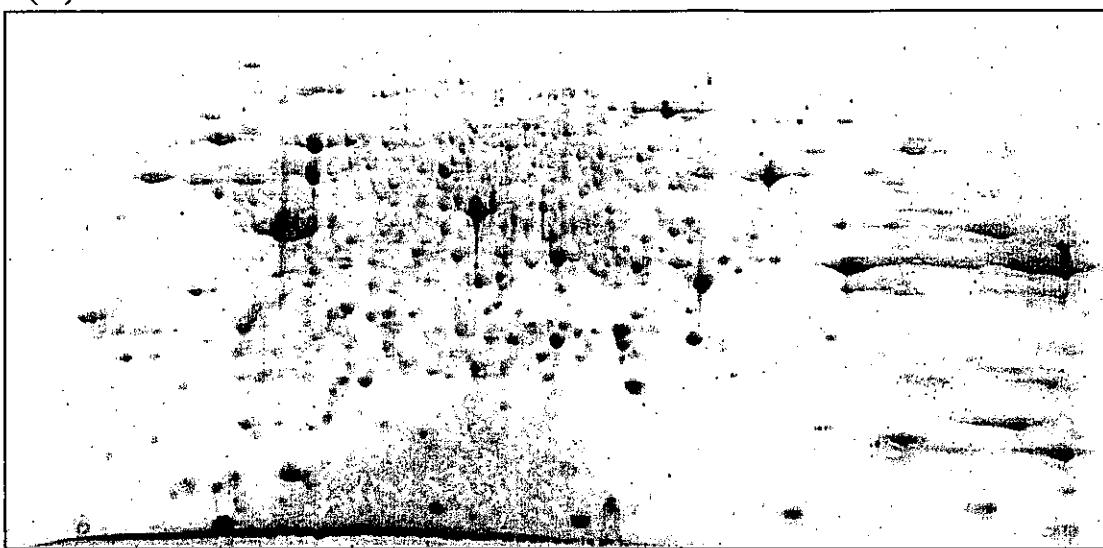


Fig. 19 Peak area ratio of oligosaccharides from CHO and CHOIII membrane

(A) 2D-PAGE



(B) Lectin blot

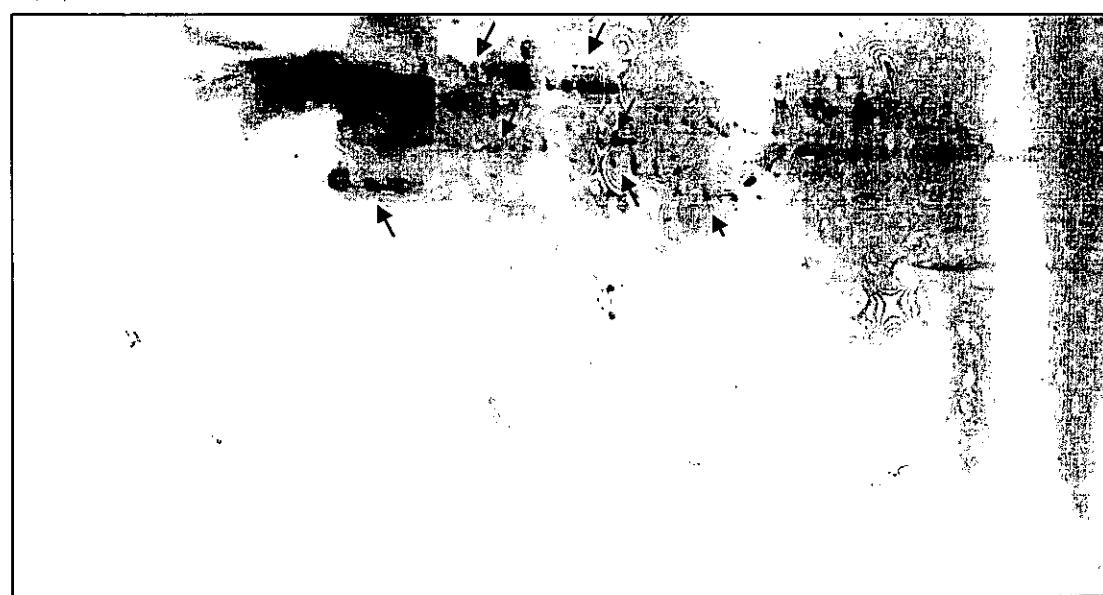
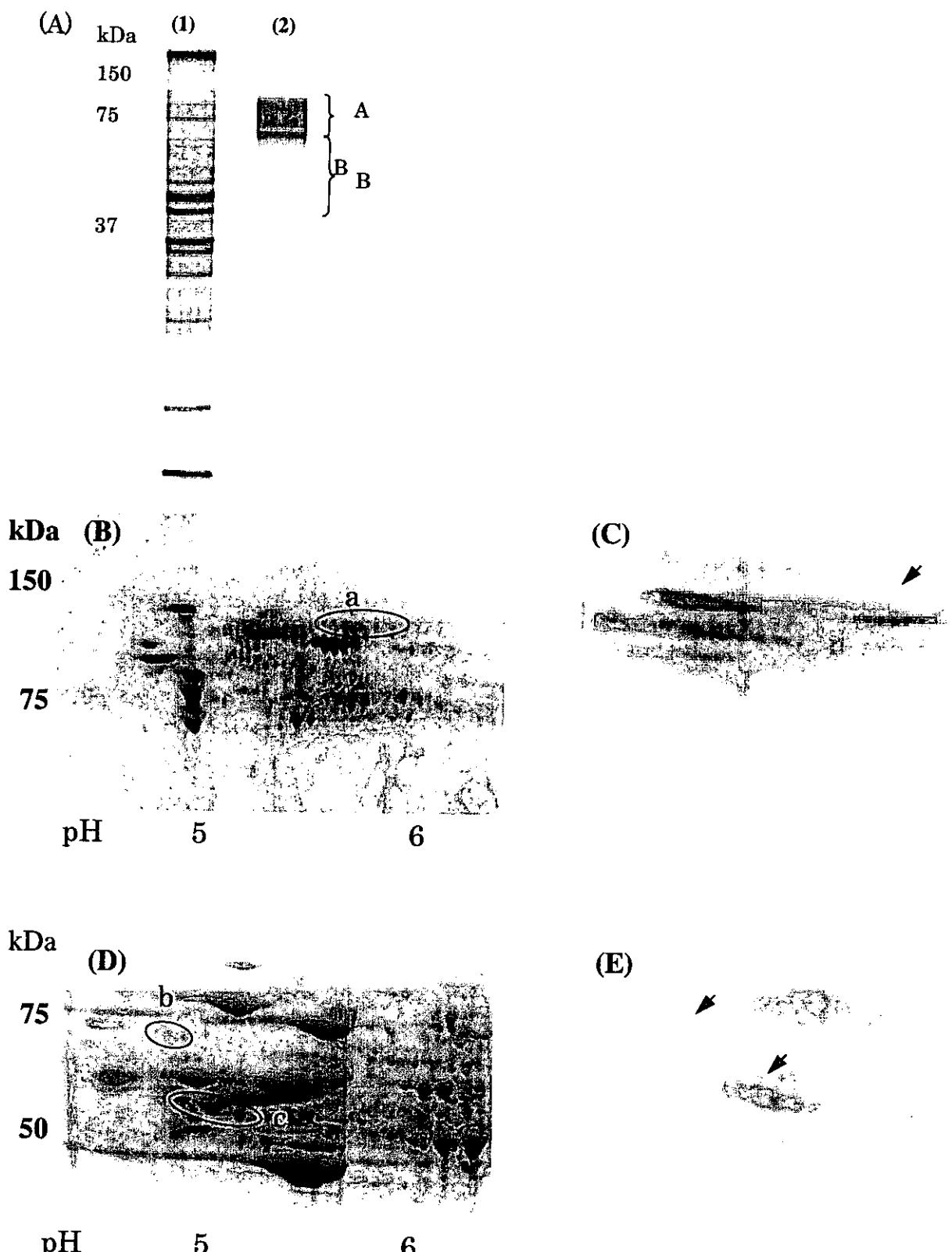


