

retrieve antigenicity based on the modified method of Shi et al<sup>19</sup>. Endogenous peroxidase activity was blocked by treatment with 3 % hydrogen peroxide. After treatment with 10 % goat serum, the sections were incubated with each primary antibody for 48 hr at 4°C. The sections were then incubated with the biotinylated anti-mouse immunoglobulin (Vector Lab.) followed by incubation with the avidin-biotin-peroxidase complex (Vector Lab.). Antibody labeling was visualized by incubation with diaminobenzidine with nickel ammonium sulfate. Normal adult human tonsil, normal adult and postnatal 10-days-old rat were used as positive controls.

*Evaluation of immunostaining.* For semiquantitative evaluation of immunohistochemical staining, staining intensity was scored on an arbitrary four-point scale: 0, no immuno-positive staining; 1+, weak immuno-positive staining intensity; 2+, moderate immuno-positive staining intensity; 3+, strong immuno-positive staining intensity. We counted the cells which were stained in the 2+ and 3+ degree as immuno-positive cells. The number of melanized nigral neurons was counted in one of the midbrain sections at the level of the oculomotor nucleus on each patient and control subject. The substantia nigra was arbitrary divided into three approximately equal parts, i.e., the medial, central and the lateral part<sup>20</sup> and the percentage of neurons

immuno-positive for Bcl-xL, Bcl-2 or Bax among the melanized neurons was calculated at each compartment. Statistical analysis was performed using the Mann-Whitney U-test.

*Double-labeling; Bax with Bcl-xL or Bcl-2.* Co-localization of Bax and Bcl-xL or Bcl-2 was analyzed by a confocal laser-scanning microscope (BIO-RAD microscopy division; MRC-1024) using antibodies labeled by rhodamine-avidin complex or fluoresceine-streptoavidine complex.

**Results.** *Bcl-xL, Bcl-2 and Bax immunohistochemistry in CTL subjects.* Bcl-xL and Bcl-2 were expressed in approximately half of the melanized nigral neurons (Fig. 1a, 1c) whereas Bax was expressed in most of the nigral neurons (Fig. 1e). Non-melanized nigral neurons also expressed Bcl-xL, Bcl-2, and Bax. These immunostaining was observed mainly in the cytosol, but Bcl-xL and Bax were also expressed in the nuclear membrane of some neurons. Bcl-xL, Bcl-2, and Bax were also expressed in most of the neurons in the oculomotor nucleus, dorsal tegmental nucleus, and the peripeduncular nucleus, but not in the neurons in the red nucleus and the superior colliculus. Some glia cells, endothelial cells, and vessel walls were positive for Bcl-xL, Bcl-2, and Bax. Pre-adsorption of antibodies studied abolished the immunostaining.

*Bcl-xL, Bcl-2, and Bax immunohistochemistry in PD patients.* Bcl-2 was expressed in approximately half of the melanized

nigral neurons (Fig. 1d) and Bax was expressed in most of the nigral neurons (Fig. 1f) as in the CTL subjects.

On the other hand, Bcl-xL was expressed in only one-fourth of the remaining nigral neurons as will be shown in the semiquantitative analysis later (Fig. 1b). The intensity of immunostaining of Bcl-xL-positive neurons itself was essentially similar to that of the control subjects. Both atrophic and apparently normal neurons showed similar immunostaining intensity. Lewy bodies were negative for Bcl-xL, Bcl-2, and Bax. No relationship was noted between the presence of Lewy bodies and expression of Bcl-xL, Bcl-2, or Bax in individual neurons. The glial expression of Bcl-xL and Bax was apparently increased in the SN of PD patients compared with the CTL subjects, but not in other midbrain areas. Expressions of Bcl-xL, Bcl-2, and Bax in other midbrain areas were essentially similar to those of the CTL subjects.

*Semiquantitative study on Bcl-xL, Bcl-2 and Bax.* In the CTL subjects, the percentages of immuno-positive nigral melanized neurons for Bcl-xL were  $51.6 \pm 9.9\%$  in the medial part,  $44.3 \pm 7.1\%$  in the central part, and  $40.4 \pm 7.1\%$  in the lateral part, whereas in the PD patients, the percentages were  $27.8 \pm 7.4\%$ ,  $22.7 \pm 6.1\%$ , and  $20.0 \pm 6.6\%$ , respectively. The number of immuno-positive neurons was significantly smaller in the medial and the lateral parts of the SN in PD (Fig. 2b; Mann-Whitney U-test;  $p < 0.05$  and  $p < 0.02$ , respectively). When the three parts of the SN were combined, Bcl-xL-positive neurons were  $46.0 \pm 5.9\%$  in the CTL and  $22.5 \pm 5.5\%$  in the PD patients (Fig. 2b; Mann-Whitney U-test;  $p < 0.02$ ).

