

plasticity and adaptation, but, its role in behavioral sensitization is still known. The MAP kinase family consists of two major subgroups, the extracellular signal-regulated kinase (ERK), which induces cell growth and proliferation (classical MAPK cascade), and the stress-activated protein kinase/c-Jun N-terminal kinase (SAPK/JNK) and p38, which induce apoptosis, the stress reaction, and gene transcription (novel MAPK cascade). MAP kinases are activated and phosphorylated by MAPKK and MAPKKK and selectively inactivated and dephosphorylated by mitogen-activated protein phosphatases (MKPs). Many kinds of MKPs are known, MKP-1 and MKP-3 were examined because of their abundant distribution in the brain^{52,53}. Rheb, a ras homologue, which is abundant in the brain and activates MAPK cascade⁵⁴, was also examined.

MKP-1 mRNA is abundantly distributed in the cerebral cortex and thalamus of the naive rat, whereas MKP-3 mRNA is restricted to the hippocampus⁵⁵ (fig 2). MKP-1 mRNA significantly increased about 60-300% in the several areas of the cortex, striatum, and thalamus, 0.5-1 h in response to acute and chronic methamphetamine administration (fig 3 and 4). MKP-3 mRNA had increased about 50% in the cortex, striatum and hippocampus 1 h after acute methamphetamine administration, but only in the hippocampus CA1 after chronic methamphetamine administration. Pretreatment with the D1 dopamine antagonist SCH 23390 completely abolished the increase in MKP-1 and

MKP-3 mRNA levels induced by acute methamphetamine administration, but it was only partially abolished by the NMDA antagonist MK-801. The MKP-1 protein level also increased in the cortex 3 h after acute and chronic methamphetamine administration. Rheb mRNA was unchanged after acute and chronic methamphetamine administration. MKPs are inactivators of MAPK^{56,57}, but they are directly activated by activation of MAPK. MKP-1 is selectively activated by SAPK/JNK and p38, and MKP-3 by ERK^{58,59}. Therefore, the increase in MKP-1 and MKP-3 mRNAs observed after methamphetamine administration must mean activation of SAPK/JNK and p38, and ERK, respectively. Taken together, these findings indicate that acute methamphetamine administration may activate both the classical and novel MAPK cascades in several areas of cortex and striatum, the novel MAPK cascade in the thalamus, and the classical MAPK cascade in the hippocampus. By contrast, chronic methamphetamine may activate selected MAPKs in restricted regions, the novel MAPK cascade in the frontal cortex and the classical MAPK cascade in the hippocampus.

6. Conclusion

The differences in regulation of several genes related to morphological plasticity after acute and subchronic methamphetamine are quite significant, and they are summarized in Table 1. The behavioral sensitization phenomenon in response to psychostimulants is considered to

consist of two major distinct processes: an early induction process and a later maintenance or expression process. Induction of sensitization has been shown to occur after administration of several doses of psychostimulants. The neurochemical changes in increased dopamine efflux and morphological changes in dendrites and spines in the accumbens and frontal cortex were demonstrated after chronic amphetamine administration. However, recent studies have shown that even a single exposure to amphetamine also induced long-term behavioral and neurochemical sensitization⁶⁰. The neural adaptation during persistent sensitization, such as synaptogenesis and neurite elongation, must begin with the first psychostimulant exposure, and thus the changes in gene expression seen after a single dose of methamphetamine and during chronic administration should correspond to the molecular mechanisms of the induction and maintenance processes of sensitization, respectively. During the induction process of sensitization, various processes involved in neural adaptation, synaptogenesis, sprouting and elongation processes, and activation of MAPK cascades begin abruptly throughout almost the entire brain, including the striatum, accumbens, frontal and several areas of cortex, and hippocampus. By contrast, no additional synaptogenesis or sprouting seemed to be required during the maintenance process, because synaptophysin and stathmin transcription returned to their basal levels. Neurite elongation indicated by the increased arc mRNA may persist

in those brain regions in response to every methamphetamine exposure, and this sustained neurite elongation during repeated methamphetamine administration may contribute to re-arrangement of the neural networks for behavioral sensitization. Activation of the novel MAPK kinase cascade in the frontal cortex and classical MAPK cascade in the hippocampus as indicated by the increase in MKP-1 and MKP-3 mRNAs, respectively, also persisted during the maintenance process of sensitization. This may imply occurrence of apoptosis in the frontal cortex and neurogenesis in the hippocampus, which has not been demonstrated in the sensitization phenomenon. The actions and roles of many other plasticity-related genes should be investigated to further clarify the molecular mechanisms in very-long lasting neural adaptation during sensitization.

Reference

1. Akiyama, K., A. Kanzaki, K. Tsuchida & H. Ujike. 1994. Methamphetamine-induced behavioral sensitization and its implications for relapse of schizophrenia. *Schizophr Res* **12**: 251-257.
2. Robinson, T.E. & J.B. Becker. 1986. Enduring changes in brain and behavior produced by chronic amphetamine administration: a review and evaluation of animal models of amphetamine psychosis. *Brain Res Rev* **11**: 157-198.
3. Sato, M., C.C. Chen, K. Akiyama & S.

- Otsuki. 1983. Acute exacerbation of paranoid psychotic state after long-term abstinence in patients with previous methamphetamine psychosis. *Biol Psychiatry* **18**: 429-440.
4. Tatetsu, S. 1963. Methamphetamine psychosis. *Folia Psychiatr Neurol Jpn Suppl* **7**: 377-380.
 5. Paulson, P.E., D.M. Camp & T.E. Robinson. 1991. Time course of transient behavioral depression and persistent behavioral sensitization in relation to regional brain monoamine concentrations during amphetamine withdrawal in rats. *Psychopharmacology (Berl)* **103**: 480-492.
 6. Ujike, H., K. Akiyama, H. Nishikawa, T. Onoue & S. Otsuki. 1991. Lasting increase in D1 dopamine receptors in the lateral part of the substantia nigra pars reticulata after subchronic methamphetamine administration. *Brain Res* **540**: 159-163.
 7. Itzhak, Y. 1994. Modulation of the PCP/NMDA receptor complex and sigma binding sites by psychostimulants. *Neurotoxicol Teratol* **16**: 363-368.
 8. Ujike, H., K. Okumura, Y. Zushi, K. Akiyama & S. Otsuki. 1992. Persistent supersensitivity of sigma receptors develops during repeated methamphetamine treatment. *Eur J Pharmacol* **211**: 323-328.
 9. Ujike, H., K. Tsuchida, K. Akiyama & S. Otsuki. 1992. Supersensitivity of sigma receptors after repeated administration of cocaine. *Life Sci* **51**: PL31-36.
 10. Ujike, H., N. Ogawa & S. Otsuki. 1988. Effects of acute and long-term treatment with methamphetamine on substance P concentration and receptor numbers in the rat brain. *Brain Res* **453**: 136-142.
 11. Nakashima, M., S. Kajita & S. Otsuki. 1989. Reduction of rat striatal thyrotropin-releasing hormone receptors produced by repeated methamphetamine administration. *Biol Psychiatry* **25**: 191-199.
 12. Wang, J.Q. & J.F. McGinty. 1995. Alterations in striatal zif/268, prodynorphin and preproenkephalin mRNA expression induced by repeated amphetamine administration in rats. *Brain Res* **673**: 262-274.
 13. Shilling, P.D., J.R. Kelsoe & D.S. Segal. 1997. Dopamine transporter mRNA is up-regulated in the substantia nigra and the ventral tegmental area of amphetamine-sensitized rats. *Neurosci Lett* **236**: 131-134.
 14. Ujike, H., K. Akiyama & S. Kuroda. 1996. Increased Gi alpha and Go alpha mRNAs in hippocampus after repeated methamphetamine administration. *Neuroreport* **7**: 2036-2040.
 15. Wang, X.B., M. Funada, Y. Imai, R.S. Revay, H. Ujike, D.J. Vandenberg & G.R. Uhl. 1997. rGbeta1: a psychostimulant-regulated gene essential for establishing cocaine sensitization. *J*