Fig. 20 2D-GE of CHO-III membrane proteins

(A) Sypro Orange, (B) Lectin blot



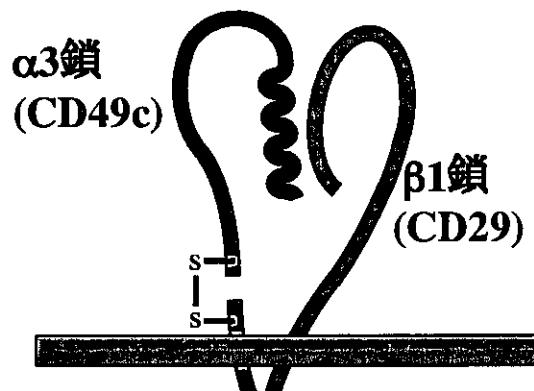
**Fig. 21 (A) SDS-PAGE of CHO-III membrane
 (B) 2D-GE of band A, (c) Lectin blot of Band A
 (D) 2D-GE of band B, (D) Lectin blot of Band B**

分布：上皮細胞

リガンド： ラミニン， フィブロネクチン，
コラーゲン

臨床：腫瘍細胞での発現異常

細胞接着の制御



1 mgpgprcapg dpgwmlgala lmvaasgrfa fafnldtrfl vvkeavnpgs lfgysvalhr
61 qterqqryll lagaprdlsv adgytNrtga vylcpltalk ddcermdise ksdpdhhie
121 dmwlqgvivas qgpagrvlvc ahrytqvrlws gmedqrrmvvg kcyvrqndlq ldpgddwqty
181 hnemcnstid ylqtgmclglg tsggftqntv yfgapgaynw kgnsymiqrk dwdlseysyk
241 gsedqgnlyi gyivqvgsav lhptyitvva gaprhqhmga vfllsqesgg dlkrkqvleg
301 tqvgayfgsa ialadlnndg wqdilvgapy yferkeevgg avyvfmnqag tfpdqpsll
361 lhgpsrsafg isiasigdin qdgfqdaiav apfeglgkv yihsssgll rqpqqivhgd
421 klglpglslsf gyslsgkmdv ddnsypdlyv gslsdhvll rarpvinilq rtivarparv
481 dpslcptisc vqvelcfayN qsagnpsyrr Nitlaytlea drdrpprll farsqsavfh
541 gflsmpehc qtlellmdn vrdklrpivi amNyslplrm pdrlkigmrs ldaypvlnqa
601 qaleNhtevh fqkecgpdnk cdsnlqmraa fvseqlqpls rlqysrdtkk lflsiNvint
661 psrerageda healltlevp pallssvrp sgtcqaNeti lcelgnpfkr nqrmelliaf
721 evigvtlhtr dlkaqlqlst sshqdnlpqm tlilqvdytl qaslslmthr lqsffggivm
781 geagmktved vgsplkyefq vspvgdglaa lgtivlglew pyevingkwI lyptcihihs
841 Neswpcqppg nlvnplnlil sdpgdkphsp qrrrr
901 vltcasgrar cvwlecpipd tsNvtNvtvk arvwNstfie dyrdfdrvrv dgwatlflrt
961 siptinmeNk ttwfsvdids elveelpaei elwlvlvavs agllllgllii illwkecffk
1021 ptryrimpk yhavriri eee rypppgstlp tkkhwtswq irdryy

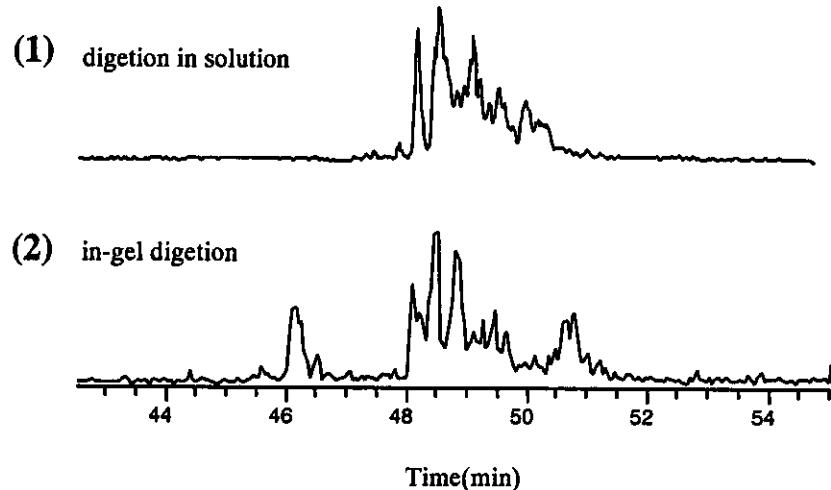
Heavy chain
(33~872)

Light chain
(876~1066)

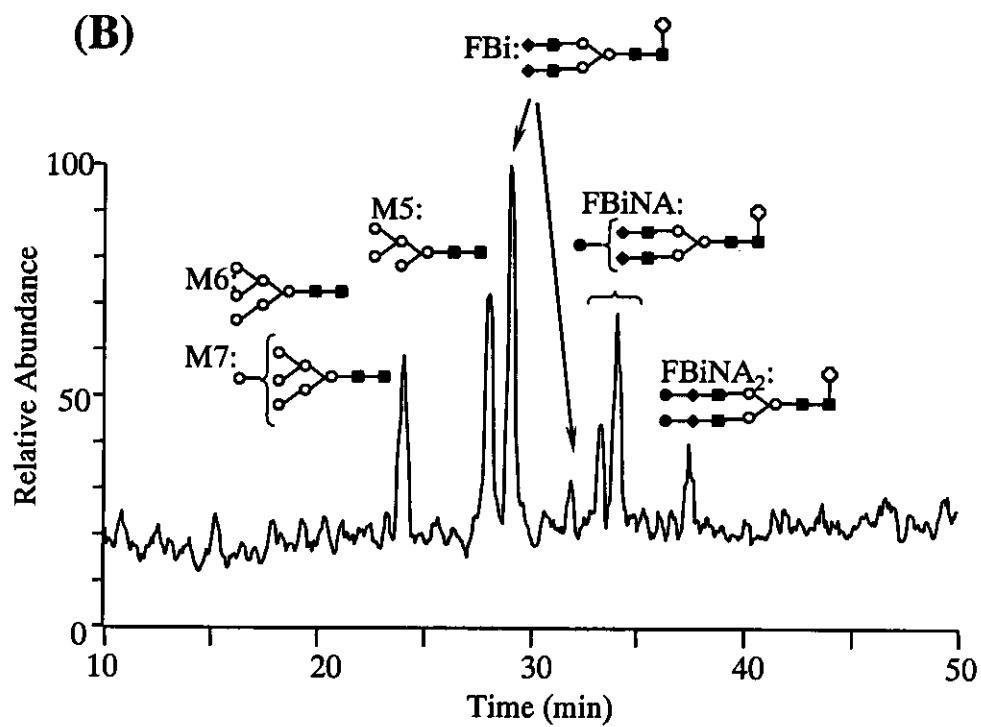
Under line : Calcium binding region, N: Glycosylation site

Fig. 22 Amino acid sequence and N-linked glycosylation sites of Integrin α-3 (CD49c)

(A)



(B)



