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IV 研究成果の刊行物・別刷

Endothelial progenitor cells: past, state of the art, and future

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Abstract

Recent evidences suggest that endothelial progenitor cells (EPCs) derived from bone marrow (BM) contribute to *de novo* vessel formation in adults occurring as physiological and pathological responses. Emerging preclinical trials have shown that EPCs home to sites of neovascularization after ischemic events in limb and myocardium. On the basis of these aspects, EPCs are expected to develop as a key strategy of therapeutic applications for the ischemic organs. Such clinical requirements of EPCs will tentatively accelerate the translational research aiming at the devices to acquire the optimized quality and quantity of EPCs. In this review, we attempt to discuss about biological features of EPCs and speculate on the clinical potential of EPCs for therapeutic neovascularization.

Keywords: endothelial progenitor cell (EPC) • vasculogenesis • angiogenesis • therapeutic neovascularization
• cardiovascular disease • cell therapy

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Introduction

The identification of EPCs derived from BM was an outstanding event of stem cell biology in the field of vascular biology. This unique cell population existing in peripheral blood mononuclear cells (PBMNCs) derived from BM shares a similar profile to that of hematopoietic stem cells (HSCs) and incorporates into foci of physiological or pathological neovascularization in response to various angiogenic growth factors. Considering the importance of blood vessel formation on embryonic organogenesis, the development of tissue and organ regeneration could not be able to be realized without understanding the biological mechanisms of vasculogenesis by EPCs. This review provides an update of EPC biology as well as highlighting their potential utility for therapeutic neovascularization.

Post-natal vasculogenesis

EPCs, HSCs related descendants, have been isolated from human adult PBMNCs [1, 2]. Flk-1 and CD34 antigens were used to detect putative EPCs [3]. This methodology was supported by former findings that embryonic HSCs and EPCs share certain antigenic determinants, including Flk-1, Tie-2, c-Kit, Sca-1, CD133, and CD34. These progenitor cells have consequently been considered to be derived from a common precursor, putatively termed 'hemangioblast'.

In vitro, EPCs differentiated into endothelial lineage cells, and in animal models of ischemia, heterologous, homologous, and autologous EPCs were shown to incorporate into sites of active neovascularization. This finding was followed by diverse identifications of EPCs by several groups [4–7] using equivalent or different methodologies. Recently, similar studies with EPCs isolated from human cord blood have demonstrated their analogous differentiation into ECs in vitro and in vivo [8, 9]. These findings, together with other recent studies [10, 11], are consistent with the notion of post-natal "vasculogenesis", which is de novo vessel formation by *in situ* incorporation, differentiation, migration, and/or proliferation of BM-derived EPCs [3] (Fig. 1).

Several studies have demonstrated that BM-derived EPCs functionally contribute to vasculogenesis during wound healing [12], limb ischemia [1, 3, 13–17], postmyocardial infarction [18, 19], endothelialization of vascular grafts [2, 12, 20, 21], or physiological cyclic organogenesis of endometrium [3] under the influence of appropriate cytokines, growth factors and/or hormones through the autocrine, paracrine, and/or endocrine systems.

These findings have raised important questions regarding fundamental concepts of blood vessel growth and development in adult subjects. Does the differentiation of EPCs *in situ* (vasculogenesis) play an important role in adult neovascularization, and would impairments in this process lead to clinical diseases? There is now a strong body of evidence suggesting that vasculogenesis does, in fact, make a significant contribution to postnatal neovascularization. Recent studies with animal bone marrow transplantation (BMT) models in which BM (donor)-derived EPCs could be distinguished have shown that the contribution of EPCs to neovessel formation may range from 5 to 25% in response to granulation tissue formation [22] or growth factor-induced neovascularization [23]. Also, in the tumor neovascularization, the range is approximately 35–45% higher than the former events [24]. The degree of EPC contribution to post-natal neovascularization is predicted to depend on each vessel formation event or disease.

More recently, Tamaki et al. reported that tissue specific stem/progenitor cells with the potency of differentiation into myocytes or ECs were isolated in skeletal muscle tissue of murine hindlimb, although the origin remains to be clarified [25]. This studies have introduced the concept that the origin of EPCs may not be limited to BM, *e.g.* tissue specific stem/progenitor cells possibly provide '*in situ* EPCs' as other sources of EPCs than BM. (Fig. 1)

Profiles of EPCs in adults

Since the initial report of EPCs [1][2], a number of groups have set out to define this cell population more profoundly. Because EPCs and HSCs share many surface markers, and no simple definition of EPCs exists, various methods of EPC isolation have

