

Drug-Drug Interactions Between Cephalosporin Antibiotics and Probenecid

There are many reports on the drug-drug interactions between cephalosporin antibiotics and probenecid (157). As both cephalosporins and probenecid interact with OAT family transporters, some of these drug-drug interactions may be due to an OAT-mediated uptake process. Most cephalosporins are excreted in the urine, which may be partly mediated by OAT family transporters. The elimination rates of cephazidone, cefazolin, cefalexin, cefradine, cefaclor, cefmetazole, cefoxitin, cefuroxime, cefmenoxime, ceftizoxime, and ceftriaxone were significantly reduced by coadministration of probenecid, which may be partly caused by the inhibition of their renal excretions (157).

Marino & Dominguez-Gil have shown that the pharmacokinetics of cefadroxil is altered by coadministration of probenecid (158). In their report, the peak concentration and half-life of cefadroxil was increased 1.4- and 1.3-fold, respectively, following coadministration of probenecid. Its urinary excretion rate constant falls by 58%, supporting the possibility of drug-drug interaction at the renal excretion. Supplemental Table 1 suggests that OAT1- and OAT3-mediated transport should be decreased to at most 25%–47% and 25%–69% of the control, and, therefore, it may be partly explained by the OAT-mediated drug-drug interaction.

Probenecid has also been shown to alter the plasma concentrations of cefamandole and ceftriaxone (159). The maximum plasma concentration and half-life of cefamandole were increased 6- and 1.8-fold by coadministration of probenecid (159). Also, 71% of cefamandole is excreted in the urine, and this was reduced to 66% of the control (159). The elimination of ceftriaxone was slightly affected by coadministration of probenecid (160). Probenecid reduced the serum clearance of ceftriaxone to 73% of the control (160). It reduced the renal and nonrenal clearance to 80% and 68% of the control, respectively, suggesting that this drug-drug interaction is, to a minor extent, due to renal excretion (160).

Drug-Drug Interaction Between Methotrexate and NSAIDs

To date, there are reports that coadministration of MTX with penicillin, probenecid, and NSAIDs cause drug-drug interactions and several potential sites for these DDI have been reported: an increase in the protein unbound fraction of MTX, a decrease in the urine flow rate resulting from the inhibition of prostaglandin synthesis, and inhibition of the renal tubular secretion of MTX (161–164). Nozaki et al. analyzed the uptake mechanism of MTX in rat kidney slices and examined the effects of NSAIDs on its uptake (165). They showed that rat Oat3 and reduced folate carrier 1 (RFC-1) equally contribute to the renal uptake (30% each), with the remaining fraction being accounted for by passive diffusion and/or adsorption, whereas rOat1 makes only a limited contribution (165). Many NSAIDs inhibited both rOat3- and RFC-1-mediated uptake of MTX, but the K_i value for Oat3 was lower than that for RFC-1 (165). At their therapeutic concentrations, they inhibited only Oat3-mediated uptake of MTX. Therefore, the effect of NSAIDs on the renal uptake of

MTX is expected to be nonextensive and partial. Many NSAIDs also inhibit human OAT3-mediated uptake of MTX with therapeutic relevant plasma concentrations of unbound drugs (26). However, also in humans, the contribution of OAT3 to the total renal uptake of MTX needs to be clarified for the identification of the mechanism of the clinically relevant DDI.

CONCLUSION

In addition to phase I and phase II enzymes, transporters also play an important role in drug elimination and distribution. Therefore, it is possible that transporter-mediated drug-drug interactions alter pharmacokinetics, and could result in severe side effects.

A large number of transporters have been characterized in rodents and humans, and the mechanism of the membrane transport of several compounds including endogenous compounds and therapeutic drugs has been clarified. However, the transport mechanism of most therapeutic drugs remains unknown. To predict a transporter-mediated drug-drug interaction, the transporters involved in the membrane transport of the drug need to be characterized. As multiple transporters have been characterized in the kidney and liver and their expression systems are available, it should be possible to predict a transporter-mediated drug-drug interaction by using these systems with the information of the contribution made by each transporter to the net transport in the kidney and liver.

We have estimated the possibility of a transporter-mediated drug-drug interaction from the R value calculated using the maximum unbound concentration of inhibitors. This method may avoid false negative predictions of drug-drug interactions. In conclusion, greater awareness of the possibility of transporter-mediated drug-drug interactions is necessary.

The *Annual Review of Pharmacology and Toxicology* is online at
<http://pharmatox.annualreviews.org>

LITERATURE CITED

1. Petzinger E. 1994. Transport of organic anions in the liver. An update on bile acid, fatty acid, monocarboxylate, anionic amino acid, cholephilic organic anion, and anionic drug transport. *Rev. Physiol. Biochem. Pharmacol.* 123:47–211
2. Oude Elferink RPJ, Meijer DKF, Kuipers F, Jansen PLM, Groen AK, Grootenhuis GMM. 1995. Hepatobiliary secretion of organic compounds; molecular mechanism of membrane transport. *Biochem. Biophys. Acta* 1241:215–68
3. Yamazaki M, Suzuki H, Sugiyama Y. 1996. Recent advances in carrier-mediated hepatic uptake and biliary excretion of xenobiotics. *Pharm. Res.* 13:497–513
4. Okudaira N, Sugiyama Y. 1996. Use of an isolated perfused kidney to assess renal clearance of drugs: information obtained in steady-state and non-steady-state experimental systems. In *Models for Assessing Drug Absorption and Metabolism*, ed RT Borchard, PL Smith, G Wilson, pp. 211–38. New York: Plenum Press

5. Giacomini KM, Hsyu PH, Gisclon JG. 1988. Renal transport of drugs: an overview of methodology with application to cimetidine. *Pharm. Res.* 5:465–71
6. van Aubel RAMH, Masereeuw R, Russel FGM. 2000. Molecular pharmacology of renal organic anion transporters. *Am. J. Physiol. Renal Physiol.* 279:F216–32
7. Kusuhara H, Sugiyama Y. 2002. Role of transporters in the tissue-selective distribution and elimination of drugs: transporters in the liver, small intestine, brain and kidney. *J. Control. Release* 78:43–54
8. Kusuhara H, Suzuki H, Sugiyama Y. 1998. The role of P-glycoprotein and canalicular multispecific organic anion transporter in the hepatobiliary excretion of drugs. *J. Pharm. Sci.* 87:1025–40
9. Suzuki H, Sugiyama Y. 2000. Transport of drugs across the hepatic sinusoidal membrane: sinusoidal drug influx and efflux in the liver. *Semin. Liver Dis.* 20:251–63
10. Inui K, Masuda S, Saito H. 2000. Cellular and molecular aspects of drug transport in the kidney. *Kidney Int.* 58:944–58
11. Dresser MJ, Leabman MK, Giacomini KM. 2001. Transporters involved in the elimination of drugs in the kidney: organic anion transporters and organic cation transporters. *J. Pharm. Sci.* 90:397–421
12. Sweet DH, Bush KT, Nigam SK. 2001. The organic anion transporter family: from physiology to ontogeny and the clinic. *Am. J. Physiol. Renal Physiol.* 281:F197–205
13. Sekine T, Cha SH, Endou H. 2000. The multispecific organic anion transporter (OAT) family. *Pflugers Arch.* 440:337–50
14. Koepsell H, Gorboulev V, Arndt P. 1999. Molecular pharmacology of organic cation transporters in the kidney. *J. Membr. Biol.* 167:103–17
15. Mizuno N, Sugiyama Y. 2002. Drug transporters: their roles and importance in the selection and development of new drugs. *Drug Metab. Pharmacokin.* 17:93–108
16. Meier PJ, Eckhardt U, Schroeder A, Ha- genbuch B, Steiger B. 1997. Substrate specificity of sinusoidal bile acid and organic anion uptake systems in rat and human liver. *Hepatology* 26:1667–77
17. Pang KS, Rowland M. 1977. Hepatic clearance of drugs I: theoretical considerations of a “well-stirred” model and a “parallel-tube” model. Influence of hepatic blood flow, plasma and blood cell binding, and the hepatocellular enzymatic activity on hepatic drug clearance. *J. Pharmacokin. Biopharm.* 5:625–53
18. Pang KS, Gillette R. 1978. Kinetics of metabolite formation and elimination in the perfused rat liver preparation: difference between the elimination of preformed acetaminophen and acetaminophen formed from phenacetin. *J. Pharmacol. Exp. Ther.* 207:178–94
19. Miyauchi S, Sugiyama Y, Sawada Y, Morita K, Iga T, et al. 1987. Kinetics of hepatic transport of 4-methylumbelliflone in rats: analysis by multiple indicator dilution method. *J. Pharmacokin. Biopharm.* 15:25–38
20. Sekine T, Watanabe N, Hosoyamada M, Kanai Y, Endou H. 1997. Expression cloning and characterization of a novel multispecific organic anion transporter. *J. Biol. Chem.* 272:18526–29
21. Hosoyamada M, Sekine T, Kanai Y, Endou H. 1999. Molecular cloning and functional expression of a multispecific organic anion transporter from human kidney. *Am. J. Physiol. Renal Physiol.* 276:F122–28
22. Sun W, Wu RR, van Poelje PD, Erion MD. 2001. Isolation of a family of organic anion transporters from human liver and kidney. *Biochem. Biophys. Res. Comm.* 283:417–22
23. Race JE, Grassl SM, Williams WJ, Holtzman EJ. 1999. Molecular cloning and characterization of two novel human renal organic anion transporters (hOAT1 and hOAT3). *Biochem. Biophys. Res. Commun.* 255:508–14
24. Cha SH, Sekine T, Kusuhara H, Yu E,