- Neurosci 17: 5993-6000.
16. Nestler, E.J., R.Z. Terwilliger, J.R. Walker, K.A. Sevarino & R.S. Duman. 1990. Chronic cocaine treatment decreases levels of the G protein subunits Gi α and Go α in discrete regions of rat brain. *J Neurochem* 55: 1079-1082.
 17. Terwilliger, R.Z., J.D. Beitner, K.A. Sevarino, S.M. Crain & E.J. Nestler. 1991. A general role for adaptations in G-proteins and the cyclic AMP system in mediating the chronic actions of morphine and cocaine on neuronal function. *Brain Res* 548: 100-110.
 18. Gnegy, M.E., G.H. Hewlett, S.L. Yee & M.J. Welsh. 1991. Alterations in calmodulin content and localization in areas of rat brain after repeated intermittent amphetamine. *Brain Res* 562: 6-12.
 19. Shimizu, Y., K. Akiyama, M. Kodama, T. Ishihara, T. Hamamura & S. Kuroda. 1997. Alterations of calmodulin and its mRNA in rat brain after acute and chronic administration of methamphetamine. *Brain Res* 765: 247-258.
 20. Iwata, S.I., G.H. Hewlett, S.T. Ferrell, L. Kantor & M.E. Gnegy. 1997. Enhanced dopamine release and phosphorylation of synapsin I and neuromodulin in striatal synaptosomes after repeated amphetamine. *J Pharmacol Exp Ther* 283: 1445-1452.
 21. Graybiel, A.M., R. Moratalla & H.A. Robertson. 1990. Amphetamine and cocaine induce drug-specific activation of the c-fos gene in striosome-matrix compartments and limbic subdivisions of the striatum. *Proc Natl Acad Sci U S A* 87: 6912-6916.
 22. Ishihara, T., K. Akiyama, K. Kashihara, H. Ujike, T. Hamamura, S. Okada & S. Kuroda. 1996. Activator protein-1 binding activities in discrete regions of rat brain after acute and chronic administration of methamphetamine. *J Neurochem* 67: 708-716.
 23. Karler, R., K.T. Finnegan & L.D. Calder. 1993. Blockade of behavioral sensitization to cocaine and amphetamine by inhibitors of protein synthesis. *Brain Res* 603: 19-24.
 24. Shimosato, K. & T. Saito. 1993. Suppressive effect of cycloheximide on behavioral sensitization to methamphetamine in mice. *Eur J Pharmacol* 234: 67-75.
 25. Fujiwara, Y., Y. Kazahaya, M. Nakashima, M. Sato & S. Otsuki. 1987. Behavioral sensitization to methamphetamine in the rat: an ontogenic study. *Psychopharmacology (Berl)* 91: 316-319.
 26. Ujike, H., K. Tsuchida, K. Akiyama, Y. Fujiwara & S. Kuroda. 1995. Ontogeny of behavioral sensitization to cocaine. *Pharmacol Biochem Behav* 50: 613-617.
 27. Ujike, H., M. Takaki & S. Kuroda. (in press). Neural plasticity-related genes and the behavioral sensitization phenomenon. *Psychiatry Clin Neurosci*.
 28. Robinson, T.E. & B. Kolb. 1997. Persistent

- structural modifications in nucleus accumbens and prefrontal cortex neurons produced by previous experience with amphetamine. *J Neurosci* **17**: 8491-8497.
29. Robinson, T.E. & B. Kolb. 1999. Alterations in the morphology of dendrites and dendritic spines in the nucleus accumbens and prefrontal cortex following repeated treatment with amphetamine or cocaine. *Eur J Neurosci* **11**: 1598-1604.
30. Marqueze-Pouey, B., W. Wisden, M.L. Malosio & H. Betz. 1991. Differential expression of synaptophysin and synaptoporin mRNAs in the postnatal rat central nervous system. *J Neurosci* **11**: 3388-3397.
31. Archer, B.T., 3rd, T. Ozcelik, R. Jahn, U. Francke & T.C. Sudhof. 1990. Structures and chromosomal localizations of two human genes encoding synaptobrevins 1 and 2. *J Biol Chem* **265**: 17267-17273.
32. Takaki, M., H. Ujike, M. Kodama, Y. Takehisa, A. Yamamoto & S. Kuroda. 2001. Increased expression of synaptophysin and staathmin mRNAs after methamphetamine administration in rat brain. *Neuroreport* **12**: 1055-1060.
33. Denovan-Wright, E.M., R.A. Newton, J.N. Armstrong, J.M. Babity & H.A. Robertson. 1998. Acute administration of cocaine, but not amphetamine, increases the level of synaptotagmin IV mRNA in the dorsal striatum of rat. *Mol. Brain Res* **55**: 350-354.
34. Iwata, S., G.H. Hewlett, S.T. Ferrell, A.J. Czernik, K.F. Meiri & M.E. Gnegy. 1996. Increased in vivo phosphorylation state of neuromodulin and synapsin I in striatum from rats treated with repeated amphetamine. *J Pharmacol Exp Ther* **278**: 1428-1434.
35. Castaneda, E., J.B. Becker & T.E. Robinson. 1988. The long-term effects of repeated amphetamine treatment in vivo on amphetamine, KCl and electrical stimulation evoked striatal dopamine release in vitro. *Life Sci* **42**: 2447-2456.
36. Himi, T., T. Okazaki, H. Wang, T.H. McNeill & N. Mori. 1994. Differential localization of SCG10 and p19/stathmin messenger RNAs in adult rat brain indicates distinct roles for these growth-associated proteins. *Neuroscience* **60**: 907-926.
37. Sobel, A. 1991. Stathmin: a relay phosphoprotein for multiple signal transduction? *Trends Biochem Sci* **16**: 301-305.
38. Tsui, C.C., N.G. Copeland, D.J. Gilbert, N.A. Jenkins, C. Barnes & P.F. Worley. 1996. Narp, a novel member of the pentraxin family, promotes neurite outgrowth and is dynamically regulated by neuronal activity. *J Neurosci* **16**: 2463-2478.
39. O'Brien, R.J., D. Xu, R.S. Petralia, O. Steward, R.L. Huganir & P. Worley. 1999. Synaptic clustering of AMPA receptors by the extracellular immediate-early gene product Narp. *Neuron* **23**: 309-323.