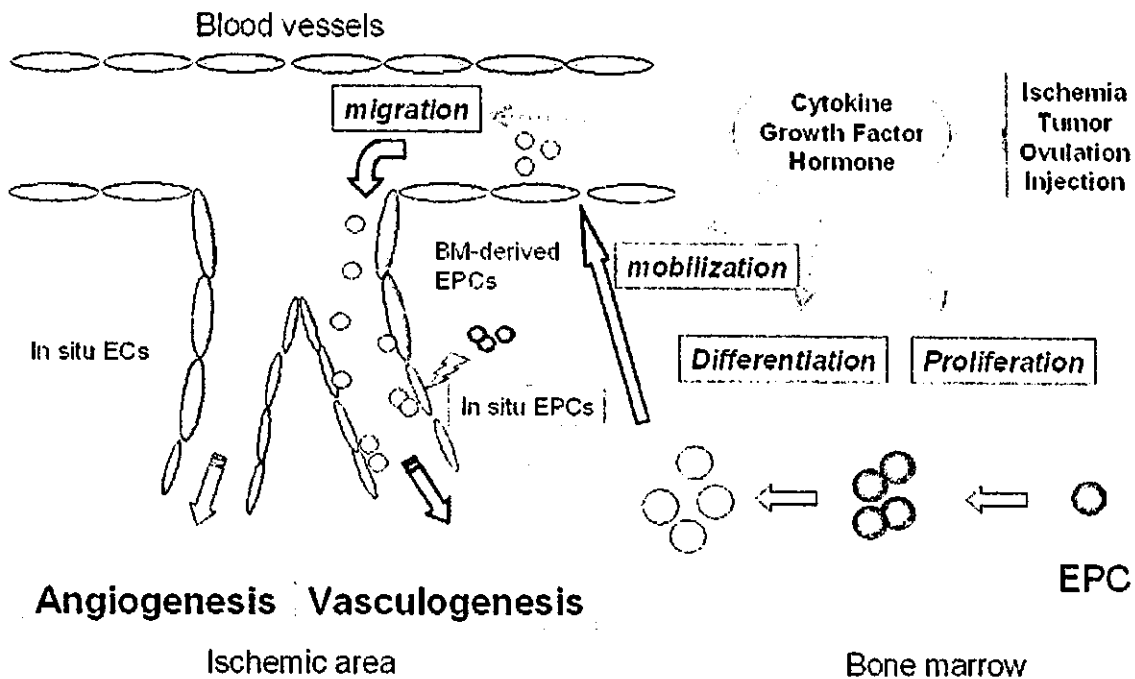


Fig. 1 Post-natal neovascularization in the physiological or pathological events is consistent with neovessel formation contributed by angiogenesis and vasculogenesis at the various rates between their two mechanisms. Angiogenesis and vasculogenesis are due to the activations of *in situ* ECs and BM-derived or *in situ* EPCs, respectively.

been reported [1, 2, 4, 6–9, 15, 16]. The term of EPC may therefore encompass a group of cells that exist in a variety of stages ranging from hemangioblast to fully differentiated endothelial cell (EC).

Under the current status, it is impossible to differentiate 'immature EPCs' from primitive HSCs, as those cells share common surface markers, i.e. CD133, CD34, or VEGFR2 (KDR). In circulation, the cell population with the capacity of differentiation to EPCs is considered to be included in the cell population expressing CD133 and VEGFR2 markers in the subset of CD34 positive cells [7]. Circulating EPCs are constitutively expressing stem/progenitor markers, i.e. CD34 or VEGFR2 except CD133, and start expressing endothelial lineage specific markers, VE cadherin or E-selectin. On the other hand, following the commitment and differentiation to hematopoietic stem/progenitor cells, the surface markers of CD133 and VEGFR2 are extinguished. Such stem/progenitor cell markers do not express on the differentiated hematopoietic cells. Alternatively, kinds of surface markers are expressed to characterize individual hematopoietic cell populations. CD133 is a marker to differ-

entiate immature EPCs or primitive HSCs from circulating EPCs. To differentiate EPCs from hematopoietic stem/progenitor cells, VE cadherin or E-selectin are useful. Accordingly, circulating EPCs may be isolated via selection by the antigenicity of CD34, VEGFR2, and/or VE cadherin and also circulating immature EPCs by CD133 (Fig. 2).

In adult human body, there is a strong evidence to suggest that impaired neovascularization results in part from diminished cytokine production. However, endogenous expression of cytokines is not the only factor leading to impaired neovascularization. Diabetic or hypercholesterolemic animals-like clinical patients-exhibit the evidence of dysfunction in mature endothelial cells. While the cellular dysfunction does not necessarily preclude a favorable response to cytokine replacement therapy, the extent of recovery in limb perfusion in these animals fails to reach that of control animals; this suggests another limitation imposed by a diminished responsiveness of EPCs/ECs. Recently Vasa *et al.* have further investigated EPC kinetics and their relationship to clinical disorders, showing that

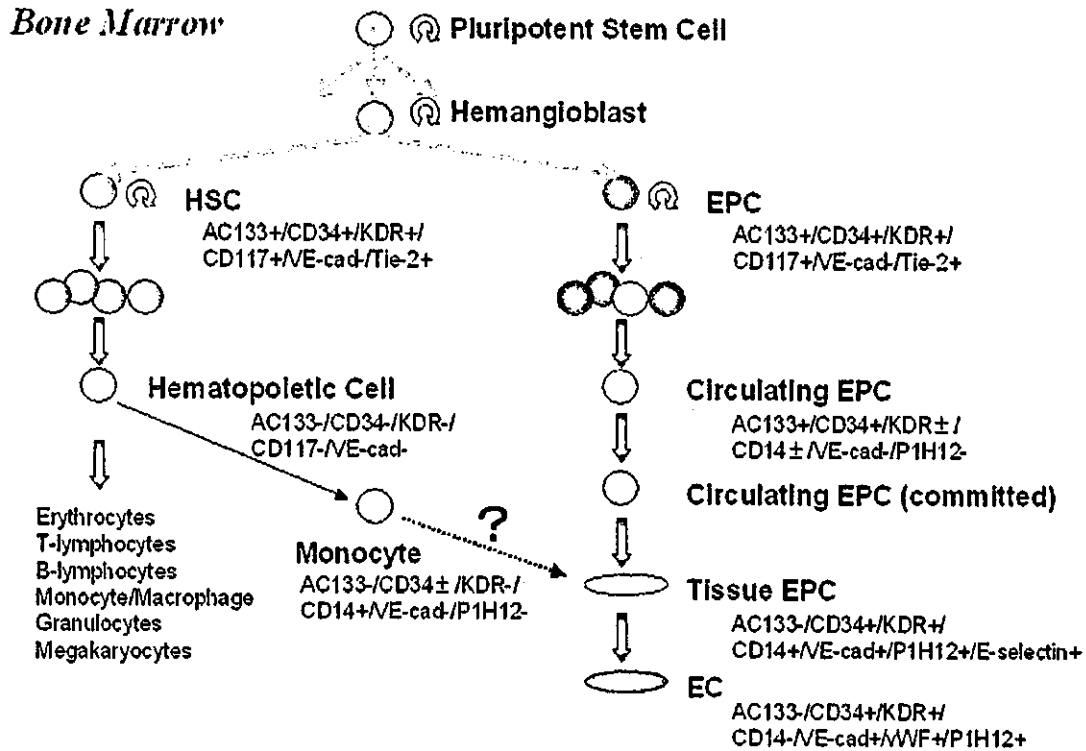


Fig. 2 Putative cascade and expressional profiles of human bone marrow-derived endothelial progenitor cell differentiation. (+: positive, -: negative).