- Kim JY, et al. 2000. Molecular cloning and characterization of multispecific organic anion transporter 4 expressed in the placenta. *J. Biol. Chem.* 275:4507-12
25. Cha SH, Sekine T, Fukushima JI, Kanai Y, Kobayashi Y, et al. 2001. Identification and characterization of human organic anion transporter 3 expressing predominantly in the kidney. *Mol. Pharmacol.* 59:1277-86
26. Takeda M, Khamdang S, Narikawa S, Kimura H, Hosoyamada M, et al. 2002. Characterization of methotrexate transport and its drug interactions with human organic anion transporters. *J. Pharmacol. Exp. Ther.* 302:666-71
27. Takeda M, Khamdang S, Narikawa S, Kimura H, Kobayashi Y, et al. 2002. Human organic anion transporters and human organic cation transporters mediate renal antiviral transport. *J. Pharmacol. Exp. Ther.* 300:918-24
28. Jung KY, Takeda M, Kim DK, Tojo A, Narikawa S, et al. 2001. Characterization of ochratoxin A transport by human organic anion transporters. *Life Sci.* 69:2123-35
29. Lecureur V, Sun D, Hargrove P, Schuetz EG, Kim RB, et al. 2000. Cloning and expression of murine sister of P-glycoprotein reveals a more discriminating transporter than MDRI/P-glycoprotein. *Mol. Pharmacol.* 57:24-35
30. Yamazaki M, Neway WE, Ohe T, Chen I, Rowe JF, et al. 2001. In vitro substrate identification studies for p-glycoprotein-mediated transport: species difference and predictability of in vivo results. *J. Pharmacol. Exp. Ther.* 296:723-35
31. Ambudkar SV, Lelong IH, Zhang J, Cardarelli CO, Gottesman MM, et al. 1992. Partial purification and reconstitution of the human multidrug-resistance pump: characterization of the drug-stimulatable ATP hydrolysis. *Proc. Natl. Acad. Sci. USA* 89:8472-76
32. Adachi Y, Suzuki H, Sugiyama Y. 2001. Comparative studies on in vitro methods for evaluating in vivo function of MDR1 P-glycoprotein. *Pharm. Res.* 18:1660-68
33. Guo A, Marinaro W, Hu P, Sinko PJ. 2002. Delineating the contribution of secretory transporters in the efflux of etoposide using Madin-Darby canine kidney (MDCK) cells overexpressing P-glycoprotein (Pgp), multidrug resistance-associated protein (MRP1), and canalicular multispecific organic anion transporter (cMOAT). *Drug Metab. Dispos.* 30:457-63
34. Walle UK, Walle T. 1998. Taxol transport by human intestinal epithelial Caco-2 cells. *Drug Metab. Dispos.* 26:343-46
35. Tanigawara Y, Okamura N, Hirai M, Yasuhara M, Ueda K, et al. 1992. Transport of digoxin by human P-glycoprotein expressed in a porcine kidney epithelial cell line (LLC-PK1). *J. Pharmacol. Exp. Ther.* 263:840-45
36. Ueda K, Okamura N, Hirai M, Tanigawara Y, Saeki T, et al. 1992. Human P-glycoprotein transports cortisol, aldosterone, and dexamethasone, but not progesterone. *J. Biol. Chem.* 267:24248-52
37. Ganapathy ME, Brandsch M, Prasad PD, Ganapathy V, Leibach FH. 1995. Differential recognition of beta-lactam antibiotics by intestinal and renal peptide transporters, PEPT 1 and PEPT 2. *J. Biol. Chem.* 270:25672-77
38. Smith DE, Pavlova A, Berger UV, Hediger MA, Yang T, et al. 1998. Tubular localization and tissue distribution of peptide transporters in rat kidney. *Pharm. Res.* 15:1244-49
39. Shen H, Smith DE, Yang T, Huang YG, Schnermann JB, et al. 1999. Localization of PEPT1 and PEPT2 proton-coupled oligopeptide transporter mRNA and protein in rat kidney. *Am. J. Physiol. Renal Physiol.* 276:F658-65
40. Tsuji A. 2002. Transporter-mediated drug interactions. *Drug. Metabol. Pharmacokin.* 17:253-74
41. Han HK, Rhee JK, Oh DM, Saito G, Hsu

- CP, et al. 1999. CHO/hPEPT1 cells over-expressing the human peptide transporter (hPEPT1) as an alternative in vitro model for peptidomimetic drugs. *J. Pharm. Sci.* 88:347–50
42. Gorboulev V, Ulzheimer JC, Akhounova A, Ulzheimer-Teuber I, Karbach U, et al. 1997. Cloning and characterization of two human polyspecific organic cation transporters. *DNA Cell. Biol.* 16:871–81
43. Zhang L, Schaner ME, Giacomini KM. 1998. Functional characterization of an organic cation transporter (hOCT1) in a transiently transfected human cell line (HeLa). *J. Pharmacol. Exp. Ther.* 286:354–61
44. Dudley AJ, Bleasby K, Brown CD. 2000. The organic cation transporter OCT2 mediates the uptake of beta-adrenoceptor antagonists across the apical membrane of renal LLC-PK1 cell monolayers. *Br. J. Pharmacol.* 131:71–79
45. Busch AE, Karbach U, Miska D, Gorboulev V, Akhounova A, et al. 1998. Human neurons express the polyspecific cation transporter hOCT2, which translocates monoamine neurotransmitters, amantadine, and memantine. *Mol. Pharmacol.* 54:342–52
46. Urakami Y, Akazawa M, Saito H, Okuda M, Inui K. 2002. cDNA cloning, functional characterization, and tissue distribution of an alternatively spliced variant of organic cation transporter hOCT2 predominantly expressed in the human kidney. *J. Am. Soc. Nephrol.* 13:1703–10
47. Tamai I, Yabuuchi H, Nezu J, Sai Y, Oku A, et al. 1997. Cloning and characterization of a novel human pH-dependent organic cation transporter, OCTN1. *FEBS Lett.* 419:107–11
48. Yabuuchi H, Tamai I, Nezu J, Sakamoto K, Oku A, et al. 1999. Novel membrane transporter OCTN1 mediates multi-specific, bidirectional, and pH-dependent transport of organic cations. *J. Pharmacol. Exp. Ther.* 289:768–73
49. Tamai I, Ohashi R, Nezu J, Yabuuchi H, Oku A, et al. 1998. Molecular and functional identification of sodium ion-dependent, high affinity human carnitine transporter OCTN2. *J. Biol. Chem.* 273:20378–82
50. Wu X, Huang W, Prasad PD, Seth P, Rajan DP, et al. 1999. Functional characteristics and tissue distribution pattern of organic cation transporter 2 (OCTN2), an organic cation/carnitine transporter. *J. Pharmacol. Exp. Ther.* 290:1482–92
51. Nezu J, Tamai I, Oku A, Ohashi R, Yabuuchi H, et al. 1999. Primary systemic carnitine deficiency is caused by mutations in a gene encoding sodium ion-dependent carnitine transporter. *Nat. Genet.* 21:91–94
52. Ohashi R, Tamai I, Yabuuchi H, Nezu JI, Oku A, et al. 1999. Na⁺-dependent carnitine transport by organic cation transporter (OCTN2): its pharmacological and toxicological relevance. *J. Pharmacol. Exp. Ther.* 291:778–84
53. Ganapathy ME, Huang W, Rajan DP, Carter AL, Sugawara M, et al. 2000. β -lactam antibiotics as substrates for OCTN2, an organic cation/carnitine transporter. *J. Biol. Chem.* 275:1699–707
54. Tamai I, Nezu J, Uchino H, Sai Y, Oku A, et al. 2000. Molecular identification and characterization of novel members of the human organic anion transporter (OATP) family. *Biochem. Biophys. Res. Commun.* 273:251–60
55. Hsiang B, Zhu Y, Wang Z, Wu Y, Sasiseville V, et al. 1999. A novel human hepatic organic anion transporting polypeptide (OATP2). Identification of a liver-specific human organic anion transporting polypeptide and identification of rat and human hydroxymethylglutaryl-CoA reductase inhibitor transporters. *J. Biol. Chem.* 274:37161–68
56. Nakai D, Nakagomi R, Furuta Y, Tokui T, Abe T, et al. 2001. Human liver-specific organic anion transporter, LST-1, mediates uptake of pravastatin by human