40. Naeve, G.S., M. Ramakrishnan, R. Kramer, D. Hevroni, Y. Citri & L.E. Theill. 1997. Neuritin: a gene induced by neural activity and neurotrophins that promotes neuritogenesis. *Proc Natl Acad Sci U S A* **94**: 2648-2653.
41. Sugiura, Y. & N. Mori. 1995. SCG10 expresses growth-associated manner in developing rat brain, but shows a different pattern to p19/stathmin or GAP-43. *Brain Res Dev Brain Res* **90**: 73-91.
42. McNeill, T.H., N. Mori & H.W. Cheng. 1999. Differential regulation of the growth-associated proteins, GAP-43 and SCG-10, in response to unilateral cortical ablation in adult rats. *Neuroscience* **90**: 1349-1360.
43. Beilharz, E.J., E. Zhukovsky, A.A. Lanahan, P.F. Worley, K. Nikolich & L.J. Goodman. 1998. Neuronal activity induction of the stathmin-like gene RB3 in the rat hippocampus: possible role in neuronal plasticity. *J Neurosci* **18**: 9780-9789.
44. Ozon, S., T. Byk & A. Sobel. 1998. SCLIP: a novel SCG10-like protein of the stathmin family expressed in the nervous system. *J Neurochem* **70**: 2386-2396.
45. Belmont, L.D. & T.J. Mitchison. 1996. Identification of a protein that interacts with tubulin dimers and increases the catastrophe rate of microtubules. *Cell* **84**: 623-631.
46. Gnegy, M.E., P. Hong & S.T. Ferrell. 1993. Phosphorylation of neuromodulin in rat striatum after acute and repeated, intermittent amphetamine. *Brain Res Mol Brain Res* **20**: 289-298.
47. Miller, F.D., C.C. Naus, M. Durand, F.E. Bloom & R.J. Milner. 1987. Isotypes of alpha-tubulin are differentially regulated during neuronal maturation. *J Cell Biol* **105**: 3065-3073.
48. Lyford, G.L., K. Yamagata, W.E. Kaufmann, C.A. Barnes, N.G. Copeland, D.J. Gilbert, N.A. Jenkins, A.A. Lanahan & P.F. Worley. 1995. Arc, a growth factor and activity-regulated gene, encodes a novel cytoskeleton-associated protein that is enriched in neuronal dendrites. *Neuron* **14**: 433-445.
49. Karler, R., L.D. Calder, I.A. Chaudhry & S.A. Turkkanis. 1989. Blockade of "reverse tolerance" to cocaine and amphetamine by MK-801. *Life Sci* **45**: 599-606.
50. Ujike, H., T. Onoue, K. Akiyama, T. Hamamura & S. Otsuki. 1989. Effects of selective D-1 and D-2 dopamine antagonists on development of methamphetamine-induced behavioral sensitization. *Psychopharmacology (Berl)* **98**: 89-92.
51. Kodama, M., K. Akiyama, H. Ujike, Y. Tanaka & S. Kuroda. 1997. Methamphetamine increases arc gene expression in rat striatum and cortex. *Abstract for Neuroscience*
52. Gass, P., A. Eckhardt, H. Schroder, R. Bravo & T. Herdegen. 1996. Transient

- expression of the mitogen-activated protein kinase phosphatase MKP-1 (3CH134/ERP1) in the rat brain after limbic epilepsy. *Brain Res Mol Brain Res* **41**: 74-80.
53. Muda, M., U. Boschert, R. Dickinson, J.C. Martinou, I. Martinou, M. Camps, W. Schlegel & S. Arkininstall. 1996. MKP-3, a novel cytosolic protein-tyrosine phosphatase that exemplifies a new class of mitogen-activated protein kinase phosphatase. *J Biol Chem* **271**: 4319-4326.
54. Yamagata, K., L.K. Sanders, W.E. Kaufmann, W. Yee, C.A. Barnes, D. Nathans & P.F. Worley. 1994. *rheb*, a growth factor- and synaptic activity-regulated gene, encodes a novel Ras-related protein. *J Biol Chem* **269**: 16333-16339.
55. Takaki, M., H. Ujike, M. Kodama, Y. Takehisa, K. Nakata & S. Kuroda. 2001. Two kinds of mitogen-activated protein kinase phosphatases, MKP-1 and MKP-3, are differently activated by acute and chronic methamphetamine treatment in the rat brain. *J Neurochem* **79**: 679-688.
56. Franklin, C.C. & A.S. Kraft. 1997. Conditional expression of the mitogen-activated protein kinase (MAPK) phosphatase MKP-1 preferentially inhibits p38 MAPK and stress-activated protein kinase in U937 cells. *J Biol Chem* **272**: 16917-16923.
57. Muda, M., A. Theodosiou, N. Rodrigues, U. Boschert, M. Camps, C. Gillieron, K. Davies, A. Ashworth & S. Arkininstall. 1996. The dual specificity phosphatases M3/6 and MKP-3 are highly selective for inactivation of distinct mitogen-activated protein kinases. *J Biol Chem* **271**: 27205-27208.
58. Bokemeyer, D., A. Sorokin, M. Yan, N.G. Ahn, D.J. Templeton & M.J. Dunn. 1996. Induction of mitogen-activated protein kinase phosphatase 1 by the stress-activated protein kinase signaling pathway but not by extracellular signal-regulated kinase in fibroblasts. *J Biol Chem* **271**: 639-642.
59. Camps, M., A. Nichols, C. Gillieron, B. Antonsson, M. Muda, C. Chabert, U. Boschert & S. Arkininstall. 1998. Catalytic activation of the phosphatase MKP-3 by ERK2 mitogen-activated protein kinase. *Science* **280**: 1262-1265.
60. Vanderschuren, L.J., E.D. Schmidt, T.J. De Vries, C.A. Van Moorsel, F.J. Tilders & A.N. Schoffelmeer. 1999. A single exposure to amphetamine is sufficient to induce long-term behavioral, neuroendocrine, and neurochemical sensitization in rats. *J Neurosci* **19**: 9579-9586.

自殺を惹起する精神疾患における Chromogranin B 遺伝子の関連解析

(分担研究者 稲田俊也 名古屋大学大学院医学系研究科精神生物学分野助教授)

研究要旨：自殺を惹起する精神疾患としては、統合失調症、覚醒剤精神病、躁うつ病などがある。統合失調症を対象とした DNA マイクロサテライトマーカーを用いたゲノムスキャンにおいて有意な差がみられた 20 番染色体上のマーカー D20S95 の最も近傍に存在する Chromogranin B 遺伝子の変異検索を行い、5'側調節領域において 5 つの変異と Exon4 内に 11 個のアミノ酸置換を伴う変異を見いだした。これらの遺伝子変異について統合失調症患者 187 名と健常対照者 192 名を対象として症例対照群間比較を行った結果、Exon4 内の 4 つ変異において、互いの連鎖不平衡、および統合失調症との関連が認められた。このほか、双極性障害および覚醒剤精神病についても健常対照者との症例群間比較を行ったが有意な関連は見いだせなかった。