The percentages of Bcl-2-positive neurons in the CTL were  $62.6 \pm 14.2\%$  in the medial part,  $59.4 \pm 15.6\%$  in the central part, and  $70.4 \pm 12.2\%$  in the lateral part, where as in the PD, they were  $64.3 \pm 9.5\%$ ,  $47.0 \pm 13.0\%$  and  $58.7 \pm 10.3$ , respectively; no essential difference was noted between the two groups (Fig. 2c). When the three parts were combined, Bcl-2 positive neurons were  $61.9 \pm 14.7\%$  in the CTL and  $59.7 \pm 10.3\%$  in the PD (Fig. 2a). The percentages of Bax-positive neurons in the CTL were  $91.0 \pm 9.9\%$  in the medial part,  $88.3 \pm 3.9\%$  in the central part, and  $92.3 \pm 1.5\%$  in the lateral part , where as in the PD, they were  $95.6 \pm 3.1\%$ ,  $89.7 \pm 5.0\%$ , and  $92.2 \pm 5.9\%$ ; no significant difference was noted between the two groups (Fig. 2d). When the three parts were combined, Bax-positive neurons were  $90.5 \pm 2.6\%$  in the CTL and  $92.5 \pm 4.2\%$  in the PD (Fig. 2a).

*Double-labeling; Bax with Bcl-xL or Bcl-2.* Figure 3 illustrates confocal lazer images of immuno-fluorescence for Bcl-xL(Fig. 3a) and Bax (Fig. 3b), and Bcl-2 (Fig. 3c) and Bax (Fig. 3d) in the SN of PD patients. Five patterns of immunostaining were detected in nigral neurons, i.e., neurons expressing only Bcl-xL, Bcl-2, or Bax, neurons expressing Bcl-xL and Bax, and Bcl-2 and Bax. Co-localization of Bcl-xL and Bcl-2 was not tested. No significant difference was note in the overall co-localization pattern between the CTL and the PD.

**Discussion.** We showed that immunostaining for Bcl-xL, but not Bcl-2 or Bax, is down-regulated in the SN neurons of PD patients compared to CTL subjects. According to the literature, Bcl-xL and Bcl-2 are expressed in a

large population of neurons during embryonic development and that Bcl-2 expression is down-regulated during maturation while that of Bcl-xL is retained in the adult CNS<sup>21-23</sup>. Therefore, Bcl-xL may play more important role than Bcl-2 in the survival of neurons in the adult CNS<sup>24</sup>. Although limitations are present in the analysis of post-mortem human specimens, down-regulation of Bcl-xL expression in PD in our study suggests increased vulnerability of nigral neurons for neuronal death by apoptosis. As Bcl-2 and Bax expression in PD did not change significantly from the CTL, the down regulation of Bcl-xL does not appear to be secondary to neuronal degeneration or loss of mitochondria; Bcl-xL, Bcl-2 and Bax are expressed on mitochondrial membrane. The down regulation of Bcl-xL may be either due to decreased synthesis or increased degradation.

Apoptosis appears to play a critical role in neuronal death in many neurodegenerative disorders including Alzheimer's disease (AD)<sup>25</sup>, Huntington disease<sup>26</sup>, PD<sup>3-4</sup>, hereditary spinocerebellar degenerations<sup>27</sup>, and ALS<sup>28</sup>. Our group<sup>4</sup>, Anglade et al.<sup>2</sup>, and Tatton et al.<sup>3</sup> reported evidence to indicate apoptosis in nigral neurons in PD TUNEL or electronmicroscopy. However, recently, Kösel et al.<sup>7</sup> and Banati et al.<sup>29</sup> reported serious doubt on the existence of apoptosis in nigral neurons in PD. Such controversies may in part be due to the lack of appropriate methods to explicitly demonstrate apoptosis in a small autopsy material like SN.

Another approach to address the question of apoptosis is the analysis of Bcl-2 family proteins. Mogi et al. reported higher level of Bcl-2 in the striatum<sup>15</sup> and SN (Mogi personal communication) in PD by enzyme-linked immunosorbant assay (ELISA). But in our study, neuronal expression of Bcl-2 was not increased in PD. Vyas et al. also reported

normal bcl-2 mRNA expression in nigral neurons of PD patients<sup>30</sup>. This discrepancy can be explained by increase in the glial expression of Bcl-2 as a whole.

Studies on Bcl-xL in human brains are limited; Shoma et al.<sup>18</sup> and Krajewski et al.<sup>31</sup> reported Bcl-xL expression in some Purkinje cells. Drache et al.<sup>24</sup> reported lower levels of Bcl-xL expression in AD brains by Western blotting but not by immunohistochemistry. We found significant down regulation of Bcl-xL expression in neurons in the lateral and the central parts; the areas which are more vulnerable in PD<sup>20</sup>.

The relationship between up- or down-regulation of anti- and pro-apoptotic proteins and the degree of neurodegeneration is not simple. In Alzheimer brains, neurons exhibiting DNA fragmentation were associated with the up-regulation of Bcl-2 expression, whereas neurons exhibiting neurofibrillary tangles showed down-regulation of Bcl-2 expression<sup>32</sup>. On the other hand, Su et al.<sup>33</sup> and MacGibbon et al.<sup>34</sup> showed positive correlation between Bax up-regulation and tangle formation in AD. In progressive supranuclear palsy, Tortosa et al.<sup>16</sup> reported normal Bcl-2 and Bax immunoreactivities in the SN neurons.

