

図1 HSV-1 及び sindbis virus の PEI 磁気ビーズによる濃縮

表5. PEIの分子量の違いによる濃縮効率の差異
(copy number $\times 10^4$)

	MW70,000	MW 1,800
-serum	4.26 \pm 0.48	1.95 \pm 0.67
+serum	3.46 \pm 0.24	1.0 \pm 0.1

5×10^4 コピー/mlのウイルス液1mlをEPI磁気ビーズに結合させた。

図2. Effect of pH on the concentration of HSV-1 by PEI-beads

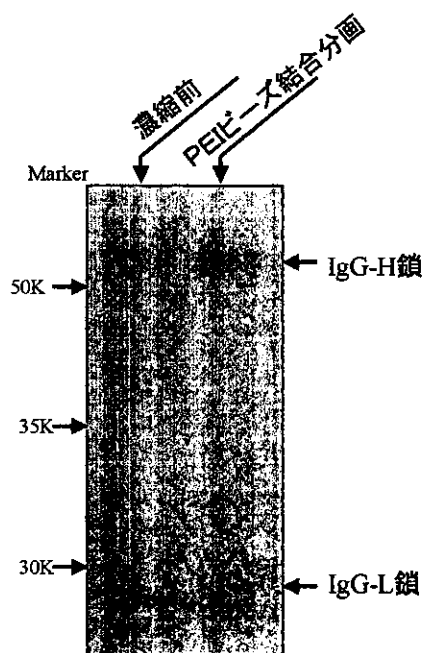
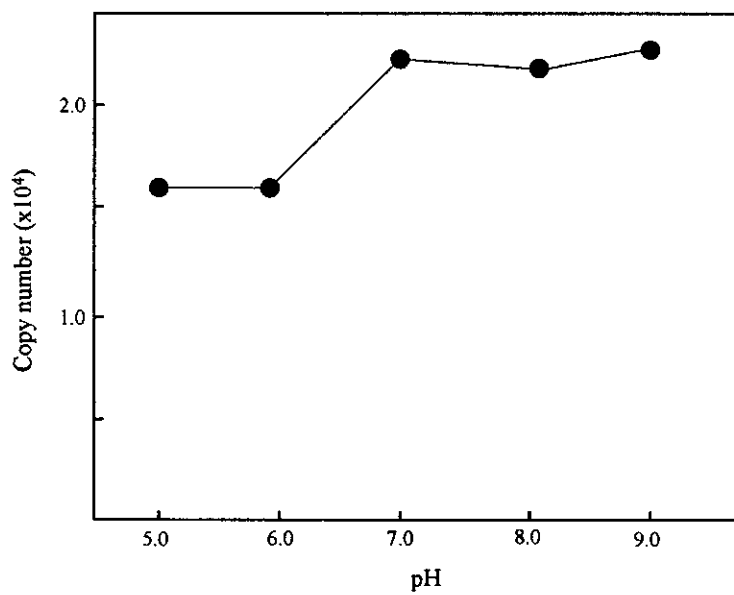


図3. PEI磁気ビーズによるIgGの濃縮

図4. 抗HSV-1抗体のPEI磁気ビーズによるウイルス濃縮への影響

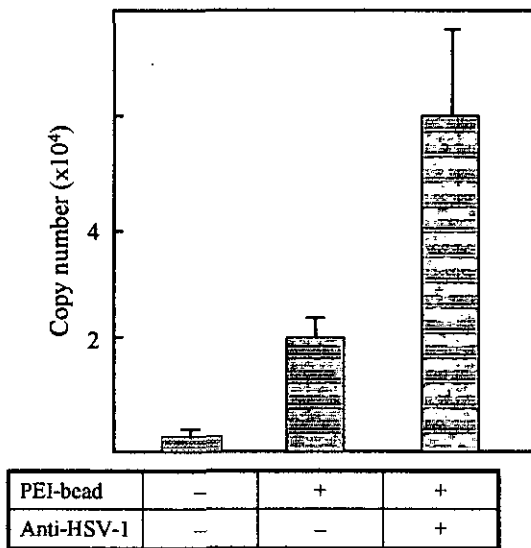


図5. PEI磁気ビーズによるHBVの濃縮と抗HBV抗体の影響

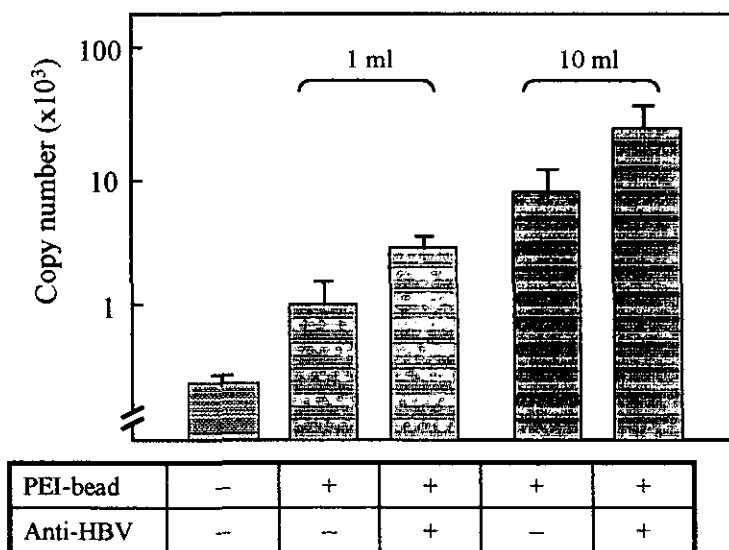


図6. PEI磁気ビーズによる HCV の濃縮

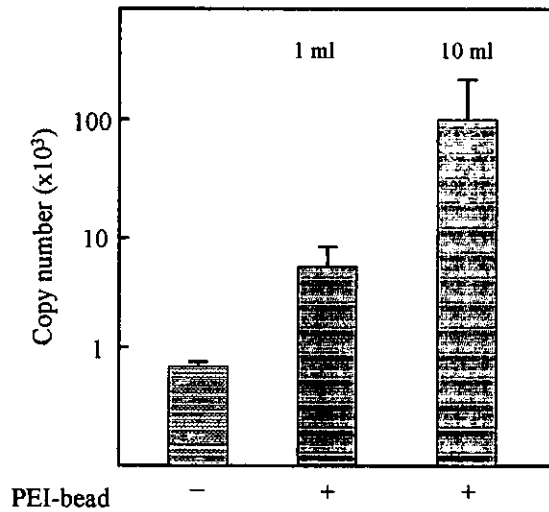


表6 PowerPlex16を用いた各種細胞株のSTRマーカーリポート数の解析結果

A. The fluorescein-labeled allele designations

	D3S1358	TH01	D21S11	D18S51	PentaE
HL60-1	16	7,8	29**, 30	14, 15**	13, 14
HL60-2	16	7,8	29**, 30	14, 15**	13, 14
HL60-RG-1	16	8	29**, 30	14, 15	13, 14
FLC-4	15**, 16	6**, 8*, 9	31.2, 32.2	13	12
WTK1	16	8, 9.3	29	11, 15	5, 7
TK6	16	8, 9.3	29	11, 16	5, 7

* smaller peak than **
 ** a half of allele peak

B. The JOE-labeled allele designations

	D5S818	D13S317	D7S820	D16S539	CSF1PO	Penta D
HL60-1	12, 13	8**, 11	11, 12	11	13, 14	10, 12
HL60-2	12, 13	8**, 11	11, 12	11	13, 14	10, 12
HL60-RG-1	12, 13	8**, 11	11, 12	11	13, 14	10, 12
FLC-4	9**, 13	11*, 12	10*, 12	12**, 13	11, 12*	9, 12*
WTK1	12, 13	11	9, 12	11, 12	11, 13	11, 12
TK6	12, 13	11	9, 11	11, 12	11, 12	11, 12