DRUG-DRUG INTERACTION INVOLVING TRANSPORTERS 715

- hepatocytes. *J. Pharmacol. Exp. Ther.* 297:861–67
57. König J, Cui Y, Nies AT, Keppler D. 2000. A novel human organic anion transporting polypeptide localized to the basolateral hepatocyte membrane. *Am. J. Physiol. Gastrointest. Liver Physiol.* 278:G156–64
58. König J, Cui Y, Nies AT, Keppler D. 2000. Localization and genomic organization of a new hepatocellular organic anion transporting polypeptide. *J. Biol. Chem.* 275:23161–68
59. Jacquemin E, Hagenbuch B, Stieger B, Wolkoff AW, Meier PJ. 1994. Expression cloning of a rat liver Na⁺-independent organic anion transporter. *Proc. Natl. Acad. Sci. USA* 91:133–37
60. Noé B, Hagenbuch B, Stieger B, Meier PJ. 1997. Isolation of a multispecific organic anion and cardiac glycoside transporter from rat brain. *Proc. Natl. Acad. Sci. USA* 94:10346–50
61. Cattori V, Hagenbuch B, Hagenbuch N, Stieger B, Ha R, et al. 2000. Identification of organic anion transporting polypeptide 4 (Oatp4) as a major full-length isoform of the liver-specific transporter-1 (rlst-1) in rat liver. *FEBS Lett.* 474:242–45
62. Shitara Y, Itoh T, Sato H, Li AP, Sugiyama Y. 2003. Inhibition of transporter-mediated hepatic uptake as a mechanism for drug-drug interaction between cerivastatin and cyclosporin A. *J. Pharmacol. Exp. Ther.* 304:610–16
63. Pang KS, Wang PJ, Chung AYK, Wolkoff AW. 1998. The modified dipeptide enalapril, an angiotensin-converting enzyme inhibitor, is transported by the rat liver organic anion transport protein. *Hepatology* 28:1341–46
64. Ishizuka H, Konno K, Naganuma H, Nishimura K, Kouzuki H, et al. 1998. Transport of temocaprilat into rat hepatocytes: role of organic anion transporting polypeptide. *J. Pharmacol. Exp. Ther.* 287:37–42
65. Kullak-Ublick GA, Ismail MG, Stieger B, Landmann L, Huber R, et al. 2001. Organic anion-transporting polypeptide B (OATP-B) and its functional comparison with three other OATPs of human liver. *Gastroenterology* 120:525–33
66. Jansen PL, Peters WH, Lamers WH. 1985. Hereditary chronic conjugated hyperbilirunemia in mutant rats caused by defective hepatic anion transport. *Hepatology* 5:573–79
67. Hosokawa S, Tagaya O, Mikami T, Nozaki Y, Kawaguchi Y, et al. 1992. A new rat mutant with chronic conjugated hyperbilirunemia and renal glomerular lesions. *Lab. Animal Sci.* 42:27–34
68. Muller M, Jansen PL. 1997. Molecular aspects of hepatobiliary transport. *Am. J. Physiol. Gastrointest. Liver Physiol.* 272:G1285–303
69. Oude Elferink RP, Meijer DK, Kuipers F, Jansen PL, Groen AK, et al. 1995. Hepatobiliary secretion of organic compounds; molecular mechanisms of membrane transport. *Biochim. Biophys. Acta* 1241:215–68
70. Stieger B, Meier PJ. 1998. Bile acid and xenobiotic transporters in liver. *Curr. Opin. Cell. Biol.* 10:462–67
71. Suzuki H, Sugiyama Y. 1998. Excretion of GSSG and glutathione conjugates mediated by MRP1 and cMOAT/MRP2. *Semin. Liver Dis.* 18:359–76
72. König J, Nies AT, Cui Y, Leier I, Keppler D. 1999. Conjugate export pumps of the multidrug resistance protein (MRP) family: localization, substrate specificity, and MRP2-mediated drug resistance. *Biochim. Biophys. Acta* 1461:377–94
73. Paulusma CC, Bosma PJ, Zaman GJ, Bakker CT, Otter M, et al. 1996. Congenital jaundice in rats with a mutation in a multidrug resistance-associated protein gene. *Science* 271:1126–28
74. Ito K, Suzuki H, Hirohashi T, Kume K, Shimizu T, et al. 1997. Molecular cloning of canalicular multispecific organic anion transporter defective in EHBR. *Am. J. Physiol. Gastrointest. Liver Physiol.* 272:G16–22

75. Buchler M, König J, Brom M, Kartemberg J, Spring H, et al. 1996. cDNA cloning of the hepatocyte canalicular isoform of the multidrug resistance protein, cMrp, reveals a novel conjugate export pump deficient in hyperbilirubinemic mutant rats. *J. Biol. Chem.* 271:15091–98
76. Taniguchi K, Wada M, Kohno K, Nakamura T, Kawabe T, et al. A human canalicular multispecific organic anion transporter (cMOAT) gene is overexpressed in cisplatin-resistant human cancer cell lines with decreased drug accumulation. *Cancer Res.* 56:4124–29
77. Borst P, Evers R, Kool M, Wijnholds J. 1999. The multidrug resistance protein family. *Biochim. Biophys. Acta* 1461:347–57
78. Kuwano M, Toh S, Uchiumi T, Takano H, Kohno K, Wada M. 1999. Multidrug resistance-associated protein subfamily transporters and drug resistance. *Anticancer Drug Des.* 14:123–31
79. Jedlitschky G, Leier I, Buchholz U, Hummel-Eisenbeiss J, Burchell B, et al. 1997. ATP-dependent transport of bilirubin glucuronides by the multidrug resistance protein MRP1 and its hepatocyte canalicular isoform MRP2. *Biochem. J.* 327:305–10
80. Niinuma K, Kato Y, Suzuki H, Tyson CA, Weizer V, et al. 1999. Primary active transport of organic anions on bile canalicular membrane in humans. *Am. J. Physiol. Gastrointest. Liver Physiol.* 276:G1153–64
81. Cui Y, Konig J, Buchholz JK, Spring H, Leier I, et al. 1999. Drug resistance and ATP-dependent conjugate transport mediated by the apical multidrug resistance protein, MRP2, permanently expressed in human and canine cells. *Mol. Pharmacol.* 55:929–37
82. Niinuma K, Kato Y, Suzuki H, Tyson CA, Weizer V, et al. 1999. Primary active transport of organic anions on bile canalicular membrane in humans. *Am. J. Physiol. Gastrointest. Liver Physiol.* 276:G1153–64
83. Paulusma CC, van Geer MA, Evers R, Heijnen M, Ottenhoff R, et al. 1999. Canalicular multispecific organic anion transporter/multidrug resistance protein 2 mediates low-affinity transport of reduced glutathione. *Biochem. J.* 338:393–401
84. Ryu S, Kawabe T, Nada S, Yamaguchi A. 2000. Identification of basic residues involved in drug export function of human multidrug resistance-associated protein 2. *J. Biol. Chem.* 275:39617–24
85. Sasaki M, Suzuki H, Ito K, Abe T, Sugiyma Y. 2002. Transcellular transport of organic anions across a double-transfected Madin-Darby canine kidney II cell monolayer expressing both human organic anion-transporting polypeptide (OATP2/SLC21A6) and multidrug resistance-associated protein 2 (MRP2/ABCC2). *J. Biol. Chem.* 277:6497–503
86. Hooijberg JH, Broxterman HJ, Kool M, Assaraf YG, Peters GJ, et al. 1999. Antifolate resistance mediated by the multidrug resistance proteins MRP1 and MRP2. *Cancer Res.* 59:2532–35
87. Tang F, Horie K, Borchardt RT. 2002. Are MDCK cells transfected with the human MRP2 gene a good model of the human intestinal mucosa? *Pharm. Res.* 19:773–79
88. Chen ZS, Kawabe T, Ono M, Aoki S, Sumizawa T, et al. 1999. Effect of multidrug resistance-reversing agents on transporting activity of human canalicular multispecific organic anion transporter. *Mol. Pharmacol.* 56:1219–28
89. Akita H, Suzuki H, Hirohashi T, Takikawa H, Sugiyma Y. 2002. Transport activity of human MRP3 expressed in Sf9 cells: comparative studies with rat MRP3. *Pharm. Res.* 19:34–41
90. Zeng H, Liu G, Rea PA, Kruh GD. 2000. Transport of amphipathic anions by human multidrug resistance protein 3. *Cancer Res.* 60:4779–84

DRUG-DRUG INTERACTION INVOLVING TRANSPORTERS 717

91. Doyle LA, Yang W, Abruzzo LV, Krogmann T, Gao Y, et al. 1998. A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc. Natl. Acad. Sci. USA* 95:15665–70.
92. Allikmets R, Schriml LM, Hutchinson A, Romano-Spica V, Dean M. 1998. A human placenta-specific ATP-binding cassette gene (ABCP) on chromosome 4q22 that is involved in multidrug resistance. *Cancer Res.* 58:5337–39.
93. Miyake K, Mickley L, Litman T, Zhan Z, Robey R, et al. 1999. Molecular cloning of cDNAs which are highly overexpressed in mitoxantrone-resistant cells. *Cancer Res.* 59:8–13.
94. Maliepaard M, Scheffer GL, Faneite IF, van Gastelen MA, Pijnenborg ACLM, et al. 2001. Subcellular localization and distribution of the breast cancer resistance protein transporter in normal human tissues. *Cancer Res.* 61:3458–64.
95. Suzuki M, Suzuki H, Sugimoto Y, Sugiyama Y. 2003. ABCG2 transports sulfated conjugates of steroids and xenobiotics. *J. Biol. Chem.* 278:22644–49.
96. Urakami Y, Nakamura N, Takahashi K, Okuda M, Saito H, et al. 1999. Gender differences in expression of organic cation transporter OCT2 in rat kidney. *FEBS Lett.* 461:339–42.
97. Hasegawa M, Kusuhara H, Sugiyama D, Ito K, Ueda S, et al. 2002. Functional involvement of rat organic anion transporter 3 (rOat3; Slc22a8) in the renal uptake of organic anions. *J. Pharmacol. Exp. Ther.* 300:746–53.
98. Hori R, Ishikawa Y, Takano M, Okano T, Kitazawa S, et al. 1982. The interaction of cephalosporin antibiotics with renal cortex of rats: accumulation to cortical slices and binding to purified plasma membranes. *Biochem. Pharmacol.* 31:2267–72.
99. Terada T, Sawada K, Ito T, Saito H, Hashimoto Y, et al. 2000. Functional expression of novel peptide transporter in renal basolateral membranes. *Am. J. Physiol. Renal Physiol.* 279:F851–57.
100. Akhteruzzaman S, Kato Y, Kouzuki H, Suzuki H, Hisaka A, et al. 1999. Carrier-mediated hepatic uptake of peptidic endothelin antagonists in rats. *J. Pharmacol. Exp. Ther.* 290:1107–15.
101. Ishigami M, Tokui T, Komai T, Tsukahara K, Yamazaki M, et al. 1995. Evaluation of the uptake of pravastatin by perfused rat liver and primary cultured rat hepatocytes. *Pharm. Res.* 12:1741–45.
102. Madan A, DeHaan R, Mudra D, Carroll K, LeCluyse E, Parkinson A. 1999. Effect of cryopreservation on cytochrome P-450 enzyme induction in cultured rat hepatocytes. *Drug Metab. Dispos.* 27:327–35.
103. Li AP, Maurel P, Gomez-Lechon MJ, Cheng LC, Jurima-Romet M. 1997. Pre-clinical evaluation of drug-drug interaction potential: present status of the application of primary human hepatocytes in the evaluation of cytochrome P450 induction. *Chem. Biol. Interact.* 107:5–16.
104. Li AP, Lu C, Brent JA, Pham C, Fackett A, et al. 1999. Cryopreserved human hepatocytes: characterization of drug-metabolizing enzyme activities and applications in higher throughput screening assays for hepatotoxicity, metabolic stability, and drug-drug interaction potential. *Chem. Biol. Interact.* 121:17–35.
105. Li AP, Gorycki PD, Hengstler JG, Kederis GL, Koebe HG, et al. 1999. Present status of the application of cryopreserved hepatocytes in the evaluation of xenobiotics: consensus of an international expert panel. *Chem. Biol. Interact.* 121:117–23.
106. Shitara Y, Li AP, Kato Y, Lu C, Ito K, et al. 2003. Function of uptake transporters for taurocholate and estradiol 17 β -D-glucuronide in cryopreserved human hepatocytes. *Drug Metab. Pharmacokin.* 18:33–41.
107. Olinga P, Hof IH, Merema MT, Smit M, de Jager MH, et al. 2001. The applicability of rat and human liver slices to the study of mechanisms of hepatic drug uptake.