**Fig. 23 Oligosaccharide profiling of gel separated proteins
(A) Erythropoietin, (B) t-PA**

Rat brain

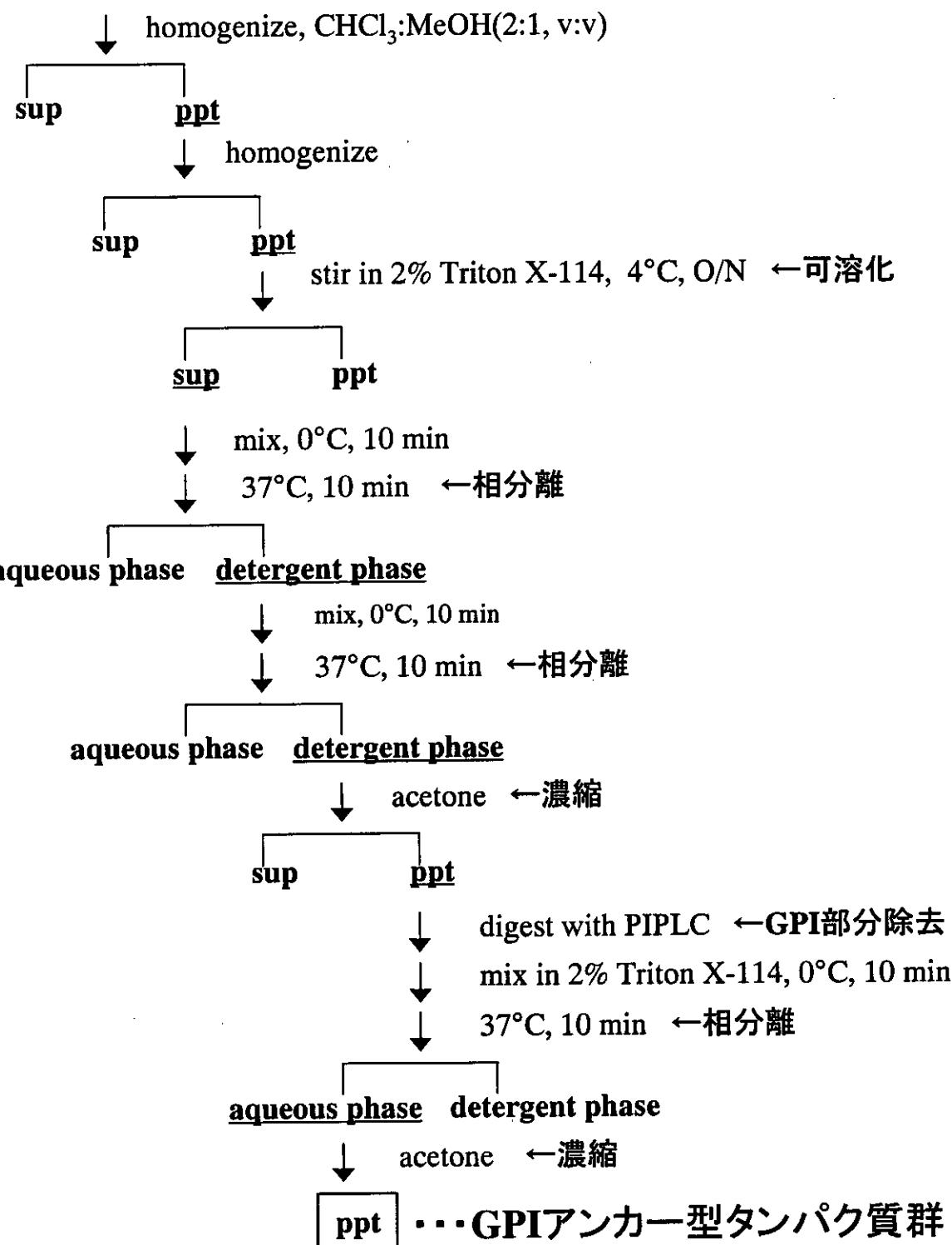


Fig. 24 Preparation of lipid-free GPI-anchored proteins from rat brain

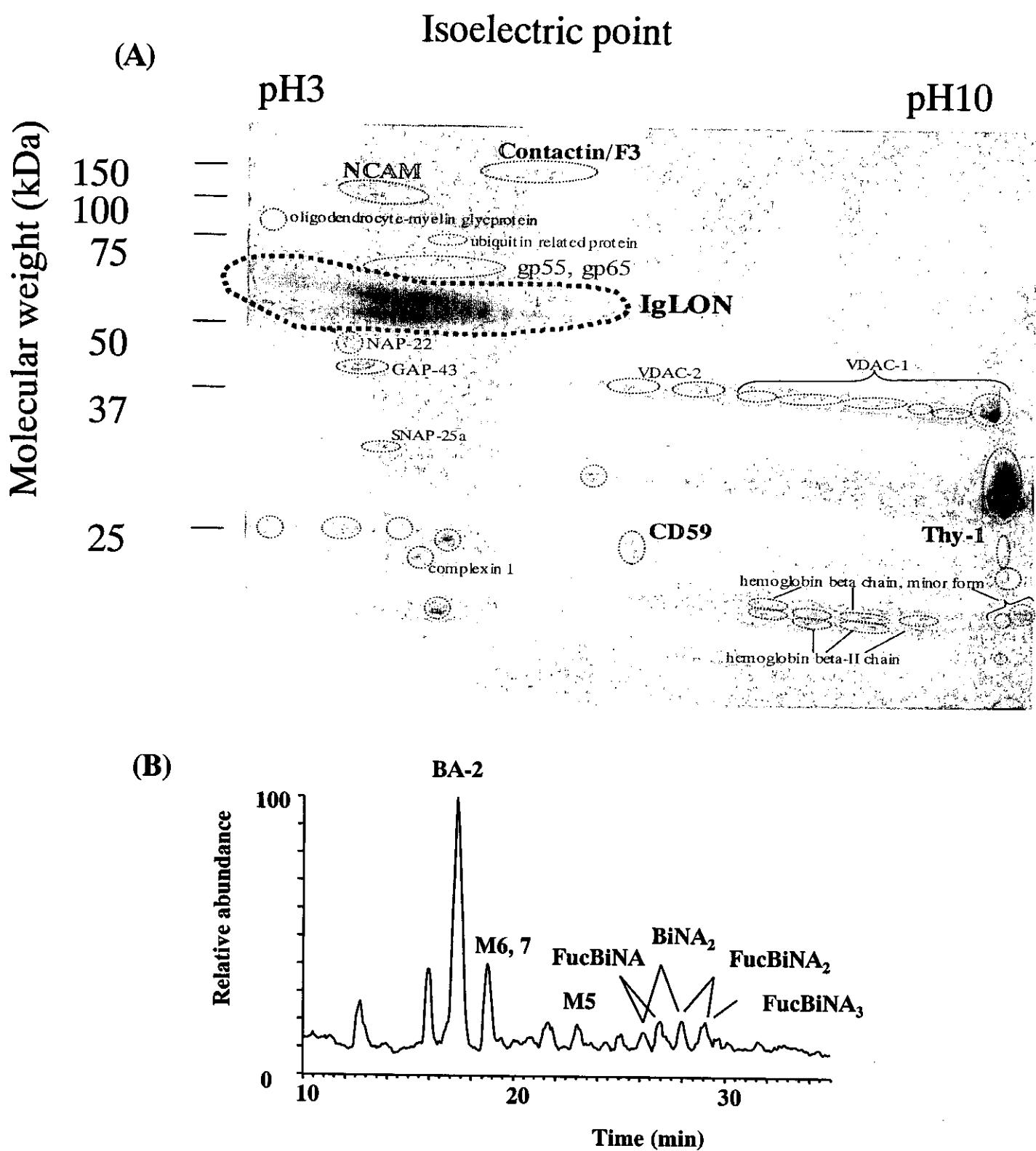


Fig. 25 (A)2D-GE of lipid-free GPI-anchored proteins from 3-week old rat brain, (B) Oligosaccharide profiling of gel separated Kilon