研究協力者：飯嶋良味¹⁾、有波忠雄²⁾、大槻露華²⁾、氏家 寛³⁾、尾崎紀夫⁴⁾、前田貴記⁵⁾、山内惟光⁶⁾、岩下 覚⁶⁾、坂元 薫⁷⁾、福永貴子⁸⁾、伊豫雅臣⁹⁾、関根吉統¹⁰⁾、原野陸正¹¹⁾、小宮山徳太郎¹²⁾、山田光彦¹³⁾、曾良一郎¹⁴⁾、中平 進¹⁵⁾、樋口輝彦¹⁶⁾

研究協力者所属施設：1)国立精神・神経センター精神保健研究所、2)筑波大学基礎医学系遺伝医学、3)岡山大学大学院医歯学総合研究科精神神経病態学分野、4)藤田保健衛生大学医学部精神医学教室、5)慶應義塾大学医学部精神神経科、6)社会福祉法人桜ヶ丘記念病院、7)東京女子医科大学神経精神科、8)東京女子医科大学第二病院心の医療科、9)千葉大学大学院医学研究院精神医学講座、10)浜松医科大学精神科、11)久留米大学医学部精神神経科、12)国立精神・神経センター武蔵病院、13)昭和大学附属烏山病院精神神経科、14)東北大学大学院医学系研究科精神神経生物学分野、15)東京高尾病院、16)国立精神・神経センター国府台病院

A. 研究目的

本研究の目的は統合失調症、覚醒剤精神病、躁うつ病などの自殺を惹起する精神疾患とクロモグラニン B 遺伝子との間に関連がみられるかどうかを検討することである。臨床遺伝学的研究から、統合失調症や双極性障害などの精神疾患には遺伝要因の関与が示されているが、全ゲノムスキャンによるそれら精神疾患の遺伝子連鎖研究では、最近急速にその報告数が増加しているにもかかわらず、一貫して連鎖陽性を示す所見が得られないことから、多くの集団に共通する Major gene は存在しないことが示

唆されている。我々は、統合失調症の発症脆弱性に関連する遺伝子座位の系統的なスクリーニング解析を行い、20番染色体上のマーカー D20S95 において症例・対照間に有意な差を見いだした(Kitao Y *et al.*, *Psychiatr Genet* 2000; 10: 139-143)。この D20S95 の最も近傍の位置的候補遺伝子は Chromogranin B 遺伝子である。Chromogranin family (A, B, C) は可溶性分泌タンパクで、種々の神経細胞や脳脊髄液中に分布し、シナプスからの伝達物質の放出をコントロールしていると考えられている。Chromogranin A および B が、統合失調症患者脳脊髄液中で有意に減少しているとの報告や(Landen M *et al.*, *Eur Neuropsychopharmacol* 1999; 9: 311-315), reserpine, clozapine, haloperidol などの抗精神病薬に反応して、脳内における mRNA の発現増加や発現の局在変化が観察されることから(Mahata SK *et al.*, *Brain Res Mol Brain Res* 1993; 19: 83-92, Kroesen S *et al. Neuroscience* 1995; 69: 881-891), 薬物反応性のマーカーとして、さらには精神疾患病態に関与する機能的候補遺伝子として重要な役割を担っていると考えられる。われわれは、統合失調症において Chromogranin B 遺伝子の変異検索および関連解析を行い、Exon4 に位置する 4 つ変異において、統合失調症との強い関連を見いだした。さらに、中国人グループからも Chromogranin B 遺伝子 Exon4 中の別の変異において、統合失調症との関連が報告された(Zhang B *et al.*, *Neurosci Lett* 2002; 323: 229-233)。また、Chromogranin B

遺伝子の位置する染色体 20pter~20p12 領域は、双極性障害においても、米国 NIMH の家系を用いた研究で、パラメトリック連鎖解析によって LOD 値 1 以上が示された領域である(Detera-Wadleigh SD *et al.*, *Am J Med Genet.* 1997; 74: 254-262)。今回われわれは、双極性障害および統合失調症と類似の精神症状を示す覚醒剤精神病について、Chromogranin B 遺伝子がそれら疾患と関連するかを検討した。

B. 研究方法

対象は東京近郊の精神科治療施設に通院または入院中の患者で、文書と口頭で本研究の目的と意義についての説明をおこない、書面での同意が得られた統合失調症患者 187 名、DSM-IV 診断基準で双極 I 型障害または双極 II 型障害と診断された 178 名および Japanese Genetics Initiative for Drug Abuse [JGIDA] (代表: 氏家 寛) の各サンプル収集施設において収集された覚醒剤依存症患者 143 名である。健常対照者は、精神疾患に関する遺伝子解析研究に自発的意志により参加を表明し、年齢・性別のマッチした 192 名である。さらに覚醒剤依存症サンプルの解析については、症例対照間の地域格差の問題をなくすため、疾患群と地域・性別・年齢をマッチさせた健常対照者を加えている。各対象者から採血した血液より DNA を抽出し、Chromogranin B 遺伝子の各変異部位を PCR-Direct Sequence 法にて増幅、ABI 3100 genetic analyzer にて遺伝子型の判定を行った。

両群の遺伝子型出現頻度をそれぞれ集計し、 2×2 , 2×3 の χ^2 検定を行った。有意水準は $p < 0.05$ とした。なお、本研究は名古屋大学医学部および国立精神・神経センター国府台地区における倫理委員会の審査で承認を得て行っている。

C. 研究結果

統合失調症患者 24 名を用いて Chromogranin B 遺伝子の変異検索をおこなったところ、5'側調節領域において 5 つの変異、さらに Exon4 内に 11 個のアミノ酸置換を伴う変異を見いだした。このうち、Exon4 内の 8 個の変異については既に dbSNP データベースに登録されているものであった。また、我々の見つけた変異のうち 12 個は中国人集団でも確認された。その他の 4 個は我々が新規に同定したものであった。統合失調症患者 187 名、健常対照者約 192 名について関連解析を行った結果、全ての変異の出現頻度は Hardy-weinberg 平衡に矛盾しなかった。中国人集団において、統合失調症との関連の見られた 433G/A, 533A/G について、本研究で用いた日本人集団における、統合失調症との関連は確認できなかった（表 1）。Exon4 内の 4 つの変異 1058G/C, 1104G/A, 1238C/T, 1250G/A, において、アリル頻度、遺伝子型頻度ともに症例対照間で有意な差が見られた（表 2）。1058G/C と 1104G/A は完全に連鎖しており、これら 4 つの変異の間では強い連鎖不平衡が見られた。また、マイクロサテライトマーカー

D20S95 とこれら変異との間にも連鎖不平衡が確認された。日本人集団において統合失調症との関連が認められた 1058G/C, 1104G/A, 1238C/T, 1250G/A 変異について、双極性障害および覚醒剤依存症との関連解析をおこなった結果、それら疾患との関連は認められなかった。