- J. Pharmacol. Toxicol. Methods 45:55–63
108. Shigematsu A, Motoji N, Momose Y, Iida A, Higashi N. 2000. Viability of liver slices exhibiting absorption, metabolism, and elimination of substrates in culture medium. *Exp. Mol. Pathol.* 69:119–43
109. Pritchard JB, Miller DS. 1993. Mechanisms mediating renal secretion of organic anions and cations. *Physiol. Rev.* 73:765–96
110. Murer H, Gmaj P, Steiger B, Hagenbuch B. 1989. Transport studies with renal proximal tubular and small intestinal brush border and basolateral membrane vesicles: vesicle heterogeneity, coexistence of transport system. *Methods Enzymol.* 172:346–64
111. Boyer JL, Meier PJ. 1990. Characterizing mechanisms of hepatic bile acid transport utilizing isolated membrane vesicles. *Methods Enzymol.* 192:517–33
112. Kinne-Saffran E, Kinne RK. Isolation of luminal and cotralumenal plasma membrane vesicles from kidney. *Methods Enzymol.* 191:450–69
113. Meier PJ, Boyer JL. 1990. Preparation of basolateral (sinusoidal) and canalicular plasma membrane vesicles for the study of hepatic transport processes. *Methods Enzymol.* 192:534–45
114. Meier PJ, St Meier-Abt A, Barrett C, Boyer JL. 1984. Mechanisms of taurocholate transport in canalicular and basolateral rat liver plasma membrane vesicles. Evidence for an electrogenic canalicular organic anion carrier. *J. Biol. Chem.* 259:10614–22
115. Cui Y, Konig J, Keppler D. 2001. Vectorial transport by double-transfected cells expressing the human uptake transporter SLC21A8 and the apical export pump ABCC2. *Mol. Pharmacol.* 60:934–43
116. Schinkel AH, Mayer U, Wagenaar E, Mol CA, van Deemter L, et al. 1997. Normal viability and altered pharmacokinetics in mice lacking mdrl-type (drug-transporting) P-glycoproteins. *Proc. Natl. Acad. Sci. USA* 94:4028–33
117. Hagenbuch B, Scharschmidt BF, Meier PJ. 1996. Effect of antisense oligonucleotides on the expression of hepatocellular bile acid and organic anion uptake systems in *Xenopus laevis* oocytes. *Biochem. J.* 316:901–4
118. Tamai I, Nakanishi T, Hayashi K, Terao T, Sai Y, et al. 1997. The predominant contribution of oligopeptide transporter PepT1 to intestinal absorption of beta-lactam antibiotics in the rat small intestine. *J. Pharm. Pharmacol.* 49:796–801
119. Kouzuki H, Suzuki H, Ito K, Ohashi R, Sugiyama Y. 1998. Contribution of sodium taurocholate co-transporting polypeptide to the uptake of its possible substrates into rat hepatocytes. *J. Pharmacol. Exp. Ther.* 286:1043–50
120. Kouzuki H, Suzuki H, Ito K, Ohashi R, Sugiyama Y. 1999. Contribution of organic anion transporting polypeptide to uptake of its possible substrates into rat hepatocytes. *J. Pharmacol. Exp. Ther.* 288:627–34
121. Hirano M, Maeda K, Shitara Y, Sugiyama Y. 2004. Contribution of OATP2 (OATP1B1) and OATP8 (OATP1B3) to the hepatic uptake of pitavastatin in humans. *J. Pharmacol. Exp. Ther.* 311:139–46
122. Shitara Y, Sugiyama D, Kusuhara H, Kato Y, Abe T. 2002. Comparative inhibitory effects of different compounds on rat oatpl (*Slc21a1*)- and Oatp2 (*Slc21a5*)-mediated transport. *Pharm. Res.* 19:147–53
123. Ito K, Iwatsubo T, Kanamitsu S, Ueda K, Suzuki H, et al. 1998. Prediction of pharmacokinetic alterations caused by drug-drug interactions: metabolic interaction in the liver. *Pharmacol. Rev.* 50:387–412
124. Ito K, Iwatsubo T, Kanamitsu S, Nakajima Y, Sugiyama Y. 1998. Quantitative prediction of in vivo drug clearance and drug interactions from in vitro data on metabolism, together with binding and

DRUG-DRUG INTERACTION INVOLVING TRANSPORTERS 719

- transport. *Annu. Rev. Pharmacol. Toxicol.* 38:461–99
125. Ueda K, Kato Y, Komatsu K, Sugiyama Y. 2001. Inhibition of biliary excretion of methotrexate by probenecid in rats: quantitative prediction of interaction from *in vitro* data. *J. Pharmacol. Exp. Ther.* 297:1036–43
126. Saito H, Masuda S, Inui K. 1996. Cloning and functional characterization of a novel rat organic anion transporter mediating basolateral uptake of methotrexate in the kidney. *J. Biol. Chem.* 271:20719–25
127. Masuda S, Saito H, Inui K. 1997. Interactions of nonsteroidal anti-inflammatory drugs with rat renal organic anion transporter, OAT-K1. *J. Pharmacol. Exp. Ther.* 283:1039–42
128. Masuda S, Takeuchi A, Saito H, Hashimoto Y, Inui K. 1999. Functional analysis of rat renal organic anion transporter OAT-K1: bidirectional methotrexate transport in apical membrane. *FEBS Lett.* 459:128–32
129. Masuda S, Ibamoto K, Takeuchi A, Saito H, Hashimoto Y, et al. 1999. Cloning and functional characterization of a new multispecific organic anion transporter, OAT-K2, in rat kidney. *Mol. Pharmacol.* 55:743–52
130. Takeuchi A, Masuda S, Saito H, Abe T, Inui K. 2001. Multispecific substrate recognition of kidney-specific organic anion transporters OAT-K1 and OAT-K2. *J. Pharmacol. Exp. Ther.* 299:261–67
131. Lu R, Kanai N, Bao Y, Wolkoff AW, Schuster VL. 1996. Regulation of renal oatp mRNA expression by testosterone. *Am. J. Physiol. Renal Physiol.* 270:F332–37
132. Yamazaki M, Tokui T, Ishigami M, Sugiyama Y. 1996. Tissue-selective uptake of pravastatin in rats: contribution of a specific carrier-mediated uptake system. *Biopharm. Drug Dispos.* 17:775–89
133. Hedman A, Angelin B, Arvidsson A, Dahlqvist R, Nilsson B. 1990. Interactions in the renal and biliary elimination of digoxin: stereoselective difference between quinine and quinidine. *Clin. Pharmacol. Ther.* 47:20–26
134. Bohme M, Jedlitschky G, Leier I, Buchler M, Keppler D. 1994. ATP-dependent export pumps and their inhibition by cyclosporins. *Adv. Enzyme Regul.* 34:371–80
135. Versantvoort CH, Broxterman HJ, Lankelma J, Feller N, Pinedo HM. 1994. Competitive inhibition by genistein and ATP dependence of daunorubicin transport in intact MRP overexpressing human small cell lung cancer cells. *Biochem. Pharmacol.* 48:1129–36
136. Chu XY, Kato Y, Niinuma K, Sudo KI, Hakusui H, et al. 1997. Multispecific organic anion transporter is responsible for the biliary excretion of the camptothecin derivative irinotecan and its metabolites in rats. *J. Pharmacol. Exp. Ther.* 281:304–14
137. Araki E, Ishikawa M, Iigo M, Koide T, Itabashi M, et al. 1993. Relationship between development of diarrhea and the concentration of SN-38, an active metabolite of CPT-11, in the intestine and the blood plasma of athymic mice following intraperitoneal administration of CPT-11. *Jpn. J. Cancer Res.* 84:697–702
138. Gupta E, Lestingi TM, Mick R, Ramirez J, Vokes EE, et al. 1994. Metabolic fate of irinotecan in humans: correlation of glucuronidation with diarrhea. *Cancer Res.* 54:3723–25
139. Horikawa M, Kato Y, Tyson CA, Sugiyama Y. 2002. The potential for an interaction between MRP2 (ABCC2) and various therapeutic agents: probenecid as a candidate inhibitor of the biliary excretion of irinotecan metabolites. *Drug Metab. Pharmacokin.* 17:23–33
140. Horikawa M, Kato Y, Sugiyama Y. 2002. Reduced gastrointestinal toxicity following inhibition of the biliary excretion of irinotecan and its metabolites by probenecid in rats. *Pharm. Res.* 19:1345–53