It has been postulated that the ratio of Bax to Bcl-xL and Bcl-2 plays a critical role in the regulation of the relative vulnerability to apoptotic cell death<sup>35</sup>. On the other hand, according to Cheng et al.<sup>36</sup> and Knudson et al.<sup>37</sup> Bax, Bcl-xL and Bcl-2 are able to regulate apoptosis independently. Our double-labeling study suggests that the interaction with may not be necessary for Bcl-xL to exert its anti-apoptotic effect, and that down regulation of Bcl-xL itself could promote apoptosis in the PD patients.

In conclusion, we showed down regulation of Bcl-xL in the SN of PD patients. This did not appear to be secondary to neuronal degeneration, as expressions of Bcl-2 and Bax were essentially normal. Further studies on the synthesis and degradation of Bcl-xL appear to contribute to the elucidation of nigral neuronal death in PD. Furthermore our results warrant studies on the genetic polymorphism of the Bcl-xL gene in comparison with Bcl-2 and Bax gene polymorphisms to explore genetic risk factors for PD.

(2193 words)

## References

1. Mochizuki H, Goto K, Mori H, Mizuno Y. Histochemical detection of apoptosis in Parkinson's disease. *J Neurol Sci* 1996;137:120-123.
2. Anglade P, Vyas S, Javoy-Agid F et al.. Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histol Histopathol* 1997;12:25-31.
3. Tatton NA, Maclean-Fraser A, Tatton GW, Perl PD, Olanow WC. A fluorescent double-labeling method to detect and confirm apoptotic nuclei in Parkinson's disease. *Ann Nuerol* 1998; 44:S142-148.
4. Loo DT, Copani A, Pike CJ, Whittemore ER, Walencewicz AJ, Cotman CW. Apoptosis is induced by beta-amyloid in cultured central nervous system neurons. *Proc Natl Acad Sci U S A* 1993;90:7951-7955.
5. Dragunow M, Faull RL, Lawlor et al.. In situ evidence for DNA fragmentation in Huntington's disease striatum and Alzheimer's disease temporal lobes. *Neuroreport* 1995;6:1053-1057.
6. Troost D, Aten J, Morsink F, de Jong JM. Apoptosis in ALS is not restricted to motoneurons: Bcl-2 expression is increased in post-central cortex, adjacent to the affected motor cortex. *J Neurol Sci* 1995;129 Suppl:79-80.
7. Kösel S, Egensperger R, Eitzen UV, Mehraein P, Graeber MB. On the question of apoptosis in the parkinsonian substantia nigra. *Acta Neuropathol* 1997;93:105-108.



8. Vaux DL, Cory S, Adams JM. Bcl-2 gene promotes haemopoietic cell survival and cooperates with c-myc to immortalize pre-B cells. *Nature* 1988;335:440-442.
9. Graeber TG, Osmanian C, Jacks T et al.. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumors. *Nature* 1996;379:88-91.
10. Hockenbery DM, Oltvai ZN, Yin XM, Milliman CL, Korsmeyer SJ. Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell* 1993;75:241-251.
11. Offen D; Beart PM; Cheung NS; Pascoe CJ; Hochman A; Gorodin S; Melamed E; Bernard R; Bernard O. Transgenic mice expressing human Bcl-2 in their neurons are resistant to 6-hydroxydopamine and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity. *Proc Natl Acad Sci USA* 1998 12;95:5789-5794.
12. Blomer U, Krafri T, Randolph-Moore L, Verma IM, Gage FH. Bcl-xL protects adult septal cholinergic neurons from axotomized cell death. *Proc Natl Acad Sci USA* 1998;95:2603-2608.
13. Clarke MF, Apel IJ, Benedict MA et al.. A recombinant bcl-x s adenovirus selectively induces apoptosis in cancer cells but not in normal bone marrow cells. *Proc Natl Acad Sci USA* 1995;92:11024-11028.
14. Bredesen ED. Neural Apoptosis. *Ann Neurol* 1995;38:839-851.
15. Mogi M, Harada M, Kondo T et al.. bcl-2 protein is increased in the brain from parkinsonian patients. *Neurosci Lett* 1996;215:137-139.

