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considerations may explain the large differences among five independent *FOB1* transformants that were derived from the same strain (e.g., mutant B) and had undergone the same transformation and subsequent subcultures (Fig. 4B, mutant B).

In passing, we note that transformants of mutant G, which received FOB1, were able to expand rDNA repeats, although apparently not to the same extent as the control FOB1 transformants. The resultant strain lacks segment G, which was originally defined as the Pol I enhancer (6), in the expanded rDNA repeats except for the single copy at the leftmost end. However, this strain was able to form colonies on glucose plates and to lose the helper plasmid. Such a strain with rDNA repeats carrying mutation G and without the helper plasmid showed only a small decrease in growth rate in glucose medium compared to the control strain with the intact enhancer in all the rDNA repeats. The role of the enhancer element in Pol I transcription is a separate subject under current study.

Relationship between rDNA repeat expansion and recombination by HOT1. The HOT1 element stimulates recombination between two nearby repeat sequences at a chromosome site outside the rDNA locus. HOTI consists of two elements, the I element, which corresponds to the Pol I promoter, and the E element, which comprises segments F and G studied here. It has been assumed that HOT1 activity is responsible for recombinational events within rDNA repeats. The discovery that FOB1 is required for both HOT1 activity (20) and rDNA repeat expansion and contraction (18) has appeared to support this assumption. However, HOT1 activity requires active transcription by Pol I (15, 35) whereas recombinational events within rDNA repeats take place in the absence of their transcription (18). In addition, the present work has demonstrated clear differences in the cis elements required for stimulation of recombination between the two systems. First, segments C, D, and E are required for rDNA repeat expansion (see above) but not for HOT1 activity (35). Second, deletion (or substitution) of segment G abolishes HOT1 activity nearly completely (35) but reduces the extent and presumably the rate of rDNA repeat expansion only weakly (see above). (It should be noted that there is one copy of the intact G segment at the left border in mutant G used in the expansion experiments described in this paper. Thus, although we think it rather unlikely, we cannot eliminate the possibility that this single copy might play a role in recombination events responsible for repeat expansion.) The main features shared by the two systems are the requirement of segment F, which contains the RFB site, and the requirement of the intact FOB1 gene as mentioned above. Thus, the previous assumption may be incorrect and elucidation of the mechanisms of rDNA sequence homogenization as well as rDNA repeat expansion and contraction may have to depend on the use of systems designed within the native rDNA repeat locus. In addition to the present FOB1-induced repeat expansion system, we have previously described experimental systems in which the effects of various factors on the expansion and contraction of rDNA repeats can be studied (18, 25). These systems should be useful in studies not only of the mechanism but also of the physiological significance of rDNA repeat expansion and contraction.

After completion of the present work, a paper by Ward et al. (37) appeared, which has demonstrated that HOT1 activity can occur in the absence of replication fork blocking, even though both HOT1 and RFB activities requires FOB1. These workers also carried out mutational analysis within the F and G segments and found that some DNA elements are shared but others are required for one activity but not for the other. Thus, their conclusion that the FOB1 function is involved in two clearly different activities, HOT1 and RFB activities, is related to our conclusion that it is also required for two clearly separable activities, HOT1 and rDNA repeat expansion. Elucidation of the function(s) of the FOB1 gene product appears to be a key to solving the intriguing problem of relationships among these three activities. In addition, consideration of these new observations made by Ward et al. (37) and by the present study raises the question whether our previous proposal is really correct, that is, whether replication fork blocking is really the first step in rDNA expansion and contraction. Although available experimental results support this proposal, they have not proven it. Detailed mutational analysis of DNA sequence elements within the F segment may be helpful to settle this question. Regardless of the answer to this question, however, the discovery of the new DNA elements that are uniquely involved in rDNA repeat expansion (and presumably also in contraction) indicates the presence of an unexplored aspect(s) of recombinational mechanisms used in rDNA repeat structures that constitute the structurally and functionally essential component of the nucleolus.

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ADDENDUM IN PROOF

We replaced the G segment, still located at the left border of rDNA repeats in mutant G. In this mutant, the FOBI-dependent expansion of rDNA took place as well. Therefore, the G segment was not required for the expansion.

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