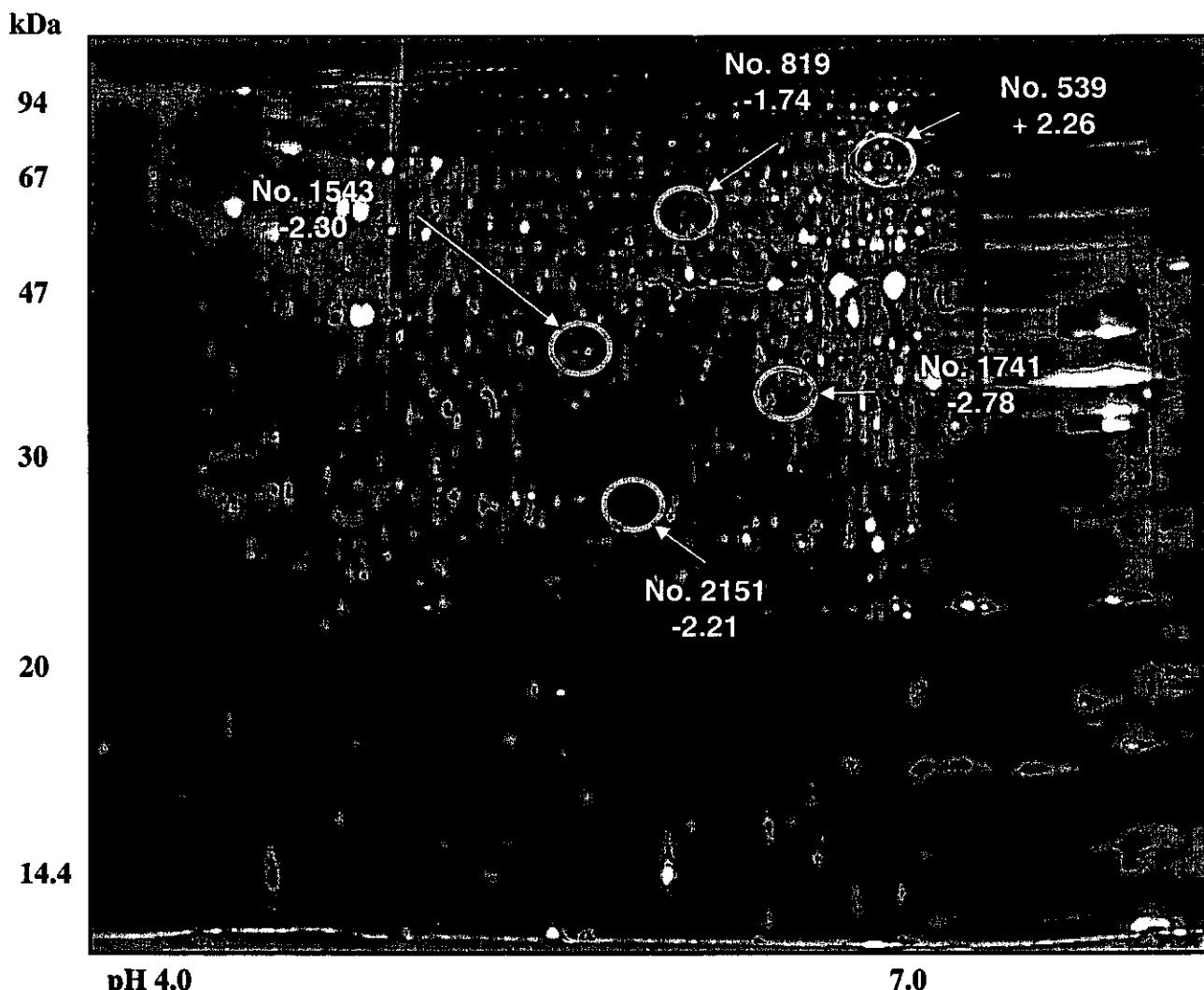


Fig.5 2D-DIGE images of Cy3 labeled HepG2 (A) and Cy5 labeled HepG2III (B) membrane fraction.

Fig. 26 2D-DIGE images of Cy3 labeled HepG2 and Cy5 labeled HepG2III membrane fraction

Table 4 Glycosylation analysis of alpha-fetoprotein

Retention time (min)	<i>m/z</i>	Charge	M.W.	Peptide Sequences	M.W.	Oligosaccharide Structures	M.W.
23	1013.43	+3	3037.3	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	2077.8
23	1519.67	+2	3037.3	k/vnftfeiqk/l	977.5	[HexNAc]5[Hex]5[Neu5Ac]1[Fuc]1	2281.4
23	1081.29	+3	3240.9	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	1931.8
23	1621.44	+2	3240.9	k/vnftfeiqk/l	977.5	[HexNAc]3[Hex]4[Neu5Ac]1	1275.9
23	1446.62	+2	2891.2	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1	1931.8
23	1118.67	+2	2235.3	k/vnftfeiqk/l	977.5	[HexNAc]3[Hex]4	
23	1264.05	2	2526.1	k/vnftfeiqk/l	977.5	[HexNAc]3[Hex]4[Neu5Ac]1	1566.6
24	1110.48	3	2526.1	k/vnftfeiqk/l	977.5	[HexNAc]3[Hex]4[Neu5Ac]1	1566.6
24	1665.44	+2	3328.9	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2369.4
24	1178.33	+3	3532.0	k/vnftfeiqk/l	977.5	[HexNAc]5[Hex]5[Neu5Ac]2[Fuc]1	2572.5
24	1061.78	3	3182.3	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2	2222.9
25	1592.17	2	3182.3	k/vnftfeiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2	2222.9
27	1236.04	+3	3705.1	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2369.4
27	1853.29	+2	3704.6	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2369.4
27	1187.17	+3	3558.5	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2	2222.8
28	1780.23	+2	3558.5	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2	2222.8
28	1303.56	+3	3907.7	k/vnftfeiqk/l	1353.7	[HexNAc]5[Hex]5[Neu5Ac]2[Fuc]1	2572.0
28	1138.84	+3	3413.5	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	2077.8
28	1206.52	+3	3616.6	k/vnftfeiqk/l	1353.7	[HexNAc]5[Hex]5[Neu5Ac]1[Fuc]1	2280.9
28	1090.14	+3	3267.4	k/vnftfeiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]1	1931.7
28	1322.75	+3	3965.3	k/vnftfeiqk/l	1353.7	[HexNAc]6[Hex]5[Neu5Ac]2	2629.6

Table 5 Glycosylation analysis of ceruloplasmin

Retention time (min)	m/z	Charge	M.W.	Peptide Sequences	M.W.	Oligosaccharide Structures	M.W.
24	1025.17	+4	4096.7	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]4[Hex]5[Neu5Ac]2	2222.9
24	1366.54	+3	4096.6	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]4[Hex]5[Neu5Ac]2+Na	2244.9
24	1373.88	+3	4118.6	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2368.9
24	1061.67	+4	4242.7	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]5[Neu5Ac]3[Fuc]2	3171.2
25	1415.23	+3	4242.7	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]5[Neu5Ac]3[Fuc]3	3025.1
25	1262.24	+4	5045.0	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3	2879.1
25	1682.65	+3	5045.0	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]1	3025.1
25	1189.21	+4	4752.8	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]2	3025.1
25	1585.27	+3	4752.8	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]3	3025.1
25	1225.72	+4	4898.9	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]1	3025.1
25	1633.96	+3	4898.9	EHEGAIYPDNTTDFQR	1891.8	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]2	3025.1
27	1093.95	+4	4371.8	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2368.8
27	1458.27	+3	4371.8	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]4[Hex]5[Neu5Ac]2	2222.7
27	1057.43	+4	4225.7	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]1	3025.0
27	1409.57	+3	4225.7	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]5[Hex]6[Neu5Ac]3	2878.9
28	1257.99	+4	5028.0	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]4[Hex]5[Neu5Ac]2	2222.7
28	1677.01	+3	5028.0	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2368.8
28	1221.48	+4	4881.9	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]2	2222.7
28	1628.30	+3	4881.9	ELHHLQEQNVSNAFLDK	2021.0	[HexNAc]5[Hex]6[Neu5Ac]3	3025.0
33	1347.54	+3	4039.6	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]1	1931.6
34	1450.24	+3	4347.7	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]2+NH3	2239.7
34	1083.68	+4	4330.7	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]2	2222.7
34	1444.56	+3	4330.7	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2368.8
34	1493.28	+3	4476.8	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1[Fuc]1	2244.7
34	1089.17	+4	4352.7	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]4[Hex]5[Neu5Ac]2+Na	2895.9
35	1668.97	+3	5003.9	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]5[Hex]6[Neu5Ac]3+NH3	2878.9
35	1663.29	+3	4986.9	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]5[Hex]6[Neu5Ac]3	2878.9
35	1247.72	+4	4986.9	ENLTAPGSDSAFFEQGTR	2126.0	[HexNAc]5[Hex]6[Neu5Ac]3[Fuc]1	2968.8
37	1282.48	+3	3844.4	AGLQAFFQQVQECNK	1639.7	[HexNAc]4[Hex]5[Neu5Ac]2	2222.7
37	1923.24	+2	3844.5	AGLQAFFQQVQECNK	1639.7	[HexNAc]4[Hex]5[Neu5Ac]2+Na	2368.8
38	1331.17	+3	3990.5	AGLQAFFQQVQECNK	1639.7	[HexNAc]4[Hex]5[Neu5Ac]2+Na	2244.7
38	1289.81	+3	3866.4	AGLQAFFQQVQECNK	1639.7	[HexNAc]4[Hex]5[Neu5Ac]2+Na	2244.7

Table 6 Ad vectors used in this study

Ad vector	Foreign gene					
	E1 deletion region		E3 deletion region		Region between E4 and 3'ITR	
	Promoter	GOI	Promoter	GOI	Promoter	GOI
AdOn-L4	TRE/CMV	Luciferase	CMV	rtTA	—	—
AdBI-rtTA-L	TRE/CMV	Luciferase+rtTA	—	—	—	—
Ad-rtTA-IRES-tTS-L	TRE/CMV	Luciferase	CMV	rtTA+tTS	—	—
Ad-tTS-BI-rtTA-L	TRE/CMV	Luciferase+rtTA	—	—	EF-1 α	tTS
Ad-rtTA-tTS-L	TRE/CMV	Luciferase	CMV	rtTA	EF-1 α	tTS
Ad-L2	CMV	Luciferase	—	—	—	—
AdOn-SEAP4	TRE/CMV	SEAP	CMV	rtTA	—	—
Ad-M2-SEAP4	TRE/CMV	SEAP	CMV	rtTA2 ^S -M2	—	—
Ad-S2-SEAP4	TRE/CMV	SEAP	CMV	rtTA2 ^S -S2	—	—
Ad-rtTA-tTS-SEAP	TRE/CMV	SEAP	CMV	rtTA	EF-1 α	tTS
Ad-M2-tTS-SEAP	TRE/CMV	SEAP	CMV	rtTA2 ^S -M2	EF-1 α	tTS
Ad-S2-tTS-SEAP	TRE/CMV	SEAP	CMV	rtTA2 ^S -S2	EF-1 α	tTS
Ad-SEAP2	CMV	SEAP	—	—	—	—

CMV = CMV intermediate-early promoter / enhancer

TRE/CMV = tet-responsive promoter containing minimal CMV promoter

EF-1 α = human elongation factor-1 α promoter

Ad-rtTA-IRES-tTS-L contains CMV promoter / Intron A / rtTA / IRES / tTS / P(A) cassette in the E3 deletion region.