16. Tortosa A, Blanco R, Terrer I. Bcl-2 and Bax protein expression in neurofibrillary tangles in progressive supranuclear palsy. *NeuroReport* 1998;9:1049-1052.
17. Mizuguchi M, Sohma O, Takashima et al.. Immunohistochemical and immunohistochemical localization of Bcl-x protein in the rat central nervous system. *Brain Res* 1996;712:281-286.
18. Sohma O, Mizuguchi M, Takashima S, Yamada M, Ikeda K, Ohta S. High expression of Bcl-x protein in the developing human cerebellar cortex. *J Neurosci Res* 1996;43:175-182.
19. Shi S, Key EM, Kalra K. Antigen retrieval in formalin-fixed, paraffin-embedded tissues: an enhancement method for immunohistochemical staining based on microwave oven heating of tissue sections. *J Histochem Cytochem* 1991;39:741-748.
20. Gibb WR, Lees AJ. Anatomy, pigmentation, ventral and dorsal subpopulations of the substantia nigra, and differential cell death in Parkinson's disease. *J Neurol Neurosurg Psychiatry* 1991;54:388-396.
21. Gonzalez Garcia M, Garcia I et al.. bcl-x is expressed in embryonic and postnatal neural tissues and functions to prevent neuronal cell death. *Proc Natl Acad Sci U S A* 1995;92:4304-4308.
22. Migheli A, Cavalla P, Piva R, Giordana MT, Schiffer D. bcl-2 protein expression in aged brain and neurodegenerative diseases. *Neuroreport* 1994;5:1906-1908.

23. Frankowski H, Missotten M, Fernandez PA et al.. Function and expression of the Bcl-x gene in the developing and adult nervous system. *Neuroreport* 1995;6:1917-1921.
24. Drache B, Diehl GE, Beyreuther K, Perlmutter LS, Konig G. Bcl-x1-specific antibody labels activated microglia associated with Alzheimer's disease and other pathological states. *J Neurosci Res* 1997;47:98-108.
25. Sugaya K, Reeves M, McKinney M. Topographic associations between DNA fragmentation and Alzheimer's disease neuropathology in the hippocampus. *Neurochem Int* 1997;31:275-281.
26. Stadelmann C, Bruck W, Bancher C, Jellinger K, Lassmann H. Trinucleotide (CAG) repeat length is positively correlated with the degree of DNA fragmentation in Huntington's disease striatum. *Neuroscience* 1998;87:49-53.
27. Igarashi S, Koide R, Shimohata T et al.. Suppression of aggregate formation and apoptosis by transglutaminase inhibitors in cells expressing truncated DRPLA protein with an expanded polyglutamine stretch. *Nature Gen* 1998;18:111-117.
28. Yoshiyama Y, Yamada T, Asanuma K, Asahi T. Apoptosis related antigen, Le(Y) and nick-end labeling are positive in spinal motor neurons in amyotrophic lateral sclerosis. *Acta Neuropathol* 1994;88:207-211
29. Banati RB, Daniel SE, Blunt SB. Glial pathology but absence of apoptotic nigral neurons in long-standing Parkinson's disease. *Mov Disord* 1998;13:221-227.

30. Vyas S, Javoy-Agid F, Herrero MT et al.. Expression of Bcl-2 in adult human brain regions with special reference to neurodegenerative disorders. *J Neurochem* 1997;69:223-231.
31. Krajewski S, Krajewska M, Shabaik A et al.. Immunohistochemical analysis of in vivo patterns of Bcl-xL expression. *Cancer Res* 1994;54:5501-5507.
32. Satou T, Cummings BJ, Cotman CW. Immunoreactivity for Bcl-2 protein within neurons in the Alzheimer's disease brain increases with disease severity. *Brain Res* 1995;697:35-43.
33. Su JH, Satou T, Anderson AJ, Cotman CW. Up-regulation of Bcl-2 is associated with neuronal DNA damage in Alzheimer's disease. *Neuroreport* 1996;7:437-440.
34. MacGibbon AG, Laelor AP, Sirimanne SE et al.. Bax expression in mammalian neuron's, and in Alzheimer disease hippocampus. *Brain Res* 1997; 750: 223-234.
35. Yin XM, Oltval ZN, Korsmeyer SJ. BH1 and BH2 domains of Bcl-2 are required for inhibition of apoptosis and heterodimerization with Bax [see comments]. *Nature* 1994;369:321-323.
36. Cheng EH, Levine B, Boise LH, Thompson CB, Hardwick JM. Bax-independent inhibition of apoptosis by Bcl-XL. *Nature* 1996;379:554-556.
37. Knudson MC and Korsmeyer SJ. Bcl-2 and Bax function independently to regulate cell death. *Nat Genet* 1997;16:358-363.

divided into three approximately equal parts, i.e., the medial, the central and the lateral, and the percentage of neurons immuno-positive for Bcl-xL (b), Bcl-2 (c), or Bax (d) is calculated at each compartment. (b) The percentage of Bcl-xL-immuno-positive neurons is significantly lower in the medial and the lateral parts of the SN in PD (\* $p < 0.05$  and \*\* $p < 0.02$  vs CTL, Mann-Whitney U-test).