* smaller peak than **
 ** a half of allele peak
 JOE = 6-carboxy-4',5'-dichloro-2',7'-dimethoxyfluorescein

C. The TMR-labeled allele designations

	Amelogenin	vWA	D8S1179	TPOX	FGA
HL60-1	X	16	12, 13	8, 11	22, 24
HL60-2	X	16	12, 13	8, 11	22, 24
HL60-RG-1	X	16	12, 13	8, 11	21*, 22, 24
FLC-4	X	14, 16**, 17	17	8, 11	22
WTK1	X, Y	16**, 17, 20	10, 13	8, 11	22, 24
TK6	X, Y	16**, 17, 20	10, 13	8, 11	22, 24

* smaller peak than **
 ** a half of allele peak
 TMR = carboxy-tetramethylrhodamine

表5 17番染色体上のSTRマーカーを用いた解析結果

STRマーカー	label	サイズ範囲		TK6		HL60/HL60-RG	
		min	max	allele-1	allele-2	allele-1	allele-2
115B3	6FAM	179	209	189-190	198-200	189	-
THRA1	6FAM	158	176	165-166	173-176	165	169
CHLC	HEX	200		195	199	196	200
AFM248yg9	TET	143	155	143	147	148	154
42D6	6FAM	154	174	138	155	138	-
AFM234xc9	HEX	114	138	109	130	116	-
AFM107yb8	TET	154	170	151-153	157-161	156	164
AFM049xc1	TET	181	207	183-185	187-189	-	192
AFM210xa5	HEX	166	188	172	180	-	-
AFM044xg3	6FAM	226	238	225	235	236	238 (RG 237)

- : no peak

(表中の数字はバンドサイズ)

図7 STRマーカーを使ったDNA多型の検出法

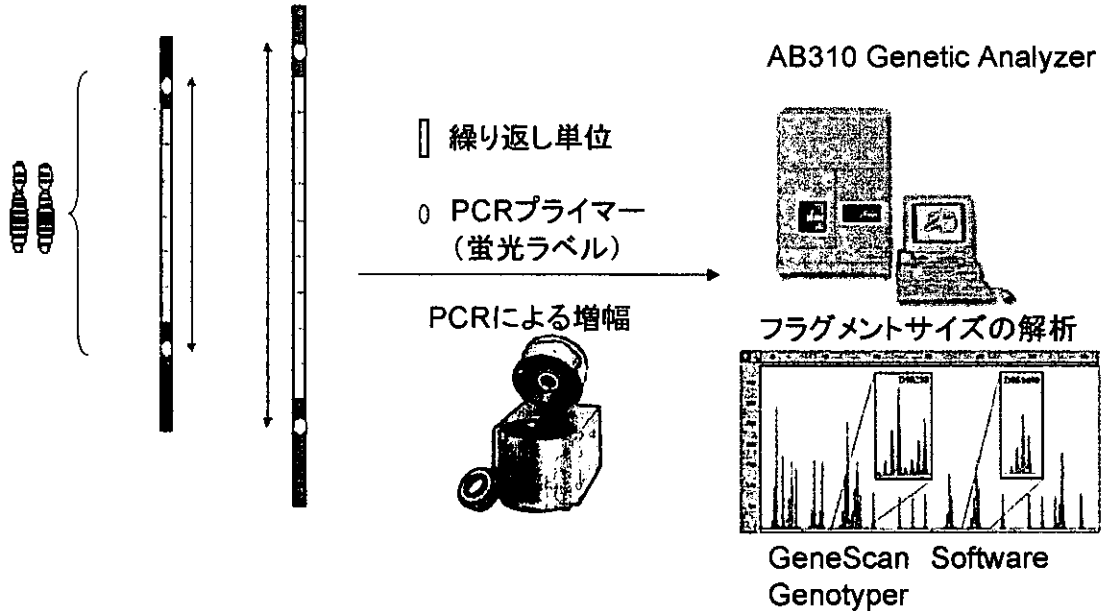


図8 STRマーカーク解析例

PowerPlex16 (Fluoresceine)

