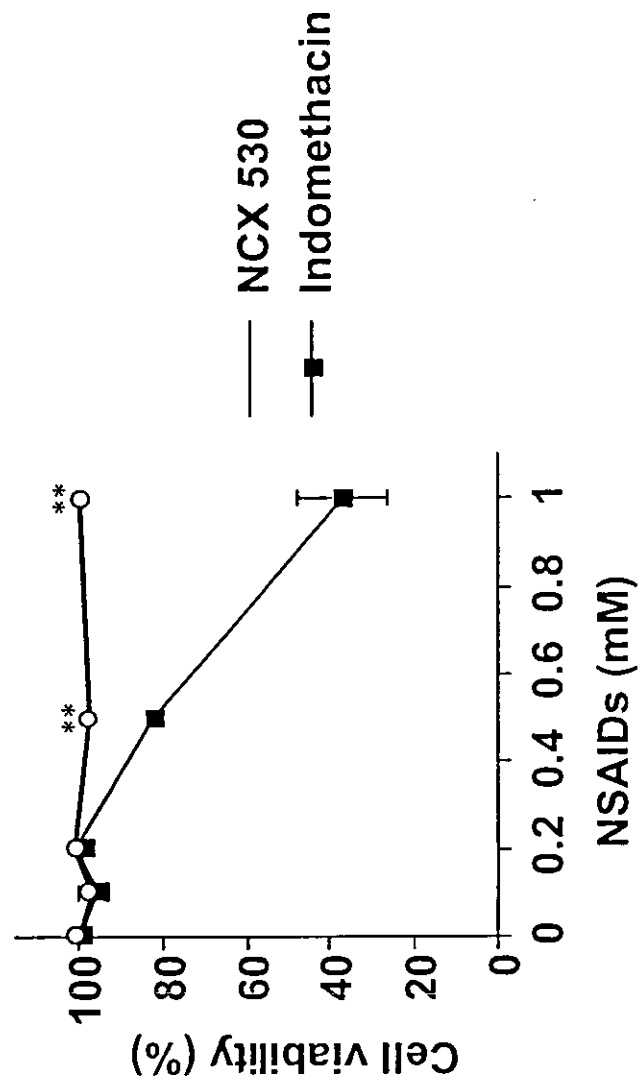
Rats were intraperitoneally administered with 7.1 mg/kg NCX 530 or 5 mg/kg indomethacin or vehicle. After 1 h, animals were administered orally with ethanol (A) or 30 mg/kg indomethacin (B) or vehicle. After 6 h, the stomach was removed and scored for hemorrhagic damage. Values are mean \pm S.E.M. (n=5 - 6). * P <0.05. n. d.; not detected