Figure 3. Double-labeling; Bax with Bcl-xL or Bcl-2. Co-localization of immuno-fluorescence for (a) Bcl-xL and (b) Bax, and (c) Bcl-2 and (d) Bax in the SN of PD patients (X150). The primary antibody for Bcl-xL in the figure (a) and for Bcl-2 in the figure (c) is recognized by green precipitates, whereas, Bax immunoreactivity is identified by red precipitates (b and d). The green arrow points to a cell immunoreactive for Bcl-xL or Bcl-2 alone, the red arrow points to a cell immunoreactive for Bax alone and the white arrow points to double-stained cells. The combinations of immunoreactivity can be classified into neurons stained for either Bcl-xL, Bcl-2, or Bax alone, or neurons stained for both, which form the majority.

## Figure legend

Figure 1. Immunohistochemical detection of Bcl-xL, Bcl-2 or Bax (X100).

Scale bar, 50  $\mu\text{m}$ . (a) Bcl-xL-, (c) Bcl-2- or (e) Bax- expression in the SN in the CTL subject; the tiny grayish spots in the cytoplasm and nuclear envelope indicate positive staining. Regarding the subcellular distribution, Bcl-xL staining is observed mainly in the cytosol, but Bcl-xL and Bax were also expressed in the nuclear membrane of some neurons. The brown spots are melanin granules in the dopaminergic neurons. (b) Bcl-xL-, (d) Bcl-2- or (f) Bax-expression in the SN in a PD patient; Bcl-xL-expression is apparently reduced, however, Bcl-2- and Bax-expression are essentially similar to those in the CTL subjects.

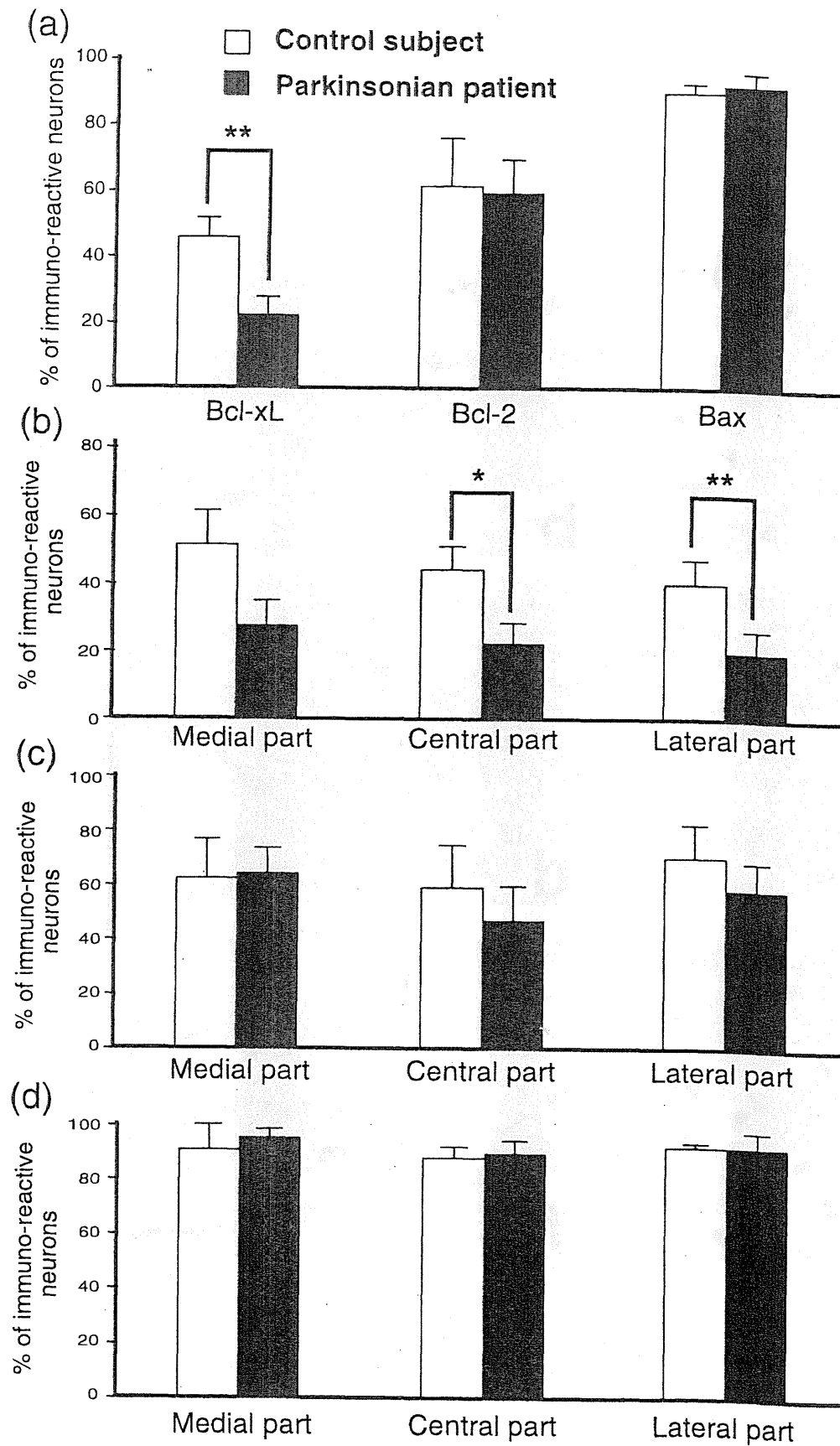
Figure 2. (a) The percentages of immuno-reactive nigral melanized neurons for Bcl-xL, Bcl-2 or Bax in the CTL subjects and in the PD patients. The ordinate indicates “%” of immuno-reactive neurons counted in one of the midbrain sections at the level of the oculomotor (mean  $\pm$  SEM, for n=7 for CTL and n=6 for PD). See the text for the details. The percentage of Bcl-xL-positive neurons among the melanized nigral neurons is significantly lower in the PD patients than in the CTL subjects (\*\*P<0.02 vs CTL, Mann-Whitney U-test). (b)(c)(d) The substantia nigra is arbitrary