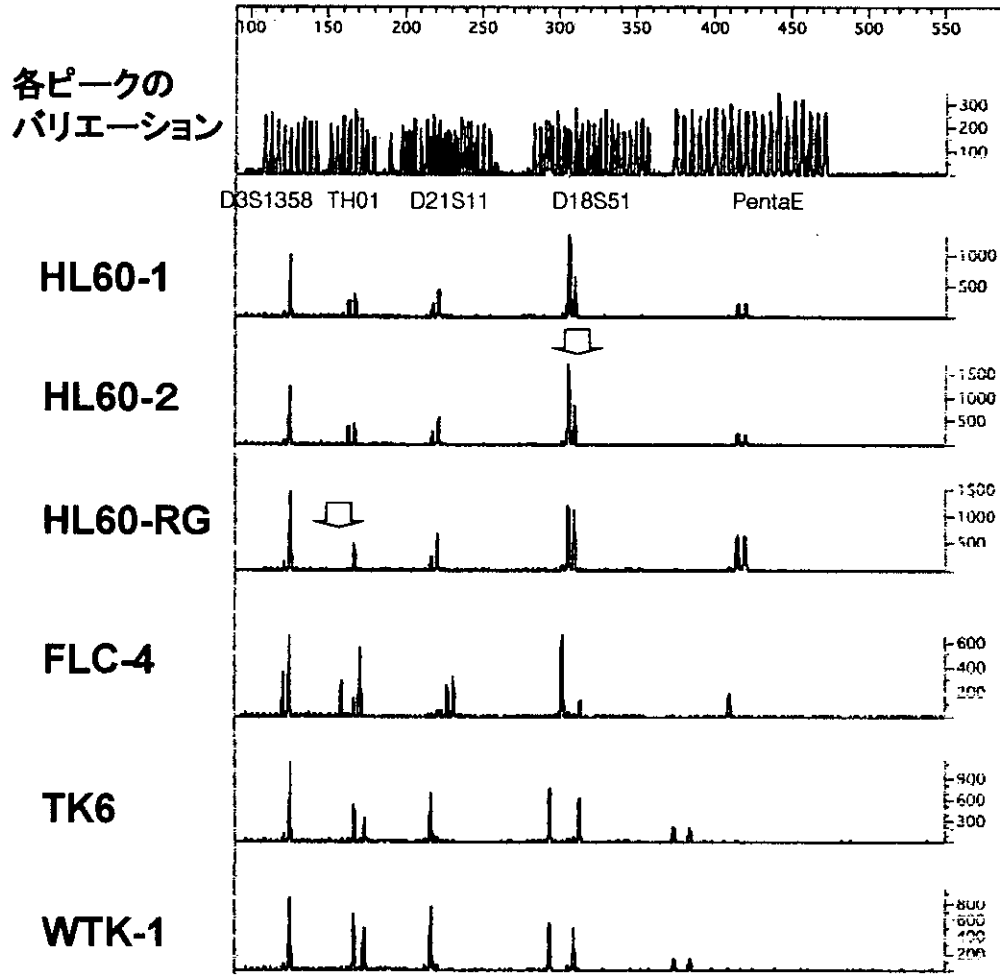


表 8. アレイ上のサイトカインの位置と名称

position	cytokines	
1A	Pos	positive control
1B	Pos	positive control
1C	Pos	positive control
1D	Pos	positive control
1E	Neg	negative control
1F	Neg	negative control
1G	ENA-78	Epithelial neutrophil-activating protein 78
1H	GCSF	Granulocyte-colony stimulating factor
1I	GM-CSF	Granulocyte-macrophage colony stimulating factor
1J	GRO	Growth Related Oncogene
1K	GRO- α	Growth Related Oncogene-Alpha
2A	I-309	I-309
2B	IL-1 α	Interleukin 1 Alpha
2C	IL-1 β	Interleukin 1 Beta
2D	IL-2	Interleukin 2
2E	IL-3	Interleukin 3
2F	IL-4	Interleukin 4
2G	IL-5	Interleukin 5
2H	IL-6	Interleukin 6
2I	IL-7	Interleukin 7
2J	IL-8	Interleukin 8
2K	IL-10	Interleukin 10
3A	IL-12	Interleukin 12
3B	IL-13	Interleukin 13
3C	IL-15	Interleukin 15
3D	IFN- γ	Interferon gamma
3E	MCP-1	Monocyte Chemoattractant Protein 1
3F	MCP-2	Monocyte Chemoattractant Protein 2
3G	MCP-3	Monocyte Chemoattractant Protein 3
3H	MCSF	Macrophage-colony Stimulating Factor
3I	MDC	Macrophage-derived Chemokine
3J	MIG	Monokine induced by gamma interferon
3K	MIP-1 β	Macrophage Inflammatory Protein 1 Beta

4A	MIP-1 δ	Macrophage Inflammatory Protein 1 Delta
		regulated upon activation, normal T-cell expressed, and presumably secreted
4B	RANTES	
4C	SCF	Stem Cell Factor
4D	SDF-1	Stromal cell-driven factor
4E	TARC	Thymus and Activation-Regulated Chemokine
4F	TGF- β 1	Transforming growth factor-beta 1
4G	TNF- α	Tumor necrosis factor-alpha
4H	TNF- β	Tumor necrosis factor-beta
4I	EGF	Epidermal growth factor
4J	IGF-I	Insulin-like growth factor-1
4K	Ang	Angiogenin
5A	OSM	Oncostatin M
5B	TPO	Thrombopoietin
5C	VEGF	Vascular Endothelial Growth Factor
5D	PDGF-B	Platelet-derived Growth Factor-B
5E	Leptin	Leptin
5F	BDNF	Brain-derived neurotrophic factor
5G	BLC	B-lymphocyte chemoattractant
5H	Ck β 8-1	Chemokine-beta-8
5I	Eotaxin	Eotaxin
5J	Eotaxin-2	MPIF-2 (Myeloid progenitor inhibitory factor-2)
5K	Eotaxin-3	MIP-4-alpha (macrophage inflammatory protein-4-alpha)
6A	FGF-4	Fibroblast growth factor-4
6B	FGF-6	Fibroblast growth factor-6
6C	FGF-7	Fibroblast growth factor-7
6D	FGF-9	Fibroblast growth factor-9
6E	Flt-3 Ligand	fms-like tyrosine kinase-3 ligand
6F	Fractalkine	Fractalkine
6G	GCP-2	Granulocyte Chemotactic Protein-2
6H	GDNF	Glial-derived Neurotrophic Factor
6I	HGF	hepatocyte growth factor
6J	IGFBP-1	Insulin-like growth factor binding proteins-1
6K	IGFBP-2	Insulin-like growth factor binding proteins-2
7A	IGFBP-3	Insulin-like growth factor binding proteins-3

7B	IGFBP-4	Insulin-like growth factor binding proteins-4
7C	IL-16	Interleukin 16
7D	IP-10	Interferon gamma inducible protein-10
7E	LIF	Leukemia Inhibitory Factor
7F	LIGHT	LIGHT
7G	MCP-4	Monocyte Chemoattractant Protein 4
7H	MIF	mesoderm inducing factor
7I	MIP-3 α	Macrophage Inflammatory Protein-3 alpha
7J	NAP-2	Neutrophil Activating Peptide 2
7K	NT-3	neurotrophin-3
8A	NT-4	neurotrophin-4
8B	Osteoprotegerin	Osteoprotegerin
8C	PARC	Pulmonary and Activation-Regulated Chemokine
8D	PIGF	Placenta growth factor
8E	TGF- β 2	Transforming growth factor-beta 2
8F	TGF- β 3	Transforming growth factor-beta 3
8G	TIMP-1	tussue inhibitor of metalloproteinases-1
8H	TIMP-2	tussue inhibitor of metalloproteinases-2
8I	Neg	negative control
8J	Pos	positive control
8K	Pos	positive control

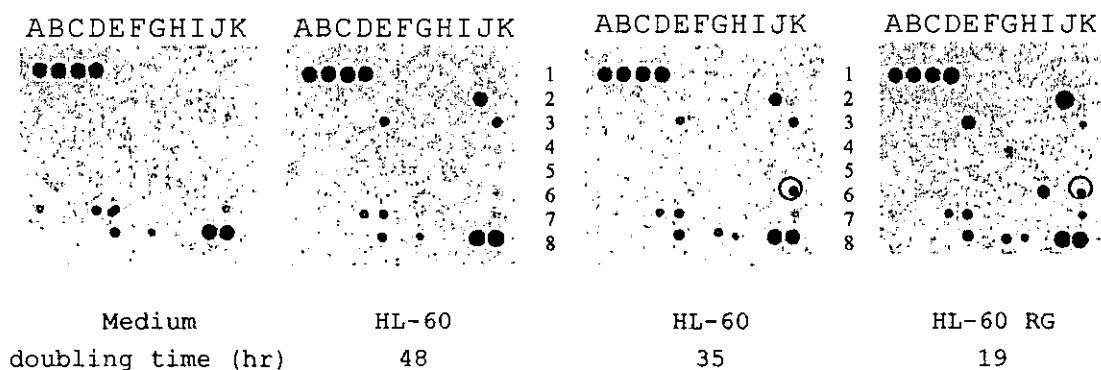


図9. HL-60 と HL-60RG の培養上清のサイトカインアレイによる解析

表4. ELISAによるHL-60細胞及びHL-60R/G細胞の産生する各種サイトカインの定量

	IL-2	IL-4	IL-5	IL-6 (pg/ml)	IL-8 (pg/ml)	IL-10	HGF (pg/ml)	VEGF (pg/ml)	TNF- α	IFN- γ	M-CSF	GM- CSF
HL-60R/G細胞	ND	ND	ND	15.8 \pm 3.2	139 \pm 30	ND	4560 \pm 672	389 \pm 26	ND	ND	ND	ND
HL-60細胞	ND	ND	ND	ND	286 \pm 12	ND	ND	185 \pm 6	ND	ND	ND	ND