141. Mück W. 2000. Clinical pharmacokinetics of cerivastatin. *Clin. Pharmacokinet.* 39:99–116
142. Mück W, Mai I, Fritzsche L, Ochmann K, Rohde G, et al. 1999. Increase in cerivastatin systemic exposure after single and multiple dosing in cyclosporine-treated kidney transplant recipients. *Clin. Pharmacol. Ther.* 65:251–61
143. Regazzi MB, Campana IC, Raddato V, Lesi C, Perani G, et al. 1993. Altered disposition of pravastatin following concomitant drug therapy with cyclosporin A in transplant recipients. *Transplant. Proc.* 25:2732–34
144. Hasunuma T, Nakamura M, Yachi T, Ariyawa N, Fukushima K, et al. 2003. The drug-drug interactions of pitavastatin (NK-104), a novel HMG-CoA reductase inhibitor and cyclosporine. *J. Clin. Ther. Med.* 19:381–89
145. Asberg A, Hartmann A, Fjeldsa E, Bergan S, Holdaas H. 2001. Bilateral pharmacokinetic interaction between cyclosporine A and atorvastatin in renal transplant recipients. *Am. J. Transplant.* 1: 382–86
146. Lennernas H. 2003. Clinical pharmacokinetics of atorvastatin. *Clin. Pharmacokinet.* 42:1141–60
147. Bruce-Joyce J, Dugas JM, MacCausland OE. 2001. Cerivastatin and gemfibrozil-associated rhabdomyolysis. *Ann. Pharmacother.* 35:1016–19
148. Mueck W, Frey R, Boix O, Voith B. 2001. Gemfibrozil/cerivastatin interaction. *AAPS PharmSci.* 3(Suppl.):3566 (Abstr.)
149. Backman JT, Kyrlund C, Neuvonen M, Neuvonen PJ. 2002. Gemfibrozil greatly increases plasma concentrations of cerivastatin. *Clin. Pharmacol. Ther.* 72:685–91
150. Shitara Y, Hirano M, Sato H, Sugiama Y. 2004. Gemfibrozil and its glucuronide inhibit the OATP2(OATP1B1: SLC21A6)-mediated hepatic uptake and CYP2C8-mediated metabolism of cerivastatin—analysis of the mechanism of the clinically relevant drug-drug interaction between cerivastatin and gemfibrozil. *J. Pharmacol. Exp. Ther.* 311:228–36
151. Schneck DW, Birmingham BK, Zalikowski JA, Mitchell PD, Wang Y, et al. 2004. The effect of gemfibrozil on the pharmacokinetics of rosuvastatin. *Clin. Pharmacol. Ther.* 75:455–63
152. Sallustio BC, Fairchild BA, Shanahan K, Evans AM, Nation RL. 1996. Disposition of gemfibrozil and gemfibrozil acyl glucuronide in the rat isolated perfused liver. *Drug Metab. Dispos.* 24:984–89
153. Sabordo L, Sallustio BC, Evans AM, Nation RL. 1998. Hepatic disposition of the acyl glucuronide 1-O-gemfibrozil β -D-glucuronide: effects of dibromosulfophthalein on membrane transport and aglycone formation. *J. Pharmacol. Exp. Ther.* 288:414–20
154. Sabordo L, Sallustio BC, Evans AM, Nation RL. 2000. Hepatic disposition of the acyl glucuronide 1-O-gemfibrozil- β -D-glucuronide: effects of clofibrate acid, acetaminophen, and acetaminophen glucuronide. *J. Pharmacol. Exp. Ther.* 295: 44–50
155. Hedman A, Meijer DK. 1998. Stereoselective inhibition by the diastereomers quinidine and quinine of uptake of cardiac glycosides into isolated rat hepatocytes. *J. Pharm. Sci.* 87:457–61
156. Olinga P, Merema M, Hof IH, Slooff MJ, Proost JH, et al. 1998. Characterization of the uptake of rocuronium and digoxin in human hepatocytes: carrier specificity and comparison with in vivo data. *J. Pharmacol. Exp. Ther.* 285:506–10
157. Brown GR. 1993. Cephalosporin-probenecid drug interactions. *Clin. Pharmacokinet.* 24:289–300
158. Marino EL, Dominguez-Gil A. 1981. The pharmacokinetics of cefadroxil associated with probenecid. *Int. J. Clin. Pharmacol. Ther. Toxicol.* 19:506–8
159. Griffith RS, Black HR, Brier GL, Wolny JD. 1977. Effect of probenecid on

DRUG-DRUG INTERACTION INVOLVING TRANSPORTERS 721

- the blood levels and urinary excretion of cefamandole. *Antimicrob Agents Chemother.* 11:809–12
160. Stoeckel K, Trueb V, Dubach UC, McNamara PJ. 1988. Effect of probenecid on the elimination and protein binding of ceftriaxone. *Eur. J. Clin. Pharmacol.* 34:151–56
161. Furst DE. 1995. Practical clinical pharmacology and drug interactions of low-dose methotrexate therapy in rheumatoid arthritis. *Br. J. Rheumatol.* 34(Suppl. 2):20–25
162. Tracy TS, Krohn K, Jones DR, Bradley JD, Hall SD, et al. 1992. The effects of a salicylate, ibuprofen, and naproxen on the disposition of methotrexate in patients with rheumatoid arthritis. *Eur. J. Clin. Pharmacol.* 42:121–25
163. Kremer JM, Hamilton RA. 1995. The effects of nonsteroidal antiinflammatory drugs on methotrexate (MTX) pharmacokinetics: impairment of renal clearance of MTX at weekly maintenance doses but not at 7.5 mg. *J. Rheumatol.* 22:2072–77
164. Frenia ML, Long KS. 1992. Methotrexate and nonsteroidal antiinflammatory drug interactions. *Ann. Pharmacother.* 26:234–37
165. Nozaki Y, Kusuvara H, Endou H, Sugiyama Y. 2004. Quantitative evaluation of the drug-drug interactions between methotrexate and nonsteroidal antiinflammatory drugs in the renal uptake process based on the contribution of organic anion transporters and reduced folate carrier. *J. Pharmacol. Exp. Ther.* 309: 226–34
166. Apiwattanakul N, Sekine T, Chairoungdua A, Kanai Y, Nakajima N, et al. 1999. Transport properties of nonsteroidal anti-inflammatory drugs by organic anion transporter 1 expressed in *Xenopus laevis* oocytes. *Mol. Pharmacol.* 55:847–54
167. Uwai Y, Saito H, Inui K. 2000. Interaction between methotrexate and nonsteroidal antiinflammatory drugs in organic anion transporter. *Eur. J. Pharmacol.* 409:31–36
168. Lu R, Chan BS, Schuster VL. 1999. Cloning of the human kidney PAH transporter: narrow substrate specificity and regulation by protein kinase C. *Am. J. Physiol. Renal Physiol.* 276:F295–303
169. Ohtsuki S, Asaba H, Takanaga H, Deguchi T, Hosoya K, et al. 2002. Role of blood-brain barrier organic anion transporter 3 (OAT3) in the efflux of indoxyl sulfate, a uremic toxin: its involvement in neurotransmitter metabolite clearance from the brain. *J. Neurochem.* 83:57–66
170. Sekine T, Cha SH, Tsuda M, Apiwattanakul N, Nakajima N, et al. 1998. Identification of multispecific organic anion transporter 2 expressed predominantly in the liver. *FEBS Lett.* 429:179–82
171. Morita N, Kusuvara H, Sekine T, Endou H, Sugiyama Y. 2001. Functional characterization of rat organic anion transporter 2 in LLC-PK1 cells. *J. Pharmacol. Exp. Ther.* 298:1179–84
172. Kusuvara H, Sekine T, Utsunomiya-Tate N, Tsuda M, Kojima R, et al. 1999. Molecular cloning and characterization of a new multispecific organic anion transporter from rat brain. *J. Biol. Chem.* 274:13675–80
173. Jariyawat S, Sekine T, Takeda M, Apiwattanakul N, Kanai Y, et al. 1999. The interaction and transport of beta-lactam antibiotics with the cloned rat renal organic anion transporter 1. *J. Pharmacol. Exp. Ther.* 290:672–77
174. Takeda M, Babu E, Narikawa S, Endou H. 2002. Interaction of human organic anion transporters with various cephalosporin antibiotics. *Eur. J. Pharmacol.* 438:137–42
175. Jung KY, Takeda M, Shimoda M, Narikawa S, Tojo A, et al. 2002. Involvement of rat organic anion transporter 3 (rOAT3) in cephaloridine-induced nephrotoxicity: in comparison with rOAT1. *Life Sci.* 70:1861–74
176. Uwai Y, Saito H, Inui K. 2002. Rat renal organic anion transporter rOAT1 mediated transport of urinary-excreted