AdBI-rtTA-L and Ad-tTS-BI-rtTA-L express luciferase and rtTA from one bidirectional tet-responsive promoter cloned in the E1 deletion region.

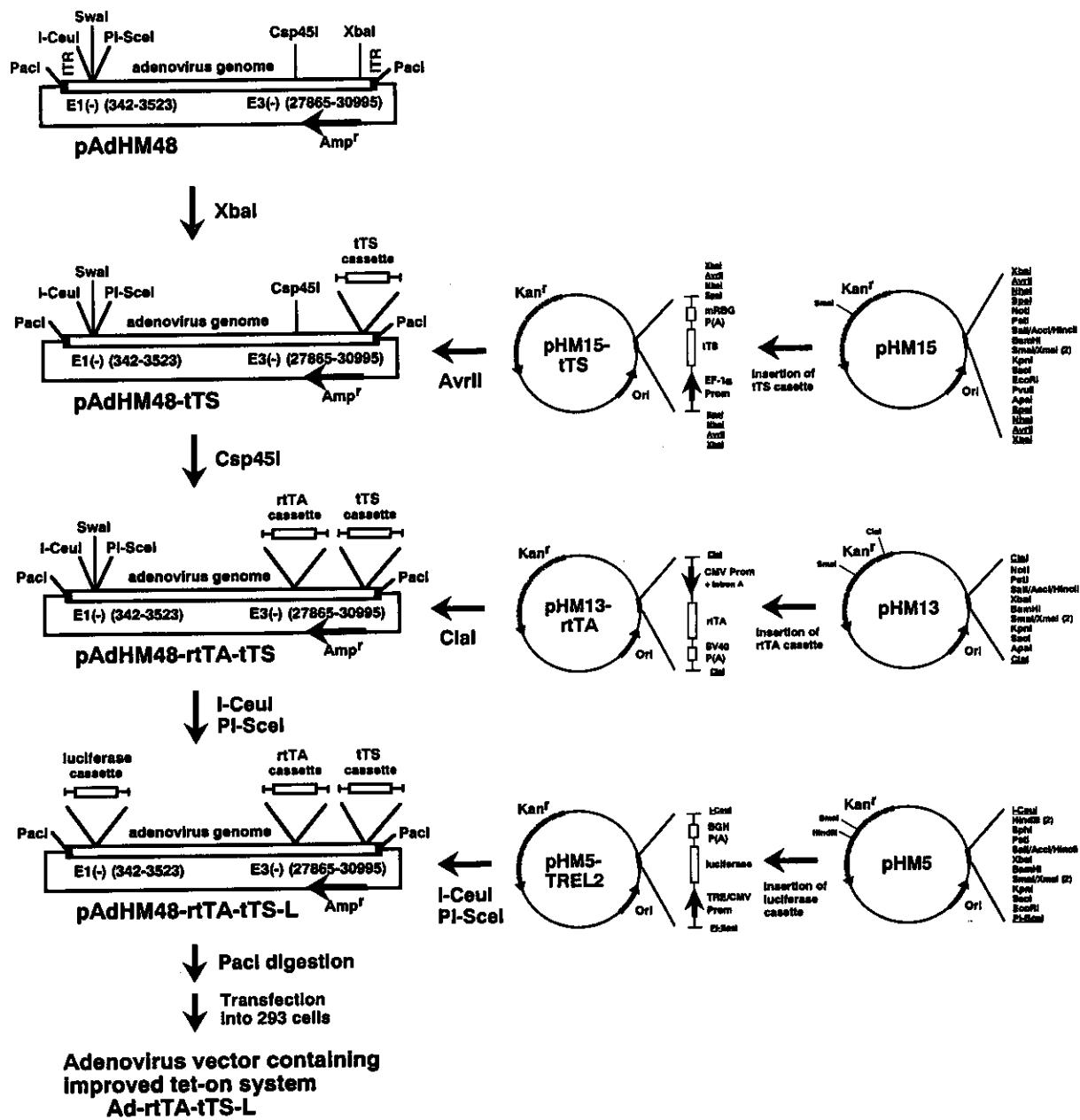


Fig.27 The construction strategy of Ad vectors containing the triple gene expression system.

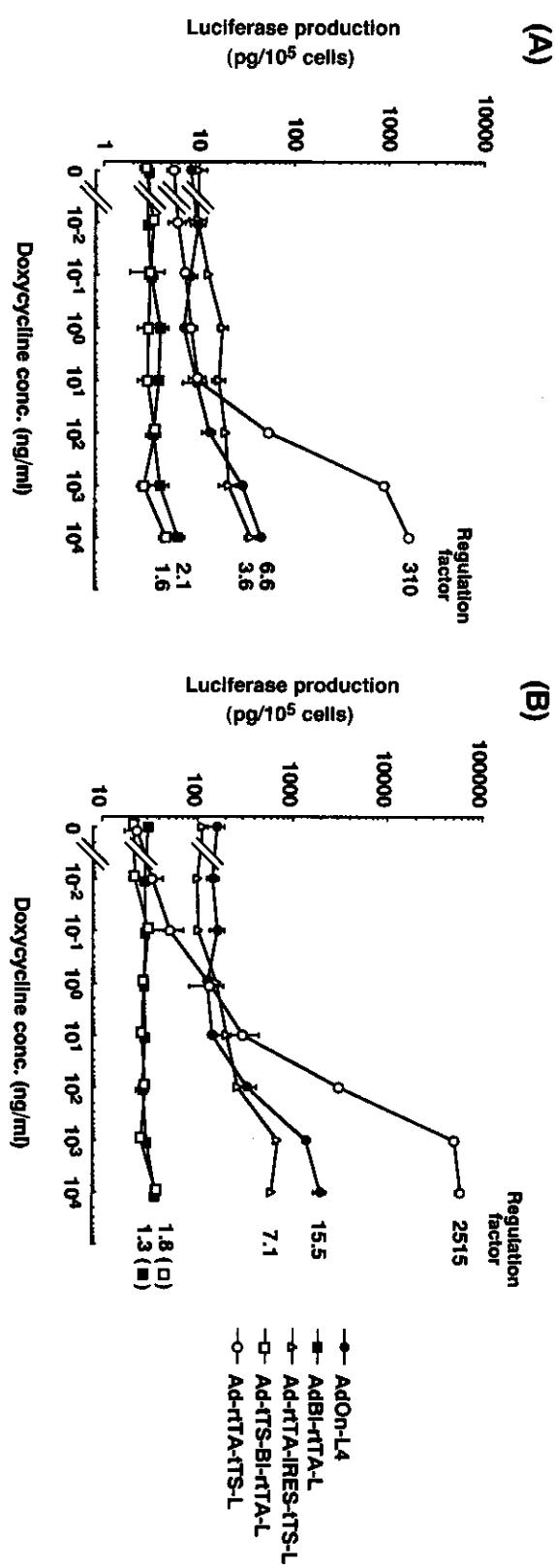


Fig.28 Regulated luciferase expression In SK HEP-1 cells transduced with various Ad-mediated tet-on systems.