the number and migratory activity of circulating EPCs inversely correlate with risk factors for coronary artery disease, such as smoking, family history and hypertension [26]. Tepper *et al.* reported that proliferation and tube formation of EPCs were down regulated in patients with type 2 diabetes compared with normal subjects [27]. Valgimigli *et al.* indicated that circulating EPCs decreased in patients with severe heart failure (HF) [28]. On the basis of these findings, monitoring of BM-derived EPC kinetics in the patients with vascular diseases is expected to be valuable in the evaluation of lesion activity and/or therapeutic efficacy.

The aging characterized by impaired neovascularization might be also associated with dysfunctional EPCs and defective vasculogenesis. Indeed, preliminary results from our laboratory indicated that the replacement of native bone marrow (including its compartment of progenitor cells) of

young mice with bone marrow transplanted from old animals leads to a marked reduction in neovascularization following corneal micropocket injury, compared with young mice transplanted with young bone marrow. These studies thus established evidence of an age-dependent impairment in vasculogenesis (as well as angiogenesis) and the origin of progenitor cells as a critical parameter influencing neovascularization. Moreover, analysis of clinical data in older patients disclosed a significant reduction in the number of circulating EPCs before and after VEGF165 gene transfer; specifically, the number of circulating EPCs of younger patients with critical limb ischemia was five times more than the number in older individuals. Impaired EPC mobilization and/or activity in response to VEGF may thus contribute to the age-dependent defect in postnatal neovascularization.

Regulation of EPC Mobilization

EPC kinetics in adults

Given the result of common antigenicity, BM has been considered the origin of EPCs as HSCs in adults. The BMT experiments have demonstrated the incorporation of BM-derived EPCs into foci of physiological and pathological neovascularization [3]. Wild-type mice were lethally irradiated and transplanted with BM harvested from transgenic mice in which constitutive LacZ expression is regulated by an EC-specific promoter: Flk-1 or Tie-2. Histological examination of the tissues in growing tumors, healing wounds, ischemic skeletal and cardiac muscles, and cornea micropocket surgery after BMT has shown localization of Flk-1- or Tie-2-expressing endothelial lineage cells derived from BM in blood vessels and stroma around vasculatures. The similar incorporation was observed in physiological neovascularization in uterus endometrial formation after induced ovulation as well as estrogen administration [3].

Previous investigators have shown that wound trauma causes mobilization of hematopoietic cells, including pluripotent stem or progenitor cells in spleen, bone marrow, and peripheral blood. Consistent with EPC/HSC common ancestry, the recent data have shown that mobilization of BM-derived EPCs constitutes a natural response to tissue ischemia. The former murine BMT model presented the direct evidence of enhanced BM-derived EPC incorporation into foci of corneal neovascularization after the development of hindlimb ischemia. Light microscopic examination of corneas excised 6 days after micropocket injury and concurrent surgery to establish hindlimb ischemia demonstrated a statistically significant increase in cells expressing α -galactosidase in the corneas of mice with, versus those without, an ischemic limb [17]. This finding indicates that circulating EPCs are mobilized endogenously in response to tissue ischemia, following the incorporation of EPCs into the foci neovascularization to promote tissue repair. Moreover, such concept were also reflected in clinical findings of EPC mobilization in patients with coronary artery bypass grafting, burns [12], and acute myocardial infarction [19].

EPC mobilization by endogenous agents

Having demonstrated the potential for endogenous mobilization of BM-derived EPCs, we considered that artificial expansion and mobilization of this putative EC precursor population might represent an effective means to augment the resident population of ECs that is competent to respond to administered angiogenic cytokines. Such a program might thereby address the issue of endothelial dysfunction or depletion that may compromise strategies of therapeutic neovascularization in older, diabetic, and/or hypercholesterolemic animals and patients. Granulocyte macrophage colony-stimulating factor (GM-CSF) is well known to stimulate hematopoietic progenitor cells and myeloid lineage cells, but has recently been shown to exert a potent stimulatory effect on EPC kinetics. The delivery of this cytokine induced EPC mobilization and enhanced neovascularization of severely ischemic tissues and de novo corneal vascularization [17].

Among other growth factors, vascular endothelial growth factor (VEGF), critical for angiogenesis in the embryo, has recently been shown to be the critical factor for vasculogenesis and angiogenesis. Our studies carried out first in mice [13] and subsequently in patients undergoing VEGF gene transfer for critical limb or myocardial ischemia [29] established that a previously unappreciated mechanism by which VEGF contributes to neovascularization is in part by mobilizing BM-derived EPCs. Similar modulation of EPC kinetics has been observed in response to other hematopoietic stimulators, such as granulocyte-colony stimulating factor (G-CSF), angiopoietin-1 [30], stroma-derived factor-1 (SDF-1) [31], and erythropoietin [32], or endogenous hormone, estrogen [33, 34].

EPC mobilization by exogenous agents

This potent therapeutic strategy of EPC mobilization has recently been implicated not only by natural hematopoietic or angiogenic stimulants but also by recombinant pharmaceuticals. The statins inhibit the activity of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, which catalyzes the synthesis of mevalonate, a rate-limiting step in cholesterol biosynthesis. The statins rapidly activate Akt signaling in ECs, thereby stimulating EC

bioactivity *in vitro* and enhancing angiogenesis *in vivo* [35]. Recently, we [36] and Dimmeler and colleagues [37] demonstrated a novel function for HMG-CoA reductase inhibitors that contributes to postnatal neovascularization by augmented mobilization of BM-derived EPCs through stimulation of the Akt signaling pathway. With regard to its pharmacological safety and effectiveness on hypercholesterolemia, one of the risk factors for atherosclerosis, the statin might be a potent medication against atherosclerotic vascular diseases.