- cephalosporins, but not of biliary excreted cefoperazone. *Drug Metab. Pharmacokin.* 17:125-29
177. Babu E, Takeda M, Narikawa S, Kobayashi Y, Enomoto A, et al. 2002. Role of human organic anion transporter 4 in the transport of ochratoxin A. *Biochim. Biophys. Acta* 1590:64-75
178. Babu E, Takeda M, Narikawa S, Kobayashi Y, Yamamoto T, et al. 2002. Human organic anion transporters mediate the transport of tetracycline. *Jpn. J. Pharmacol.* 88:69-76
179. Uwai Y, Saito H, Hashimoto Y, Inui K. 2000. Interaction and transport of thiazide diuretics, loop diuretics, and acetazolamide via rat renal organic anion transporter rOAT1. *J. Pharmacol. Exp. Ther.* 295:261-65
180. Uwai Y, Saito H, Hashimoto Y, Inui K. 2000. Inhibitory effect of anti-diabetic agents on rat organic anion transporter rOAT1. *Eur. J. Pharmacol.* 398:193-97
181. Nagata Y, Kusuvara H, Endou H, Sugiyama Y. 2002. Expression and functional characterization of rat organic anion transporter 3 (rOat3) in the choroid plexus. *Mol. Pharmacol.* 61:982-88
182. Takeda M, Narikawa S, Hosoyamada M, Cha SH, Sekine T, et al. 2001. Characterization of organic anion transport inhibitors using cells stably expressing human organic anion transporters. *Eur. J. Pharmacol.* 419:113-20
183. Enomoto A, Takeda M, Shimoda M, Narikawa S, Kobayashi Y, et al. 2002. Interaction of human organic anion transporters 2 and 4 with organic anion transport inhibitors. *J. Pharmacol. Exp. Ther.* 301:797-802
184. Cvetkovic M, Leake B, Fromm MF, Wilkinson GR, Kim RB. 1999. OATP and P-glycoprotein transporters mediate the cellular uptake and excretion of fexofenadine. *Drug Metab. Dispos.* 27:866-71
185. Tokui T, Nakai D, Nakagomi R, Yawo H, Abe T, et al. 1999. Pravastatin, an HMG-CoA reductase inhibitor, is transported by rat organic anion transporting polypeptide, oatp2. *Pharm. Res.* 16:904-8
186. Kullak-Ublick GA, Fisch T, Oswald M, Hagenbuch B, Meier PJ, et al. 1998. Dehydroepiandrosterone sulfate (DHEAS): identification of a carrier protein in human liver and brain. *FEBS Lett.* 424:173-76
187. Bossuyt X, Muller M, Hagenbuch B, Meier PJ. 1996. Polyspecific drug and steroid clearance by an organic anion transporter of mammalian liver. *J. Pharmacol. Exp. Ther.* 276:891-96
188. Sugiyama D, Kusuvara H, Shitara Y, Abe T, Meier PJ, et al. 2001. Characterization of the efflux transport of 17 β -estradiol-D-glucuronide from the brain across the blood-brain barrier. *J. Pharmacol. Exp. Ther.* 298:316-22
189. Sugiyama D, Kusuvara H, Shitara Y, Abe T, Sugiyama Y. 2002. Effect of 17 β -estradiol-D-17 β -glucuronide on the rat organic anion transporting polypeptide 2-mediated transport differs depending on substrates. *Drug Metab. Dispos.* 30:220-23
190. Abe T, Unno M, Onogawa T, Tokui T, Kondo TN, et al. 2001. LST-2, a human liver-specific organic anion transporter, determines methotrexate sensitivity in gastrointestinal cancers. *Gastroenterology* 120:1689-99
191. Vavricka SR, van Montfoort J, Ha HR, Meier PJ, Fattinger K. 2002. Interactions of rifamycin SV and rifampicin with organic anion uptake systems of human liver. *Hepatology* 36:164-72
192. Fattinger K, Cattori V, Hagenbuch B, Meier PJ, Stieger B. 2000. Rifamycin SV and rifampicin exhibit differential inhibition of the hepatic rat organic anion transporting polypeptides, Oatp1 and Oatp2. *Hepatology* 32:82-86
193. Ichimaru N, Takahara S, Kokado Y, Wang J-D, Hatori M, et al. 2001. Changes in lipid metabolism and effect of simvastatin in renal transplant recipients induced by cyclosporine or tacrolimus. *Atherosclerosis* 158:417-23

DRUG-DRUG INTERACTION INVOLVING TRANSPORTERS 723

194. Arnadottir M, Eriksson LO, Thysell H, Karkas JD. 1993. Plasma concentration profiles of simvastatin 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase inhibitory activity in kidney transplant recipients with and without cyclosporin. *Nephron* 65:410–13.
195. Goldberg R, Roth D. 1996. Evaluation of fluvastatin in the treatment of hypercholesterolemia in renal transplant recipients taking cyclosporine. *Transplantation* 62:1559–64.
196. Kyrlund C, Backman JT, Kivistö KT, Neuvonen M, Latila J, et al. 2001. Plasma concentrations of active lovastatin acid are markedly increased by gemfibrozil but not by bezafibrate. *Clin. Pharmacol. Ther.* 69:340–45.
197. Backman JT, Kyrlund C, Kivistö KT, Wang J-S, Neuvonen PJ. 2000. Plasma concentrations of active simvastatin acid are increased by gemfibrozil. *Clin. Pharmacol. Ther.* 68:122–29.
198. Kyrlund C, Backman JT, Neuvonen M, Neuvonen PJ. 2003. Gemfibrozil increases plasma pravastatin concentrations and reduces pravastatin renal clearance. *Clin. Pharmacol. Ther.* 73:538–44.
199. Spence JD, Munoz CE, Hendricks L, Latchinian L, Khouri HE. 1995. Pharmacokinetics of the combination of fluvastatin and gemfibrozil. *Am. J. Cardiol.* 76: 80A–83.
200. Mathew P, Cuddy T, Tracewell WG, Salazar D. 2004. An open-label study on the pharmacokinetics (PK) of pitavastatin (NK-104) when administered concomitantly with fenofibrate or gemfibrozil in healthy volunteers. *Clin. Pharmacol. Ther.* 75:P33.

遺伝子多型と薬物の効果の個人差

前田和哉, 杉山雄一

Katsuya Maeda, Yuichi Sugiyama

東京大学大学院薬学系研究科分子薬物動態学教室

ポイント

- 薬効の個人差を生み出す要因は、薬物動態的な要因と薬力学的な要因の2つに大別され、それぞれを決定するタンパク質の遺伝子多型が重要と考えられる。
- 体内動態にかかわる分子には、代謝酵素・トランスポーターがあり、それぞれ遺伝子多型によって生じる機能変化が薬物動態を大きく変え、ひいては効果・副作用を規定する事例が数多く報告されている。
- 薬効・副作用の感受性の個人差は、直接薬のターゲットタンパク質の遺伝子多型によるケース以外に、薬効標的と関連のないタンパク質の多型とリンクするケースがあり、今後広範囲な遺伝子多型と感受性の関連解析、原因の追究が望まれる。

はじめに

臨床において、同じ量の薬物を投与しても、適正な治療効果を示す人もあるが、まったく効果のない人や副作用ができる人もある。その手がかりとして、患者集団での薬物血中濃度のヒストограмに多峰性が観察されたり、2種の薬物を個々の患者に投与したときに、それぞれの代謝物の総量に相関が認められたりすること〔たとえば、デブリソキンとスバルテイン¹⁾（CYP2D6の多型に起因する）〕、また一卵性双生児と二卵性双生児ペアに薬物を投与して動態をペア間で比較したところ一卵性双生児の方が有意にペア間の変動が小さいことなどから、薬物動態の個人差の原因として遺伝的要因が関与することが示唆されてきた。現在では、これらは薬物代謝酵素の遺伝子多型で説明がついている。

一般的に、薬効・副作用の個人間変動に影響する過程は、投与後の体内での薬物動態の個人差の部分（pharmacokinetics）と、薬物の標的

部位との相互作用、シグナル伝達の感受性など薬力学的な個人差の部分（pharmacodynamics）に大別できる。前者は、薬物の吸収、分布、代謝、排泄を決定する代謝酵素・トランスポーターの遺伝子多型が主に関与し、投与設計、投与量の個別化で個人差を克服できる。一方、後者は、薬物の標的となるレセプター、転写因子や、その下流のシグナル伝達を行うアダプター分子群の遺伝子多型が該当し、個人差の克服には薬剤選択の個別化が必要となる。またより厳密には、薬効発現の程度は標的分子近傍の薬物濃度の関数として表されることから、細胞内にターゲットをもつ場合は、血中濃度のみならず、標的細胞内濃度を決定する要因も考慮に入れる必要がある（たとえば耐性癌細胞における排出トランスポーター）（図1）。

薬物の体内動態にかかわる分子の遺伝子多型

一般に薬物の排泄は、① 血中から臓器細胞へ

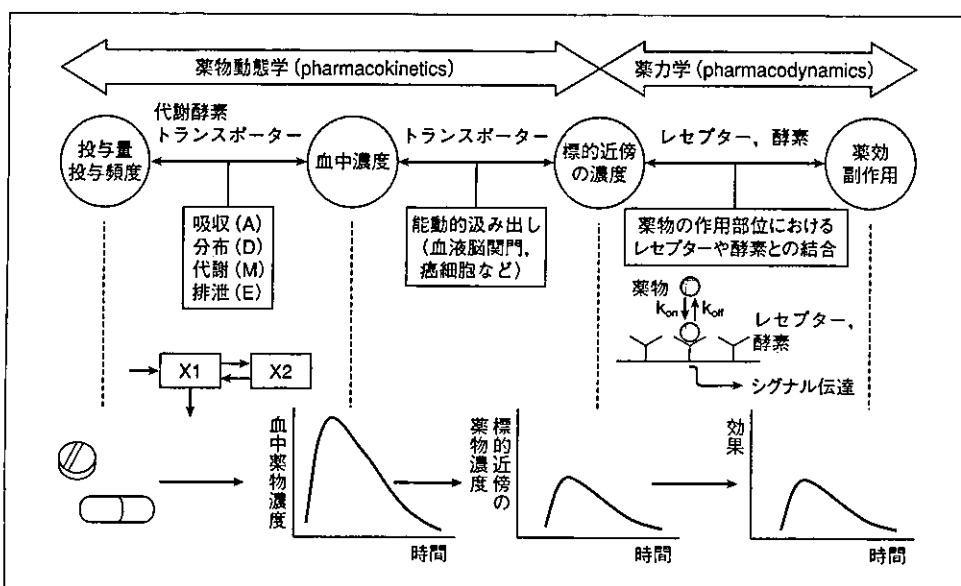


図1 薬物が効果・副作用を発現するまでの過程

(杉山雄一ほか編：ファーマコキネティクス 演習による理解、p249、南山堂、2003²⁾より改変)