D. 考察

Chromogranin は、種々の神経細胞に分布している可溶性分泌蛋白であり、褐色細胞種や神経内分泌腫瘍においては、カテコールアミン等と共に過剰に分泌されるため、診断の有用なマーカーであると考えられているが、生理的意義についてはまだ不明な点が多い。最近、統合失調症患者において、脳脊髄液中における Chromogranin A および B の有意な減少 (Landen ら, 1999) や海馬歯状回における Chromogranin A 陽性細胞数の有意な減少 (Shibata ら, 2000) が報告されており、今回のわれわれの結果も、Chromogranin B 遺伝子と統合失調症との間に有意な関連を示した。Chromogranin B 遺伝子は自殺を惹起する精神疾患の中でも統合失調症の病態生理に関連する有力な候補遺伝子の一つであると考えられ、現在さらに例数を増やして解析を進めていく予定である。双極性障害および覚醒剤依存症については、症例を増やして再検討するとともに、症例をサブタイプに分けて解析をおこなったり、別の部位についての検討も行っていく予定である。

E. 参考文献

- 1) Kitao Y, Inada T, Arinami T, Hirotsu C, Aoki S, Iijima Y, Yamauchi T, Yagi G (2000) A contribution to genome-wide association studies: search for susceptibility loci for schizophrenia using DNA microsatellite markers on chromosomes 19, 20, 21 and 22. *Psychiatr Genet* 10: 139-143.
- 2) Landen M. *et al.* (1999) Reduction of chromogranin A and B but not C in the cerebrospinal fluid in subjects with schizophrenia. *Eur Neuropsychopharmacol* 9: 311-315.
- 3) Mahata SK, Mahata M, Fischer-Colbrie R, Winkler H (1993) Reserpine causes differential changes in the mRNA levels of chromogranin B, secretogranin II, carboxypeptidase H, alpha-amidating monooxygenase, the vesicular amine transporter and of synaptin/ synaptophysin in rat brain. *Brain Res Mol Brain Res* 19: 83-92.
- 4) Kroesen S, Marksteiner J, Mahata SK, Mahata M, Fischer-Colbrie R, Saria A, Kapeller I, Winkler H (1995) Effects of haloperidol, clozapine and citalopram on messenger RNA levels of chromogranins A and B and secretogranin II in various regions of rat brain. *Neuroscience* 69: 881-891.
- 5) Zhang B *et al.* (2002) Polymorphisms of chromogranin B gene associated with schizophrenia in Chinese Han population. *Neurosci Lett* May 3;323(3):229-33.
- 6) Detera-Wadleigh SD, Badner JA, Yoshikawa T, Sanders AR, Goldin LR, Turner G, Rollins DY, Moses T, Guroff JJ, Kazuba D *et al.* (1997) Initial

genome scan of the NIMH genetics initiative bipolar pedigrees: chromosomes 4, 7, 9, 18, 19, 20, and 21q. *Am J Med Genet.* 74: 254-262.

G. 研究発表

1. 稲田俊也, 樋口輝彦, 上島国利, 中込和幸, 岡島由佳, 三村 將, 磯野 浩, 大坪天平, 山田光彦, 稲本淳子, 岩波 明, 平島奈津子, 篠田淳子, 松尾幸治, 大溪俊幸, 三宮正久, 中川種栄, 西岡玄太郎, 加藤忠史, 山田和夫, 田島 治, 神庭重信, 岡崎祐士, 長沼英俊: Young Mania Rating Scale 日本語版の信頼性についての予備的検討. *臨床精神薬理* 5(4): 425-431, 2002.
2. Furukawa T, Inada T, Adams CE., McGuire H, Inagaki A, Nozaki S: Are the Cochrane group registers comprehensive? A case study of Japanese psychiatry trials. *BMC Medical Research Methodology* 2: 6, 2002.
3. Inada T, Yagi G, Miura S: Extrapyramidal symptom profiles in Japanese patients with schizophrenia treated with olanzapine or haloperidol. *Schizophr Res* 57(2-3): 227-238, 2002.
4. Obata T, Aomine M, Inada T, Kinemuchi H: Nicotine suppresses 1-methyl-4-phenylpyridinium ion (MPP+) induced hydroxyl radical generation in rat striatum. *Neurosci Lett* 330(1): 122-124, 2002.
5. Kokai M, Inada T, Ohara K, Shimizu M, Iwado H, Morita Y: Inter-rater and test-retest reliability of the Japanese version of the subjective deficit syndrome scale. *Hum Psychopharmacol Clin Exp* 18: 145-149, 2003.
6. Inada T, Beasley C, Tanaka Y, Walker D: Extrapyramidal Symptom Profiles Assessed with DIEPSS: Comparison with Western Scales in the Clinical Double-Blind Studies of Schizophrenic Patients Treated with either Olanzapine or Haloperidol. *Int Clin Psychopharmacol* 18(1): 39-48, 2003.

7. Suzuki E, Kitao Y, Ono Y, Iijima Y, Inada T: Cytochrome P450 2D6 Polymorphism and Character Traits. *Psychiatr Genet*, in press.
8. Hori K, Inada T, Tominaga I: "Awakening" in demented patients. *Psychiatr Clin Neurosci*, in press.
9. Inada T, Senoo H, Iijima Y, Yamauchi T, Yagi G: Cytochrome P450IID6 gene polymorphism and the neuroleptic-induced extrapyramidal symptoms in schizophrenic patients. *Psychiatr Genet*, in press.
10. 堀宏治, 千貫 悟, 稲田俊也, 竹下裕行, 平井慎二, 池田正行, 山崎 慶, 富永 格, 織田辰郎, 女屋光基, 堀 一郎, 浅岡俊泰, 金 廣一, 寺元弘, 鹿島晴雄: 塩酸ドネペジルの日常臨床における課題. 周辺症状の観点から. *老年精神医学雑誌* 13(増刊号): 44-48, 2002.
11. Inada T, Nakamura A, Iijima Y: Catechol-O-Methyltransferase (COMT) Polymorphism and Schizophrenia: Possible relation with the treatment-resistant subgroup. *Am J Med Genet (Neuropsychiatr Genet)*, in press.
12. The Japanese Schizophrenia Sib-pair Linkage Group (JSSLG)(Arimami T, Ishiguro H, Minowa Y, Ohtsuki T, Tsujita T, Imamura A, Yoshikawa T, Toyota T, Yamada K, Shimizu H, Yoshitsugu K, Shibata H, Fujii Y, Fukumaki Y, Tashiro N, Inada T, et al.): Initial genome-wide scan for linkage with schizophrenia in the Japanese Schizophrenia Sib-pair Linkage Group (JSSLG) families. *Am J Med Genet (Neuropsychiatr Genet)*, in press.
13. Hori K, Inada T, Sengan S, Ikeda M: Is Charles Bonnet syndrome an early manifestation of dementia? *Acta Neuropsychiatr*, in press.
14. Hori K, Oda T, Tominaga I, Inada T: "Awakenings" in demented patients. *Psychiatr Clin Neurosci*, in press.