**Table1** Clinical characteristics

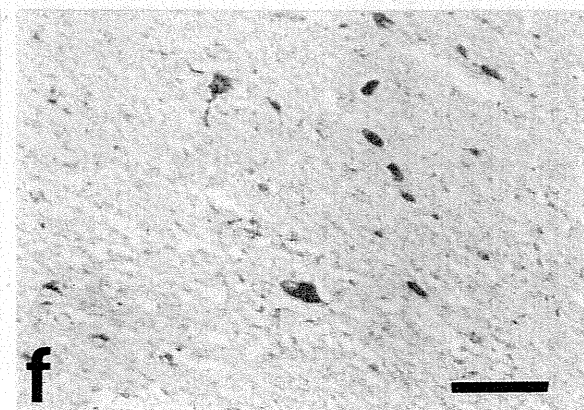
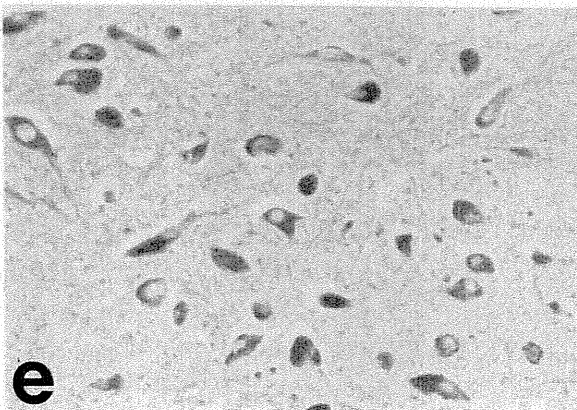
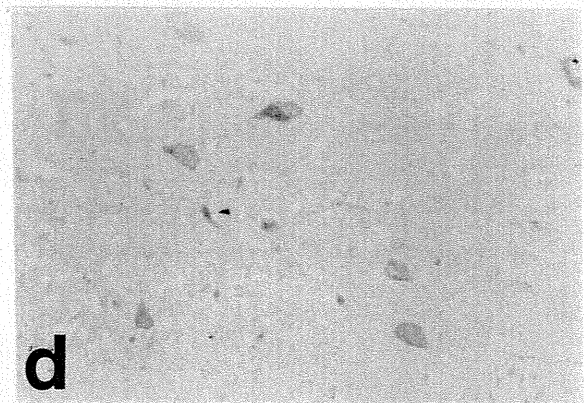
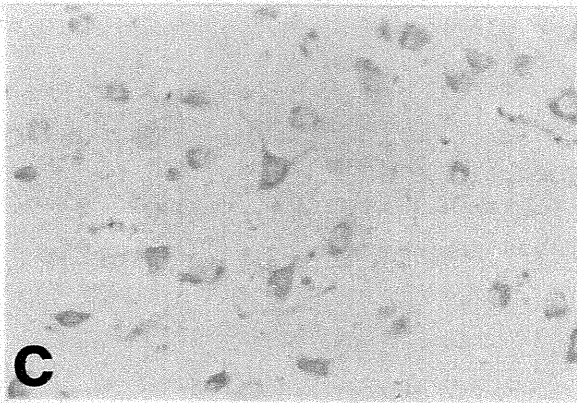
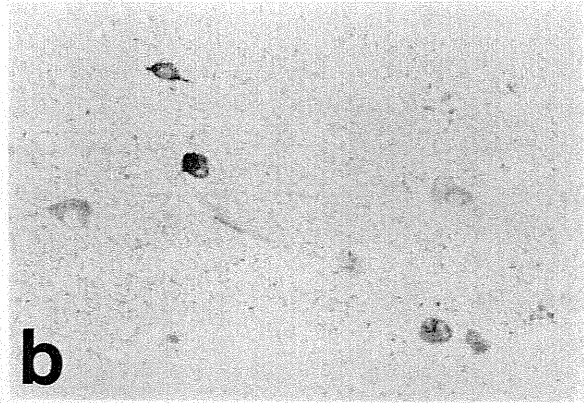
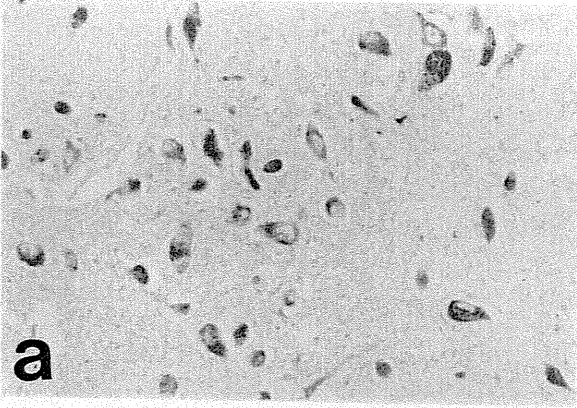
Patient number	Diagnosis	Age (yr)	Sex	Duration of the disease (yr)	Cause of Death	Postmortem interval (hr)
1	CTL	41	M		pneumonia	4.5
2	CTL	52	M		pneumonia	1.0
3	CTL	53	F		myocardial infarction	2.5
4	CTL	67	M		leukemia	3.5
5	CTL	84	M		heart failure	3.1
6	CTL	93	F		heart failure	4.0
7	CTL	76	M		heart failure	2.0
8	PD	63	F	7	pneumonia	4.5
9	PD	66	F	10	sudden death	8.5
10	PD	75	M	11	pneumonia	1.6
11	PD	76	F	18	pneumonia	3.8
12	PD	78	M	15	heart failure	8.5
13	PD	81	F	7	pneumonia	2.5