On the other hand, some antiangiogenic agents, *i.e.* angiostatin or soluble flk-1, have been shown to inhibit BM-derived EPC kinetics, leading to tumor regression, as BM-derived EPC kinetics is a critical factor for tumor growth, in terms of tumor neovascularization [38].

Therapeutic potential of EPC transplantation

The regenerative potential of stem cells is presently under intense investigation. *In vitro*, stem and progenitor cells possess the capability of self-renewal and differentiation into organ-specific cell types. When placed *in vivo*, these cells are then provided with the proper milieu that allows them to reconstitute organ systems. We therefore considered a novel strategy of EPC transplantation to provide a source of robust ECs that might supplement fully differentiated ECs thought to migrate and proliferate from preexisting blood vessels according to the classic paradigm of angiogenesis developed by Folkman and colleagues.

Although it is not known whether local administration of exogenous EPCs may augment tumor neovascularization, this issue should be carefully considered for clinical application of EPC cell therapy to treat cardiovascular diseases.

Indications of EPC transplantation

Three kinds of clinical states could be currently applied to indications of EPC transplantation. (1) Critical limb ischemia such as arteriosclerosis obliterans (ASO) or Burger disease, (2) Post myocardial infarction which is excluded from percutaneous

catheter intervention (PCI) or coronary artery bypass grafting (CABG). (3) Vascular graft as a means of improving biocompatibility.

(1) Our studies indicated that cell therapy with *ex vivo* expanded EPCs could successfully promote neovascularization of ischemic tissues, even when administered as 'sole therapy,' *i.e.* in the absence of angiogenic growth factors. Such a 'supply-side' version of therapeutic neovascularization in which the substrate (EPCs/ECs) rather than ligand (growth factor) comprises the therapeutic agent, was first demonstrated by intravenously transplanting human EPCs to immunodeficient mice with hindlimb ischemia [15]. Not only did the heterologous cell transplantation improve neovascularization and blood flow recovery, but also led to important biological outcomes-notably, the reduction of limb necrosis and auto-amputation by 50% in comparison with controls. Murohara *et al.* reported similar findings in which human cord blood-derived EPCs also augmented neovascularization in a hindlimb ischemic model of nude rats, followed by *in situ* transplantation [9]. In addition, Shatteman *et al.* [16] conducted local injection of freshly isolated human CD34⁺ MNCs into diabetic nude mice with hindlimb ischemia and showed an increase in the restoration of limb flow. These findings provided novel evidence that exogenously administered EPCs rescue impaired neovascularization in an animal model of critical limb ischemia.

(2) A similar strategy with limb ischemia applied to a model of myocardial ischemia in the nude rat demonstrated that transplanted human EPCs localize to areas of myocardial neovascularization, differentiate into mature ECs and enhance neovascularization. These findings were associated with preserved left ventricular (LV) function and diminished myocardial fibrosis [39]. Kocher *et al.* attempted intravenous infusion of freshly isolated human CD34⁺ MNCs into nude rats with myocardial ischemia, and found preservation of LV function associated with inhibition of cardiomyocyte apoptosis [40]. These strategies resulted in preservation of LV function associated with inhibition of cardiomyocyte apoptosis. These experimental findings obtained using immunodeficient animals suggest that both cultured and freshly isolated human EPCs have therapeutic potential in peripheral and coronary artery diseases.

(3) EPCs have recently been applied to the field of tissue engineering as a means of improving biocompatibility of vascular grafts. Artificial grafts first seeded with autologous CD34⁺ cells from canine bone marrow and then implanted into the aorta were found to have increased surface endothelialization and vascularization compared with controls [20]. Similarly, when cultured autologous ovine EPCs were seeded onto carotid interposition grafts, the EPC-seeded grafts achieved physiological motility and remained patent for 130 days vs. 15 days in nonseeded grafts [21].

Cell source and modification of EPC for transplantation

A critical limitation for the therapeutic application of postnatal EPCs is their low number in the circulation. Especially patients with cardiovascular risk factors, aging, or HF who are the candidate for cell therapy have been considered to possess lower EPCs.

Ex vivo expansion of EPCs cultured from PBMCs of healthy human volunteers typically yields 5.0×10^6 cells per 100 ml of blood on day 7. Our animal studies [15] suggest that heterologous transplantation requires systemic injection of $0.5\text{--}2.0 \times 10^4$ human EPCs/g body weight of the recipient animal to achieve satisfactory reperfusion of an ischemic hindlimb. Rough extrapolation of these data to human suggests that a blood volume of as much as 12 l may be necessary to obtain adequate numbers of EPCs to treat critical limb ischemia in patients.

Considering autologous EPC therapy, certain technical improvements that may help to overcome the primary scarcity of a viable and functional EPC population should include: (1) local delivery of EPCs, (2) adjunctive strategies (e.g. growth factor, cytokine, or drugs) to promote BM-derived EPC mobilization [13, 17], (3) enrichment procedures, *i.e.* leukapheresis or BM aspiration, or (4) enhancement of EPC function by gene transduction, (5) *ex vivo* expanded EPCs from self-renewable primitive stem cells in BM or other tissues, (6) allogenic EPCs derived from umbilical cord blood (Fig. 3).

These approaches of EPC modification to acquire the ideal quality and quantity of EPCs for

EPC therapy have already been applied to clinical patients in some institutions and preliminary results are expected to come out in the near future.