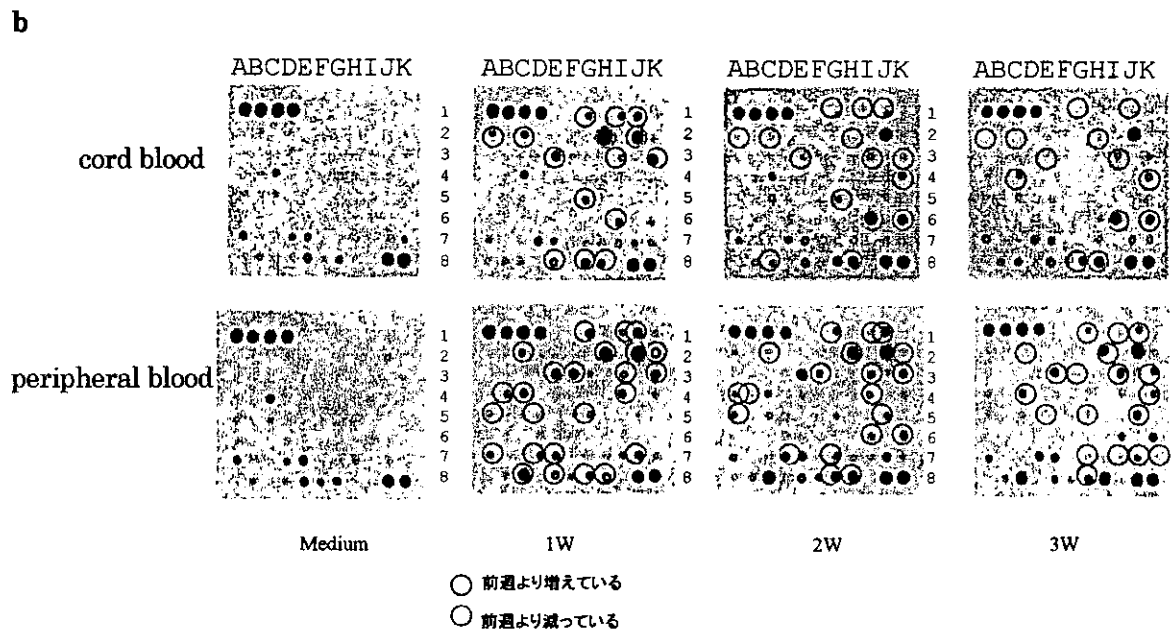
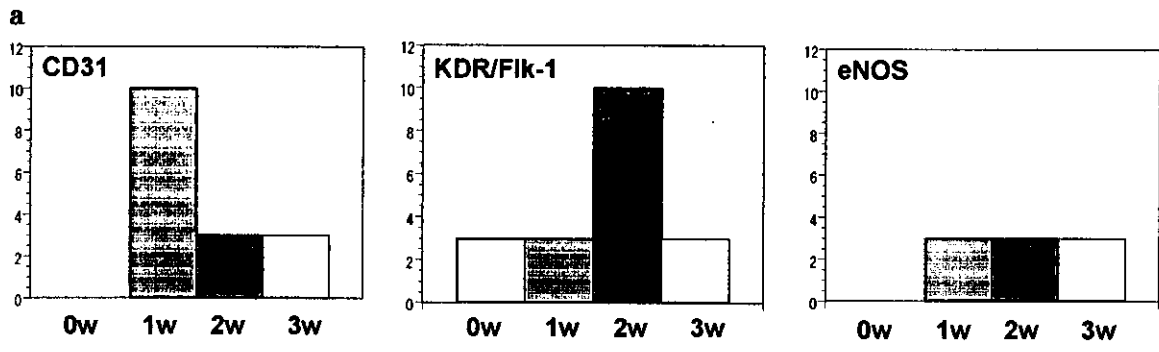


図 10. 臍帯血、末梢血由来の AC133 陽性細胞の培養上清のサイトカインアレイによる解析

mkwvesifli	fllnftesrt	lhrneygias	ildsyqetae	isladlatif
faqfvqeaty	kevskmvkda	ltaiekptgd	eqssgclenq	lpafleelch
ekeilekygh	sdccsqseeg	rhncflahkk	ptpasiplfq	vpepvtscea
yeedretfmn	kfiyeiarrh	pflyaptill	waarydkiip	scckaenave
cfqtkaatvt	kelresslln	qhacavmknf	gtrtfqaitv	tklsqkftkv
nfteiqklvl	dvahvhehcc	rgdvldclqd	gekimsyics	qgdtlsnkit
eccklttler	gqcihaend	ekpeglspl	nrflgdrdfn	qfssgeknif
lasfvheysr	rhpqlavsvi	lrvakgyqel	lekcfqtenp	lecqdkgeee
lqkyiqesqa	lakrscglfq	klgeyylna	flvaytkkap	qltsselmai
trkmaataat	ccqlsedkll	acgegaadii	ighlcirhem	tpvnpgvgqc
ctssyanrrp	cfsslvdet	yvppafsddk	fifhkdleqa	qgvalqtmkq
eflinlvkqk	pqiteeqlea	viadfsglle	kccqgqegev	cfaeegqli
sktraalgv				

Fig. 11 Amino acid sequence and N-glycosylation sites of alpha-Fetoprotein
n: N-glycosylation site

mkililgifl	flcstpawak	ekhyyigiie	ttwdyasdhg	ekklisvdte
hsniylqngp	drigrlykka	lylqytdetf	rttiekpowl	gflgpiikae
tgdkvyvhlk	nlasrpytfh	shgityykeh	egaiypd ^h tt	dfqraddkvy
pgeqytymll	ateeqspgeg	dgncvtriyh	shidapkdia	sgligpliic
kkdsldkeke	khidrefvvm	fsvvde ^h fsw	yledniktyc	sepekvdkdn
edfquesrmy	svngytfgsi	pglsmcaedr	vkwyifgmgn	evdvhaaffh
gqaltknknyr	idtinlfpat	lfdaymvaqn	pgewmlscqn	lnhlkaglqa
ffqvqec ^h ks	sskdnirgkh	vrhyyiaaee	iiwnyapsgi	diftke ^h lta
pgsdsavffe	qgttriggysy	kklyreytd	asftnrkerg	peeehlgilg
pviwaevgdt	irvtfhnkga	yplsiepigv	rfnknnegty	yspnynpqsr
svppsashva	ptettyewt	vpkevgptna	dpvclakmyy	savdptkdif
tgligpmkic	kkgsllhangr	qkdvdkefyl	fptvfde ^h es	lllednirmf
ttapdqvdke	dedfquesnm	hsmngfmygn	qpgltmckgd	svvwyifsg
neadvhgiyf	sgntylwrge	rrdtanlfpq	tsllhmwpd	tegtfnvecl
ttdhytggmk	qkytvnqerr	qsedstfyig	ertyyiaave	vewdyspqre
wekelhhllqe	q ^h vsnafldk	gefyigskyk	kvvyrqytds	tfrvpverka
eeehlgilgp	qlhadvgdkv	kiifknmatr	pysihahgvq	tesstvtptl
pgetltyvwk	ipersgagte	dsacipwayy	stvdqvkdy	sgligplivc
rrpylkvfnp	rrklefallf	lvfde ^h eswy	lddniktysd	hpekvnkdde
efiesnkmha	ingrmfgnlq	gltmhvgdev	nwyimmgne	idlhtvhfhg
hsfqykhrgv	yssdvfdifp	gtyqtlemp	rtpgiwllhc	hvtddihagm
ettytvlqne	dtksg			

Fig. 12 Amino acid sequence and N-glycosylation sites of ceruloplasmin (CP)