Tomisato et al. Fig. 1



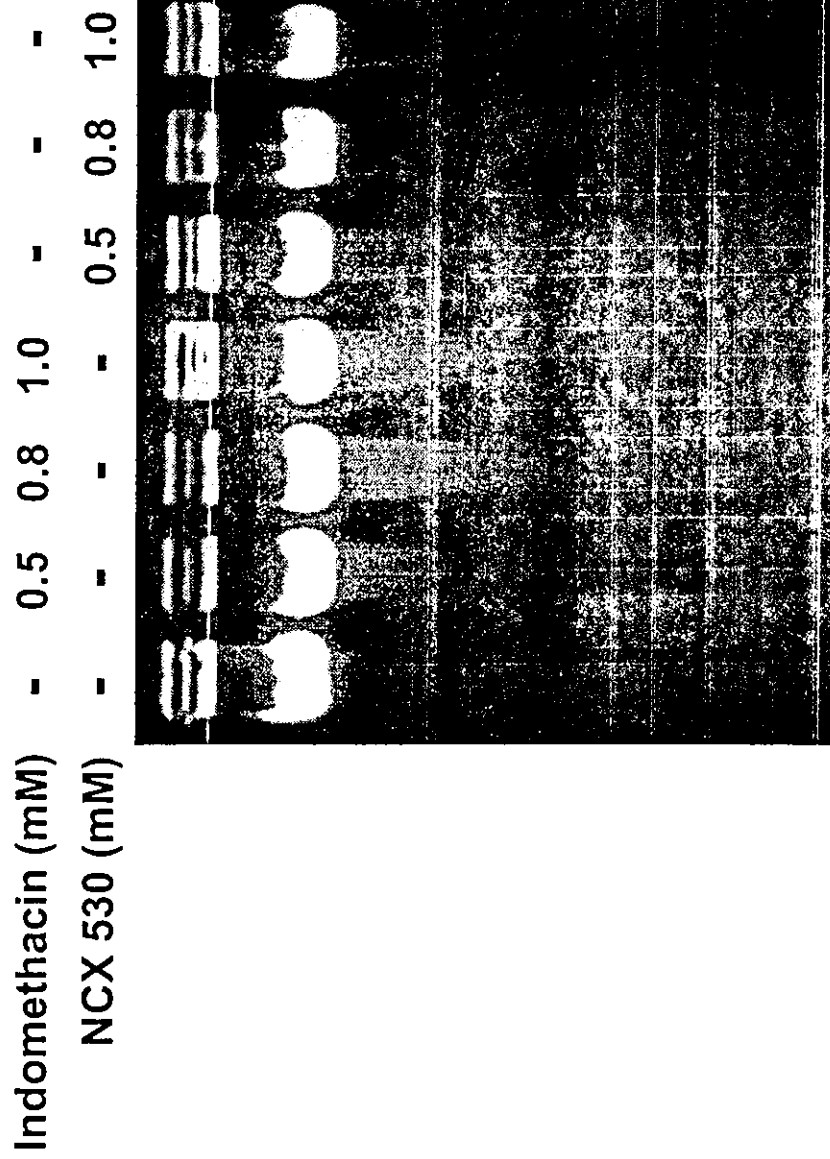
Tomisato et al. Fig. 1

B

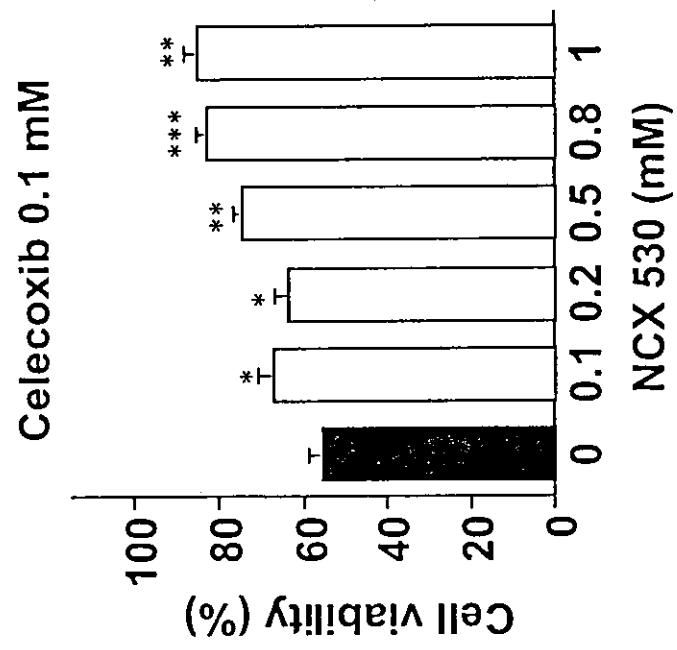


Tomisato et al. Fig. 1

C

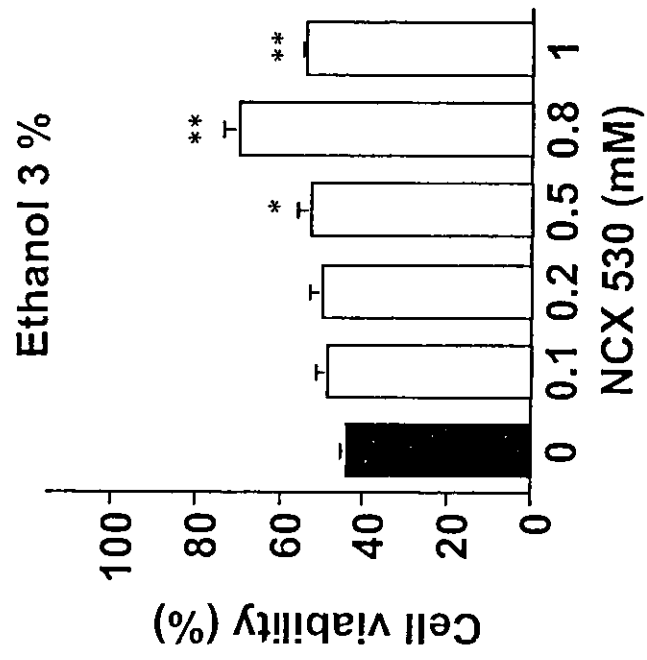


A



Tomisato et al. Fig. 2

B



Tomisato et al. Fig. 2

C