表 1 統合失調症とクロモグラニンB遺伝子多型 (433G/A および 533A/G) との
関連

433G/A	GG	GA	AA	n	Gratio	Aratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control (Chinese)	71	70	29	170	62.0%	38.0%		(vs Cont)	0.110		(vs Cont)
(Japanese)	73	90	24	187	63.1%	36.9%			0.963		
Schizo (Chinese)	103	74	16	193	73.0%	27.0%	1.170 - 2.180	1.600	0.600	0.0044	0.016
(Japanese)	77	85	23	185	64.6%	35.4%	0.695 - 1.264	0.937	1.000	0.672	0.878
533A/G	AA	AG	GG	n	Aratio	Gratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control (Chinese)	51	85	34	170	55.0%	45.0%		(vs Cont)	0.900		(vs Cont)
(Japanese)	51	90	47	188	51.1%	48.9%			0.918		
Schizo (Chinese)	94	74	25	193	68.0%	32.0%	1.280 - 2.340	1.730	0.093	0.0017	0.005
(Japanese)	41	100	45	186	48.9%	51.1%	0.689 - 1.223	0.918	0.738	0.559	0.439

表2 統合失調症とクロモグラニンB遺伝子多型 (1058G/C, 1104G/A, 1238C/T, 1250G/A および 1499G/A) との関連

1058G/C	GG	GC	CC	n	Gratio	Cratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control	54	89	49	192	51.3%	48.7%	(vs Cont)		0.801	(vs Cont)	
Schizophrenia	87	82	21	190	67.4%	32.6%	0.381 - 0.684	0.510	0.973	0.000	0.000
Bipolar	51	84	43	178	52.2%	47.8%	0.721 - 1.285	0.963	0.892	0.797	0.955
Control for MAP	69	127	65	261	50.8%	49.2%	(vs Cont)		0.954	(vs Cont)	
MAP user	37	72	43	152	48.0%	52.0%	0.841 - 1.481	1.116	0.900	0.448	0.734
1104G/A	GG	GA	AA	n	Gratio	Aratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control	54	89	49	192	51.3%	48.7%	(vs Cont)		0.801	(vs Cont)	
Schizophrenia	91	76	20	187	69.0%	31.0%	0.352 - 0.637	0.474	0.891	0.000	0.000
Bipolar	51	84	43	178	52.2%	47.8%	0.721 - 1.285	0.963	0.892	0.797	0.955
Control for MAP	71	126	65	262	51.1%	48.9%	(vs Cont)		0.925	(vs Cont)	
MAP user	38	72	43	153	48.4%	51.6%	0.843 - 1.482	1.118	0.900	0.440	0.736
1238C/T	CC	CT	TT	n	Cratio	Tratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control	155	36	1	192	90.1%	9.9%	(vs Cont)		0.821	(vs Cont)	
Schizophrenia	147	38	2	187	88.8%	11.2%	0.724 - 1.832	1.152	0.995	0.550	0.766
Bipolar	142	35	1	178	89.6%	10.4%	0.655 - 1.703	1.056	0.821	0.823	0.974
Control for MAP	212	50	1	263	90.1%	9.9%	(vs Cont)		0.577	(vs Cont)	
MAP user	119	33	1	153	88.6%	11.4%	0.748 - 1.853	1.177	0.819	0.481	0.754
1250G/A	GG	GA	AA	n	Gratio	Aratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control	71	84	37	192	58.9%	41.1%	(vs Cont)		0.670	(vs Cont)	

Schizophrenia	42	98	47	187	48.7%	51.3%	1.132 - 2.011	1.509	0.897	0.005	0.008
Bipolar	60	88	29	177	58.8%	41.2%	0.749 - 1.346	1.004	0.976	0.979	0.502
Control for MAP	95	123	45	263	59.5%	40.5%	(vs Cont)		0.937	(vs Cont)	
MAP user	60	73	20	153	63.1%	36.9%	0.644 - 1.150	0.860	0.970	0.310	0.528
1499G/A	GG	GA	AA	n	Gratio	Aratio	95% CI	Odds ratio	GTvsHW	AF freq	GT freq
Control	160	29	2	191	91.4%	8.6%	(vs Cont)		0.839	(vs Cont)	
Schizophrenia	166	18	2	186	94.1%	5.9%	0.380 - 1.163	0.665	0.754	0.150	0.270
Bipolar	141	25	1	167	91.9%	8.1%	0.547 - 1.582	0.930	1.000	0.789	0.895
Control for MAP	225	34	3	262	92.4%	7.6%	(vs Cont)		0.849	(vs Cont)	
MAP user	131	20	1	152	92.8%	7.2%	0.550 - 1.621	0.944	1.000	0.843	0.887

Relationship between catechol-O-methyltransferase polymorphism and treatment-resistant schizophrenia

Toshiya Inada¹⁾, Ataru Nakamura²⁾, Yoshimi Iijima³⁾

1) Department of Psychiatry and Psychobiology, Nagoya University, Graduate School of Medicine, Nagoya, Japan; 2) Inokashira Hospital, Mitaka-shi, Tokyo, Japan; 3) National Institute of Mental Health, National Center of Neurology and Psychiatry, Chiba, Japan

Abstract: Catechol-O-methyltransferase (COMT) plays a crucial role in the regulation of central dopaminergic systems. We examined the allelic association of a functional polymorphism of the COMT gene with the clinical manifestations and the response to antipsychotics of 100 schizophrenic patients and 201 healthy controls from the general Japanese population. No statistically significant difference was observed in the allele and genotype frequencies between the schizophrenic patients and the healthy controls. The daily neuroleptic dosage that patients received during their maintenance therapy was significantly higher in patients with the L/L genotype than in the other patients ($p < 0.05$). The present results suggest that the presence of the COMT genotype does not help in evaluating the susceptibility to the development of schizophrenia, but that it may help in the estimation of treatment-resistant features of schizophrenia.

Key words: COMT, dopamine, antipsychotic drug, extrapyramidal symptom, gene

INTRODUCTION

Polymorphisms at gene loci related to central dopaminergic systems have been extensively examined as candidate loci for schizophrenia because antipsychotic drugs, which act as dopamine antagonists, are effective in the treatment of psychiatric symptoms seen in schizophrenic patients.

Although the results are still controversial, a number of studies have shown a positive association between schizophrenia as a whole or subgroups of this disease with some specific polymorphic sites of the gene loci related to central dopaminergic transmission. However, whether specific mutations in the dopamine-related genes can contribute significantly to the clinical features of schizophrenia still remains to be demonstrated.

Catechol-O-methyltransferase (COMT) metabolizes catecholamines such as dopamine, and catechol drugs such as L-DOPA. The COMT

gene, which lies on the cytogenetic band of 22q11.2 on chromosome 22, has been considered to be one of the candidate genes for schizophrenia because it inactivates dopamine. There is an amino acid alteration that determines the activity of the COMT enzyme (Lotta et al., 1995; Lachman et al., 1996; Sander et al., 1997). It has been shown that a G-A transition exists at codon 158 of the COMT gene that results in a valine-to-methionine substitution. The association with this functional polymorphism has been examined extensively in the general population of schizophrenic patients or in subgroups with various clinical features. For example, relationships with this polymorphism have been reported in schizophrenia (Ohmori et al., 1998), violence in patients with schizophrenia or schizoaffective disorders (Lachman et al., 1998), homicidal schizophrenia (Kolter et al., 1999), suicidal behavior in patients with schizophrenia (Nolan et al., 2000) and aggressive behavior in

patients with schizophrenia (Jones et al., 2001). This polymorphism is reportedly associated with specific subgroups of various psychiatric diseases such as type-I alcoholism (Tiihonen et al., 1999), higher alcohol consumption (Kauhanen et al., 2000), delirium tremens in alcoholics (Nakamura et al., 2001), polysubstance abuse (Vandenberg et al., 1997), obsessive-compulsive disorder (Karayiorgou et al., 1997), rapid-cycling mood disorder (Papolos et al., 1998), female bipolar disorder (Mynett-Johnson et al., 1998), and depressive disorder (Ohara et al., 1998). However, there are also some reports of a lack of significant association with schizophrenia (Daniels et al., 1996; Strous et al., 1997; Wei et al., 1999; de Chaldee et al., 2001; Liou et al., 2001), affective disorders (Kunugi et al., 1997) and alcoholics (Ishiguro et al., 1991), suggesting that the exact role of COMT polymorphism in determining the phenotype in terms of clinical psychiatry and human behavior is not yet fully elucidated.