の細胞膜を介した取り込み、②細胞内での主にシトクロム P-450 (CYP) による酸化反応などの種々の物質変換、③グルクロン酸、グルタチオン、硫酸基など水溶性を上げる官能基を付加する抱合反応、④細胞内からの排出の各過程からなっており、それぞれの段階において多くの分子が機能している。①、④の段階においては、脂溶性が低く容易に膜透過できない物質について多くのトランスポーターが効率良い膜輸送を担っていることが明らかとなっており、これらの遺伝子多型も現在解析が進みつつある。

1. CYP の遺伝子多型と薬物動態の個人差

CYP は薬物の排泄にかかわる分子としていち早く遺伝子多型解析と体内動態との個人差の相関研究が行われた領域であり、臨床でも多くの事例が集積している。また多型のハプロタイプごとに体系化された命名法が確立している³⁾。

CYP2D6 は、肝臓内における CYP 含量としては比較的少ないが、薬物代謝への寄与は、CYP3A4 に次ぎ薬物全体の約 20 % を占めるほ

ど重要な酵素である⁴⁾。現在までに、79 種類もの変異アレルの報告がある。多型の頻度には大きな人種差が認められている。欧米人には、機能欠損した遺伝子（主に*3, *4, *5）を両アレルにもつ poor metabolizer (PM) が 5~10 % 存在するのに対し、日本人では、PM の頻度は 1 % 未満（主に*5）である。さらに発現量と結合親和性の低下を示す⁵⁾は、東洋人にはのみ高頻度に出現し、PM と extensive metabolizer (EM, 野生型) の中間の性質を示す intermediate metabolizer (IM, *5/*10, *10/*10) の分類が存在する。また、頻度は少いものの、2D6 の遺伝子重複による発現の上昇がみられる ultra rapid metabolizer (UM) も存在する。2D6 の基質薬物には、β遮断薬（プロプラノロール、メトプロロール）、抗うつ薬（ノルトリプチリン、フルオキセチン）、抗不整脈薬（プロバフェノン）など治療域の狭い重要な薬物が多く、実際に PM, IM など遺伝子型に対応した血中クリアランスの変動がみられている^{5, 6)}。また、向精神薬の副作用発現と 2D6

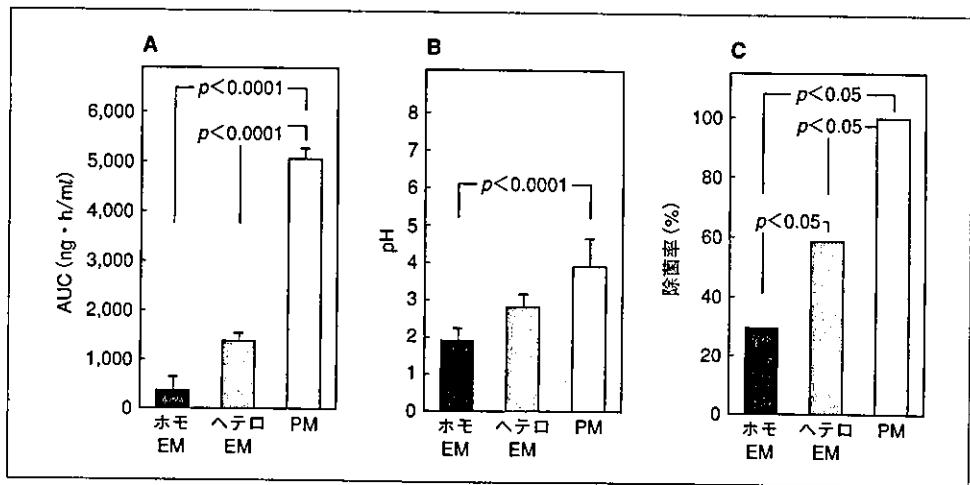


図2 CYP2C19の遺伝子多型がオメプラゾール(OMZ)に与える影響

A: OMZ 20 mg 経口投与時の AUC

B: OMZ 20 mg 経口投与後 24 時間の平均胃内 pH

C: OMZ 20 mg +アモキシリン 2.0 g、2 週間投与後の *H.pylori* の除菌率

EM: extensive metabolizer, PM: poor metabolizer

(大河内秀昭: 月刊薬事 43, 473-7, 2001¹⁴⁾ より)

の EM の割合との間にも相関が認められるとの報告がある⁷⁾ ことから、特に初期の投与量設定において多型診断は臨床上重要な役割を示すと考えられる。

CYP2C19 は、PM (主に*2, *3) の頻度は東洋人 (20 %) が欧米人 (10 %) より高い⁸⁾。2C19 はプロトンポンプ阻害薬 (PPI) を基質とすることから、2C19 の多型と、PPI の薬物動態・効果、また PPI は胃十二指腸潰瘍時の *H. pylori* 除菌療法に併用されるため除菌成績との関連が詳細に観察されている⁹⁻¹³⁾。2C19 の PM では、オメプラゾール経口投与後の AUC (薬物血中濃度時間曲線下面積) が EM の 10 倍まで上昇する¹¹⁾。それに伴い、投与 24 時間後の平均胃内 pH の上昇も PM の方が有意に高い¹¹⁾。また、抗生素質アモキシリンとの併用による除菌成績も PM の方が有意に高く、胃内 pH 上昇による抗生素質の安定化と *H. pylori* の増殖による感受性上昇によるものと考えられている¹⁰⁾。実際に、オメプラゾールの投与量を増加すると除菌効率の上昇がみられること¹²⁾、

また 2C19 による代謝の寄与が比較的小さいラベプラゾールについては、上記のような変動は小さかったこと^{9, 13)} からも 2C19 の遺伝子型による PPI の投与設計が有効であると考えられる (図2)¹⁴⁾。

CYP2C9 にも、*3 を保有する *1/*3 群が 2 ~3 % 程度存在する。2C9 はきわめて治療域の狭い薬剤であるフェニトインやワルファリンの主要代謝酵素で、血中濃度の変動と中枢毒性や抗凝固能の変化といった重篤な副作用との相関が報告されており^{15, 16)}、多型の頻度は低いものの、遺伝子型による投与量の層別化が必要であろうと思われる。一方で、2C9 を主代謝経路にもつジクロフェナクは、*3 変異による代謝能の変化がみられないとの報告¹⁷⁾ があり、多型の影響が薬物により異なることが示唆されている。

2. CYP 以外の代謝酵素 (抱合酵素) の遺伝子多型と薬物動態の個人差

1950 年代に、抗結核薬イソニアジドの N-

アセチル化代謝活性に個人差がみられることがすでに示唆されており、現在ではその原因がNAT (*N*-acetyltransferase) 2にあることがわかっている。すなわち、通常は加水分解産物のヒドラジンがNAT2によりすみやかにアセチル化され解毒されるが、変異によりNAT2活性の低下があると、ヒドラジンが肝臓内に蓄積し、肝障害を発現すると考えられている¹⁸⁾。また、抗不整脈薬プロカインアミドや潰瘍性大腸炎治療薬スルファサラジンについてはNAT2変異と全身性エリテマトーデス様の副作用発現の間に関連が認められている^{19, 20)}。

CPT-11 (カンプトテシン) の活性代謝物SN-38は、UGT (UDP-glucuronosyltransferase) 1A1により肝臓で抱合を受け解毒・胆汁排泄される。UGTの変異は、もともと高ビリルビン血症であるCrigler-Najjar症候群およびGilbert症候群の解析から多数同定されてきた²¹⁾。UGT1A1プロモーター領域のTAリピートが通常より1回多い変異では転写活性が半分に低下しており、UGTの活性低下からSN-38の消失遅延につながる可能性が考えられている²²⁾。また、TPMT (thiopurine S-methyltransferase) はプリン代謝拮抗薬メルカブトプリン (6-MP) の活性本体6-チオグアニンヌクレオチドの不活化を担っている酵素であり、特に*2, *3変異体は、発現量の低下による活性体の消失遅延、それに伴う骨髄抑制など重篤な副作用発現を引き起こす²³⁾。ドナーの赤血球ライセートと6-MPを混合して活性体の生成をみる方法もある²⁴⁾が、遺伝子多型判定が簡便になれば、TPMTの多型による投与量の層別化も今後可能になると思われる²⁵⁾。また、5-FU (5-fluorouracil) の代謝律速酵素であるDPD (dehydroxyuridine dehydrogenase) も活性の低下する多型があり、5-FUの神経毒性が低活性の人で有意に多くみられている²⁶⁾。

3. トランスポーターの遺伝子多型と薬物動態の個人差

近年、ヒトにおいても非常に多様なトランスポーターの発現が認められており、容易に膜透過できない化合物の濃縮的な取り込み・排出機構として働いていることが知られている。一般にトランスポーターは、きわめて広範な構造の化合物を認識することが知られており、それゆえ、トランスポーターの遺伝子多型による輸送活性、発現変動は、多くの基質化合物の輸送を変え、体内動態に影響を与えることが考えうる。