^hn: Potential N-glycosylation sites

n: N-glycosylation sites

(A) TIC for the full scan m/z 700-2000

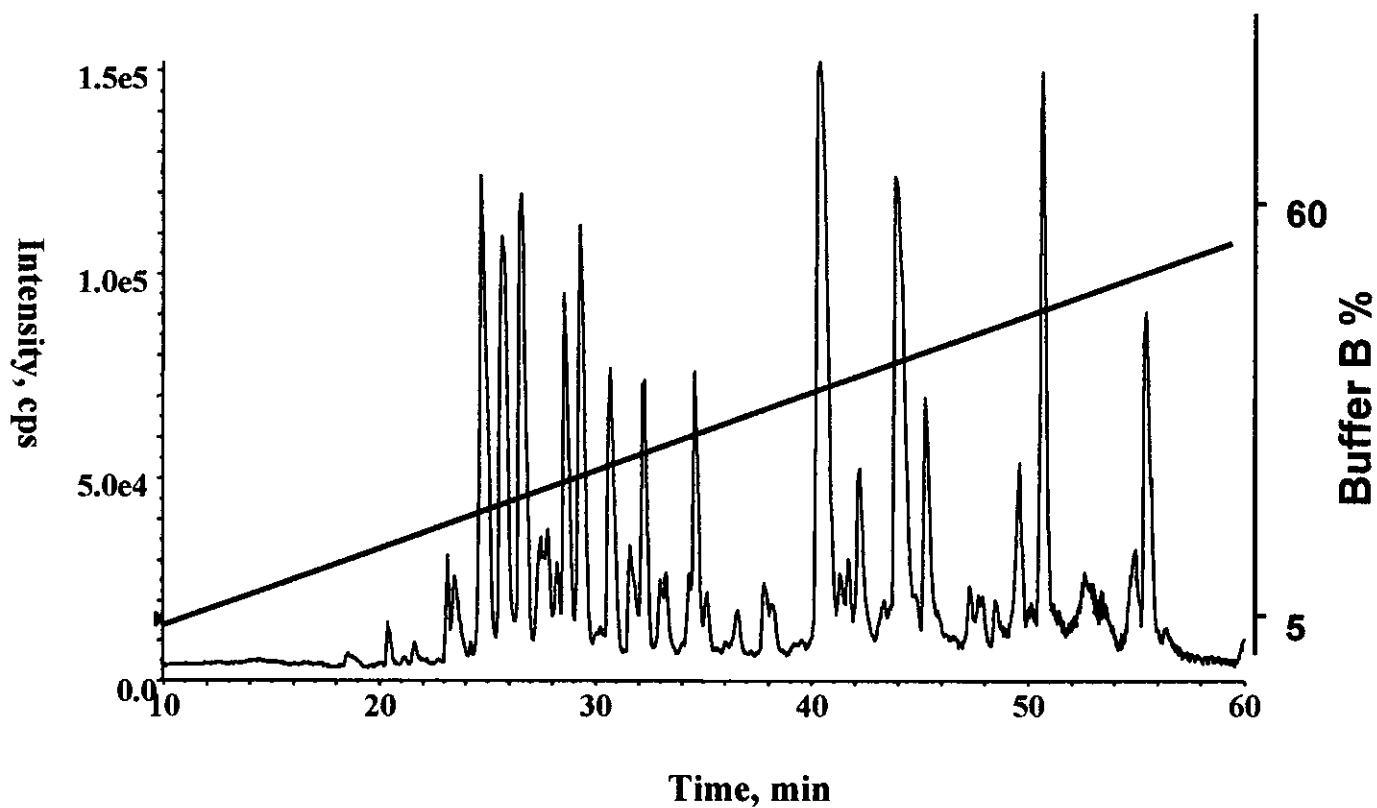


Fig. 13 LC-MS/MS of tryptic digest of α -fetoprotein

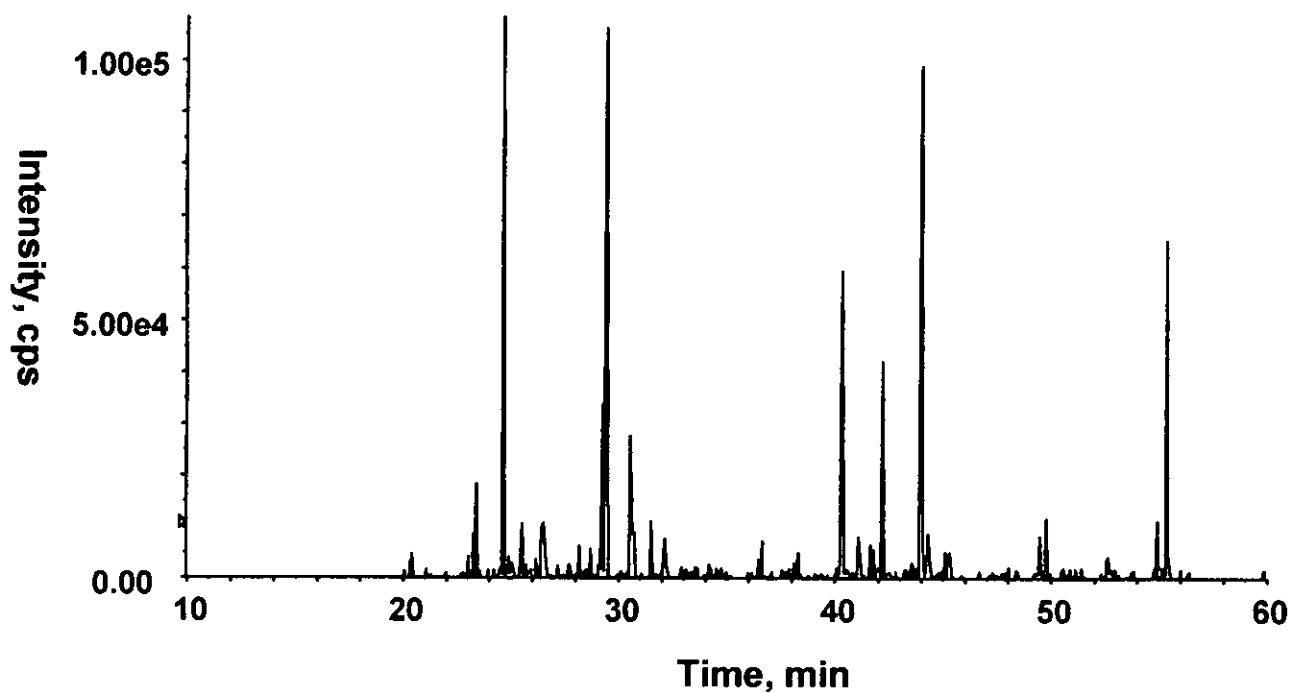
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3 μ , 0.2*50 mm, 2 μ l/min