The present study examined the relationship between COMT and schizophrenia in the Japanese population, focusing on the allelic association of COMT polymorphism with the clinical manifestations and antipsychotic features of schizophrenia.

METHODS

Ethical considerations: This study had the approval of the ethics committee of the Kohnodai Area, National Center of Neurology and Psychiatry, Chiba, Japan. Written informed consent was obtained from all subjects who participated.

Subjects: The subjects for the present study were chronic inpatients who met the criteria of the DSM-III-R diagnosis (American Psychiatric Association, 1987) for schizophrenia, who had been hospitalized and had been receiving antipsychotic therapy for at least one year during their current hospitalization. Volunteer control subjects were mostly medical staff with no history of psychosis or substance abuse, or of receiving antipsychotic medication. All subjects were Japanese and had Japanese parents.

Status of extrapyramidal symptoms: Extrapyramidal symptoms (EPS) induced by antipsychotic medication were basically evaluated using DIEPSS (Inada et al., 2002): we assumed that acute EPS had been present within 3 months of the initial neuroleptic therapy if the clinical records clearly described EPS and a subsequent reduction in the neuroleptic dosage or a

subsequent addition of antiparkinsonian drugs. Patients who showed no signs of acute EPS in spite of receiving neuroleptic therapy for more than 6 months were regarded as not having acute EPS. Tardive dyskinesia (TD) was assessed with the Japanese version of the Abnormal Involuntary Movement Scale (AIMS). The diagnosis of TD was made according to the criteria of Schooler and Kane (1982). The inclusion criteria used to subclassify patients with or without TD were those reported by Inada et al. (1997). Briefly, patients who had been suffering from TD with at least one item persistently rated 3 on the AIMS for more than 1 year or patients who had been suffering from persistent TD for more than 1 year that had developed within 5 years after the first neuroleptic exposure were classified as having TD. Patients who had never developed TD despite receiving neuroleptics for more than 10 years were regarded as not having TD. In rating EPS and TD, suspected cases were videotaped for later evaluation. When information about the presence of acute EPS in the initial stage of neuroleptic therapy for the first episode of schizophrenia was incomplete, the patient was excluded from the acute EPS study. When the TD status could not be obtained, the patient was omitted from the TD study.

Neuroleptic dosage: Information about the neuroleptic therapy that schizophrenic patients had been receiving was obtained from their clinical records. The daily neuroleptic dosage was calculated from the most recent 1-year neuroleptic prescription history. The chlorpromazine-equivalent daily dose administered to each patient was calculated from a table developed specifically for Japanese patients (Inagaki et al., 1998).

Definition of treatment-resistant schizophrenia: Schizophrenic patients were diagnosed as having treatment-resistant schizophrenia (TRS) when they had been hospitalized for more than 1 year and had been receiving antipsychotic therapy at dosages of at least 1,000 mg/day chlorpromazine equivalents for more than 1 year.

Experimental procedure: Genomic DNA was extracted from samples of whole blood obtained from the subjects. The COMT polymorphism is generated by the presence of a G or A encoding a valine or methionine at codon 158. The 210-bp target segment was amplified by the polymerase chain reaction (PCR) method according to the standard protocol (Daniels et al., 1996), using the primers 5'-ACT GTG GCT ACT CAG CTG TG and 5'-CCT TTT TCC AGG TCT GAC AA. The

Table 1. Genotype and allele frequencies of catechol-O-methyltransferase H/L polymorphism in schizophrenic patients and controls

Group	Genotype distribution			<i>p</i> value (vs controls)	Allele frequency		<i>p</i> value (vs controls)
	H/H	H/L	L/L		H	L	
Controls (<i>n</i> =201)	43.8%	49.8%	6.5%	-----	68.7%	31.3%	-----
Schizophrenia (<i>n</i> =100)	41.0%	51.0%	8.0%	0.88	66.5%	33.5%	0.59
Delusion and hallucination (<i>n</i> =64)	51.6%	43.8%	4.7%	0.53	73.4%	26.6%	0.31
Bizarre behavior (<i>n</i> =70)	50.0%	44.3%	5.7%	0.67	72.1%	27.9%	0.44
Disorganization (<i>n</i> =72)	44.4%	51.4%	4.2%	0.77	70.1%	29.9%	0.74
Negative symptoms (<i>n</i> =54)	55.6%	38.9%	5.6%	0.30	75.0%	25.0%	0.20
Positive first-degree family history (<i>n</i> =42)	47.6%	45.2%	7.1%	0.87	70.2%	29.8%	0.78
Onset at 21 years or younger (<i>n</i> =45)	44.4%	48.9%	6.7%	0.99	68.9%	31.1%	0.97

For comparison between controls and schizophrenic patients as a whole or their subgroups listed in this table, two-tailed chi-squared tests for 2×2 and 2×3 contingency tables were performed.

PCR products were digested with 5 units of *Nla* automated and manual staining of DNA separated on polyacrylamide gels with a DNA silver staining kit (Pharmacia Biotech, Tokyo, Japan).

Statistics: Allele and genotype frequencies were compared by using the chi-squared test for 2×2 and 2×3 contingency tables. The association between neuroleptic-induced EPS (the status of TD and acute EPS) and the polymorphism was also assessed by the chi-squared test. Comparison of the daily neuroleptic dosage between the L/L and the other genotypic subgroups was performed using the Mann-Whitney U test. The probability values are listed in the tables. The computer package SPSS for Windows (release 11.0J, SPSS Japan, Tokyo, Japan) was used for the statistical analyses.

RESULTS

The COMT genotype (H/H, H/L, L/L) and allele frequency in schizophrenic subjects are shown in Table 1. No significant differences in allele frequencies were observed between the healthy controls and whole schizophrenic patients or any of the subgroups classified according to the psychiatric symptoms seen at the patient's first episode.

Table 2 shows the genotype and allele

III for 2 h at 37°C, and electrophoresed using frequencies of the COMT H/L polymorphism and the status of TD and acute EPS in schizophrenic patients. No significant differences were observed in the susceptibility to neuroleptic-induced extrapyramidal side effects among three subgroups of the COMT genotype.

The characteristics of neuroleptic treatment among the three subgroups showing COMT H/L polymorphism are shown in Table 3. The daily neuroleptic dosage received during maintenance therapy was significantly higher in patients with the L/L genotype than in the other patients (Mann-Whitney U test, $z = -2.248$, $p = 0.0246$). The rate of TRS tended to be higher in patients with the COMT L/L genotype than in the other patients, although this is a marginal difference statistically expressed as a significant trend level (chi-squared = 3.782, $df = 1$, $p = 0.052$). The odds ratio of L/L for TRS was 4.392 (95% confidence interval: 0.894–21.588).