In some cases, nonselected total BM cells or BM-MNCs including immature EPC population have also been investigated for their potential to induce neovascularization. Several experiments have reported that autologous BM administration into hindlimb ischemic model and myocardial ischemic model, and could augment neovascularization in ischemic tissue mainly through the production of angiogenic growth factors and less through the differentiation of a portion of the cells into EPCs/ECs *in situ*. Although there are no long-term safety and efficacy data for local delivery of such cell population mostly composed of inflammatory leukocytes, these strategies have already been investigated in some institutions.

Gene modified EPC therapy

A strategy that may alleviate potential EPC dysfunction in ischemic disorders is considered reasonable, given the findings that EPC function and mobilization may be impaired in certain disease states. Genetic modification of EPCs to overexpress angiogenic growth factors, to enhance signaling activity of the angiogenic response, and to rejuvenate the bioactivity and/or extend the life span of EPCs, can constitute such potential strategies.

We have recently shown for the first time that gene-modified EPCs rescue impaired neovascularization in an animal model of limb ischemia [14]. Transplantation of heterologous EPCs transduced with adenovirus encoding human VEGF165 not only improved neovascularization and blood flow recovery, but also had meaningful biological consequences, *i.e.* limb necrosis and auto-amputation were reduced by 63.7% in comparison with controls. Notably, the dose of EPCs needed to achieve limb salvage in these *in vivo* experiments was 30 times less than that required in the previous experiments involving unmodified EPCs [15]. Thus, EPC cell therapy combined with gene (*i.e.* VEGF) transduction may be one option to overcome the limited number and function of EPCs that can be isolated from peripheral blood in patients.

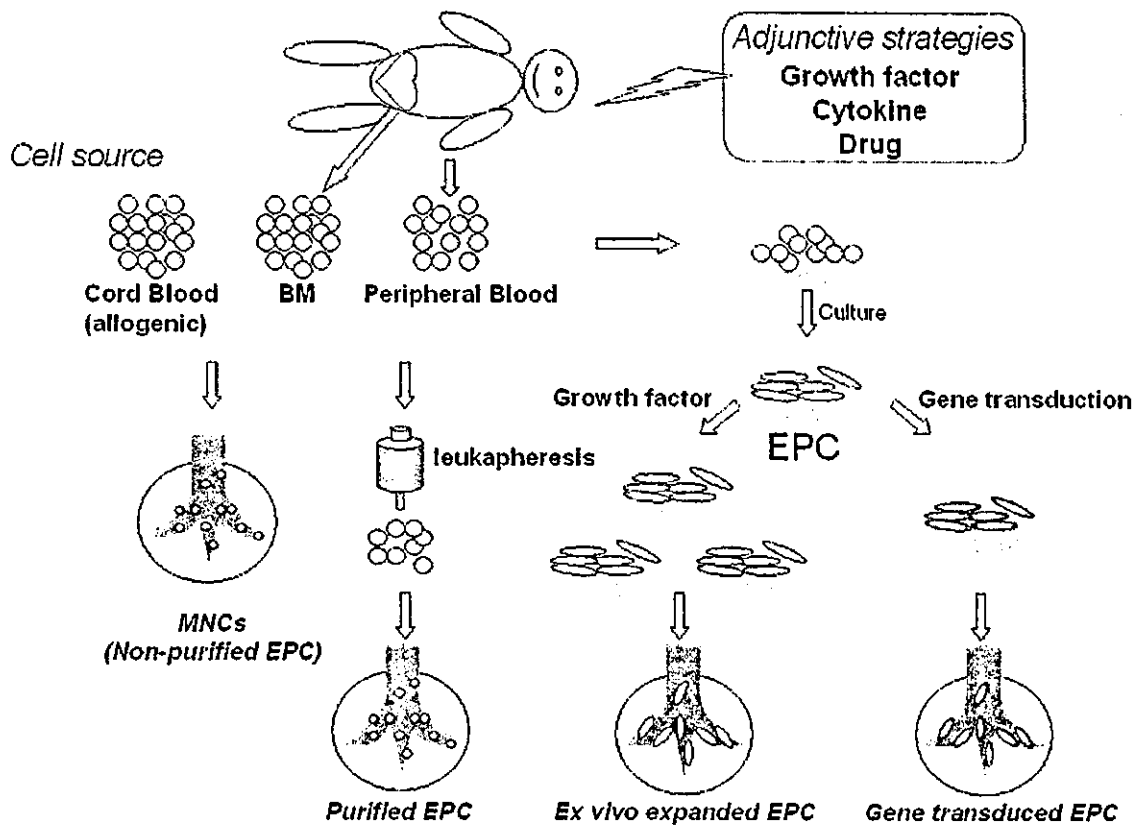


Fig. 3 Therapeutic application of EPCs for neovascularization.

EPC preview

EPCs have also been investigated in the cerebrovascular field. Embolization of the middle cerebral artery in Tie2/lacZ/BMT mice disclosed that the formation of new blood vessels in the adult brain after stroke involves vasculogenesis/EPCs. Similar data were reported using gender-mismatched wild-type mice transplanted with BM from green fluorescent protein-transgenic mice. However, whether autologous EPC transplantation would augment cerebral revascularization has yet to be examined.

To date, the role of EPCs in tumor angiogenesis has been demonstrated by several groups. Davidoff et al. showed that BM-derived EPCs contribute to tumor neovasculature and that BM cells transduced with an anti-angiogenic gene can restrict tumor growth in mice. Lyden et al. recently used angiogenic defective, tumor resistant Id-mutant mice and showed the restoration of tumor angiogenesis with

BM (donor)-derived EPCs throughout the neovessels following the transplantation of wild-type BM into these mice. These data demonstrate that EPCs are not only important but also critical to tumor neovascularization. Given the findings, 'anti-tumor EPC mediated gene therapy' by transplantation of EPCs transferred genes to inhibit tumor growth may be developed in the near future.

Pulmonary hypertension might also be included into EPC therapy candidates. Nagaya *et al.* [41] reported that transplantation of vasodilator gene-transduced EPCs derived from umbilical cord blood ameliorates pulmonary hypertension in rats.