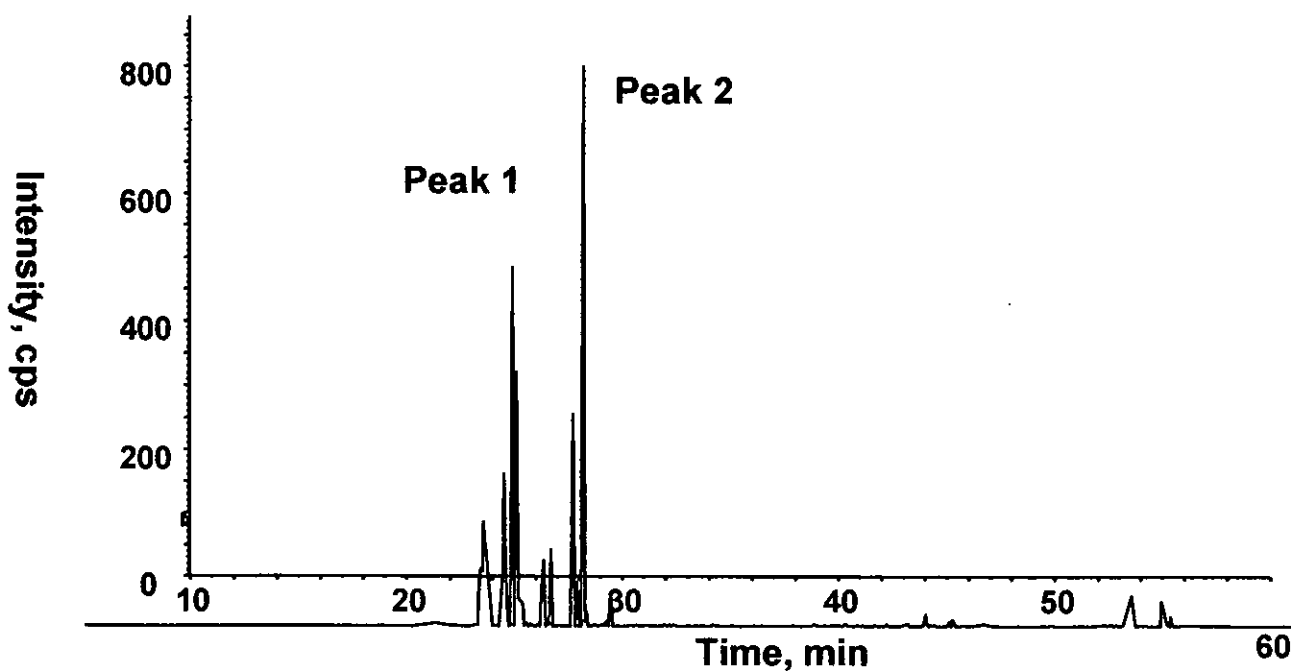
A: 2 % CH_3CN + 0.1 % formic acid

B: 90 % CH_3CN + 0.1 % formic acid

(B) TIC for the product ion scan at m/z 100 - 2000



(C) Product ion scan at m/z 204



Residue	Mass	b	b-NH3	y	y-NH3
V	99.07	100.08	83.05	978.53	961.50
N	114.04	214.12	197.09	879.46	862.43
F	147.07	361.19	344.16	765.41	748.39
T	101.05	462.23	445.21	618.35	601.32
E	129.04	591.28	574.25	517.30	500.27
I	113.08	704.36	687.33	388.26	371.23
Q	128.06	832.42	815.39	275.17	258.14
K	128.10	960.51	943.49	147.11	130.09

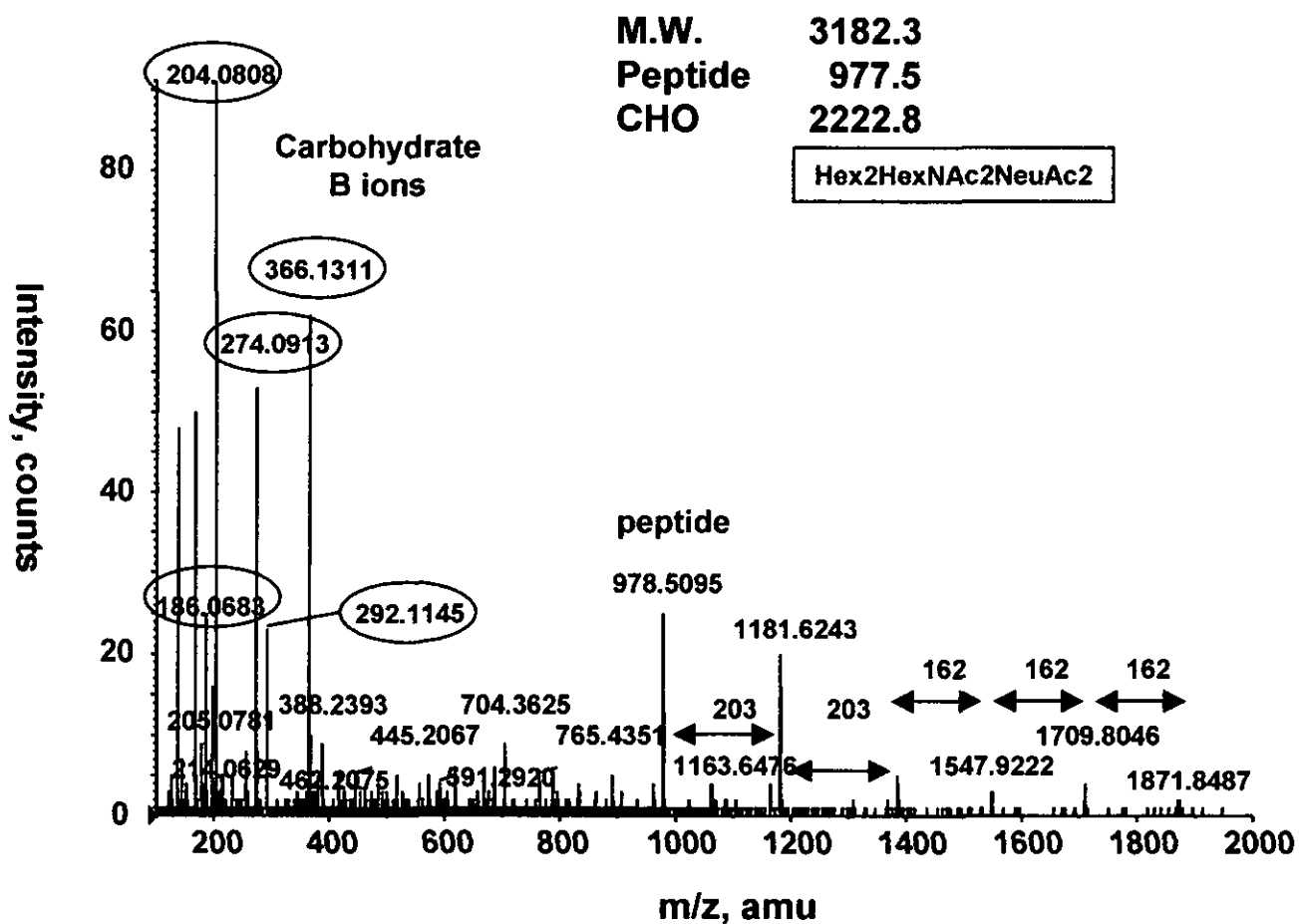


Fig. 14 Product ion spectrum of $M^+(m/z1061.8^{3+})$ at 25 min

Table 10 Glycosylation analysis of alpha-fetoprotein

Retention time (min)	m/z	Charge	M.W.	Peptide Sequences	M.W.	Oligosaccharide Structures	M.W.
23	1013.43	+3	3037.3	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	2077.8
23	1519.67	+2	3037.3	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	2077.8
23	1081.29	+3	3240.9	Kvvnfteiqk/l	977.5	[HexNAc]5[Hex]5[Neu5Ac]1[Fuc]1	2281.4
23	1621.44	+2	3240.9	Kvvnfteiqk/l	977.5	[HexNAc]5[Hex]5[Neu5Ac]1[Fuc]1	2281.4
23	1446.62	+2	2891.2	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]1	1931.8
23	1118.67	+2	2235.3	Kvvnfteiqk/l	977.5	[HexNAc]3[Hex]4	1275.9
23	1264.05	2	2526.1	Kvvnfteiqk/l	977.5	[HexNAc]3[Hex]4[Neu5Ac]1	1566.6
	1110.48	3	2526.1	Kvvnfteiqk/l	977.5	[HexNAc]3[Hex]4[Neu5Ac]1	1566.6
24	1665.44	+2	3328.9	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2369.4
24	1178.33	+3	3532.0	Kvvnfteiqk/l	977.5	[HexNAc]5[Hex]5[Neu5Ac]2[Fuc]1	2572.5
24	1061.78	3	3182.3	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2	2222.9
25	1592.17	2	3182.3	Kvvnfteiqk/l	977.5	[HexNAc]4[Hex]5[Neu5Ac]2	2222.9
27	1236.04	+3	3705.1	Kvvnfteiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2[Fuc]1	2369.4
27	1853.29	+2	3704.6	Kvvnfteiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]2	2222.8
27	1187.17	+3	3558.5	Kvvnfteiqk/l	1353.7	[HexNAc]5[Hex]5[Neu5Ac]2[Fuc]1	2572.0
28	1780.23	+2	3558.5	Kvvnfteiqk/l	1353.7	[HexNAc]5[Hex]5[Neu5Ac]2[Fuc]1	2572.0
28	1303.56	+3	3907.7	Kvvnfteiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]1[Fuc]1	2077.8
28	1138.84	+3	3413.5	Kvvnfteiqk/l	1353.7	[HexNAc]5[Hex]5[Neu5Ac]1[Fuc]1	2280.9
28	1206.52	+3	3616.6	Kvvnfteiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]1	1931.7
28	1090.14	+3	3267.4	Kvvnfteiqk/l	1353.7	[HexNAc]4[Hex]5[Neu5Ac]1	1931.7
28	1322.75	+3	3965.3	Kvvnfteiqk/l	1353.7	[HexNAc]6[Hex]5[Neu5Ac]2	2629.6