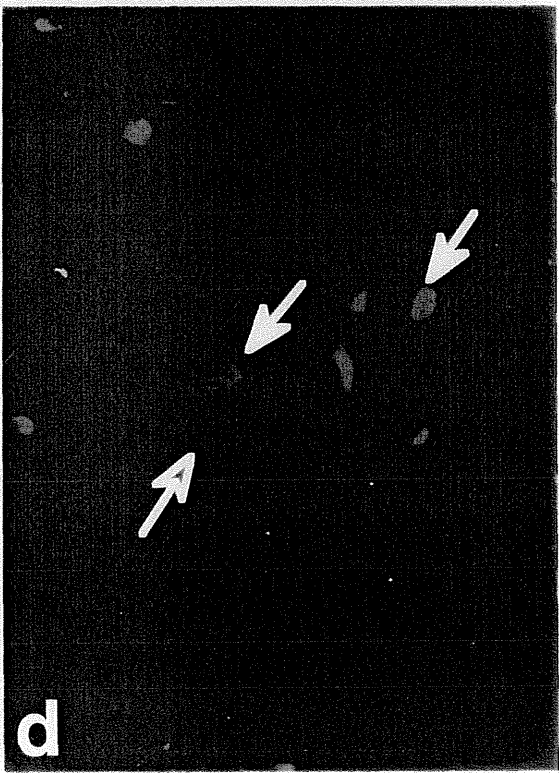
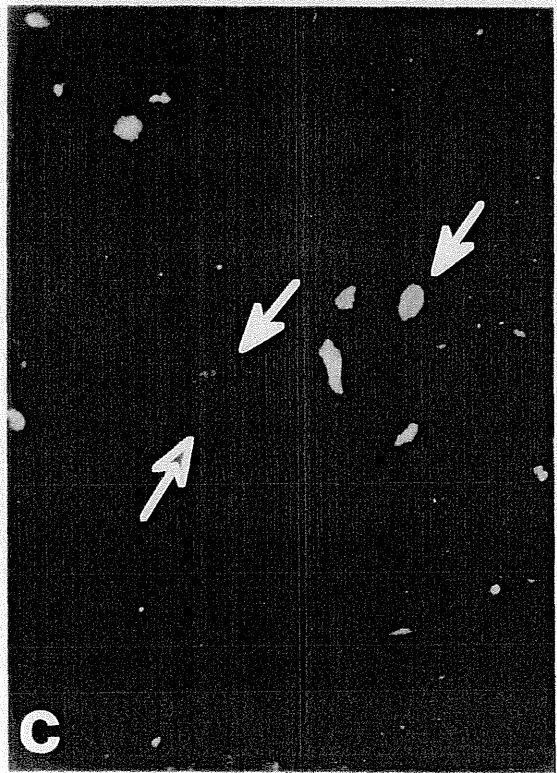
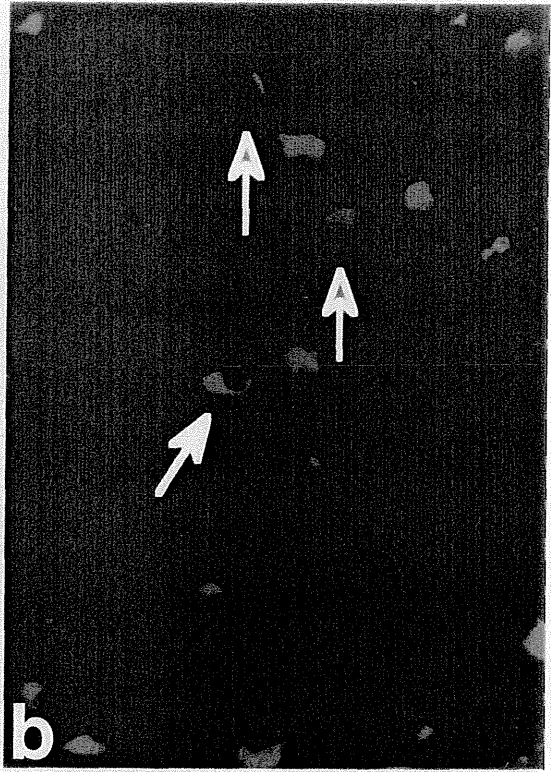
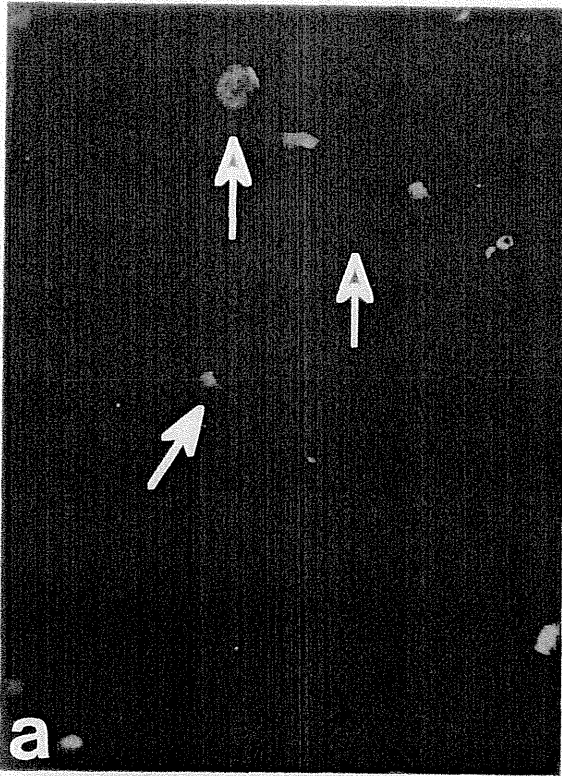
CTL=Control subjects, PD=Parkinson's Disease