Conclusion

BM-derived EPCs in adults possess numerous potentials as clinical tools for cardiovascular dis-

ease, tissue engineering, tumor, and so on. To acquire the more optimized quality and quantity of EPCs, several issues remain to be addressed in this research field. Some of the future perspectives are as follows: (1) identification of a specific marker for EPC with which other lineage cells do not share; (2) evaluation of EPC transdifferentiation in vitro and in physiological, pathological, and iatrogenic regeneration of tissues and organs; (3) methodological optimization of EPC purification, expansion, gene transfer, and administration to improve the efficacy of EPC transplantation; and (4) comparison of the therapeutic impact between purified EPCs and total bone marrow MNCs.

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Review

Post-natal endothelial progenitor cells for neovascularization in tissue regeneration

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Abstract

The isolation of endothelial progenitor cells (EPCs) derived from bone marrow (BM) was an outstanding event in the recognition of 'de novo vessel formation' in adults occurring as physiological and pathological responses. The finding that EPCs home to sites of neovascularization and differentiate into endothelial cells (ECs) in situ is consistent with 'vasculogenesis', a critical paradigm well described for embryonic neovascularization, but proposed recently in adults in which a reservoir of stem or progenitor cells contributes to vascular organogenesis. EPCs have also been considered as therapeutic agents to supply the potent origin of neovascularization under pathological conditions. This review provides an update of EPC biology as well as highlighting their potential use for therapeutic regeneration.

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1. Introduction

Tissue regeneration by somatic stem/progenitor cells has been recognized as a maintenance or recovery system of many organs in adult. The isolation and investigation of these somatic stem/progenitor cells has described how these cells contribute to postnatal organogenesis. On the basis of the regenerative potency, these stem/progenitor cells are expected to develop as a key strategy of therapeutic applications for the damaged organs.

Recently endothelial progenitor cells (EPCs) have been isolated from adult peripheral blood (PB). EPCs are considered to share common stem/progenitor cells with hematopoietic stem cells and have been shown to derive from bone marrow (BM) and to incorporate into foci of

physiological or pathological neovascularization. The finding that EPCs home to sites of neovascularization and differentiate into endothelial cells (ECs) in situ is consistent with 'vasculogenesis', a critical paradigm well described for embryonic neovascularization, but recently proposed in adults in which a reservoir of stem/progenitor cells contributes to post-natal vascular organogenesis. The discovery of EPCs has therefore drastically changed our understanding of adult blood vessel formation. The following review provides an update of EPC biology as well as highlighting their potential utility for therapeutic vascular regeneration.

2. Post-natal neovascularization

Through the discovery of EPCs in PB [1,2], our understanding of post-natal neovascularization has been expanded from angiogenesis to angio/vasculogenesis. As previously described [3], post-natal neovascularization was

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originally recognized to be constituted by the mechanism of 'angiogenesis', which is neovessel formation, operated by in situ proliferation and migration of preexisting endothelial cells. However, the isolation of EPCs resulted in the addition of the new mechanism, 'vasculogenesis', which is de novo vessel formation by in situ incorporation, differentiation, migration, and/or proliferation of BM-derived EPCs [4] (Fig. 1). More recently, tissue specific stem/progenitor cells with the potency of differentiation into myocytes or ECs were isolated in skeletal muscle tissue of murine hindlimb, although the origin remains to be clarified [5]. This finding suggests that the origin of EPCs may not be limited to BM, e.g. tissue specific stem/progenitor cells possibly provide 'in situ EPCs' as other sources of EPCs than BM.

In the event of minor scale neovessel formation, i.e. slight wounds or burns, 'in situ preexisting ECs' causing post-natal angiogenesis may replicate and replace the existing cell population sufficiently, as ECs exhibit the ability for self-repair that preserves their proliferative activity. Neovascularization through differentiated ECs, however, is limited in terms of cellular life span (Hayflick limit) and their inability to incorporate into remote target sites. In the case of large scale tissue repair, such as patients who experienced acute vascular insult secondary to burns, coronary artery bypass grafting (CABG), or acute myocardial infarction [6,7], or in physiological cyclic organogenesis of endometrium [4], BM-derived or in situ EPC kinetics are activated under the influence of appropriate cytokines, hormones and/or growth factors through

the autocrine, paracrine, and/or endocrine systems. Thus the contemporary view of tissue regeneration is that neighboring differentiated ECs are relied upon for vascular regeneration during a minor insult, whereas tissue specific or BM-derived stem/progenitor cells bearing EPCs/ECs are important when an emergent vascular regenerative process is required (Fig. 1).

3. Profiles of EPCs in adults

3.1. The evidence of circulating EPCs in adults

In the embryo, evidence suggests that hematopoietic stem cells (HSCs) and EPCs [8,9] are derived from a common precursor (hemangioblast) [10,11]. During embryonic development, multiple blood islands initially fuse to form a yolk sac capillary network [12], which provides the foundation for an arteriovenous vascular system that eventually forms following the onset of blood circulation [8]. The integral relationship between the cells which circulate in the vascular system (the blood cells) and those principally responsible for the vessels themselves (ECs) is suggested by their spatial orientation within the blood islands; those cells destined to generate hematopoietic cells are situated in the center of the blood island (HSCs) while EPCs or angioblasts are located at the periphery of the blood islands. In addition to this arrangement, HSCs and EPCs share common antigens, including CD34, Vascular