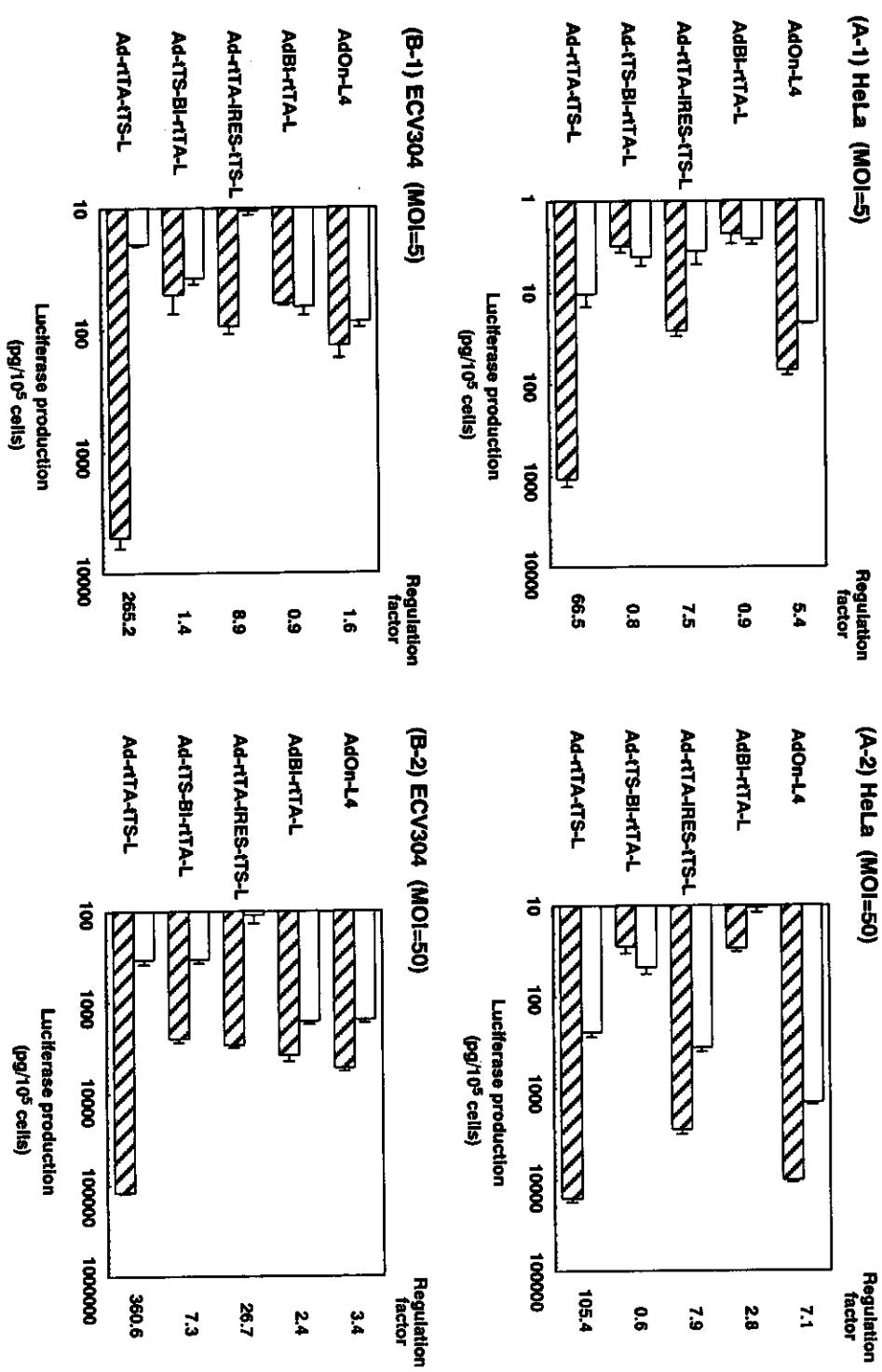
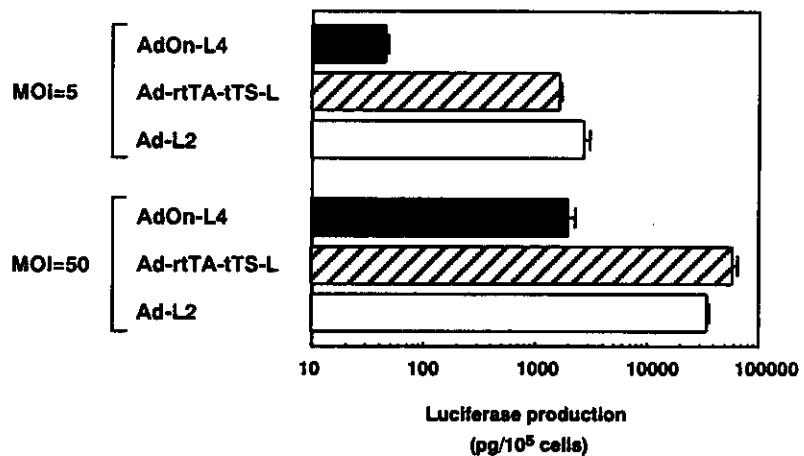
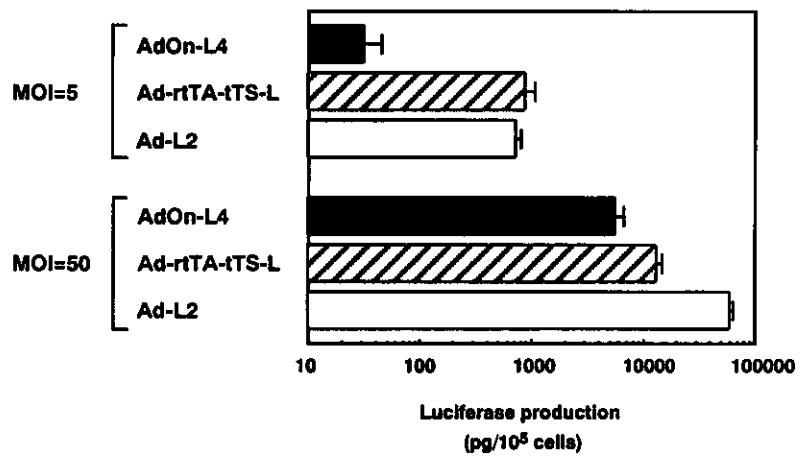


Fig.29 Regulated luciferase expression in HeLa and ECV304 cells transduced with various Ad-mediated tet-on systems.

(A) SK HEP-1



(B) HeLa



(C) ECV304

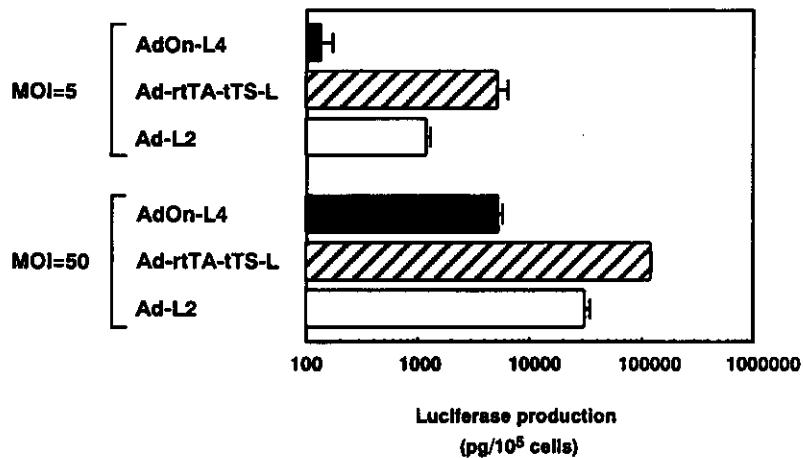


Fig.30 Comparison of induced (maximum) luciferase production in cells transduced with AdOn-L4, Ad-rtTA-tTS-L, or Ad-L2.