Fig.2









Submitted

99-0079  
H. Shimura et al.

# Increase of 8-oxo-dGTPase (*hMTH1*) in Mitochondria of Nigrostriatum of Parkinsonian Brain

Hideki Shimura, M. D.,<sup>1</sup> Nobutaka Hattori, M. D.,<sup>1</sup>  
Dongchon Kang, M. D.,<sup>2</sup> Ken-ichi Miyako, M. D.,<sup>3</sup>  
Yusaku Nakabeppu, Ph. D.,<sup>4</sup>  
and Yoshikuni Mizuno, M. D.<sup>1</sup>

<sup>1</sup> Department of Neurology, Juntendo University school of Medicine 2-1-1  
Hongo, Bunkyo, Tokyo 113-0033, Japan

<sup>2</sup> Department of Clinical Chemistry and Laboratory Medicine, Faculty of  
Medicine, Kyushu University, Fukuoka 812-8582, Japan

<sup>3</sup> Department of Biochemistry, Kyushu University, Fukuoka 812-8582, Japan

<sup>4</sup> Medical Institute of Bioregulation, Kyushu University, Fukuoka 812-8582,  
Japan

Correspondence to Nobutaka Hattori, M. D.

Department of Neurology, Juntendo University School of Medicine  
2-1-1 Hongo, Bunkyo, Tokyo 113-0033, Japan

Tel.: ++81-3-5802-1072

Fax: ++81-3-3813-7440

E-mail: nhattori@med.juntendo.ac.jp

page 1

RECEIVED  
JAN 11 1999  
PROCEEDINGS OFFICE

**ABSTRACT**

There is growing evidence that oxidative stress and mitochondrial respiratory failure with an attendant decrease of energy output are involved in nigral neuronal cell death in Parkinson's disease (PD). 8-oxo-dGTPase (8-oxo-7,8-dihydrodeoxyguanosine triphosphatase: *hMTH1*) played an important role in the control of spontaneous mutagenesis, the production of including 8-oxo-dG (8-oxo-2,8-dihydroguanine). 8-oxo-dGTPase was increased in the mitochondria of the substantia nigra in PD patients. In the study of 8-oxo-dG, while nuclear DNA showed no modification in all subjects, mitochondrial DNA showed increased modification in PD patients. These results support the concept in oxidative mitochondrial stress in PD. Moreover, an increase of *hMTH1* appears to be a useful molecular marker of oxidative stress that can be employed to explore the relationship between such stress and genomic instability.

Key words: 8-oxo-7,8-dihydrodeoxyguanosine triphosphatase, *hMTH1*, 8-dihydrodeoxyguanosine, oxidative stress, Parkinson's disease