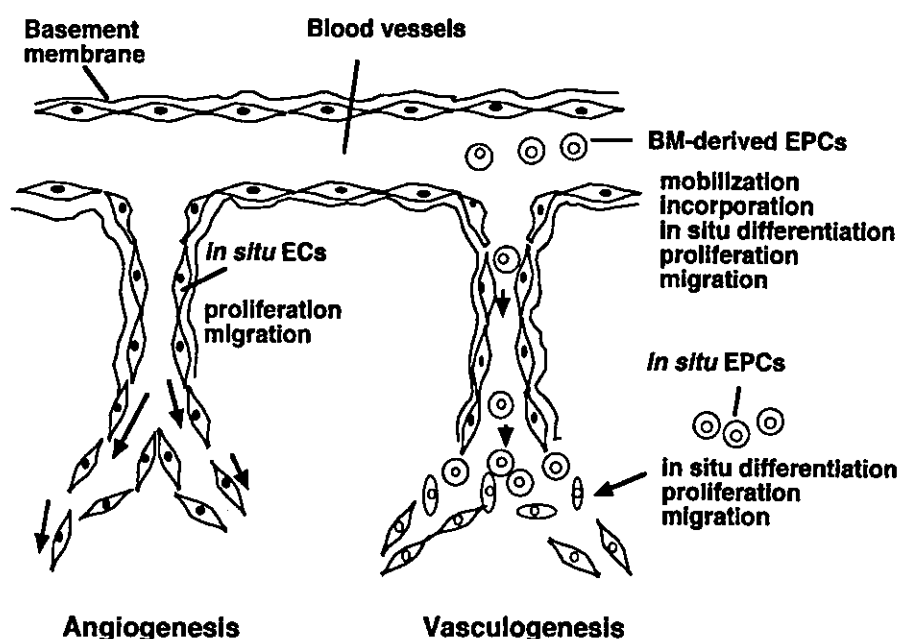


Fig. 1. Post-natal neovascularization in the physiological or pathological events is consistent with neovessel formation contributed by angiogenesis and vasculogenesis at the various rates between their two mechanisms. Angiogenesis and vasculogenesis are due to the activations of in situ ECs and BM-derived or in situ EPCs, respectively.

endothelial growth factor receptor-2 (VEGFR2), Tie-2, CD117, and stem cell antigen-1 (Sca-1) [13].

The existence of HSCs in the PB and BM, and the demonstration of sustained hematopoietic reconstitution with HSC transplantation led to the idea that a closely related cell-type, namely EPCs, may also exist in adult tissues. Recently, EPCs were successfully isolated from circulating mononuclear cells (MNCs) using VEGFR2, CD34, and CD133 antigens shared by both embryonic EPCs and HSCs [1,14,15]. In vitro, these cells differentiate into endothelial lineage cells, and in animal models of ischemia, heterologous, homologous, and autologous EPCs have been shown to incorporate into the foci of neovascularization, contributing to neovascularization. Recently, similar studies with EPCs isolated from human cord blood have demonstrated their analogous differentiation into ECs in vitro and in vivo [16–19].

These findings have raised important questions regarding fundamental concepts of blood vessel growth and development in adults. Does the differentiation of EPCs in situ (vasculogenesis) play an important role in adult neovascularization, and would impairments in this process lead to clinical diseases? There is now a strong body of evidence suggesting that vasculogenesis in fact significantly contributes to postnatal neovascularization. Recent studies with animal BM transplantation (BMT) models in which BM (donor)-derived EPCs could be distinguished have shown that the contribution of EPCs to neovessel formation may range from 5 to 25% in response to granulation tissue formation [20] or growth factor-induced neovascularization [21]. Also, in the tumor neovascularization, the range is approximately 35–45% higher than the former events [22]. The degree of EPC contribution to post-natal neovascularization is predicted to depend on each neovascularizing event or disease.

3.2. Isolation of EPCs in circulation

Under the current status, it is impossible to differentiate 'immature EPCs' from primitive HSCs, as those cells share common surface markers, i.e. AC133, CD34, or VEGFR2 as described above. In circulation, the cell population with the capacity of differentiation to EPCs is considered to be included in the cell population expressing AC133 and VEGFR2 markers in the subset of CD34 positive cells [15]. Circulating EPCs are constitutively expressing stem/progenitor markers, i.e. CD34 or VEGFR2 except AC133, and start expressing endothelial lineage specific markers, VE cadherin or E-selectin. On the other hand, following the commitment and differentiation to hematopoietic stem/progenitor cells, the surface markers of AC133 and VEGFR2 are extinguished. Such stem/progenitor cell markers do not express on the differentiated hematopoietic cells. Alternatively, kinds of surface markers are expressed to characterize individual hematopoietic cell populations. AC133 is a marker to

differentiate immature EPCs or primitive HSCs from circulating EPCs. To differentiate EPCs from hematopoietic stem/progenitor cells, VEGFR2, VE cadherin, or E-selectin are useful. Also, circulating EPCs do not express monocyte or myeloid markers, such as CD14 or CD15. Accordingly, circulating EPCs may be isolated via selection by the antigenicity of CD34, VEGFR2, and/or VE cadherin and also circulating immature EPCs by AC133 (Fig. 2).

3.3. Diverse identification of human EPCs and their precursors

Since the initial report of EPCs [1,2], a number of groups have set out to define this cell population better. Because EPCs and HSCs share many surface markers, and no simple definition of EPCs exists, various methods of EPC isolation have been reported [1,2,15–18,23–31]. The term EPC may therefore encompass a group of cells that exist in a variety of stages ranging from hemangioblasts to fully differentiated ECs. Although the true differentiation lineage of EPCs and their putative precursors remains to be determined, there is overwhelming evidence in vivo that a population of EPCs exists in human.

Lin et al. cultivated peripheral MNCs from patients receiving gender-mismatched BMT and studied their growth in vitro. In this study, they identified a population of BM (donor)-derived ECs with high proliferative potential (late outgrowth); these BM cells likely represent EPCs [24]. Gunsilius et al. investigated a chronic myelogenous leukemia model and disclosed that BM-derived EPCs contribute to postnatal neovascularization in human [26]. Interestingly, in the report, BM-derived EPCs could be detected even in the wall of quiescent vessels without neovascularization events. This finding suggests that BM-derived EPCs may relate even to the turnover of ECs consisting of quiescent vessels.