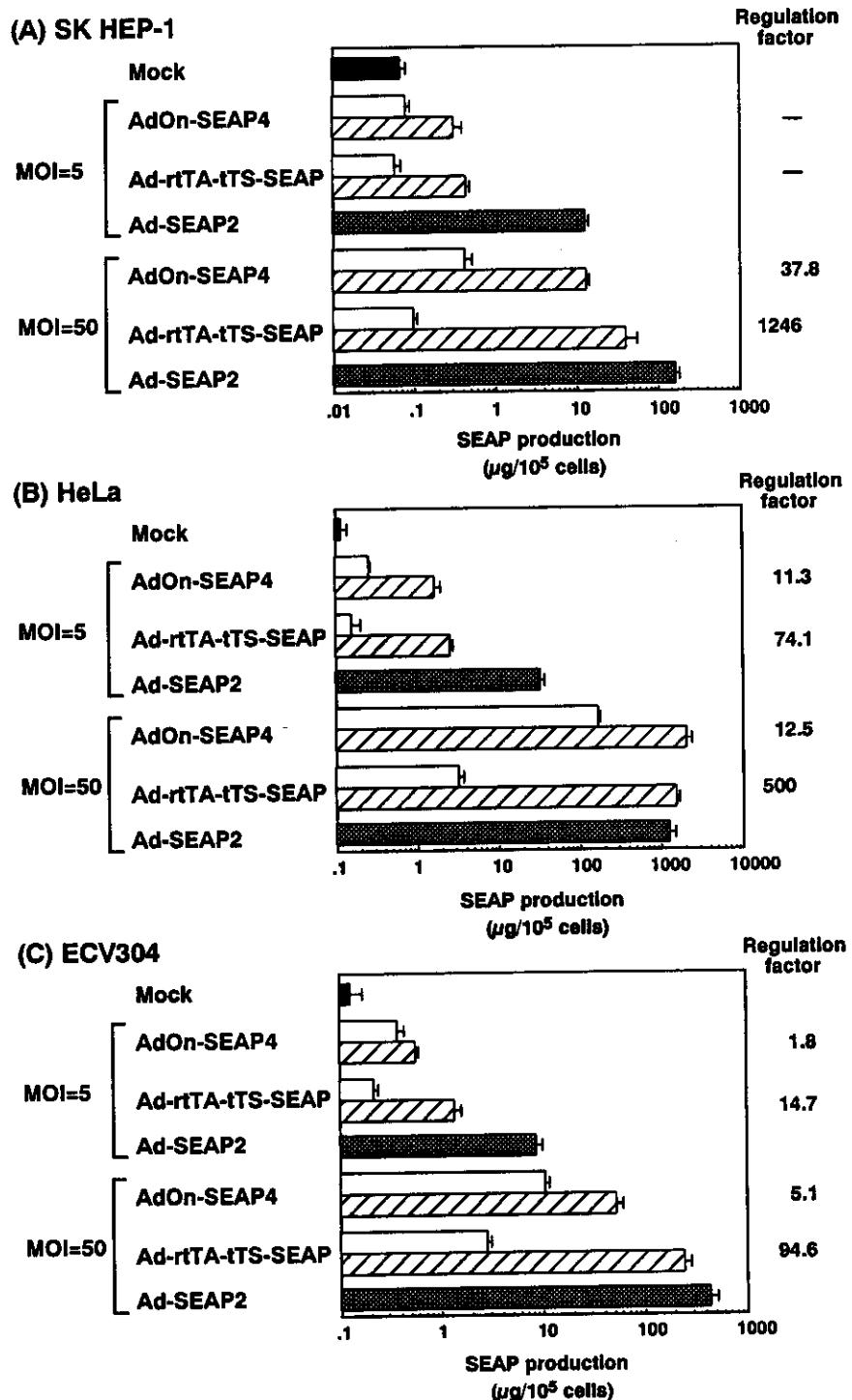


Fig.31 Regulated SEAP expression in SK HEP-1, HeLa, and ECV304 cells transduced with AdOn-SEAP4 or Ad-rtTA-tTS-SEAP.

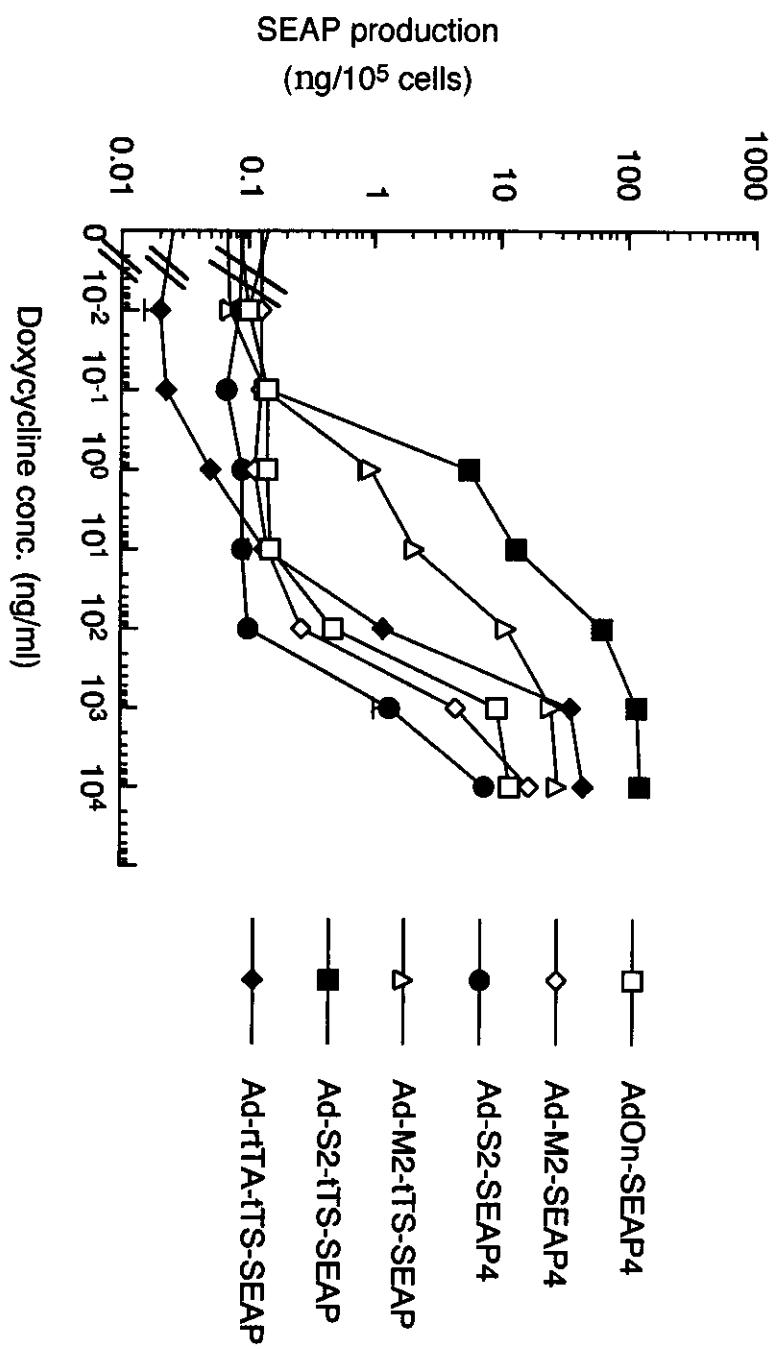


Fig.32 Regulated SEAP expression in SK HEP-1 cells transduced with various Ad-mediated tet-on systems.