DISCUSSION

In the present study, no significant association was observed between COMT polymorphism and schizophrenic patients as a whole or any of the subgroups classified according to the psychiatric symptoms seen at the

Table 2. Genotype and allele frequencies of Catechol-O-Methyltransferase H/L polymorphism and the status of TD and acute EPS in schizophrenic patients

Group	Genotype			p value	Allele frequency		p value
	H/H	H/L	L/L		H	L	
<i>Status of TD</i>							
Patients with TD (n=33)	42.4%	51.5%	6.1%	0.79	68.2%	31.8%	0.60
Patients without TD (n=46)	34.8%	58.7%	6.5%		64.1%	35.9%	
<i>Status of acute EPS</i>							
Acute EPS present (n=30)	43.3%	50.0%	6.7%	0.67	68.3%	31.7%	0.44
Acute EPS absent (n=45)	33.3%	57.8%	8.9%		62.2%	37.8%	

For comparison of the status of TD and acute EPS, two-tailed chi-squared tests for 2x2 and 2x3 contingency tables were performed. Abbreviations: TD, tardive dyskinesia; EPS, extrapyramidal symptom.

patient's first episode, suggesting that this polymorphism is unlikely to play an essential role in the development of schizophrenia. The lack of association with the general population of schizophrenic subjects is consistent with previous reports of French (de Chaldee et al., 2001), Taiwanese (Liou et al., 2001), Turkish (Herken and Erdal, 2001), and Caucasian (Wei et al., 1999) schizophrenic populations, but contradictory to the report of Ohmori et al. (1998). Although most of the studies failed to detect significant relationships between this polymorphism and schizophrenic subjects as a whole (Chen et al., 1999; Semwal et al., 2001), relationships with this polymorphism have been reported in certain subgroups of schizophrenia (Herken and Erdal, 2001; Liou et al., 2001). Recently, Egan et al. (2001) reported that the COMT low-activity genotype slightly increases the risk for schizophrenia by its effect of increasing prefrontal dopamine catabolism.

Here we have used a definition of TRS that is modified from that proposed by Kane et al. (1988), which was defined based on observations of monotherapy antipsychotic treatment. The main reason for modifying the definition of TRS is that Japanese psychiatrists generally prefer polypharmacy in routine clinical practice: if patients relapse during their antipsychotic maintenance therapy, Japanese psychiatrists often prescribe

another antipsychotic agent in addition to the original one, instead of switching to it, so that it is quite rare for patients in Japanese routine psychiatric practice to be diagnosed as having TRS as proposed by Kane et al. (1988), even when the schizophrenia is considered treatment-resistant. In the present study, a significantly higher daily neuroleptic dosage was observed in patients with the L/L genotype (average: 1,226 mg/day) than in those with the other genotypes. In addition, the rate of TRS tended to be higher in patients with the COMT L/L genotype than in the other patients. These findings suggest that the low-activity COMT genotype is more common in TRS. This is consistent with the results of Herken and Erdal (2001), who reported that Turkish schizophrenics with the L/L genotype may have clinical signs that are much more severe. Here we assumed that the patients with persistent and severe condition of psychoses usually receive high doses of antipsychotics for the long period. Base on this assumption, we did not examine the direct observation of 'current and persistent positive symptoms of psychosis and at least moderate overall severity of current illness' in the present study, although they are included in the Kane's criteria of TRS. Therefore, this is a limitation of this study: some of the patients who were identified as TRS in this study might contain the

Table 3. Characteristics of neuroleptic treatment among three subgroups showing catechol-O-methyltransferase H/L polymorphism

	Catechol-O-methyltransferase H/L polymorphism		
	H/H	H/L	L/L
Daily neuroleptic dosage (mg / day)	728 ± 655	685 ± 621	1,226 ± 1,069*
(n=78)	(n=32)	(n=40)	(n=6)
Rate of treatment-resistant cases	25.0%	22.0%	57.1%

Data are expressed as mean ± standard deviation. Daily neuroleptic dosage was calculated from the most-recent 1-year neuroleptic therapy as chlorpromazine equivalent dose using the table of Inagaki et al (1998). *p<0.05 when compared to the (H/H + H/L) subgroups (Mann-Whitney U tests).

mere chronic schizophrenic patients with poor positive symptoms (Conley et al., 1997), although they had been hospitalized for more than 1 year and had been receiving high doses of antipsychotic therapy for more than 1 year.

As for the susceptibility to neuroleptic-induced EPS, no significant differences were observed among three subgroups of the COMT genotype. If the low-activity COMT genotype contributed to the high dopamine concentration in the synaptic cleft due to the decrease in COMT activity, then EPS would be less likely to develop in patients with the COMT L/L genotype. On the other hand, the patients with the COMT L/L genotype are observed to receive significantly higher doses of antipsychotics in the present study, which suggests that EPS would be more likely to develop in these patients. Our present results of no differential characteristics of neuroleptic-induced EPS among three subgroups of the COMT genotype may be the consequence of offset of these two factors discussed above. To clarify the exact role of the COMT H/L polymorphism for the susceptibility to neuroleptic-induced EPS, the comparison should be done under the condition that the neuroleptic dosage patients receive is strictly controlled.

Since the COMT H/L polymorphism examined in the present study has been demonstrated to alter enzyme activity (Lotta et al., 1995; Lachman et al., 1996; Sander et al., 1997), the polymorphism is considered to affect the clinical manifestations related to central dopaminergic systems. However, it should be noted that the present conclusion was drawn

mainly from the findings in only eight patients with the COMT L/L genotype, which represents a small sample size. Further research using a larger sample set is required to clarify the exact role of this polymorphism; that is, whether alteration of enzyme activity could modify the phenotype of various psychiatric diseases and human behavior, including the characteristics of TRS suggested in this study.

REFERENCES

- American Psychiatric Association. 1987. Diagnostic and Statistical Manual of Mental Disorders. 3rd rev. Ed. American Psychiatric Association, Washington DC.
- Chen CH, Lee YR, Chung MY, Wei FC, Koong FJ, Shaw CK, Yeh JI, Hsiao KJ. 1999. Systematic mutation analysis of the catechol O-methyltransferase gene as a candidate gene for schizophrenia. *Am J Psychiatry* 156: 1273-1275.
- Conley RR, Buchanan RW. 1997. Evaluation of treatment-resistant schizophrenia. *Schizophr Bull* 23: 663-674.
- Daniels JK, Williams NM, Williams J, Jones LA, Cardno AG, Murphy KC, Spurlock G, Riley B, Scambler P, Asherson P, McGuffin P, Owen MJ. 1996. No evidence for allelic association between schizophrenia and polymorphism determining high or low catechol-O-methyltransferase activity. *Am J Psychiatry* 153: 268-270.
- de Chaldee M, Corbex M, Campion D, Jay M, Samolyk D, Petit M, Thibaut F, Laurent C, Mallet J. 2001. No evidence for linkage