Reyes et al. have recently isolated multipotent adult progenitor cells (MAPCs) from BM MNCs, differentiated them into EPCs and proposed MAPCs as an origin of EPCs [22]. These studies therefore provide evidence to support the presence of BM-derived EPCs that take part in neovascularization. Also, as described above, the existence of 'in situ EPCs' as derived from tissue specific stem/progenitor cells in murine skeletal muscle remains to be investigated also in the other tissues [5] (Fig. 2).

4. EPC kinetics in adults

4.1. EPC kinetics effected by endogenous agents

The incorporation of BM-derived EPCs into foci of physiological and pathological neovascularization has been demonstrated through various animal experiments. One well-established model that allows the detection of BM-

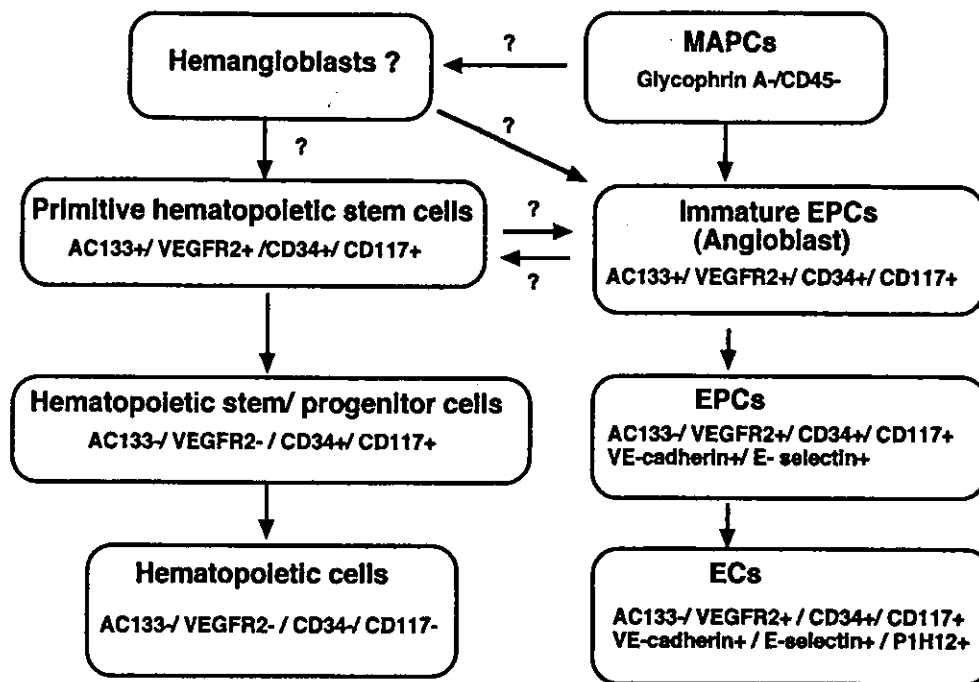


Fig. 2. Origin and differentiation of EPCs in adult BM. EPCs are thought to differentiate not only from putative hemangioblasts, common precursor cells with HSCs, as previously described, but also from MAPCs. Representative antigenicities to stem/progenitor cells are shown (+, positive; -, negative).

derived EPCs includes transplanting wild-type mice with BM cells harvested from transgenic mice in which LacZ expression is regulated by an EC lineage-specific promoter, *flk-1* or *Tie-2* (*flk-1/lacZ/BMT*, *Tie-2/lacZ/BMT*). Using such mice, *flk-1*- or *Tie-2*-expressing endothelial lineage cells derived from BM (EPCs) have been shown to localize to vessels during tumor growth, wound healing, skeletal and cardiac ischemia, corneal neovascularization, and endometrial remodeling following hormone-induced ovulation [4].

Tissue trauma causes mobilization of hematopoietic cells as well as pluripotent stem or progenitor cells from the hematopoietic system [32]. Consistent with the notion that EPCs and HSCs share a common ancestry, recent data from our laboratory have shown that mobilization of BM-derived EPCs constitutes a natural response to tissue ischemia. The aforementioned murine BMT model also provided direct evidence of enhanced BM-derived EPC incorporation into foci of corneal neovascularization following the development of hindlimb ischemia [33]. This finding indicates that circulating EPCs are mobilized endogenously in response to tissue ischemia and can incorporate into neovascular foci to promote tissue repair. These results in animals were recently confirmed by human studies illustrating EPC mobilization in patients following burns [6], CABG, or acute myocardial infarction [7].

As previous studies demonstrated, endogenous mobiliza-

tion of BM-derived EPCs, we considered exogenous mobilization of EPCs as an effective means of augmenting the resident population of EPCs/ECs. Such a strategy is appealing for its potential to overcome the endothelial dysfunction or depletion that may be associated with older, diabetic, or hypercholesterolemic patients. Granulocyte macrophage colony-stimulating factor (GM-CSF) is well known to stimulate hematopoietic progenitor cells and myeloid lineage cells, but has recently been shown to exert a potent stimulatory effect on EPC kinetics. The delivery of this cytokine induced EPC mobilization and enhanced neovascularization of severely ischemic tissues and *de novo* corneal vascularization [33].

The exact mechanism by which EPCs are mobilized to the peripheral circulation remains unknown, but may mimic aspects of embryonic development. Vascular endothelial growth factor (VEGF), critical for angio/vasculogenesis in the embryo [34–36], has recently been shown to be an important stimulus of adult EPC kinetics. Our studies carried out first in mice [37] and subsequently in patients undergoing VEGF gene transfer for critical limb or myocardial ischemia [38] established that a previously unappreciated mechanism by which VEGF contributes to neovascularization is in part by mobilizing BM-derived EPCs. Similar modulation of EPC kinetics has been observed in response to other hematopoietic stimulators, such as granulocyte-colony stimulating factor (G-CSF) and stroma-derived factor-1 (SDF-1) [39].