臨床において、遺伝子多型と薬物動態の個人差について関連解析が最も進んでいるのは、MDR1 [P-gp (P糖タンパク質), ABCB1] であると思われる。MDR1は、肝臓の胆管側や小腸の管腔側、血液脳関門の血管側に発現が認められ、ATP水解活性を駆動力として細胞内からの排出を担うタンパク質である。現在では多くの遺伝子多型が同定されているが、なかでもアミノ酸変化を伴わない変異であるC3435Tは、興味深いことに、この変異の保有者の十二指腸におけるMDR1の発現量は有意に低く、経口投与したジゴキシンのAUCの上昇が認められている²⁷⁾。別の機能変化、発現変動を引き起こしうる変異との連鎖が想定され、実際、G2677 (T, A), C1236Tとの間に連鎖不平衡が観察されている²⁸⁾。ただし *in vitro* 発現系を用いた実験においては、2677, 3435位の変異を考慮した多型体の機能解析では差異がみられないことが報告されている²⁹⁾。また連鎖不平衡ブロックの統計解析から、既知の変異の周辺40~80 kb以内に別の未知の変異がリンクする可能性が示唆されており²⁸⁾、今後の解析が待たれる。最近のハプロタイプ解析により、先に述べた3か所の変異保有者のみならず、7か所のイントロンのみの変異の保有者においてもSN-38の腎クリアランスの低下がみられており³⁰⁾、きわめて興味深い。しかし、多くの臨

床試験のなかで、先のジゴキシンの結果に矛盾する結果も報告されており³¹⁾、今後情報の蓄積が望まれる。

また最近、肝臓に選択的な発現が認められる取り込みトランスポーター OATP2 (SLC21A6, OATP1B1) についても多型に関して報告がされてきた。たとえば *in vitro* 実験で、抗結核薬リファンピシンの輸送能力が多型により変動し、そのため PXR (pregnane X receptor) を介した転写誘導能にも影響を与える可能性を示唆する報告もある³²⁾。日本人においては、特に*15 (Asn130Asp, Val174Ala) が約 10 % 存在しており、HMG-CoA (hydroxymethylglutaryl-CoA) 還元酵素阻害薬プラバスタチンの腎外クリアランス (すなわち肝クリアランス) が*15 保有者で有意に低下することが臨床事例として初めて報告された (図 3)³³⁾。一方、筆者らは *in vitro* 実験系を用いて*15 変異が単位タンパク質あたりの輸送量を低下させることを示し、腎外クリアランス低下の程度と半定量的に一致することを見出している (投稿中)。OATP2 は基質認識性が非常に広範であることから、多様な薬物の肝クリアランスの第 1 ステップを担う取り込みトランスポーターの変異が体内動態に影響を与える可能性は十分に考えられ、今後のさらなる臨床試験が待たれる。

薬物の効果の感受性にかかわる分子の遺伝子多型

近年、多くの薬物に関して薬効の標的分子やその後のシグナル伝達、転写制御のカスケードが明らかになるにつれ、それらの遺伝子多型と効果との関連を示す報告が相次いでいる。特にレセプター分子や酵素との結合を介して効果を発揮する薬物にとって、直接相互作用する分子の遺伝子多型は薬効に影響を与える可能性が強く、解析が進んでいる。最近の主な事例を簡単に表 1 にまとめた³⁴⁻³⁶⁾。

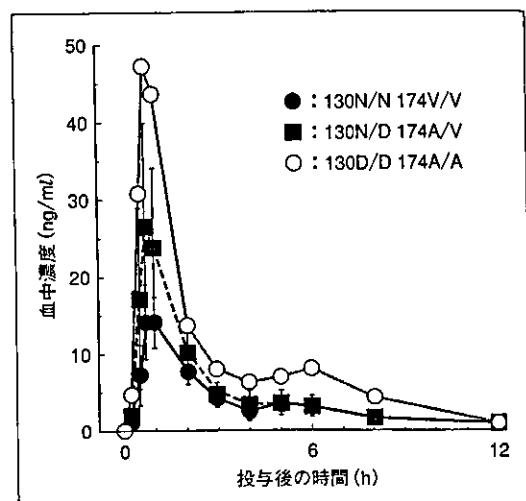


図 3 OATP2 遺伝子多型とプラバスタチン血中濃度推移との関連

(Nishizato Y, et al : *Clin Pharmacol Ther* 73, 554-65, 2003³³⁾ より)

この領域で最も報告の多い β_2 アドレナリンレセプターの遺伝的多型は、特に頻度が高い Arg16Gly, Gln27Glu の 2 か所について多くの検討がなされている³⁷⁾。 *in vitro* によるイソプロテノールを用いた検討では、Arg16Gly, Gln27Glu それぞれ単独の変異体においては、レセプターと薬物の親和性やアデニル酸シクラーゼの活性には影響ないが、前者では脱感作の亢進がみられたのに対し、後者では逆にまったくみられなかった³⁸⁾。ヒトにおける変異解析の結果から、Arg16-Gln27, Gly16-Glu27 の間で強い連鎖不平衡がみられ³⁹⁾、一方、Gly16-Glu27 両方の変異を入れたタンパク質は、脱感作の亢進 (Gly16 の性質) がみられた³⁸⁾ことから、*in vitro* 実験においてもハプロタイプを考慮した実験デザインの重要性がうかがえる。薬効との関連では、喘息患者に β_2 刺激薬アルブテロールを経口投与した後、血中濃度は多型により変化しないが、喘息の改善効果の指標となる FEV₁ (forced expiratory volume in the first second) の値の変化は、Arg16/Arg16 が、Gly16/Arg16, Gly16/Gly16 群に比べて有意に大きな改善がみられている (図 4)⁴⁰⁾。ほかに

表1 薬物の効果・副作用に遺伝子多型が関連しうる報告のある遺伝子群

遺伝子名	薬剤のカテゴリー・代表的薬物名	影響がみられる事象
1. 直接薬効ターゲットに関係する遺伝子		
アンジオテンシン変換酵素 (ACE)	ACE阻害薬 (エナラブリル)	血圧の低下、腎保護作用など
β_2 アドレナリンレセプター	β_2 刺激薬 (アルブテロール)	気管支拡張作用、心血管作用など
アラキドン酸5-リポキシゲナーゼ	ロイコトリエン阻害薬	FEV ₁ の改善
ドーパミンレセプター (D ₂ , D ₃ , D ₄)	抗精神病薬 (プロムベリドール)	抗精神病作用
エストロゲンレセプター α	エストロゲン	骨密度上昇、HDL上昇など
グリコプロテインIIIaサブユニット	アスピリン、グリコプロテインIIb/IIIa阻害薬	抗血小板作用
セロトニントランスポーター (5-HT)	抗うつ薬 (クロザビン)	抗うつ作用
アンジオテンシンレセプター (AT1R)	AT1R拮抗薬 (カンデサルタン)	腎血液循環改善作用
スルホニルウレアレセプター (SUR1)	SU系糖尿病薬 (トルバタミド)	インスリン分泌促進作用
ビタミンDレセプター	活性型ビタミンD ₃	くる病改善効果
2. 副作用に関係する遺伝子		
プラジキニンB2レセプター	ACE阻害薬 (エナラブリル)	(ACE阻害薬により誘発される) から咳
ドーパミンレセプター (D ₂ , D ₃ , D ₄)	抗精神病薬 (プロムベリドール)	運動障害、静座不能
HLA	アバカビル (抗HIV薬)	過敏症反応
K ⁺ チャネルおよび関連タンパク質 (HERG, KvLQT1, Mink, MiRP1)	エリスロマイシン、テルフェナジン、シサブリド、クラリスロマイシンなど	薬剤誘導性のtorsade de pointesのリスク上昇
プロトロンビンおよび第V因子	経口避妊薬	血栓症のリスク上昇
過敏症反応	カルバマゼピン (抗痙攣薬)	過敏症反応
3. 間接的に効果に関係する遺伝子 (作用が明らかでないものも含む)		
アンジオテンシン変換酵素 (ACE)	HMG-CoA還元酵素阻害薬	血中LDL減少、動脈硬化進展など
adducin	利尿薬	心筋梗塞の危険因子
アボリボタンパク質E	タクリン (Alzheimer病治療薬)	病状の改善
コレステロールエステル転送タンパク質 (CETP)	HMG-CoA還元酵素阻害薬	生存率上昇
メチルグアニンメチル転移酵素 (MGMT)	カルムスチン (抗腫瘍薬)	動脈硬化進展の遅延
Parkin	デボドバ	gliomaに対する効果
stromelysin-1	HMG-CoA還元酵素阻害薬	Parkinson病の治療効果改善
リボタンパク質リバーゼ	HMG-CoA還元酵素阻害薬	心血管イベントの減少など
インターロイキン6	HMG-CoA還元酵素阻害薬	動脈硬化進展の遅延
Toll-like レセプター4	HMG-CoA還元酵素阻害薬	血中LDL減少
レブチンレセプター	HMG-CoA還元酵素阻害薬	心血管イベントの減少など
		血中コレステロール低下

(Evans WE & McLeod HL: *N Engl J Med* 348, 538-49, 2003³⁴⁾を中心まとめた)

もさまざまな臨床研究がなされているが、同じ指標で評価しているにもかかわらず、結果が互いに相反する事例が多くみられる。その原因の一つとして、多くの研究が単独のSNPを対象としており、対象集団のなかで連鎖する他の変異の影響を考慮していないために、的確な層別化が図れていない可能性が示唆される。実際、77人の13か所の変異についてハプロタイプ解析を行ったところ、わずか12種類に分類され、アルブテロール投与後のFEV₁の変化を層別化

して評価したところ、単独のSNPとはいずれも相関が認められなかったのに対し、ハプロタイプ別の解析では有意な相関がみられたとする報告がある³⁹⁾。

また、先天的QT延長症候群の発症に関与する候補遺伝子の多型に関しては数多くの報告があり、タイプごとに適した治療法が提案されてきたが、一方、多種の薬剤について、薬剤誘導性のQT延長が臨床上重篤な副作用として問題視されている。現に、これが原因で上市されな