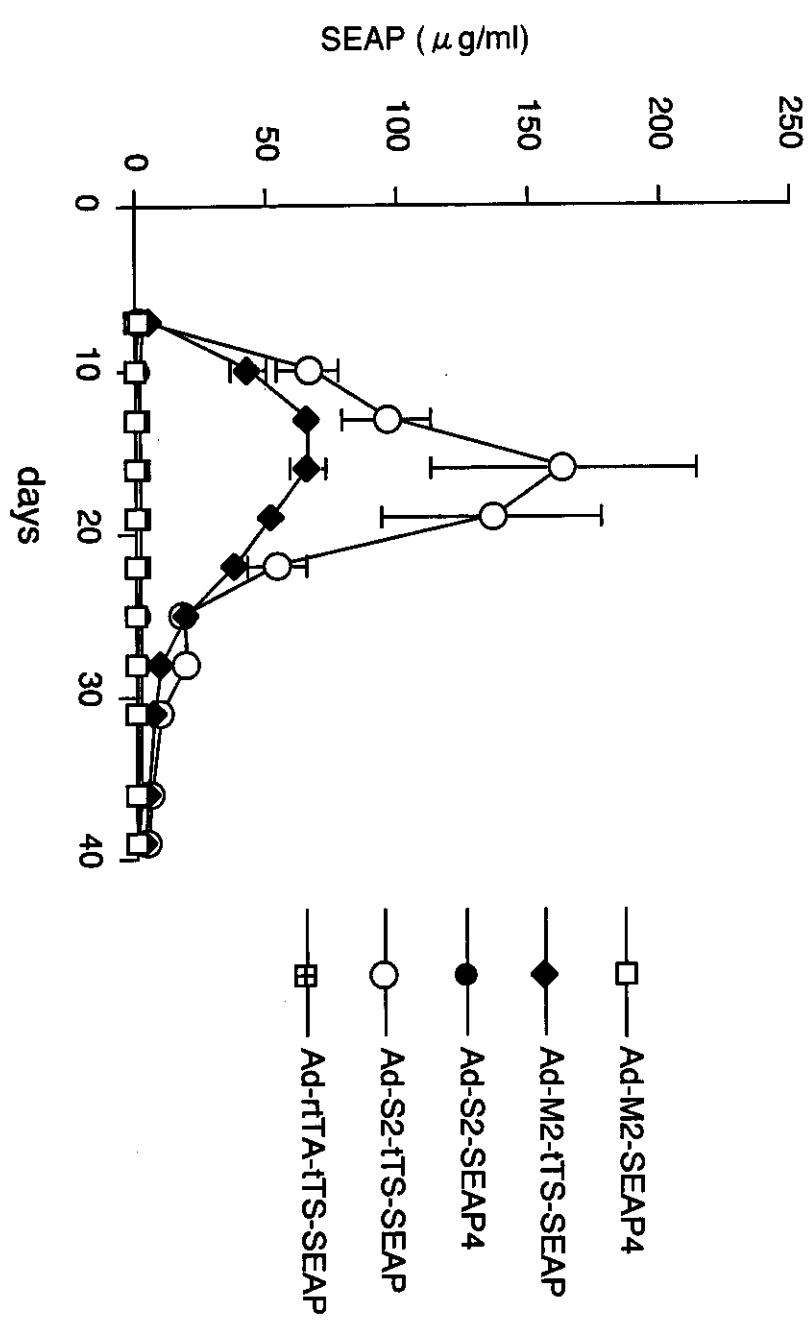


Fig. 33 Regulated SEAP expression in Balb/c nude mice.

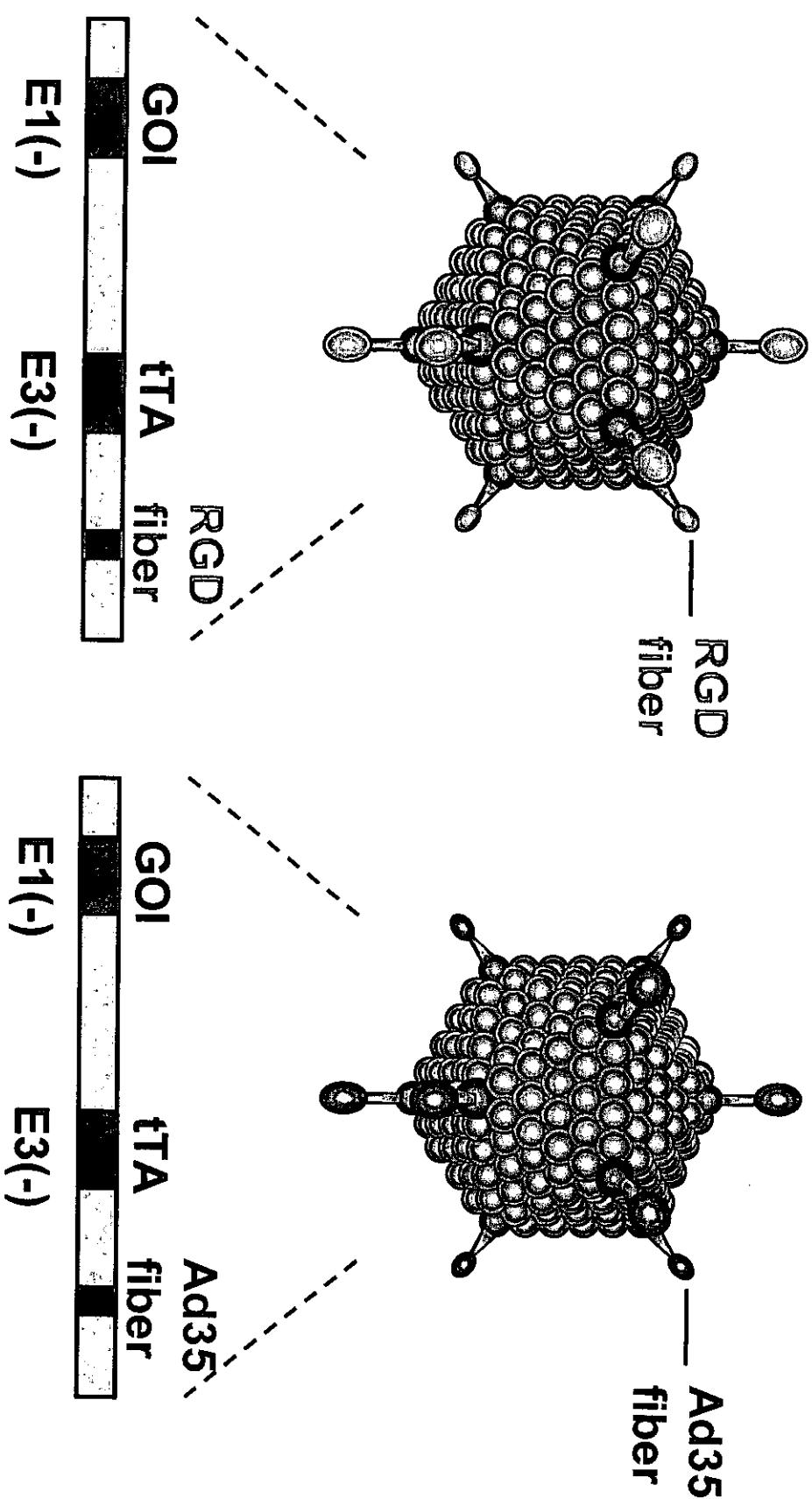
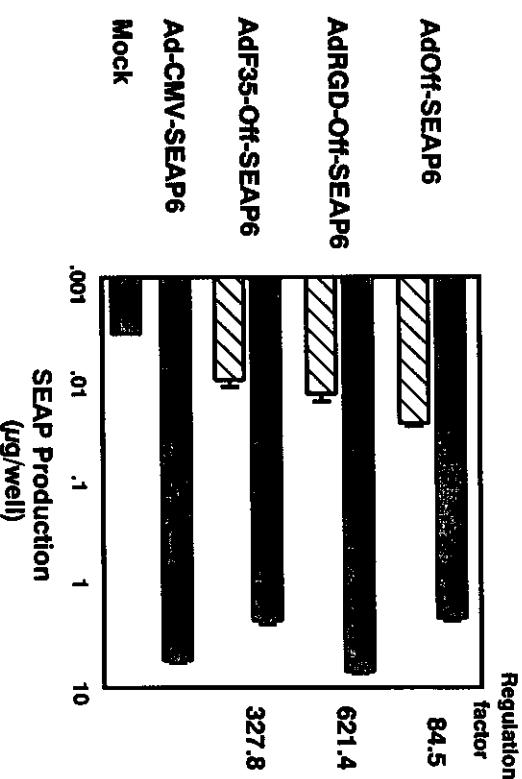


Fig.34 Capsid-modified single adenovirus vectors containing tet-off system

(A) HeLa; CAR(+), av integrin(+), CD46(-)



(C) NIH3T3; CAR(-), av integrin(+), CD46(-)

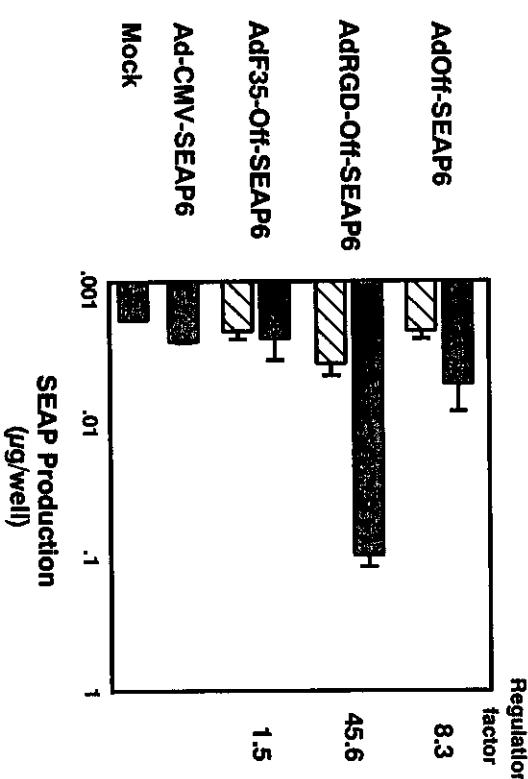


Fig. 35 Capsid-modified single adenovirus vectors containing tet-off system