(A) TIC for the full scan m/z 1000-2000

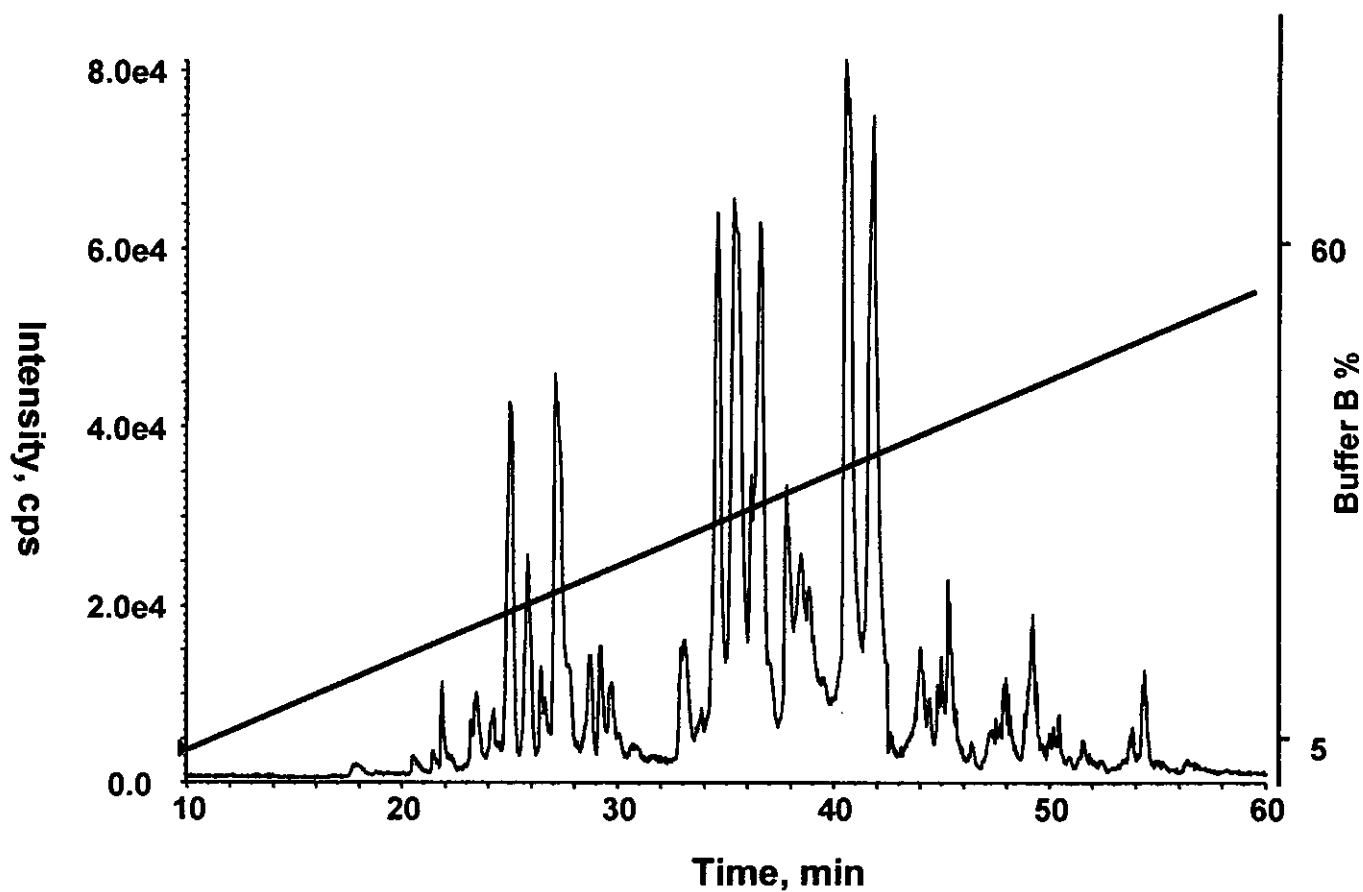


Fig. 15 LC-MS/MS of tryptic digest of ceruloplasmin

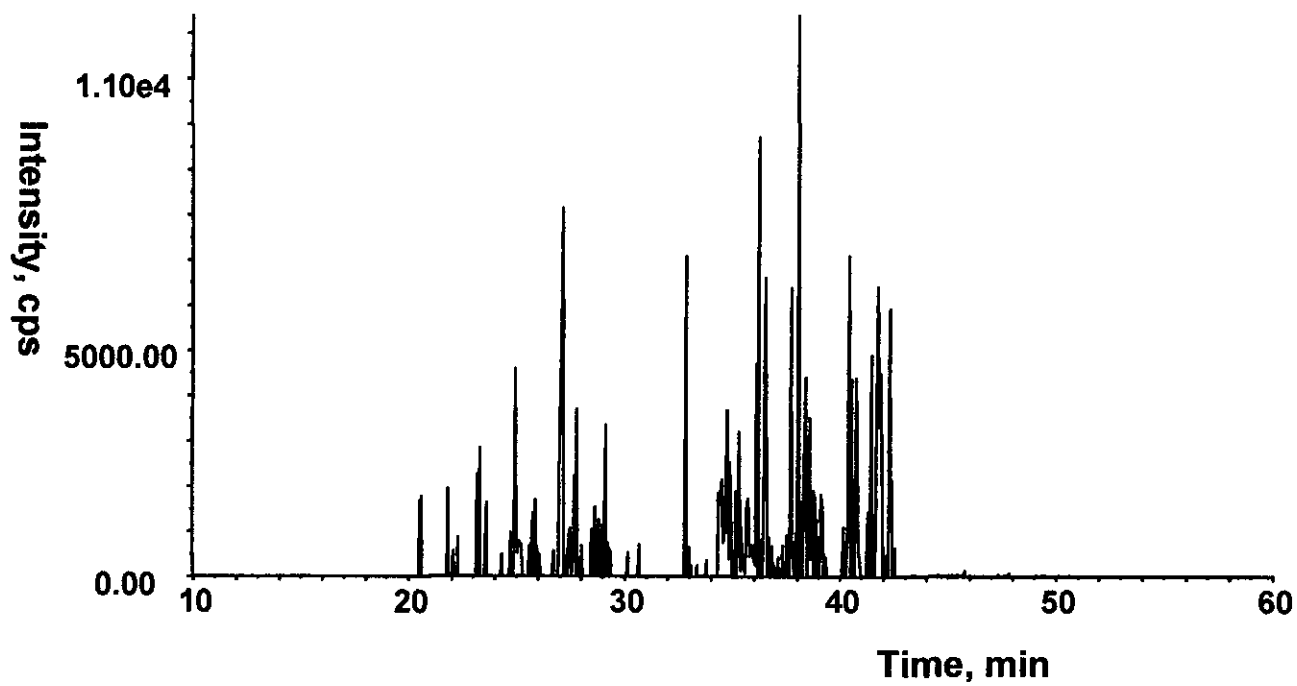
Column Magic C18

3 μ , 0.2*50 mm, 2 μ l/min

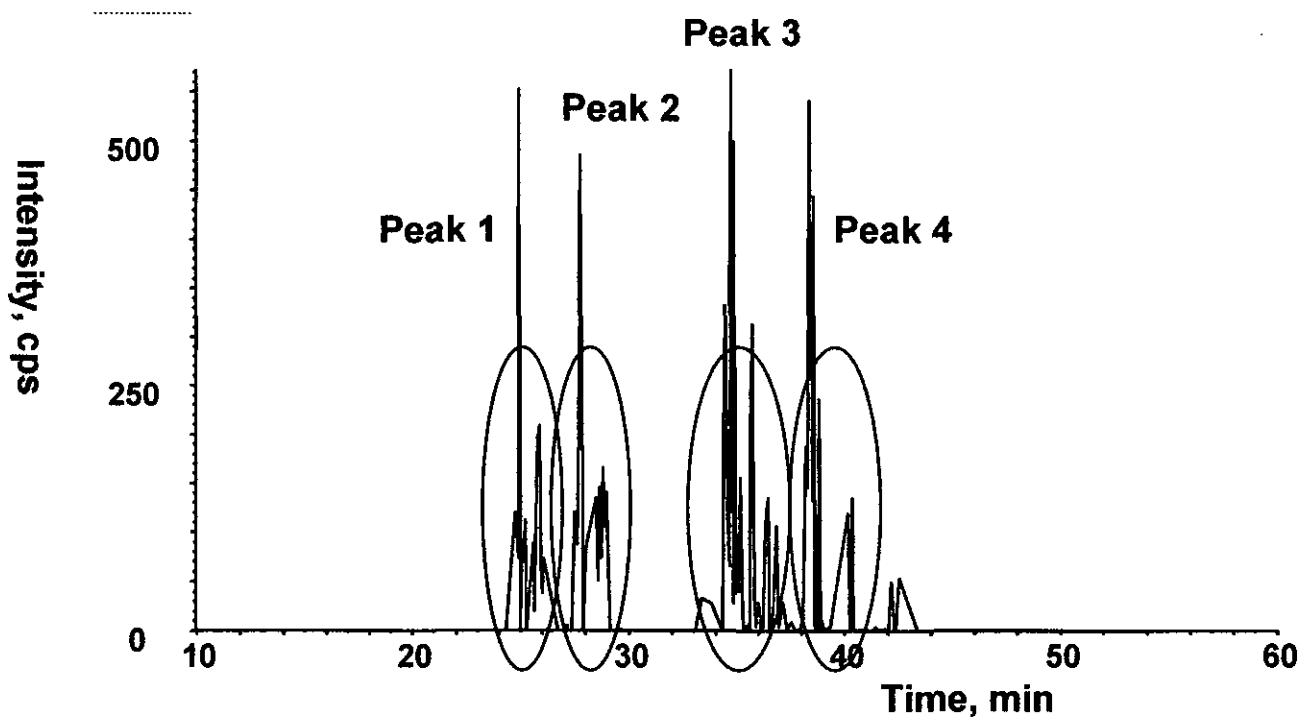
A: 2 % CH_3CN + 0.1 % formic acid

B: 90 % CH_3CN + 0.1 % formic acid

(B) TIC for the product ion scan at m/z 100 - 2000



(C) Product ion scan at m/z 204



M.W. 4096.6
Peptide 1891.8
CHO 2222.8

Hex₂HexNAc₂NeuAc₂

Residue	Mass	b	b-NH3	y	y-NH3
E	129.04	130.05	113.02	1892.84	1875.81
H	137.06	267.11	250.08	1763.80	1746.77
E	129.04	396.15	379.12	1626.74	1609.71
G	57.02	453.17	436.15	1497.70	1480.67
A	71.04	524.21	507.18	1440.68	1423.65
I	113.08	637.29	620.27	1369.64	1352.61
Y	163.06	800.36	783.33	1256.55	1239.53
P	97.05	897.41	880.38	1093.49	1076.46
D	115.03	1012.44	995.41	996.44	979.41
N	114.04	1126.48	1109.45	881.41	864.38
T	101.05	1227.53	1210.50	767.37	750.34
T	101.05	1328.58	1311.55	666.32	649.29
D	115.03	1443.60	1426.58	565.27	548.25
F	147.07	1590.67	1573.64	450.25	433.22
Q	128.06	1718.73	1701.70	303.18	286.15
R	156.10	1874.83	1857.80	175.12	158.09

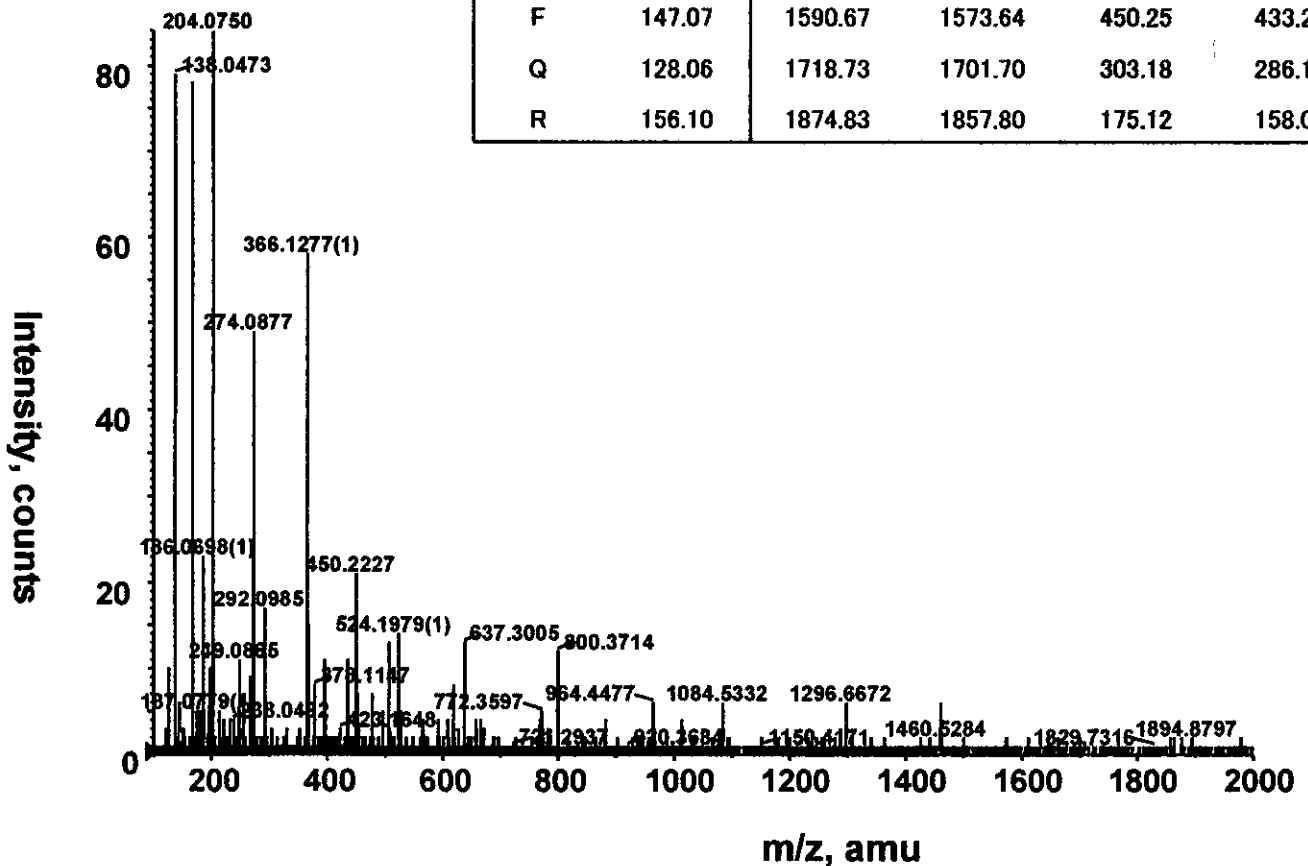


Fig. 16 Product ion spectrum of M⁺(m/z1366.6³⁺) at 25 min